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Drosophila as a model to study the role of blood cells in inflammation, innate immunity and cancer

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Abstract

Drosophila has a primitive yet effective blood system with three types of haemocytes which function throughout different developmental stages and environmental stimuli. Haemocytes play essential roles in tissue modelling during embryogenesis and morphogenesis, and also in innate immunity. The open circulatory system of Drosophila makes haemocytes ideal signal mediators to cells and tissues in response to events such as infection and wounding. The application of recently developed and sophisticated genetic tools to the relatively simple genome of Drosophila has made the fly a popular system for modelling human tumourigenesis and metastasis. Drosophila is now used for screening and investigation of genes implicated in human leukaemia and also in modelling development of solid tumours. This second line of research offers promising opportunities to determine the seemingly conflicting roles of blood cells in tumour progression and invasion. This review provides an overview of the signalling pathways conserved in Drosophila during haematopoiesis, haemostasis, innate immunity, wound healing and inflammation. We also review the most recent progress in the use of Drosophila as a cancer research model with an emphasis on the roles haemocytes can play in various cancer models and in the links between inflammation and cancer.
Abbreviations

AMP, antimicrobial peptide; AML, acute myeloid leukaemia; ALL, acute lymphoblastic leukaemia; Bsk, Basket; Chn, Charlatan; CML, chronic myeloid leukaemia, CNS, central nerve system; Col, Collier; Dif, Dorsal related immunity; Dom, Domeless; ECM, extracellular matrix proteins; EGF, epidermal growth factor, FOG, Friend of GATA; GADD45, growth arrest and DNA damage-inducible gene 45; Gcm, Glial-cells-missing; Hg, Hedgehog; Hml, Hemolectin; HSC, haematopoietic stem cells; JNK, Jun N-terminal kinase; LG, lymph gland; Lgl, lethal giant larvae; Lz, Lozenge; MARCM, mosaic analysis with a repressible cell marker system; PDGF, platelet-derived growth factor; PI3K, phosphatidylinositol 3-kinase; PPAE, Prophenoloxidase activating enzyme; PPO, Prophenoloxidase; PSC, posterior signalling centre; ROS, reactive oxygen species; Ser, Serrate; Smo, Smoothened; Srp, Serpent; TF, Transglutaminase; Upd, Unpaired; Ush, U-shaped; VEGF, vascular endothelial growth factor; Wnt, Wingless.
Introduction

*Drosophila* has undoubtedly been a powerful model organism for the study of nearly all essential and fundamental biological processes. What we have learned from the fruit fly has expanded our knowledge in life science at an unprecedented speed. This is in particular due to the recent availability of the complete annotated genome, a versatile array of genomic modifying techniques and powerful life imaging tools. Cellular and molecular mechanisms underlying many basic biological processes have been discovered to be highly conserved between *Drosophila* and mammals. For example, the Notch, Hedgehog (Hg) and Wingless (Wnt) pathways first identified in *Drosophila* embryogenesis and the Runt and Hippo signalling pathways conserved in the *Drosophila* haematopoiesis and tissue growth are also implicated in the progression of various human cancers (Harvey *et al.*, 2013; Geissler & Zach, 2012;). Indeed the past decade has witnessed a rapidly emerging trend for *Drosophila* to be used in modelling human tumour growth, progression, invasion and metastasis and as a test-bed for therapeutic discovery (reviews in Gonzalez, 2013; Hsu, 2012; Crozatier & Vincent, 2011; Miles *et al.*, 2011; Harris, 2005).

Most forms of human cancers progress step by step from mutations in the oncogene, the tumour suppressor gene and signalling molecules and can eventually kill the host by spreading uncontrollable immortal growth of mutant malignant tissues into different organs. On the route to spread and invade, cancer cells can influence their microenvironment via the interaction with the infiltrated blood cells, gradually disabling the host immunosurveillance and finally breaking the stromal barrier to become invasive and metastatic (Dunn *et al.*, 2004). It is at the metastatic stage that many lives would be claimed. Therefore the outcome from the tug of war in the tumour microenvironment between malignant cancerous cells that undergo constant somatic mutations and surrounding blood cells plays a vital role in the prevention and intervention of tumourigenesis. In addition, chronic inflammation has been well-
documented as contributing to and promoting the initiation and progression of various cancers (Aggarwal, et al., 2009; Mantovani et al., 2008, Coussens & Werb, 2002). It is now generally accepted that an inflammatory microenvironment is necessary for tumour progression and metastasis (Grivennikov et al. 2010; Wu & Zhou, 2009). Macrophages in particular have been reported to facilitate many aspects of this process in different cancers and also to intervene in the anti-cancer therapies (Lee, et al., 2013; De Palma & Lewis, 2013).

Apart from the role of macrophages in cancer development, they have been for many years subjected to extensive research as the key player in inflammatory responses which accompany infection, tissue damage and wound healing (Mantovani, et al., 2013; Novak & Koh, 2013). Therefore inflammation, immunity and cancer are inter-linked and any imbalance can result in serious health issues. Blood cells such as macrophages appear to be the link and have a crucial role in influencing and maintaining the equilibrium between protection (immunity and inflammation) and regeneration/tissue homeostasis (where cancer can be considered a malignant proliferative and invasive tissue). Animal models such as mice have revealed invaluable insights into the multi-step interaction of mammalian innate immunity with associated inflammatory responses in defining the cancer microenvironment. These innate immune responses can include the complement pathways (Ricklin & Lambris, 2013), pro-inflammatory cytokine and chemokine production (Sethi, et al. 2012; Candido & Hagemann, 2013). However, the multi-layered interaction in the context of a generally slow progression of the human cancer has created fragmentary and controversial results in the mouse model and thus inevitably slows down our progress to understand the disease. *Drosophila*, on the contrary, as a simply-formed and genetically tractable multi-cellular organism, has been used to dissect processes of development (tissue homeostasis) and innate immunity with such precision that the time is now ripe for us to look into the active dialogues between these fundamental processes in the context of mammalian inflammation and cancer.
Drosophila has a primitive open blood circulation system with only three types of blood cells or haemocytes circulating in the haemolymph during a fly’s life span. The majority of the circulating haemocytes in the haemolymph are macrophage-like cells that engulf and degrade apoptotic cells and invading pathogens. Haemocytes perform vital roles through their contribution both to cellular and humoral immune responses in the fly. In combination with the currently well-developed Drosophila tumour models, the roles of haemocytes in tumour regression and/or progression can be explored and important clues can be obtained to understand further the inflammatory responses in relation to tumours; the focus can be directed more at the molecular and cellular level by use of sophisticated genetic manipulation and live imaging tools. Tian Xu and colleagues have done pioneering work in this direction by using a Drosophila tumour model to investigate the role of haemocytes in the tumour growth control during a systemic inflammatory response (Pastor-Pareja et al., 2008). In this review, we first give a brief overview of Drosophila haematopoiesis. This is followed by a discussion of the roles of haemocytes in Drosophila at different developmental stages. Next we consider how haemocytes function in tissue injury and wound healing. Drosophila leukaemia model and interaction between immunity and tumourigenesis are also discussed. Finally, perspectives for possible future research opportunities in the interplay of inflammation, immunity and cancer revolving around the blood cells are discussed.

Drosophila Haematopoiesis

The blood system of Drosophila is rather primitive compared to the great complexity in vertebrates. The fruit fly does not have a vascular network to separate the blood cells from other tissues and organs and its internal organs are bathed in haemolymph. Meanwhile vertebrates have many different types of blood cells. Each type has evolved to perform specialised functions during million years of evolution. On the contrary only three major types of blood cells, collectively termed haemocytes, have been identified in the fruit fly and
none of them has acquired the capability to undergo DNA rearrangement and somatic hypermutation to generate a vast repertoire for immunological memory in the B-and T-lymphocytes. Therefore *Drosophila* relies on a very simple system to fulfil basically all the roles that vertebrate blood cells can play. However there is extensive conservation in the molecular mechanisms of haematopoiesis in both *Drosophila* and mammals.

