

## **Supplementary Material**

# **Complete genome sequence of *Vibrio gazogenes* PB1: An estuarine bacterium capable of producing prodigiosin from starch or cellulose**

***Bincy Baby<sup>1,2</sup>, Dhanya Vijay<sup>1</sup>, Pretty S. Philip<sup>3</sup>, Afra A. Alnuaimi<sup>1</sup>, Hessa M. Almansoori<sup>1</sup>, Sarah O. Areidat<sup>1</sup>, Gulfaraz Khan<sup>3,5</sup>, Ranjit Vijayan<sup>2,4,5\*</sup> and M. Kalim Akhtar<sup>1\*</sup>***

<sup>1</sup>*Department of Chemistry, College of Science, United Arab Emirates University, Al Ain, United Arab Emirates*

<sup>2</sup>*Department of Biology, College of Science, United Arab Emirates University, Al Ain, United Arab Emirates*

<sup>3</sup>*Department of Medical Microbiology and Immunology, College of Medicine and Health Sciences, United Arab Emirates University, Al Ain, United Arab Emirates*

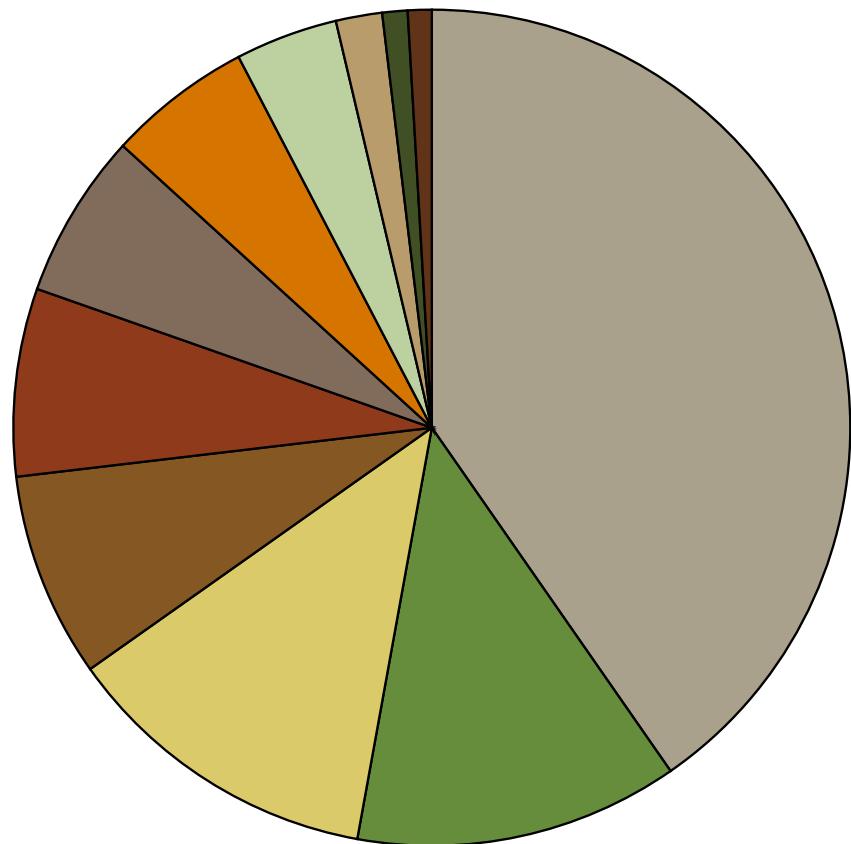
<sup>4</sup>*The Big Data Analytics Center, United Arab Emirates University, Al Ain, United Arab Emirates.*

<sup>5</sup>*Zayed Center for Health Sciences, United Arab Emirates University, Al Ain, United Arab Emirates.*

**\*Corresponding authors:**

E-mails: [ranjit.v@uaeu.ac.ae](mailto:ranjit.v@uaeu.ac.ae), [mk.akhtar@uaeu.ac.ae](mailto:mk.akhtar@uaeu.ac.ae)

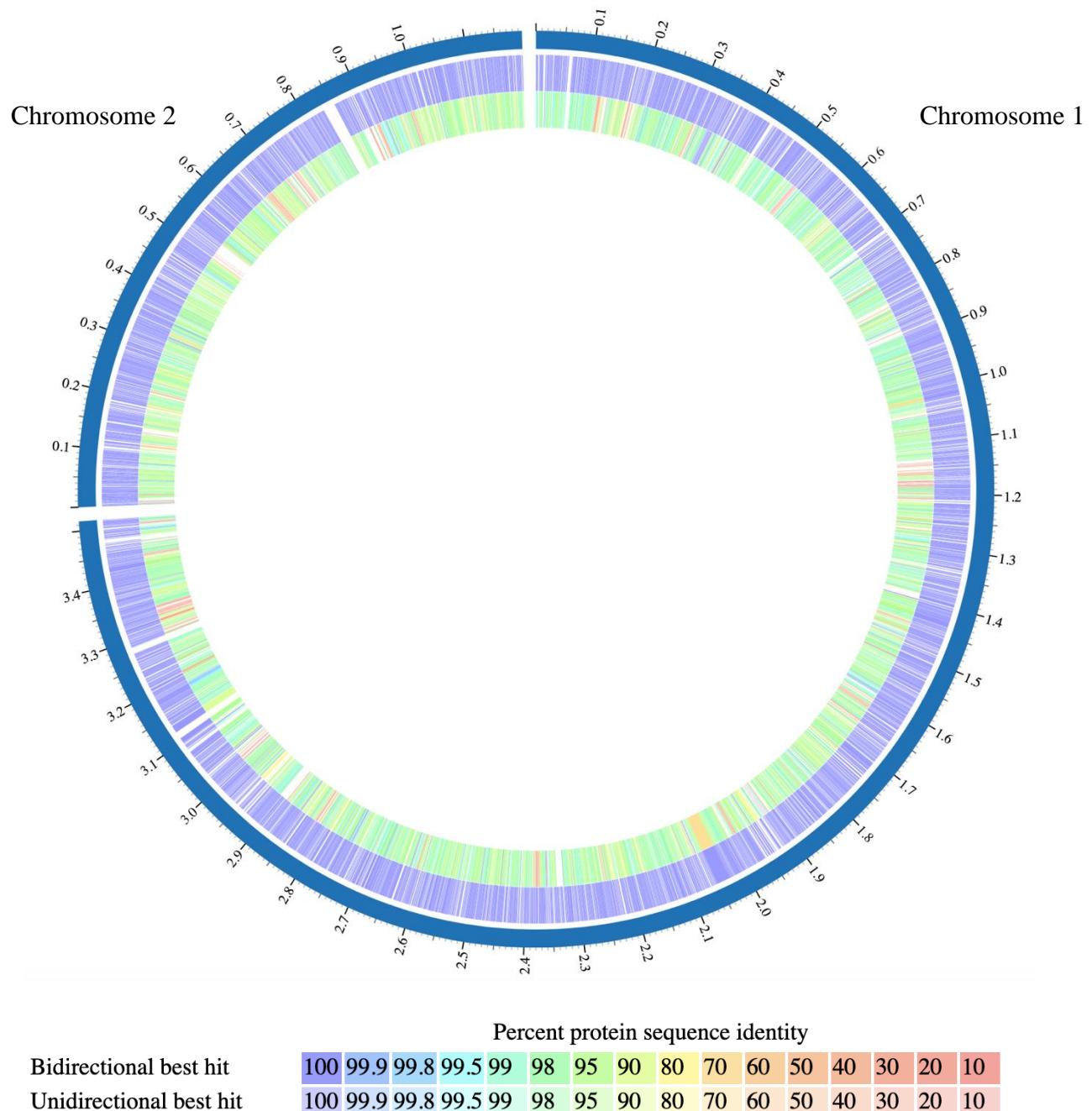
**Figure S1:** Subsystems identified in the *V. gazogenes* PB1 genome. The subsystem classification was produced by RAST annotation in PATRIC.



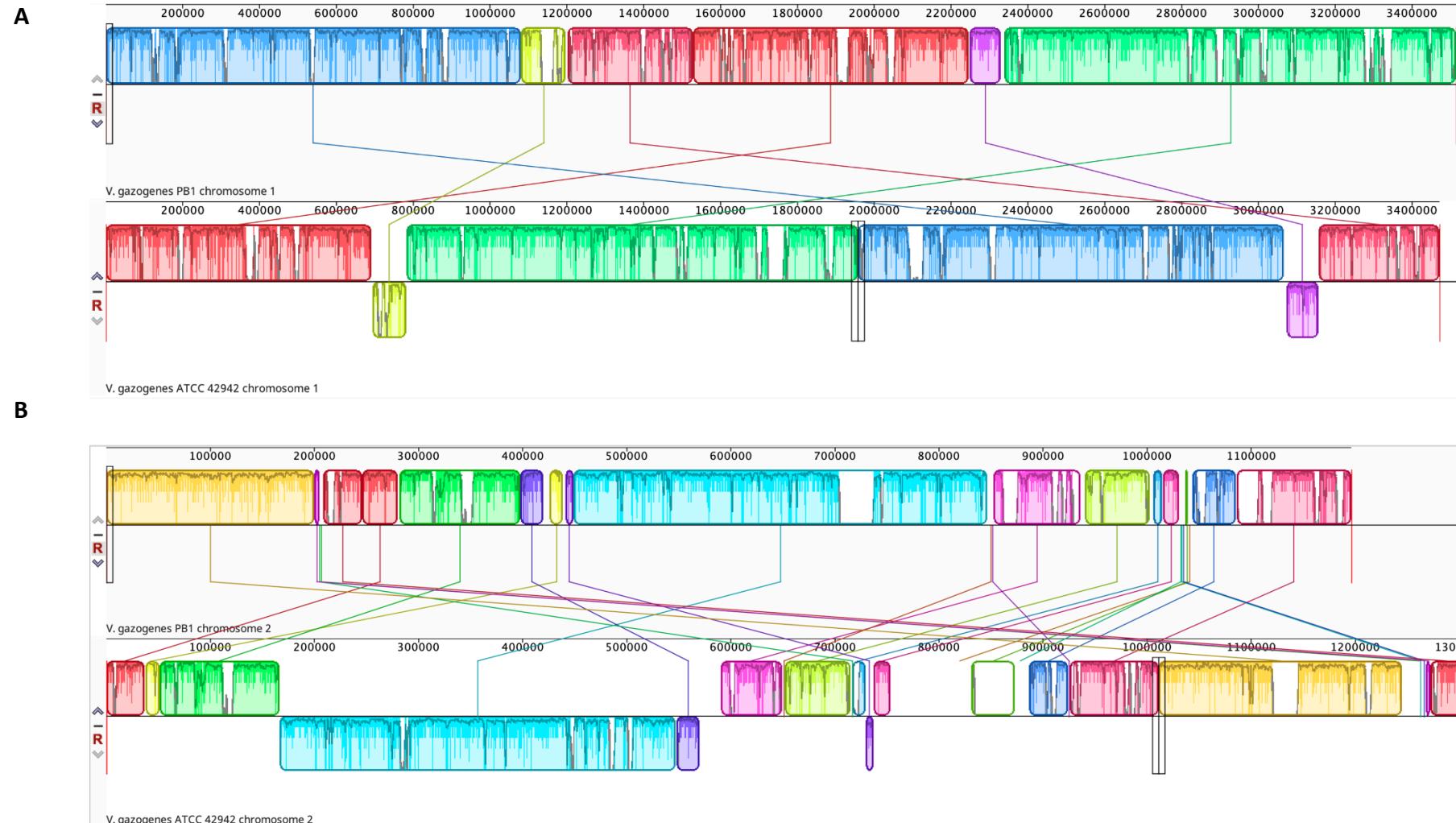
#### Subsystem Counts (Subsystems, Genes)