As in vertebrates, *Drosophila* haematopoiesis takes place in two phases: primitive haematopoiesis and definitive haematopoiesis (for more detailed review see Krzemien *et al.*, 2010, Crozatier & Meister, 2007; Evans, *et al.*, 2003). Briefly, the site for primitive haematopoiesis resides in the precephalic mesoderm which gives rise to the early wave of haemocyte generation in the embryo (Fig.1A). At the end of embryogenesis, a specialised organ termed Lymph Gland (LG) originating from the lateral mesoderm starts to appear along the dorsal vessel and becomes fully mature during the first half of larval development (Fig. 1B). Definite haematopoiesis initiates in the LG (Fig. 1B & Fig. 2B) and generates terminally differentiated haemocytes at the onset of metamorphosis; the cells are released during the pupal stage with disintegration of the LG. After the disappearance of LG, no haematopoiesis will occur in the pupa or in the adult fly. Haemocytes persisting through the whole life stages of the fly therefore have either embryonic or larval lineage (Holz *et al.*, 2003).

LG forms at the end of embryogenesis with a single pair of lobes called the anterior or primary lobes. Larval haematopoiesis takes place primarily in the primary lobes to temporally and spatially regulate the haemocyte differentiation. The primary lobes can be physically divided into three compartments (Fig.2B): the cortical and the medullary zones and the Posterior Signalling Centre (PSC) (Jung *et al.*, 2005). PSC is essential in controlling haemostasis in healthy larvae by a direct cell-cell communication via filopodia, thin cytoplasmic extensions (Krzemień *et al.*, 2007; Mandal *et al.*, 2007). This cellular contact
provides a platform for the interplay of a network of key signalling pathways required for normal larval haematopoiesis (Fig. 2A).

Based on different morphological features three major types of haemocytes can be identified throughout the life cycle of the fruit fly, namely, plasmatocytes, crystal cells and lamellocytes (Fig.2A) (Hartenstein, 2006; Lanot et al., 2001). Plasmatocytes are the dominating haemocyte population during all Drosophila developmental stages. They are macrophage-like cells that are primarily responsible for the removal of apoptotic debris, phagocytosis of invading microbes and repair of damaged tissues. Crystal cells are larger in size than plasmatocytes and are named from the paracystalline inclusions in the cytoplasm. The crystal cell inclusions are believed to contain large quantities of components involved in a process called melanisation involving a cascade of serine proteases leading to melanin synthesis (Jiravanichpaisal et al., 2006). Melanin is important to prevent haemolymph loss in wound sites, immobilise microbial pathogens and facilitate wound healing (see below). In addition, its free radical oxidative by-product can directly kill microorganisms. Although they constitute only a small proportion of 5% of the total population of Drosophila haemocytes in the embryo and larvae, crystal cells are major executioners in Drosophila innate immunity. Lamellocytes are physically distinctive from both plasmatocytes and crystal cells. They are flat and adhesive and are the largest haemocytes observed in Drosophila (Lanot et al., 2001). They do not appear in the embryo or in healthy larvae but can be induced quickly to differentiate from the LG or sessile population along the border of larval segments (see below) to engulf foreign particles larger than those that can be phagocytosed by plasmatocytes, such as the parasitoid eggs (Lee et al., 2009; Sorrentino et al., 2002). This process is termed encapsulation. Lamellocytes can also launch a melanisation cascade to kill the parasitic invaders with the aid of crystal cells and thus are essential in the Drosophila immunity against parasite infection (Krzemien et al., 2010).
The key players including primarily transcription factors and the corresponding molecular mechanisms are briefly summarized and illustrated in Figure 2.

**Haemocytes in immunity**

In the wild, *Drosophila* feeds on rotting fruits and lives in a microorganism-enriched environment so fruit flies constantly face the danger of physical injury and gastrointestinal infection. The selection pressure from the hostile environment must be one of the driving forces for *Drosophila* to develop multi-layered defence responses so that it can survive and propagate. Not surprisingly the cellular and molecular mechanisms in the various facets of *Drosophila* innate immunity have been phylogenetically conserved. *Drosophila* can mount an array of cellular and humoral responses when challenged by pathogens such as bacteria, fungi, viruses and parasites. The cellular responses include direct engulfment of small objects such as the bacteria, as in phagocytosis, and encapsulation of larger objects such as parasitoid eggs. The humoral responses take place primarily in the haemolymph and three major events can occur during an immune challenge depending on the nature of the invading pathogen: 1) direct killing by Antimicrobial peptide (AMP) released into haemolymph from rapid de novo synthesis in the haemocyte and fat body; 2) direct killing by Hydrogen Peroxide (H$_2$O$_2$) or Nitric Oxide (NO) agents produced during melanisation; and 3) immobilisation of opportunistic pathogens by blood coagulation at an open wound (for more in depth reviews see Kounatidis & Ligoxygakis, 2012; Lemaitre & Hoffmann, 2007). In this review, we focus on the roles of haemocytes in the *Drosophila* host defence and the key signalling pathways in orchestrating the different strategies deployed against a wide range of pathogens.

**Molecular basis of haemocyte migration & motility in embryos**

*Drosophila* haemocytes respond to numerous signals during development or following injury and/or infection (Will Wood & Jacinto, 2007). These signals can include migrating cues
during embryogenesis, inflammatory and stress chemoattractants from the injury site and pathogen invasion in the haemolymph or in the tissue. During development, embryonic plasmatocytes are highly motile cells and they migrate from the precephalic mesoderm around stage 10 of the embryogenesis to start to disperse the entire embryo (Tepass et al., 1994). Haemocytes migrate along several invariant main routes throughout the embryo: towards the tail, along the ventral nerve cord, along the dorsal vessel and the gut primordium. The cell migration is guided by Pvr/Pvf signalling. The receptor tyrokinase Pvr is the homologue of the vertebrate Platelet-derived growth factor (PDGF) and Vascular endothelial growth factor (VEGF) receptors. The Pvr has three ligands: PDGF- and VEGF-related factor (Pvf)-1, Pvf-2 and Pvf-3. Haemocytes express Pvr and are attracted by Pvf2 and Pvf3 expressed in different tissues of the embryo. For example, the nerve cord expresses Pvf2 and Pvf3 spatially and temporally in its different compartments to attract haemocytes to move along the Central Nerve System (CNS) (Will Wood et al., 2006; Cho et al., 2002). The impressively fixed migrating patterns for haemocytes to populate the entire embryo from anterior to posterior and from dorsal to ventral rely on sustained motility and cell polarity and highly organised cell shape to enable smooth and rapid movement along the tissue surface. By use of live confocal microscopy, plasmatocytes were discovered to move with large, polarized and actin-rich filopodia and lamellopodia during the embryonic migration (Wood et al., 2006). Towards the end of embryogenesis, these cytoplasmic protrusions become highly dynamic and continually extend and retract to survey the surrounding microenvironment. The Rho family small GTPase Rac, Rac1 and Rac2, function redundantly to control the lamellopodia formation and thus the successful dispersal of haemocytes in the embryo (Paladi & Tepass, 2004). A Drosophila PDZ guanine-nucleotide exchange factor (PDZ-GEF) Dizzy was also identified to be required for the embryonic haemocyte migration (Huelsmann et al., 2006). In the absence of Dizzy, the cytoplasmic protrusions are reduced in size and thus slow
down the migration rate of the cells. Overexpression of Dizzy in haemocytes generates cells
with abnormally extended protrusions. Dizzy is believed to act upstream of the Ras
superfamily member of small GTPase Rap1 to regulate integrin dependent adhesion of
haemocytes to the epithelia and to maintain their cellular ‘microspikes’ throughout migration
(Huelsmann et al., 2006). Siekhaus et al discovered that during embryonic haemocyte
migration Drosophila haemocytes invade an epithelial barrier as they move into the tail
despite an open blood system (Siekhaus et al., 2010). A mutant of RhoL, another Drosophila
GTPase homologue specifically expressed in haemocytes blocks this epithelia invasion but
not other aspects of guided migration. RhoL interferes with Rap1 mediated integrin adhesion
by moving Rap1 away from a concentration in the cytoplasm to the leading edge during
invasive migration. RhoL therefore functions as a regulator for integrin adhesion and Rap1
localization during the invasion. Inhibition of integrin-based adhesion is necessary to regulate
the cadherin interactions that allow plasmatocytes to transmigrate from the head region,
through the epithelium, to the posterior of the embryo. These findings revealed a striking
similarity of the stepwise migratory process during Drosophila development with vertebrate
immune cell transmigration during inflammation.

Apart from moving along fixed developmental migratory patterns, embryonic plasmatocytes
respond to the epithelia wound by migrating rapidly to the site of injury. This process also
shares many physiological relevancies with the vertebrate inflammation. Similar to the
developmental migration, this deployment to the injury site also requires Rac-mediated
lamellopodia formation (Stramer et al., 2005). Stramer and co-workers showed that Rho
signalling is necessary for haemocytes to retract from sites of cell matrix and disengage from
cell-cell contact. During the migration to the wound, CDC42 is required to maintain the
plasmatocyte polarity. In contrast to the developmental dispersal of haemocytes in response
to Pvr/Pvf cues, the chemotaxic signals from the injury site activate a different mechanism to
mobilise plasmatocytes. This *Drosophila* inflammation induced cell migration depends on a phosphatidyl-inositol 3-kinase (PI3K) signalling which is also used by the mammalian neutrophils in response to chemotactic cues (Stramer *et al.*, 2005). Therefore, it can be concluded that actin protrusion formation controlled by the Rac signalling in the cell motility is essential for plasmatocyte migration in these two different processes. Ena is another player identified recently to regulate the actin protrusions in the embryonic haemocyte (Tucker *et al.*, 2011). Ena is the *Drosophila* homologue of Mena, member of the evolutionarily conserved Ena/VASP family of actin cytoskeletal regulators. Mena promotes metastasis and invasive motility of breast cancer cells *in vivo*. Tucker *et al* study found that Ena stimulates lamellipodial dynamics and positively regulates the number and length of filopodia. Overexpression of Ena in the haemocyte results in dramatic increase in the migration rate. One of the phenotypes can be also observed from overexpression of Mena in the mammalian fibroblast.

**Postembryonic haemocyte migration**

In the larva, overexpression of Rac in the haemocyte disrupts the sessile haemocytes population and causes a large migration of haemocytes into the circulation. Sessile haemocyte activation and mobilisation require the Jun N-terminal kinase (JNK) Basket (Bsk) and Rac1. Bsk is also found to regulate the turnover of focal adhesions in the circulating haemocyte in the larva (Williams *et al.*, 2006). These findings suggest that the Rho and JNK signalings are conserved, underlining their roles in the formation of cytoplasmic protrusions and actin focal adhesions for proper plasmatocyte mobility and migration to support their cellular roles in development and immunity. Two very recent studies on the postembryonic haemocyte migration have shed more light on the cellular dynamics and molecular basis of this process. An *ex vivo* culturing system using the primary larval or pre-pupal haemocytes has been developed to allow a real time analysis and manipulation to examine the roles of
cytoskeleton dynamics in plasmatocyte migration (Sampson & Williams, 2012). From this system, it was found that larval circulating haemocytes are less motile than the pre-pupal haemocytes and thus unable to migrate. The extending and retracting rates of the protrusions appear dormant while the prepupal haemocytes have normal dynamic protrusions potentially required in the morphogenesis. The same study also reinforced the role of Rho family members: Rac1 and Rac2 and CDC42 to sustain the size of the filopodia and lamellipodia in the prepulal haemocytes. Absence of these genes caused a static phenotype of pre-pupal haemocytes similar to what was observed in haemocytes from third instar larvae. Though an in vivo assay still awaits to confirm these findings, the importance of Rho signalling in the actin cytoskeleton shaping has been strengthened in the ‘walking’ of the haemocyte along the extracellular matrix.

Another in vivo study based on MARCM (Mosaic Analysis with a Repressible Cell Marker System) investigated the integrin adhesion activation and maturation in the migration of the sessile haemocyte population in the late larval stage into pupal stage (Moreira et al., 2013). The Drosophila βPS integrin myosperoid and integrin containing adhesion regulators such as Rhea and Fermitin were found to be required in pupal haemocyte migration.

**Cellular immunity mediated by plasmatocytes**

Plasmatocytes represent around 90% of the total circulating haemocyte population in all developmental stages of Drosophila. They are professional macrophages which function as sentinels to maintain cell and tissue homeostasis and to recognise pathogen entry for subsequent immune reactions.

In the embryo, mature plasmatocytes function primarily as scavengers to remove apoptotic cell debris during embryogenesis. The clearance of apoptotic cell debris is dependent on scavenger receptors: CD36 homologue Croquemort (Franc et al., 1996), Draper (Manaka et
al., 2004) and NimC4/Simu (Kurant et al., 2008). In the larval stage, recognition and rapid engulfment of invading microbes such as bacteria rely on the cell surface receptors Eater (Kocks et al., 2005) and NimC1 (Kurucz et al., 2007) and in the adult fly on Draper also (Cuttell et al., 2008). Loss of function of mutants in those receptors results in functional deficiency of phagocytosis of Gram-positive bacteria such as *Staphylococcus aureus* and Gram-negative bacteria such as *Escherichia coli*. Eater, NimC1, NimC4 and Draper together with CED in *C. elegans* belong to a large protein family conserved across the metazoan animal kingdom including other insects such as *Anopheles* and humans (Kurucz et al., 2007).

EGF (Epidermal Growth Factor)-like repeats are abundantly found in the extracellular domains of these bacterial phagocytosis receptors and there is evidence to show the direct interaction and binding of the EGF-like repeats with bacteria in Eater (Kocks et al., 2005) and Draper (Hashimoto et al., 2009). These interesting findings suggest that proteins with GF-like repeats may play an evolutionary conserved role in phagocytosis in the entire animal kingdom (Table 1).

Another receptor that binds directly microorganisms and participates in phagocytosis is Dscam, which is a member of the Ig superfamily with an essential function in neuron interconnection. By alternative splicing as many as 18,000 isoforms can be theoretically generated in the haemocyte and fat body (Watson et al., 2005). Haemocyte-specific Dscam silencing reduces the phagocytic uptake of bacteria. The existence of a potential extensive repertoire of thousands of Ig-domain-containing proteins in the recognition of a variety of pathogens in *Drosophila* and other invertebrates has opened up a new avenue in exploring the possibility of an ‘adaptive’ immunity across animal phyla that have been considered as having only innate responses (Schmucker & Chen, 2009; Watson et al., 2005).

Haemocyte-mediated humoral response
In response to pathogens that manage to gain entry into the haemocoel *Drosophila* can mount a robust systemic immune humoral response. The hallmark of this response is the rapid synthesis of a broad spectrum of AMPs against bacteria and fungi both in the haemocyte and in the fat body. AMPs are secreted into the haemolymph and directly kill the microbes at an optimal concentration. Although the fat body - the *Drosophila* equivalent of mammalian liver - is the prominent site of AMP synthesis, plasmatocytes play important roles in triggering the AMP production as haemocyte ablation can abolish the AMP expression in larval fat body (Shia *et al*., 2009). The roles plasmatocytes can play in the systemic immune response can be achieved by signalling between the site of infection and the fat body or by degradation of invading pathogens. To date, a haemocyte-released cytokine, Unpaired-3 (Upd-3) has been proposed to activate the JAK/STAT pathway in response to septic injury by binding to the fat body Domeless (Dom) receptor (Agaisse *et al*., 2003). Nevertheless the precise role of this pathway and its overall contribution to the host defence remains to be established. Spätzle is another cytokine secreted by haemocytes, processed by a serine protease cascade in the haemolymph and required for the Toll signalling pathway controlled AMP synthesis in the fat body (Shia *et al*., 2009). The Toll signalling has been well characterised in the Dorsal-Ventral patterning during embryogenesis and AMP production against mainly fungi, Gram positive bacteria, *Pseudomonas aeruginosa* (a Gram negative bacterium) and also stress/danger signals (reviews in Kounatidis & Ligoxygakis, 2012; Valanne *et al*., 2011). The core Toll signalling event is the degradation of *Drosophila* NFκB Inhibitor homologue Cactus followed by the activation and translocation of the *Drosophila* NFκB transcription factors Dorsal or Dorsal related immunity factor (Dif) into the nucleus. Dorsal and Dif are homologues of mammalian p50 and p65. Apart from these secreted cytokines, a cytoplasmic lysosomal protein called Psidin has been found to be the link between the haemocyte phagocytosis and AMP activation in the fat body in the larval immune response (Brennan *et al*., 2007). Psidin
is required both for the phagocytic degradation of internalized bacteria and for the induction of one of the AMPs, Defensin, in the fat body. This interesting finding suggests a likely ‘antigen’ presentation mechanism of the haemocyte to the fat body for the activation of AMP. Contrary to these findings, plasmatocyte ablation does not affect the antimicrobial responses upon systemic infection in the adult fly (Defaye et al., 2009; Charroux & Royet 2009). This might suggest that tissue specific humoral responses, such as local expression of antimicrobial peptide and cytokines independent of haemocytes in the gut or in the trachea, play dominant roles in the adult immunity against pathogens.