□	METABOLISM	(92, 2978)
□	PROTEIN PROCESSING	(44, 924)
□	ENERGY	(32, 912)
□	STRESS RESPONSE, DEFENSE, VIRULENCE	(36, 585)
□	CELLULAR PROCESSES	(18, 536)
□	MEMBRANE TRANSPORT	(22, 472)
□	DNAPROCESSING	(21, 412)
□	RNAPROCESSING	(14, 292)
□	CELL ENVELOPE	(5, 132)
□	MISCELLANEOUS	(6, 72)
□	REGULATION AND CELL SIGNALING	(4, 68)

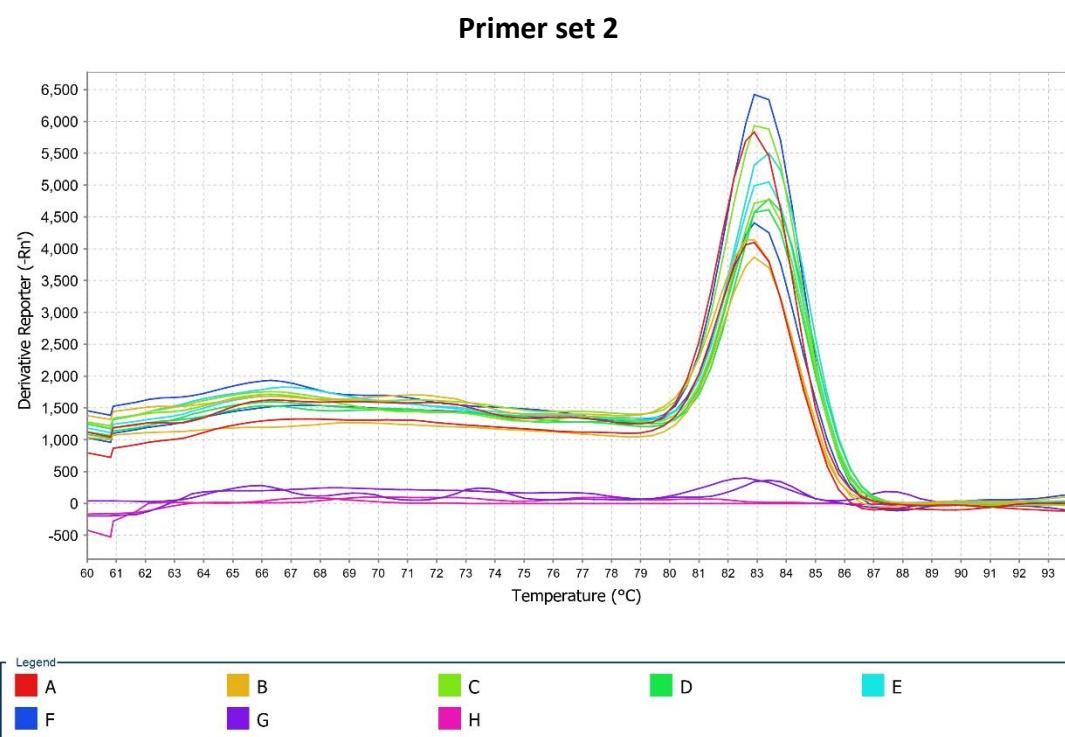
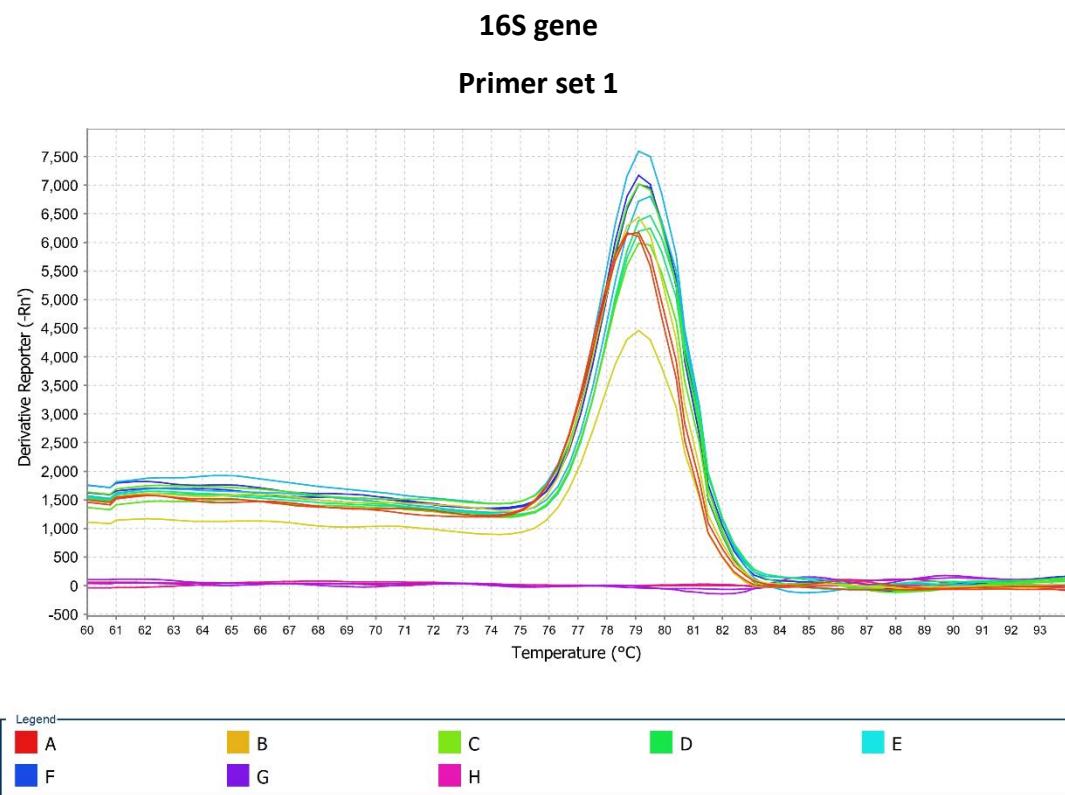
**Figure S2:** Comparison of the proteome of *V. gazogenes* PB1 and *V. gazogenes* ATCC 43942 using PATRIC. From outside to inside the tracks represent the genome size, including the two chromosomes, *V. gazogenes* PB1 proteins and the orthologous *V. gazogenes* ATCC 43942 protein.