**AMP production in haemocytes**

AMP production plays a vital role throughout the life cycle of *Drosophila*. Many tissues that have direct contact with the microorganisms such as the trachea, the gut and malpighian tubules have the capability to synthesize AMP and kill the microbes locally and efficiently. For microbes that manage to gain entry into the circulation via an open wound or the digestive or reproductive tracts, the fly can mount a systemic humoral response to produce large amounts of AMP from mainly the fat body into the haemolymph. Although haemocytes are not the major organ in the fly for systemic AMP production, the signalling pathways in control of AMP synthesis are activated in haemocytes like in the other tissues during a concerted immune response, in particular in the embryonic haemocytes (reviewed by Kounatidis & Ligoxygakis, 2012; Lemaitre & Hoffmann, 2007).

**Melanisation**

Melanisation in arthropods is generally believed to play an important and central role in arthropod defence reactions such as wound healing, encapsulation, microbe immobilisation and the production of toxic intermediates that are speculated to kill invading microorganisms (Cerenius & Söderhäll, 2004). As described briefly above, crystal cells are the major
haemocytes responsible for the melanisation reaction in the larva. Melanisation can be
immediately induced at the site of cuticular injury or on the surface of parasites invading the
haemocoel. It involves formation of black pigmentation resulting from de novo synthesis
and deposition of melanin. Prophenoloxidase (PPO) is the enzyme required in melanisation to
catalyze the oxidation of mono- and di-phenols to ortho-quinones, which polymerise into
melanin. PPO in normal physical conditions is enzymatically inert. A serine protease known
as prophenoloxidase activating enzyme (PPAE) acts upstream to cleave and turn PPO into
active phenoloxidase (PO). Like PPO, PPAE also exists as an inactive zymogen and it is
processed by a tightly regulated serine protease cascade in a step wise way into enzymatically
functional form leading to the final melanin formation. As in other invertebrates, a recent in vitro study on the Drosophila PPOs suggested a direct binding of PPOs to bacteria and fungi
which might play a role to initiate their activation (Yang et al., 2013). The Drosophila
genome encodes three PPOs: DoxA1, DoxA3 and CG8193 (Irving et al., 2005). Crystal cells
express DoxA1 and CG8193 while lamellocytes express exclusively DoxA3, a strong
indication that Dox3 participates in the encapsulation that accompanies melanisation.

Melanisation is diminished in the domino mutant that lacks haemocytes (Braun et al., 1998)
and in the Black cells (Bc) mutant with aberrant crystal cells (Corbo & Levine, 1996; Rizki et
al., 1980) and the Lz knockout, which is devoid of crystal cells (Peeples et al., 1969). One
serine protease Sp7 has been reported to be involved in PPO activation and expressed also in
the crystal cells (Castillejo-López & Häcker, 2005). In the absence of crystal cells in the adult
fly, melanisation perhaps relies on the activation of proteolytic cascades in the haemolymph
including PPAE, PPO and serine proteases. The cascades are tightly regulated by serine
protease inhibitors in the haemolymph such as Serpin27 A to restrict the reaction to the site of
injury and to prevent the spread of systemic melanisation (Ligoxygakis et al., 2002; De
Gregorio et al., 2002). Two serine proteases MP1 and MP2 are reported to activate the
cascade in response to different microbial changes (Tang, et al., 2006). This pathogen-
specific activation of melanisation can be attributed to PGLP-LC and PGRP-LE expressed
both by haemocytes and the fat body (Schmidt et al., 2008; Takehana et al., 2004). However
the connection between other types of pathogen receptors (for example, Gram-positive
bacteria and fungi etc) has not been linked to melanisation triggering. To date only one PPAE
has been identified in the melanisation cascade of the adult fly (Leclerc et al., 2006)

Encapsulation

Lamellocytes are the major executioner of encapsulation during parasite infection in the
Drosophila larva. Encapsulation involves three key steps with coordinated actions from both
plasmatocytes and lamellocytes. Firstly circulating plasmatocytes sense and recognise the
entry of parasitoid eggs in the haemocoel and attach to the egg chorion. Secondly a massive
proliferation and differentiation of sessile compartments and of haemocytes in the LG to
lamellocytes is induced via unknown signalling molecules within a few hours to appear in the
circulation where the lamellocytes form a multi-layered capsule around the eggs. Eventually
the lamellocytes, like the crystal cells, release their cellular content such as PPO to activate
the melanisation process and kill the parasites, possibly by the cytotoxic by products from the
localised melanisation reaction (Nappi et al., 1995). To date genes that have been reported to
play a role in encapsulation process are involved primarily in cell-cell interaction such as
αPS4/βPSintegrins (myospheroid) (Irving et al., 2005; Wertheim et al., 2005) and in
cytoskeleton remodelling for motility and migration, such as RhoGTPase protein family
member Rac1(Williams et al., 2006), Rac2 and CDC42 (Williams et al., 2005). Rac2 and
CDC42 are activated by the Drosophila homologue of Rho guanine nucleotide exchange
factor (RhoGEF), Zir (Sampson et al., 2012; Sampson & Williams, 2012). Rac1 and Rac2
function in a non-redundant manner. Interaction between Rac1 and myospheroid has recently
been reported to be required in the directed localisation of β-integrin on the cell surface of lamellocytes in response to parasitoid eggs (Xavier & Williams, 2011). Rac1 requires the JNK pathway component Bsk to regulate the formation of actin- and focal adhesion kinase (FAK)-rich placodes in haemocyte migration and both are required for the proper encapsulation of wasp eggs (Williams et al., 2006). A recent screen to target genes involved in the cell adhesion and shape change not only strengthened the previous findings but also discovered more conserved components in these cellular processes that participate in the encapsulation reaction, for example the extracellular matrix proteins (ECM) (Howell et al., 2012). Loss of function of ECM components results in failure to encapsulate. In correlation with the previous discovery on the encapsulation of mechanically damaged self-tissue, it is plausible that exposure of ECM by foreign particle intrusion or deposition of ECM on the eggs can be the initiative signal in encapsulation (Howell et al., 2012; Rizki & Rizki, 1980). Genome-wide analysis of the transcriptional profiles in haemocytes after parasitoid infection has offered many interesting and promising candidate genes that are differentially regulated in the encapsulation reaction (Wertheim et al., 2005). This study also reinforced the importance of the Toll and JAK/STAT signalling pathways in the differentiation and proliferation of lamellocytes in the LG (Sorrentino et al., 2004). In addition, the haemocyte-specific transmembrane protein Hemese has been reported to play a modulatory role to keep lamellocyte proliferation in check from overacting during parasitoid egg infection (Kurucz et al., 2003). Despite these findings, the molecular nature of the signals sent from plasmatocytes for lamellocyte differentiation in the LG remains elusive. It has been proposed that a signal delivered to the PSC initiates lamellocyte differentiation as the PSC-restricted expression of Collier (Col), the *Drosophila* homologue of human Early B cell factor, is required upon parasite invasion (Crozatier et al., 2004).
Blood Coagulation

Haemocytes have essential roles in blood coagulation, not only to maintain haemostasis but also to defend against pathogens. It has been found that Hemolectin (Hml) expressed mainly by plasmatocytes is required in blood coagulation. Blood coagulation led by plasmatocytes is independent of both melanin production and phenoloxidase activity, which is also part of the wound healing process (Goto et al., 2003). By proteomics and pull out analysis, important components of the blood clot have been isolated and subjected to detailed genetic and cellular investigations (Karlsson et al., 2004; Scherfer et al., 2004). Among these the best-characterised clotting factors are Drosophila Transglutaminase (TF) and Fondue. Drosophila TF is the only mammalian blood coagulation factor homologue (Factor XIIIa) found in the fly and uses Fondue as its substrate to form the blood clot. Unlike hemolectin (hml) mutants shown to affect only coagulation (Goto et al., 2003), ubiquitous silencing of fondue also results in cuticle defects in the pupa as well as in the clot forming in larvae (Scherfer et al., 2006). Hml is expressed mainly by plasmatocytes and contains domains found in coagulation factors (Goto et al., 2003; Beck & Kitagawa, 2001). It is suggested that TF/Fondue acts more actively in cross-linking of fibres formed by Hml, reacting promptly to bleeding and injury (Lindgren et al., 2008; Scherfer et al., 2006). Interestingly TF is most likely expressed in haemocytes (Johansson et al., 2005) while Fondue is expressed in the fat body under control of the Toll signalling pathway (Scherfer et al., 2006). This strongly suggests that cellular and humoral factors are required in blood coagulation with contributions from both haemocytes and the fat body. Therefore, the lack of a signal sequence in TF gene (like PPO, another enzyme expressed in haemocytes) suggests that its release from haemocytes may be a key step in the initiation of coagulation. Most recently Wang et al proposed a conserved innate immune mechanism based on TF’s ability to use a potential microbial surface substrate to sequestrate and immobilise bacteria to the clot formed in blood coagulation (Wang et al.,
This interesting piece of work provides direct evidence for the blood coagulation factor to directly bind to microbes in the process of blood clot formation.