**Figure S3:** Whole chromosome alignments of *V. gazogenes* PB1 and *V. gazogenes* ATCC 43942 **(A)** chromosome 1 and **(B)** chromosome 2 generated using progressive Mauve. Homologous blocks are shown in the same colour.

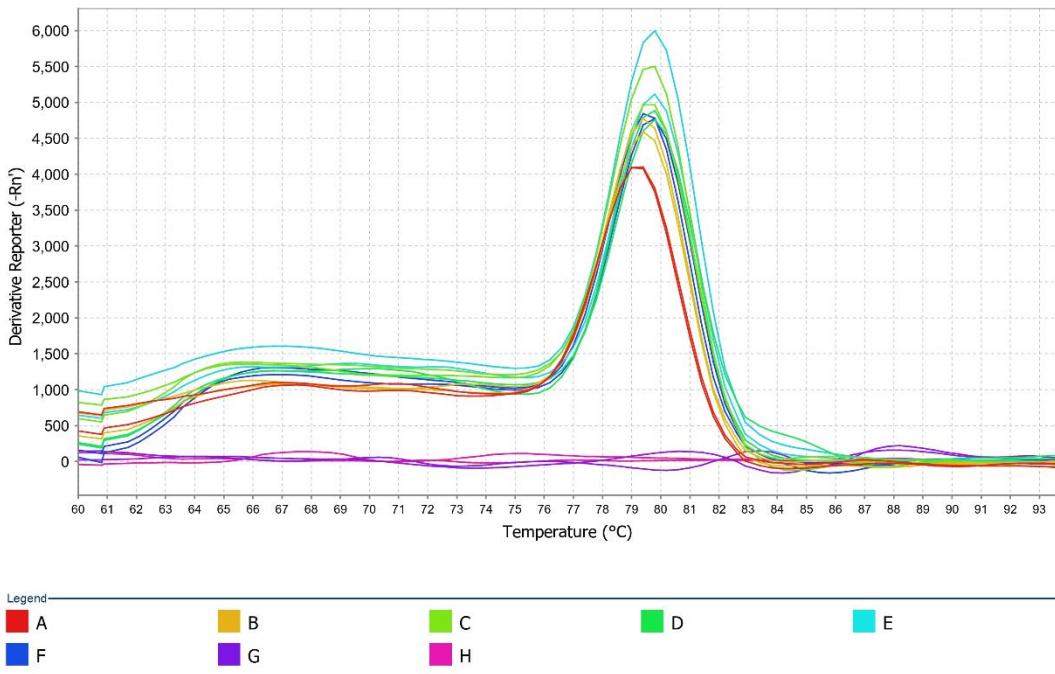


**Figure S4:** RT-qPCR melt curves for target genes related to the metabolism of starch and cellulose.

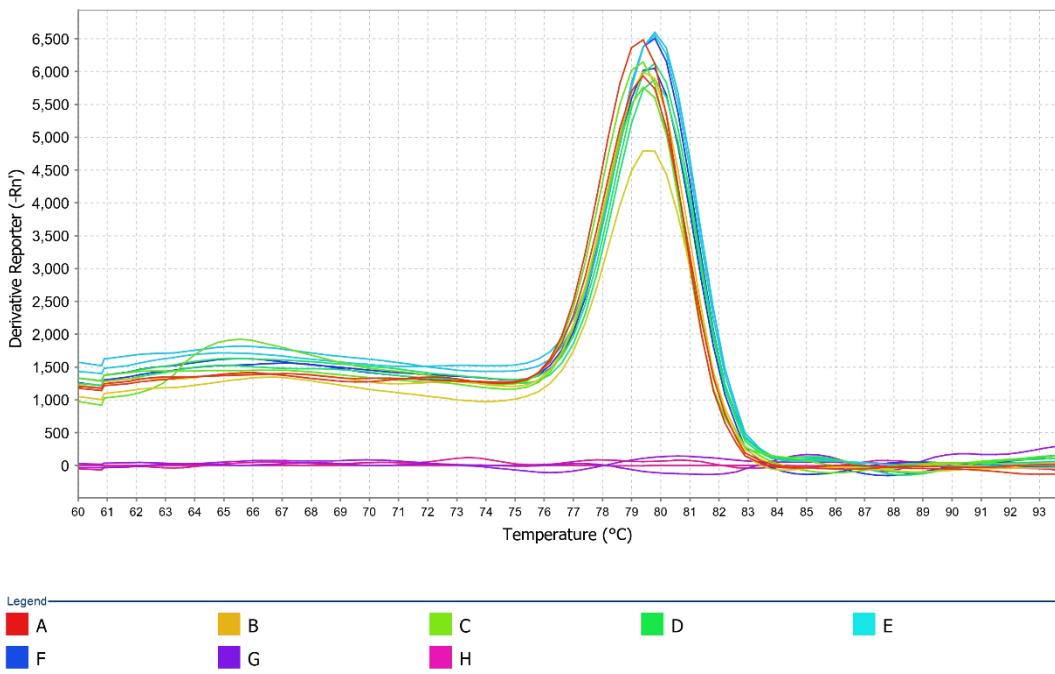


## USP13629.1 $\alpha$ -glucosidase

### Primer set 1

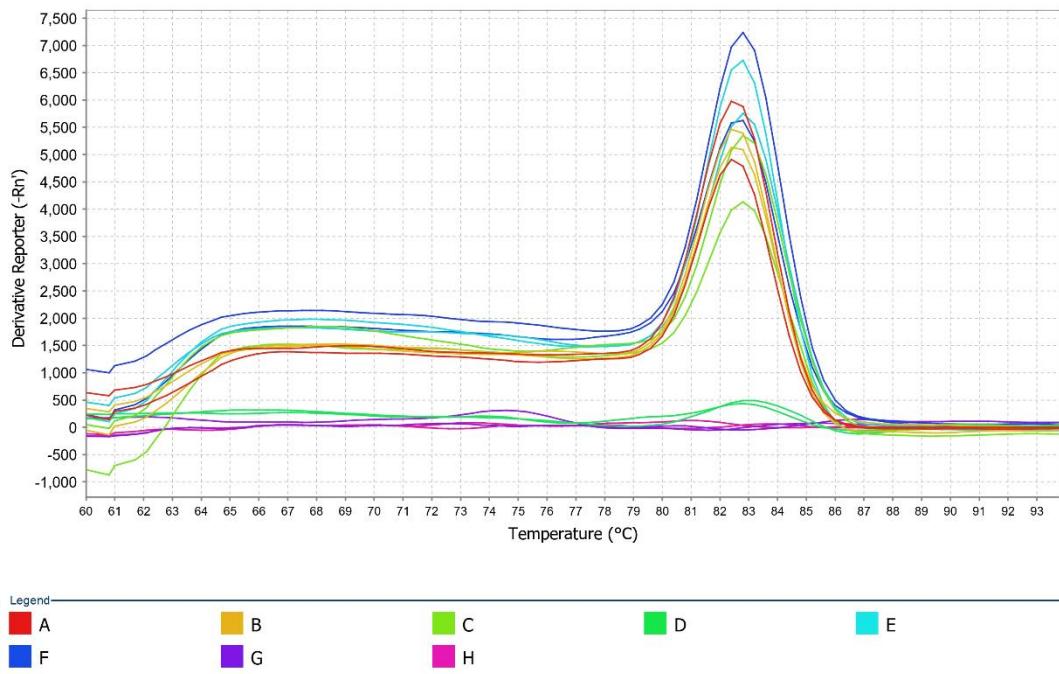


### Primer set 2

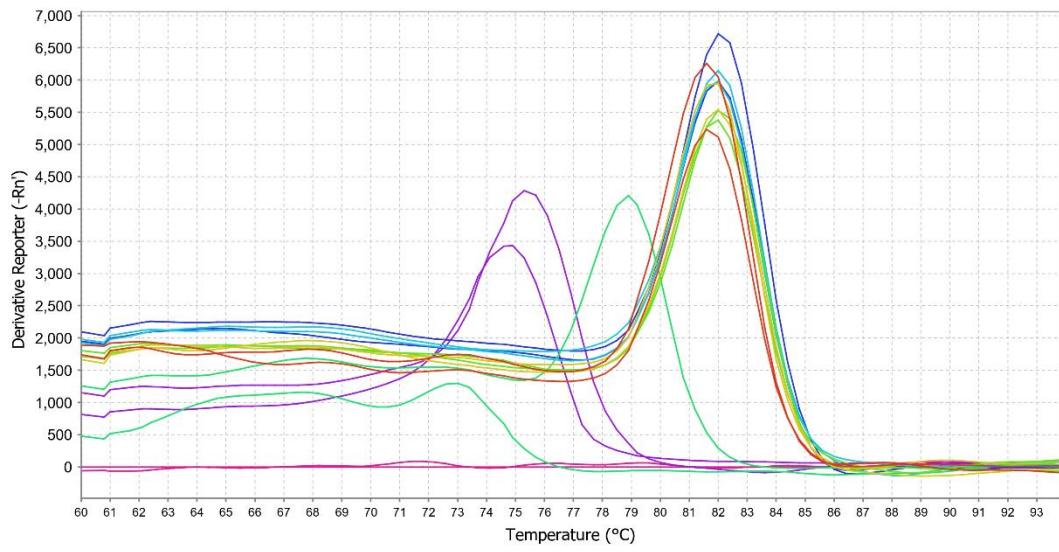


## USP14760.1 exoglucanase

### Primer set 1

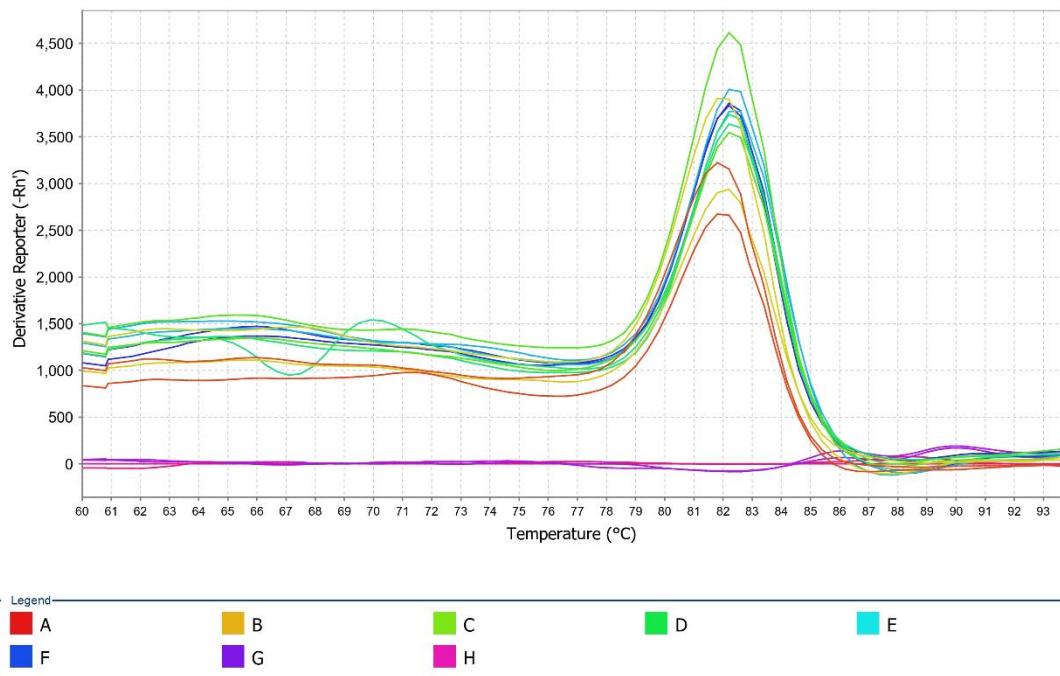


### Primer set 2

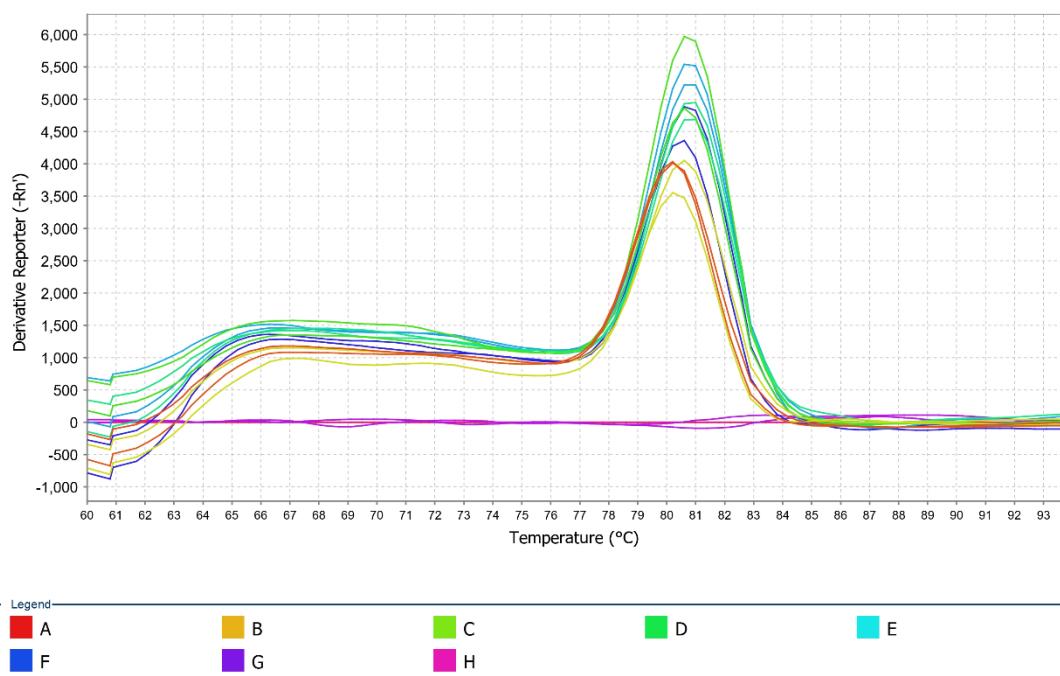