The *Drosophila* ‘inflammatory response’

All organisms have developed various mechanisms to maintain structural and physiological integrity in response both to external injury and to internal disruption. Wound healing, tissue repair and regeneration are essential processes for multi-cellular organisms to survive and proliferate against constant environmental or physical assaults. Therefore it is highly feasible to hypothesize that wound healing is an ancient process that evolved before the divergence of insects and mammals and this view can be supported by evidence from extensive research conducted in various model organisms on the pathways underlying this basic process.

Wound healing depends on complex molecular and cellular networks involving different types of cells and tissues. This complexity increases with the basic mechanisms varying in a tissue-specific, developmental stage dependent, and damage related manner. For example, in fruit fly embryos wound healing occurs rapidly via actin cable assembly and filopodial extension by cells at the wound margin, and proceeds without blood clot formation (Wood *et al.*, 2002; Kiehart *et al.*, 2000). Again, despite the substantial structural differences between *Drosophila* and mammalian epidermis, embryonic wound healing in mammalian embryos appears to be similar to that in *Drosophila*. It is also a rapid process involving actin cable formation without apparent haemostatic or inflammatory response (Martin & Lewis, 1992).

Haemocytes in embryonic wound healing

Although blood clots are not formed and required during the embryonic wound healing, microarray analysis comparing the transcriptional profile of wild type and haemocyte-absent embryos still revealed interesting haemocyte signature genes involved specifically in wound
healing. These include phospholipase A2 conserved also in the mammalian inflammatory response (Stramer et al., 2008). From this study, a Drosophila orthologue of a novel mouse inflammatory-responsive gene Growth Arrest and DNA Damage-inducible gene 45 (GADD45) (Takekawa & Saito, 1998) was found to be induced in the damaged epidermal cells. This finding reinforces the idea that inflammatory responses are ancient processes for organisms to respond to danger signals. JNK signalling has also been reported to be essential in the epithelial wound healing in the embryo (Wood et al., 2002) The earliest signal that triggers the haemocyte attraction to a wound in embryos has been recently identified as the calcium wave from the damaged epithelial cells following immediate laser wounding. Blocking this calcium flash inhibits H$_2$O$_2$ synthesis which relies on the activation of an NAPDH oxidase, DUOX (Razzell et al., 2013). In response to H$_2$O$_2$ that transiently outcompetes developmental migrating cues, haemocytes are quickly recruited to the injury site in the embryo (Moreira et al., 2010). The establishment of calcium flux-induced H$_2$O$_2$ production in the inflammatory response associated with wound healing from Drosophila will certainly give more insights into the signalling events taking place in wound induced inflammation in mammals.

**Haemocytes in tissue injury and wound repair in the larva**

The mammalian epithelial tissue can summon a set of humoral and cellular reactions lasting from days to months in response to tissue damage until the damage cap is properly closed and the injured cells or tissues are removed and replaced. The reactions typically include the rapid formation of a blood clot at the injury site and recruitment of inflammatory blood cells followed by spreading of the damaged epithelium across the wound gap to restore tissue integrity (Of & Healing, 1999; Singer & Clark, 1999). Likewise, Drosophila larval wound healing shares many similarities to postembryonic wound healing in mammals. By developing an aseptic puncture wounding in the third instar larva in combination with *in vivo*
life imaging, Galko and Krasnow (Galko & Krasnow, 2004) have established a system to study the process of *Drosophila* postembryonic wound healing. They have characterised the wound healing process by three key stages: 1) primary clot formation during blood coagulation: larvae bleed following the puncture wounding and the primary clot forms in the wound gap; 2) scab and syncytium formation: the primary clot is further cross-linked and hardened by melanisation to form a scab while epidermal cells surrounding the primary clot migrate toward it and then fuse to form a syncytium; and 3) central syncytium formation: more epidermal cells are attracted to the initial syntecium and a larger central syncytium forms. JNK pathway is activated in the epidermal cells of the syncytium in a gradient manner to emanate signals for the epidermal cells to move along or through the wound clot to rebuild a continuous epithelium with its basal lamina and apical cuticle lining. Crystal cells are the haemocyte required in the formation of the scab. The scab stabilises the wound site, establishes a physical barrier to the external microbes, prevents the over-activation of JNK pathway which can result in chronic wounding and provides a scaffold for re-epithelialisation. Scab forming and wound closure are controlled by independent genetic and signalling pathways as re-epithelialisation can still be activated at the wound gap though it never heals in the absence of a scab (Galko & Krasnow, 2004). Presumably, multiple signals must be produced to spatiotemporally co-ordinate this dynamic flow of cellular events involving crystal cells, epidermal cells and also plasmatocytes to remove cell debris for tissue remodelling in addition to phagocytosis of invading microbial pathogens from the open wound. For example: the signals from damaged sites to initiate blood coagulation as discussed in the previous section are still to be identified. What also remains unknown at present is the signal to attract crystal cells to the primary clot, the signals during the formation of a scab (perhaps from crystal cells or plasmacytes) to negatively modulate the JNK activity and the signal to recruit plasmatocytes to the wound site. These signals may be
able to behave like mammalian chemokines or cytokines and possess distinct characteristics in terms of the range and the different signalling pathways they can activate. In addition, there could also be mitogenic signals from apoptotic cells to stimulate cell migration and regeneration (Bergmann & Steller, 2010). Although the identity of these invertebrate inflammatory signalling molecules remains largely unknown, in combination with studies on other arthropods the interconnection between inflammatory responses and wound healing seems to be phylogenetically conserved (Eleftherianos & Revenis, 2011; Theopold et al. 2004). Recent studies have begun to reveal the molecular identities of some of these signals.

A blood borne Pvf1 ligand has been found to be expressed by epidermal cells at the wounding edge and to function in an autocrine manner to activate the motility of epidermal cells in wound closure (Wu et al., 2009). In a study by mutant screening using crystal cell rupture and melanisation as the readout, Bidla et al. reported that the rapid rupture of crystal cells and subsequent local melanisation in the clot at injury depended on the JNK pathway and on Eiger, the Drosophila homologue of tumour necrosis factor (Bidla et al., 2007). The most interesting finding is that endogenous signals such as Eiger released from crystal cells and plasmatocytes undergoing apoptosis followed by secondary necrosis can function independently of microbial elicitors in triggering the PPO activation, which can support the idea that endogenously induced ‘death signals’ initiate inflammatory and repair responses. Nevertheless, the molecular nature of the signal to initiate JNK pathway by the mechanical stress for the crystal cells to rupture still remains to be discovered to date.

In a study to reveal the genetic and molecular networks in control of systemic wound response after physical wounding in Drosophila larvae and adult flies, Nam et al. reported that a redox signal released from proPPO activation via the blood borne serine protease Hayan is required for the downstream activation of JNK signalling to protect remote internal tissues from systemic wound response induced by local physical trauma (Nam et al., 2012).
Forced expression of Hayan in the haemocyte, but not in other tissues, rescued the wound induced mortality in the *hayan* loss of function mutant suggesting that a redox dependent mechanism communicates between circulating haemocytes (most probably plasmatocytes) and the remote internal tissues in a process similar to systemic inflammation in mammals. The cellular and genetic basis of wound healing has recently been studied in detail by using a *Drosophila* embryo laser wounding model and it is revealed not surprisingly that a coordinated process exists involving myosin, E-cadherin, Echinoid, the plasma membrane, microtubules and the CDC42 small GTPase which respond dynamically during wound repair (Abreu-Blanco *et al.*, 2012; Abreu-Blanco *et al.*, 2011). The wound healing mechanism in *Drosophila* larvae was also explored by the development of a targeted large scale *in vivo* RNAi screen in the larval epidermis. Likewise, in the embryo, components in the JNK pathway and genes involved in the remodelling of actin cytoskeleton also actively participate in the larval wound healing (Lesch *et al.*, 2010). Key genes in the haemocyte mediated innate immunity and in the *Drosophila* inflammatory response are summarised in Table 1. For recent reviews on the topic of wound healing, see Ríos-Barrera & Riesgo-Escovar, 2013; Belacortu & Paricio, 2011 and Belacortu & Paricio, 2011.

**Haemocytes in infection induced inflammation**

The last decade has witnessed a rapid growth of research in gut immunity in *Drosophila* (reviews in Kounatidis & Ligoxygakis, 2012; Royet, 2011). In 2009 a genome-wide RNAi screen revealed a large numbers of genes in both haemocytes and the fat body to be regulated following intestinal *Serratia macescens* infection (Cronin *et al.*, 2009). By ontology enrichment analysis, this study found a strong enrichment of genes in haemocytes implicated in processes including phagocytosis, responses to external stimuli and vesicle trafficking. The critical role of JAK/STAT signalling in the gut immunity was reinforced to function through regulation of intestinal stem cell proliferation which controls the gut epithelial cell
haemostasis. Though potential signalling pathways were not the focus of this genome-wide
analysis, the large number of genes to be either upregulated or downregulated in haemocytes
suggests important modulatory roles that haemocytes can play in organ to organ
communication in response to internal infection or inflammation. Recently, Juang and
colleagues found that intestinal ROS signal triggers a systemic AMP expression in the fat
body following oral feeding of Ecc15 (Erwinia carotovaro subsp.) to larvae. The ROS stress
in the gut induces NO expression and transduces the signal to haemocytes by a NO dependent
pathway (Wu et al., 2012). NO-dependent signalling mediated by haemocytes has been also
observed following gastrointestinal infection by Candida albicans (Glittenberg et al., 2011).
In the absence of haemocytes, the AMP production was greatly reduced but not completely
abolished in the fat body. Though ROS has long been recognised to be involved in the
initiation of inflammatory bowel diseases in humans (Rezaie et al., 2007), this recent finding
from Drosophila can offer further insight into the potential role of macrophages in triggering
a systemic inflammation during inflammatory bowel diseases. In addition, gut-associated
macrophage-like cells were also found in the larval gut and their number was regulated by the
PI3K signalling pathway (Zaidman-Rémy et al., 2012). This situation closely resembles the
mouse colitis model (for review see Lin and Hackam, 2011). Taken together, research in
Drosophila has revealed many similarities in the important signalling roles macrophage-like
blood cells can fulfil during either local tissue-specific or systemic inflammation following
internal infection. On the other hand, more recently, Panayidou & Apidianakis have given a
comprehensive review on using Drosophila as the model to study interlinking mechanisms
underlying intestinal cell proliferation, differentiation and maintenance during bacterial
infection and intestinal stress. The authors therefore proposed a regenerative inflammation
phenomenon independent of haemocytes conserved between Drosophila and mammals in
cancer progression (Panayidou & Apidianakis 2013).
Cancer and immunity in *Drosophila*

*Drosophila* as the model to study haematopoiesis and its associated leukaemia

In adult mammals such as humans and mice, bone marrow is the organ that houses haematopoietic stem cells (HSC) which give rise to both the myeloid and lymphoid lineage. Mammalian HSC possesses the ability to self-renew and the pluripotency to differentiate into a great variety of blood cells in response to signals from its microenvironment, which has been termed the HSC niche (review in Wang & Wagers, 2011). The HSC niche has been a subject for vigorous research since the concept has been fully accepted (Lensch, 2012; Wang & Wagers, 2011). The dynamic communication between HSC and its niche has been shown to be fundamental in the control and regulation of haematopoietic process in vertebrates. Any dysfunction in the genetic and cellular mechanism underlying the HSC and niche interaction can result in blood borne cancers such as AML (Oh & Humphries, 2012). However, the structural and cellular complexity of the bone marrow niche has hindered the progress of fully understanding the basic genetic and molecular events fundamental to the haematopoietic process and their application in potential human diseases. The *Drosophila* larval PSC as discussed previously functions as a primitive niche to instruct the different fate that the prohaemocyte would adopt or to help the prohamemocyte to maintain its stem cell status (Crozatier & Meister, 2007). The LG primary lobes represent a very simplified HSC and niche model compared to its mammalian counterpart. Although there are obvious restraints such as the limited number of differentiated blood cell types and the complete absence of lymphocytes, this simplicity can further our understanding of the basic signalling and cellular communication mechanisms involved in particular between the HSC and its microenvironment (review in Crozatier & Vincent, 2011).
Research in *Drosophila* haematopoiesis has revealed a number of pathways in control of prohaemocyte proliferation and differentiation. Overexpression of the *Drosophila* JAK gene *hop<sup>Tum-1</sup>* causes proliferation of prohaemocytes and leads to melanotic tumour formation in the LG (Harrison *et al.*, 1995). This discovery preceded the demonstration that mutated constitutive activation of JAK/STAT signalling could result in human leukaemia (Lacronique *et al.*, 1997). Compared to the vertebrate system, the *Drosophila* JAK/STAT pathway is much simplified and shows nearly complete absence of genetic redundancy. The *Drosophila* genome encodes only three upstream ligands of JAK/STAT pathway Unpaired (Upd1-3) while the mammalian JAK/STAT can be activated by a large group of cytokines and growth factors. Dom is the only transmembrane receptor upstream of one JAK kinase (Hop) and the one STAT transcription factor (STAT92E). Therefore the misexpression of a dominant-active form of STAT92E can also promote tumourigenesis in the eye of the *Drosophila* adult flies and melanotic tumour formation in the larva (Ekas *et al.*, 2010). A systematic genome-wide RNAi screening for genes required for JAK/STAT pathway activity in cultured *Drosophila* haemocyte-like cells also identified interacting genes that can function as suppressors of leukaemia-like blood cell tumours in humans (Müller *et al.*, 2005). Increasing evidence from clinical research in human AML has pinpointed a role for JAK/STAT signalling pathway to be implicated in AML pathogenesis. In particular an activating mutation on the human JAK2 has been discovered to be responsible for various forms of AML (Vainchenker & Constantinescu, 2013; Lee *et al.*, 2013).

Apart from the JAK/STAT signalling pathway, the Notch, Hg, Wnt, and JNK pathways have all been identified as regulators of prohaemocyte fate (Mandal *et al.*, 2007; Owusu-Ansah & Banerjee, 2009; Sinenko *et al.*, 2009). Hg signalling in the PSC has been identified to maintain the undifferentiated fate of prohaemocytes in the larval medullary zone of the...
primary lobes in the LG (Mandal et al., 2007). In a mouse B cell lymphoma model, Hg signalling from the stromal cells was also shown to provide an important survival signal for B- and plasma-cell malignancies in vitro and in vivo (Dierks et al., 2007). The Hg signalling pathway was also discovered to be required in the maintenance of cancer stem cells of chronic myeloid leukemia (CML) (Zhao et al., 2009). Loss of Smoothened (Smo) the downstream transmembrane G protein coupled receptor in the JAK/STAT pathway causes depletion of CML stem cells whereas constitutively active Smo augments CML stem cell number and accelerates the disease. These studies are reminiscent of Hg signalling implicated in the communication between PSC niche and the prohaemocyte in Drosophila haematopoiesis.

Notch signalling pathway has been well conserved in vertebrate haematopoiesis, in particular in lymphoid cell commitment (Tanigaki & Honjo, 2007; Radtke et al., 2005). In Drosophila LG, the Serrate-mediated Notch signalling from the PSC is required to maintain normal levels of Col transcription and thus PSC cell identity (Krzemień et al., 2007). In addition, a non-canonical and ligand-independent activation of Notch signalling has also been reported to determine crystal cell fate in the LG. The Drosophila orthologue of mammalian hypoxia-inducible factor–a (HIF-a), Sima, activates full length Notch receptor under conditions of normal oxygen availability and commits the prohaemocyte to the crystal cell lineage (Mukherjee et al., 2011). In fact, the first human Notch was originally identified from human Acute lymphoblastic leukemia (ALL) and since then Notch signalling has been discovered to be involved in many forms of human leukaemia (review in Pancewicz & Nicot, 2011).