## USP16020.1 isoamylase

### Primer set 1

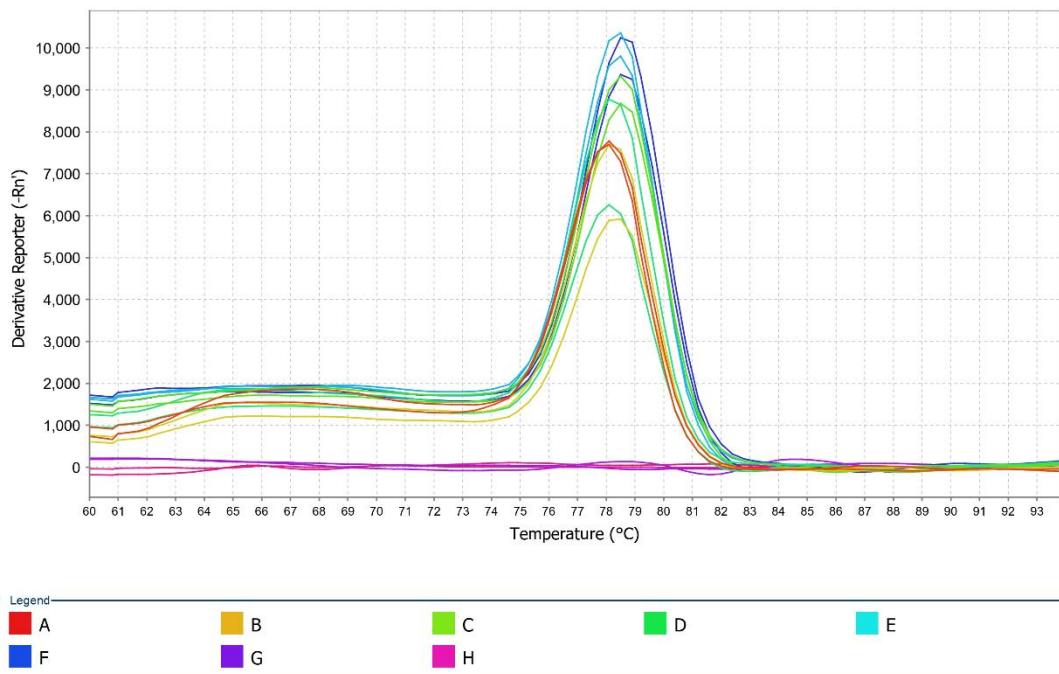


### Primer set 2

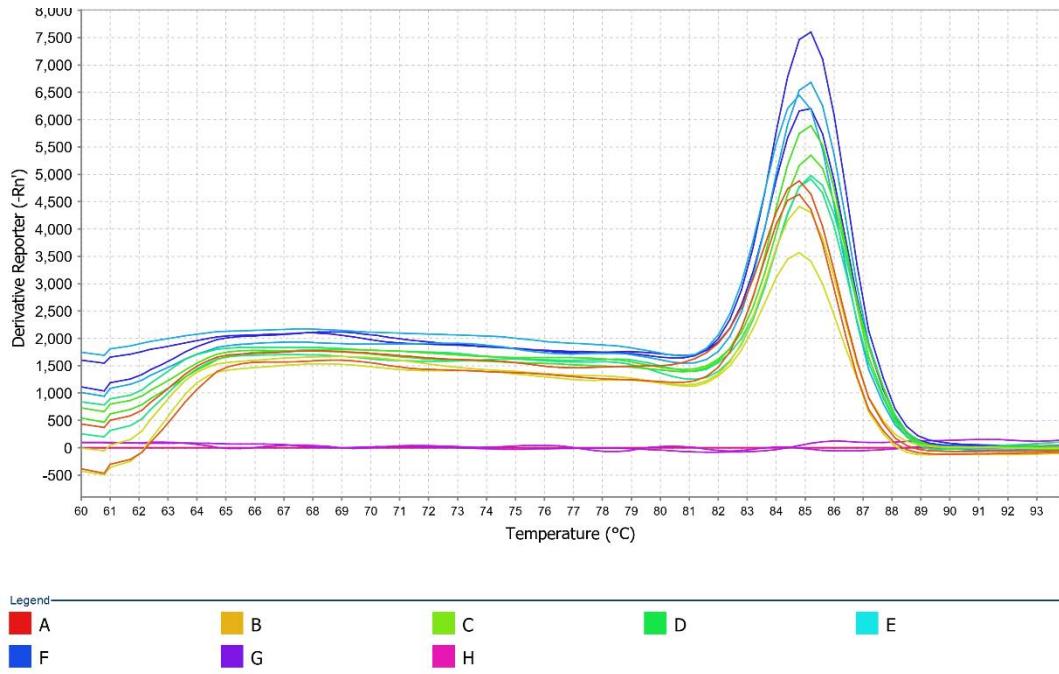


### USP15696.1 $\alpha$ -amylase

#### Primer set 1

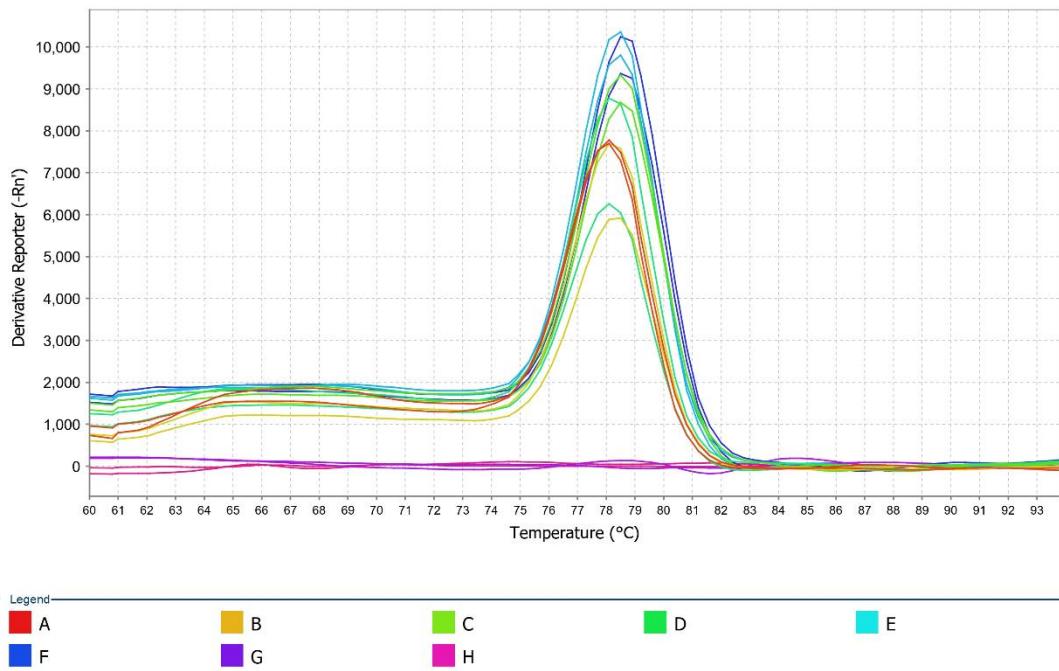


#### Primer set 2

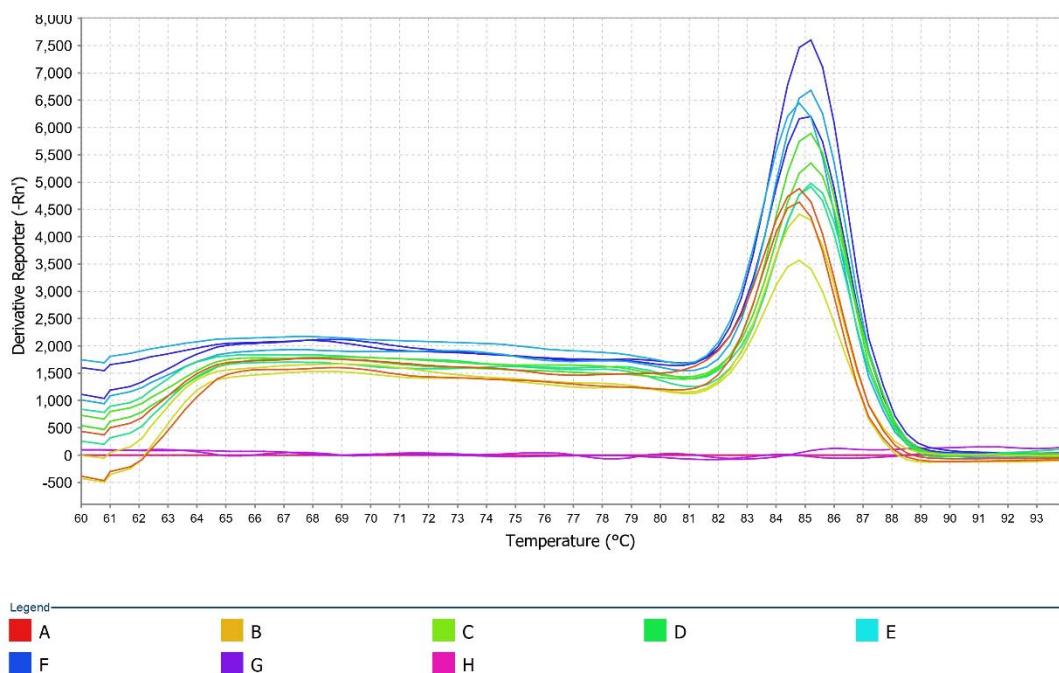


## USP15545.1 $\beta$ -glucosidase

### Primer set 1



### Primer set 2



**Table S1:** Genome attributes of *V. gazogenes* PB1

Attribute	Value
Genome size	4,712,479 bp
GC %	45.3 %
Coverage	293X
Number of chromosomes	2 (3,516,273 bp and 1,196,206 bp)
NCBI Accessions	Chromosome 1: CP092587/ NZ_CP092587 Chromosome 2: CP092588/NZ_CP092588
Genes (total)	4,178
Genes (coding)	3,988
Genes (RNA)	114
rRNAs	5S: 8; 16S: 7; 23S: 7
tRNAs	87
ncRNAs	5
Pseudogenes	76