Wnt signalling has been shown to promote proliferation of prohaemocytes and prevent differentiation at the same time by controlling the PSC niche (Sinenko et al., 2009). In addition, Wnt signalling also positively regulates the proliferation and maintenance of PSC cells while inhibition of Wnt signalling results in fewer PSC cells than observed in control
flies. Likewise in vertebrates, Wnt signalling has been reported to play important roles in the HSC homeostasis and maintenance of its microenvironment for self renewal (for the most recent review see Seke Etet et al., 2013) and thus has been implicated in many forms of haematologic malignancy such as AML (Gandillet et al., 2011) and CML (Nagao et al., 2011). To elucidate the controversial role of reactive oxygen species (ROS) in the haematopoietic system, work done by Owusu-Ansah and Banerjee set out to make use of the Drosophila haematopoietic model to study levels of ROS in the in vivo proliferation and differentiation of prohaemocytes. It was found that increased levels of ROS promote the differentiation of prohaemocytes while inhibition of the ROS level delays the differentiation of prohaemocytes into mature haemocytes. Interestingly, through a downstream signalling pathway that involves JNK and FoxO activation as well as Polycomb downregulation, increasing the haematopoietic progenitor ROS beyond their basal level triggers premature differentiation of prohaemocytes into all three mature haemocytes found in Drosophila. Therefore a moderately high level of ROS can be the developmental signal for the population of haemocyte progenitor to commit to lineage differentiation (Owusu-Ansah & Banerjee, 2009) (Fig.2B). In mammals, higher ROS level was also observed during the common myeloid progenitor differentiation in response to oxidative stress (Tothova et al., 2007). A recent study by Dragojlovic-Munther and Martinez-Agosto demonstrated that the tumour suppressors TSC and PTEN also have important roles in controlling blood progenitor proliferation through a common TOR- and 4EBP-dependent pathway in the LG. Loss of function of Tsc2 or Pten in prohaemocytes increases TOR signalling and causes overgrowth of the LG by haemocyte hyper-proliferation accompanied by a higher level of ROS. This study illustrates further how TSC and PTEN influence TOR function in response to physical stress such as starvation, hypoxia or increased ROS level during infection (Dragojlovic-
Interestingly, the PTEN/mTOR signalling pathway has indeed been the therapeutic target in the treatment of human leukaemia (Martelli et al., 2011). Collectively these studies demonstrate the strength of Drosophila as an excellent model to study the HSC and its microenvironment interaction and shed light on the potential for therapeutic prevention of various haematological malignancies by dissecting its underlining genetic and cellular mechanisms.

**Drosophila human leukaemia model**

Apart from the elucidation of basic molecular signalling pathways in the HSC and its niche interaction, Drosophila can also be used directly to model human leukaemia. In addition to overactivation of JAK/STAT signalling, human AMLs can result from the chromosomal translocation of the transcription factor AML1, a RUNX domain protein, to form a protein fusion product with ETO (Hatlen et al., 2012). Targeted expression of human AML1-ETO fusion transcription factor in the haemocyte lineage cells by using the UAS/GAL4 system caused human leukaemic-like phenotypes such as hyper-proliferation of the circulating haemocytes resulting from the expansion of prohaemocytes in the LG (Sinenko et al., 2010). The successful establishment of the AML1-ETO leukaemia model in Drosophila allowed a rapid tissue-specific genetic screening to identify suppressors for the hyperproliferation phenotype. The authors thus were able to show that ROS is a signalling factor promoting maintenance of normal as well as aberrant haemocyte precursors which suggested the importance of antioxidant enzymes and their regulators as targets for further study in the context of leukaemia (Sinenko et al., 2010). In another independent genetic screening for modifiers in the AML1-ETO Drosophila model, Osman et al identified calpainB as required for AML1-ETO-induced blood cell disorders in Drosophila by using an in vivo RNAi-based screen for suppressors of AML1-ETO. Remarkably, calpain was also found to interact with
AML1-ETO in the human leukemic blood cell line Kasumi-1 (Osman et al., 2009). Therefore these studies have paved new avenues in our understanding of the pathogenesis of haematopoietic associated leukaemia by developing and recapitulating the fundamental features of the disease in Drosophila and will contribute significantly to more precise and effective AML therapy.

_Drosophila and human solid tumour_

During the past two decades, the completion of the _Drosophila_ genome and the development of advanced genome editing tools have given unprecedented stimulus and fast expansion on the use of Drosophila as a model for cancer. In particular, _Drosophila_ has been instrumental for the discovery of three fundamental mechanisms involved in tumour progression and metastasis: 1) the role of Hippo signalling pathway in control of cell growth and survival together with Scrib/Dlg/Lgl signalling pathway in cell polarity to regulate organ sizes; (reviews in Harvey et al., 2013; Enomoto & Igaki, 2011; Martin-Belmonte & Perez-Moreno, 2012); 2) the _in situ_ cell competition mechanism for morphogens during the formation of epithelium where slow growing cells are outcompeted and removed (review in Levayer & Moreno, 2013); and 3) apoptosis induced mitogenic signals in compensatory proliferation to replace surrounding damaged tissues (review in Fan & Bergmann, 2008). All these pathways underline the basic cellular communication, shaping and tissue organisation which if disrupted can lead to tumourigenesis and facilitate tumour metastasis.

Despite a very short life span, cancer research to investigate directly the tumour progression and invasion in _Drosophila_ can be dated back to nearly a century ago. It was observed that _Drosophila_ can naturally develop hereditary tumours; carcinogens such as X-rays have been used to discover mutants with abnormal growth (Salomon & Jackson, 2008; Bridges & Brehme, 1944). The first fly strain carrying a mutation in a tumour suppressor gene called
lethal giant larvae (lgl) was isolated 70 years ago. However only recently has Lgl been identified as a component of a signalling circuit in the regulation of apico-basal polarity in epithelial cells: Scrib/Dlg/Lgl (Humbert et al., 2008). Loss of function of the genes in this pathway can lead to the formation of neoplasma in the brain and imaginal discs in the developing larva. Based on the neoplastic phenotype observed in the scrib signalling mutants, Gateff was one of the pioneers who carried out more screenings to isolate recessive lethal mutations that could promote neoplastic overgrowth in the brain, imaginal discs or haematopoietic organ (Gateff, 1978). A tissue transplantation technique was developed from Gateff’s study to assess the malignancy and invasive capacity of the neoplastic clones arising from various tissues. This technique involves the implantation of cancerous cell clones into the abdomen of wild type adult flies. Flies implanted with malignant cell clones usually die within 2 weeks and histological examination usually observes a massive invasion of cancerous cells into various tissues in the adult fly. This method resembles the tail veil intravenous injection of tumour cell lines into the mouse to model cancer in small rodents. This pioneering work fully established the potential for Drosophila to be used as a whole organism model to recapitulate key stages in cancer pathogenesis. The in vivo lac-Z reporter gene expression system can be utilized to quantify the proliferation rate and metastatic index of donor tumours in a tissue specific context in the normal wild type host (Beaucher et al., 2007). By using mutant clones of labelled cells such as GFP within a specific tissue, the invasive and metastatic behaviour of cancer cells can be observed in situ by in vivo live imaging (Brumby & Richardson, 2003; Pagliarini & Xu, 2003). The recent development of MARCM system allows the establishment of large homozygous mutant cells clones within a normal or heterozygous tissue. A similar system is the FLP/FRT site directed recombination that can also generate genetic mosaics in the targeted organ or tissue. The mutant cells usually overexpress a UAS-tagged transgene fused with a reporter gene like GFP to allow
imaging. These systems have been used by two independent groups to identify genes in cooperation with known oncogenes to induce tumour growth and invasion in the context of normal tissue (Brumby & Richardson, 2003; Pagliarini & Xu, 2003). By overexpressing an activated Ras (Ras^{v12}) in the *Drosophila* eye imaginal disc and screening for the entire *Drosophila* genome, Pagliarini and Xu identified Scrib as the promoting factor for the metastatic transformation of the otherwise benign tumour caused by overexpression of Ras oncogene alone (Pagliarini & Xu, 2003). Similar work by using the *Drosophila* eye as an *in vivo* ‘test tube’ to investigate genetic interactions during tumour progression has proved to be extremely fruitful in dissecting the Scrib tumour suppressor signalling pathway and Hippo pathway and human oncogenes and tumour suppressors such as Ras and PTEN. Critical reviews of the most recent progress in the use of *Drosophila* in cancer modelling and therapeutic potentials can be found elsewhere (Gonzalez, 2013; Tipping & Perrimon, 2013; Miles *et al.*, 2011; Stefanatos & Vidal, 2011).

**Innate immunity in *Drosophila* tumour progression and tumour invasion model**

Like other aspects of tumour progression that can be modelled and studied comparatively in *Drosophila*, the interplay of innate immunity mediated inflammation and tumour growth and invasion can be investigated accordingly. Of note is the establishment of *Drosophila* intestinal tumour model to study the molecular and cellular mechanisms linking inflammation and cancer pathogenesis (reviewed in Christofi & Apidianakis, 2013). The *Drosophila* cytokines Upd1-3 have been reported to activate the JAK/STAT signalling in promoting intestinal stem cell proliferation upon enteric infection or JNK-mediated stress response and thus the innate immunity plays essential roles in gut tissue homeostasis (Jiang *et al.*, 2009). Moreover, overexpression of *Drosophila* Ras oncogene in association with bacterial infection can result in the formation of intestinal dysplasia (Apidianakis *et al.*, 2009). More recently it
is revealed that both the activation of Ras and bacterial induced IMD signalling can activate JNK pathway, which culminates in the up-regulation of matrix metalloproteinase 1 and thus cell invasion and migration (Bangi et al., 2012). Two studies have also emerged to explore the roles of haemocytes in the tumour progression and both relied on induced Ras\textsuperscript{v12}Scrib\textsuperscript{-/-} model in the Drosophila eye discs, which has been used to offer complementary views of systemic innate immunity to tumour growth and invasion. Under the condition of established tumour, the Drosophila haemocyte could be recruited to the tumour site and tumour associated haemocytes are the major source of Drosophila TNFa, Eiger, to promote the growth and invasion of the tumour cells into other tissues (Cordero et al., 2010), while a TNF-dependent mechanism in Drosophila eliminates cells deficient for the polarity tumour suppressors Scrib or Dlg to maintain the tissue homeostasis and keep any malignant growth in check (Brumby & Richardson, 2003). In an earlier study by Tian Xu and co-workers, the same eye imaginal disc derived Ras\textsuperscript{v12}Scrib\textsuperscript{-/-}tumour was found to induce a systemic proliferation of haemocytes via the JNK-JAK/STAT signalling cross-talk conserved also in response to tissue injury. The disrupted tumour basal membrane recruited circulating haemocytes in a manner reminiscence of wound healing (Pastor-Pareja et al., 2008). This study also modelled for the first time in Drosophila the anti-tumour effect of a systemic inflammation response induced by mechanical injury and provided critical insight into the cross-talk between the different signalling pathways in regulating the multiple step progression of tumour.

**Perspective**

Cancer has a complex biology. As a rationalisation, Hanahan and Weinberg identified six major successive changes in human tumour development (Hanahan & Weinberg, 2011). The increasing awareness of this daunting complexity has turned more scientists to develop Drosophila as the complementary genetic tool to dissect each of the hallmark processes and
to offer fundamental insights into the underlying genetic and cellular basis of the disease. In future, the Drosophila model can continue to be used as the reductionist system to investigate more of the cross-talks between these fundamental cancer biological processes. For example, the contrasting roles of immunity to control tumour growth or to be hijacked by the tumour can be modelled in the fly in a temporal and spatial manner. The unique advantage of Drosophila to be used as a whole organism has started to show promising potential in cancer drug screening and testing recently (Dar et al., 2012). This advantage can be fully explored by modelling simultaneously or sequentially physiological processes such as wound or infection induced innate immune/inflammatory response in the progression of cancer (Fig. 3). The study of the role of haemocytes in response to wounding and invasive tumours will shed some light to the fundamentals of macrophages in immunity, inflammation and tumour microenvironment in human cancer.

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References


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Figure legends.

**Figure 1** *Drosophila* haematopoiesis. A) The embryonic haematopoiesis. Two waves of haemocyte generation take place in *Drosophila*. During embryogenesis, haemocytes originate from the precephalic mesoderm as indicated in red in the region of mesoderm anlage (shown in brown). The pink region denotes the embryonic origin that will give rise to lymph gland haemocytes (LGH) in the larva. B) Larval haematopoiesis. The LG (in pink) composes of the primary and secondary lobes and is located in the anterior end of the larva along the dorsal aorta. The sessile haemocyte population distributes diffusely along the segmental borders of the larva and consists of functional differentiated haemocytes and a few prohaemocytes with an embryonic origin (shown in the same red colour). Until the end of the third star, circulating haemocytes including plasmatocytes and crystal cells (small black circles) are derived from the embryonic haemocytes.

**Figure 2.** A) The major transcriptional network in the lineage commitment of *Drosophila* prohaemocytes (Waltzer et al., 2010; Evans et al., 2003; Lebestky et al., 2000). Prohaemocytes express the early GATA transcription factor, Serpent (Srp) for haematopoietic development (Lebestky et al., 2000). The expression of Srp defines the haemocyte anlage from embryogenesis. Later on the Srp expressing prohaemocytes turn on the transcription of the *Drosophila* friend of GATA, U-shaped (Ush). Ush together with Srp activates the expression of another transcription factor Gilial cell missing (GCM) and its isoform GCM2. Activation of GCM/GCM2 turns on the expression of plasmatocyte cell markers and thus commits prohaemocytes into plasmatocyte specification (Fossett et al., 2001). In a small population of prohaemocytes, the expression of a *Drosophila* Runx family member of transcription factors, Lozenge (Lz) antagonises Ush and also inhibits the expression of Gcm/Gcm2. Cells expressing Lz adopt the crystal cell fate (Bataillé et al., 2005). Lamellocytes do not appear in normal circumstances but can be induced rapidly from the LG or sessile haemocytes by parasitic infection in the larva. Most recent lineage tracing studies have pointed out a direct differentiation of lamellocytes from plasmatocytes by the upregulation of Srp and Charlatan (Chn) and downregulation of Ush to suppress the plasmatocyte transcriptional profile (Stofanko et al., 2010). However direct differentiation from the LG prohaemocytes may also contribute to the total population of lamellocytes in particular by the over-activation of JAK/STAT and Toll signalling. B) Compartments of the primary lobes of the LG and key signalling pathways in larval haemocyte specification (Krzemień et al., 2007). The primary lobes are the major sites for the larval haemocyte differentiation. The primary lobe can be divided into three major compartments: the Cortical Zone (CZ), the Medullary Zone (MZ) and the Posterior Signalling Centre (PSC) (Jung et al., 2005). There is also a region that contains a population of intermediate haemocytes from undifferentiated prohaemocytes. This region is sometimes termed Intermediate Cortical Zone (ICZ). The stem cell-like fate of prohaemocytes is regulated by communication between the PSC cells and MZ via filopodia of the PSC cells. Activation of JAK/STAT and Hg signalling by the PSC cells maintains the undifferentiated status of prohaemocytes (Krzemień et al.,...
Meanwhile the transcription factor Collier (Col) expression defines PSC cell identity and is controlled by the Serrate/Notch signalling (Lebestky et al., 2003). Toll signalling is also important in the survival and proliferation of prohaemocytes while increased ROS level and Pvr/Pvf signalling can also contribute to the differentiation of prohaemocytes to plasmatocytes (Qiu et al., 1998; Kockel et al., 2004).

Figure 3. A simple illustration of roles of haemocytes in the Drosophila tumour model. Both tumour (Ras\textsuperscript{v12}Scrib\textsuperscript{\textminus\textminus}) and wounding (as illustrated by a needle) can trigger proliferation of circulating haemocytes in the larva. This is mediated by cytokines (most probably Upd 1-3 shown in purple dots) induced by JNK signalling in response to wounding or the invasive tumour itself. In responses to the cytokines, JAK/STAT signalling in haemocytes or the fat body (not shown) can be activated and thus can further amplify the cytokine expression to promote haemocyte proliferation. Meanwhile disruption of basal membrane in the tumour attracts the adherent of haemocytes and limits the tumour growth perhaps by the synthesis of Drosophila TNFa, Eiger. This cartoon delineates that a step wise proliferation of haemocytes induced by wounding can be used to counter tumour growth. Modelling of these two processes (tumour growth and inflammation) in the Drosophila larva points out the potential beneficial effect that systemic inflammation can exert on tumourigenesis and will open more avenues for the basic research on innate immunity in particular macrophages’ role in human tumour (see text for more detail).
Figure 1: TIFF

A

EH anlage
LGH anlage
mesoderm anlage

B

circulating haemocytes

crystal cells

LG

sessile population