**Table S2:** *V. gazogenes* PB1 genes predicted by PATRIC to code for virulence factors

Gene	Gene Product	Source	Source ID	Source Organism	Classification	References
<i>arcA</i>	Aerobic respiration control response regulator ArcA	Victors	218927655	<i>Yersinia pestis</i> CO92	Regulation	Merrell et al., 2002
<i>atpA</i>	ATP synthase alpha chain	Victors	16767151	<i>Salmonella enterica</i> subsp. <i>enterica</i> serovar <i>Typhimurium</i> str. LT2	Metabolism	Merrell et al., 2002
<i>bioB</i>	Biotin synthase	Victors	147673516	<i>Vibrio cholerae</i> O395	Nutritional/Metabolic factor	Chen et al., 2005
<i>cap</i>	Cyclic AMP receptor protein	PATRIC_VF	SL1344_3433	<i>Salmonella enterica</i> subsp. <i>enterica</i> serovar <i>Typhimurium</i> str. SL1344		
<i>carA</i>	Carbamoyl-phosphate synthase small chain	PATRIC_VF	Z0037	<i>Escherichia coli</i> O157:H7 str. EDL933	Nutritional/Metabolic factor	Chen et al., 2005
<i>carB</i>	Carbamoyl-phosphate synthase large chain	Victors	147673654	<i>Vibrio cholerae</i> O395	Nutritional/Metabolic factor	Chen et al., 2005
<i>deoC</i>	Deoxyribose-phosphate aldolase	Victors	147674428	<i>Vibrio cholerae</i> O395		
<i>dksA</i>	RNA polymerase-binding transcription factor DksA	PATRIC_VF	S0140	<i>Shigella flexneri</i> 2a str. 2457T	Regulation	Merrell et al., 2002
<i>fabG</i>	3-oxoacyl-[acyl-carrier protein] reductase , FabG	Victors	147675453	<i>Vibrio cholerae</i> O395	Metabolism	Merrell et al., 2002
<i>fbp</i>	Fructose-1,6-bisphosphatase, type I	Victors	147674062	<i>Vibrio cholerae</i> O395		
<i>frdC</i>	Fumarate reductase subunit C	Victors	147674812	<i>Vibrio cholerae</i> O395		
<i>fur</i>	Ferric uptake Regulation protein FUR	Victors	15642106	<i>Vibrio cholerae</i> O1 biovar El Tor str. N16961	Regulation	Chen et al., 2005
<i>glmM</i>	Phosphoglucosamine mutase	Victors	15640659	<i>Vibrio cholerae</i> O1 biovar El Tor str. N16961	Immune modulation	Chen et al., 2005

<i>glyA-1</i>	Serine hydroxymethyltransferase	Victors	227081115	<i>Vibrio cholerae</i> M66-2		
<i>gmhA</i>	D-sedoheptulose 7-phosphate isomerase	PATRIC_VF	SL1344_0306	<i>Salmonella enterica</i> subsp. <i>enterica</i> serovar Typhimurium str. SL1344	Immune modulation	Chen et al., 2005
<i>grxD</i>	Monothiol glutaredoxin GrxD	PATRIC_VF	E2348C_1740	<i>Escherichia coli</i> O127:H6 str. E2348/69		
<i>guaA</i>	GMP synthase [glutamine-hydrolyzing], amidotransferase subunit / GMP synthase [glutamine-hydrolyzing], ATP pyrophosphatase subunit	PATRIC_VF	S2725	<i>Shigella flexneri</i> 2a str. 2457T		
<i>hcp-2</i>	T6SS component Hcp	VFDB	VFG007138	<i>Vibrio cholerae</i> O1 biovar El Tor str. N16961	Effector delivery system	Chen et al., 2005
<i>hfq</i>	RNA-binding protein Hfq	PATRIC_VF	S4595	<i>Shigella flexneri</i> 2a str. 2457T		
<i>kdsA</i>	2-Keto-3-deoxy-D-manno-octulosonate-8-phosphate synthase	VFDB	VFG013465	<i>Haemophilus influenzae</i> Rd KW20	Immune modulation	Chen et al., 2005
<i>luxS</i>	S-ribosylhomocysteine lyase @ Autoinducer-2 production protein LuxS	VFDB	VFG018241	<i>Vibrio cholerae</i> O1 biovar El Tor str. N16961	Biofilm	Chen et al., 2005
<i>lysA</i>	Diaminopimelate decarboxylase	Victors	147674363	<i>Vibrio cholerae</i> O395	Lysine synthesis	Merrell et al., 2002
<i>nqrE</i>	Na(+)-translocating NADH-quinone reductase subunit E	Victors	147674521	<i>Vibrio cholerae</i> O395	Immune modulation	Chen et al., 2005
<i>nqrF</i>	Na(+)-translocating NADH-quinone reductase subunit F	Victors	147675137	<i>Vibrio cholerae</i> O395	Immune modulation	Chen et al., 2005
<i>ompR</i>	Two-component system response regulator OmpR	PATRIC_VF	SEN3328	<i>Salmonella enterica</i> subsp. <i>enterica</i> serovar Enteritidis str. P125109		
<i>prfC</i>	Peptide chain release factor 3	Victors	165975488	<i>Actinobacillus pleuropneumoniae</i> serovar 3 str. JL03		
<i>pta</i>	BioD-like N-terminal domain / Phosphate acetyltransferase	Victors	147675254	<i>Vibrio cholerae</i> O395		

<i>purD</i>	Phosphoribosylamine--glycine ligase	Victors	147673352	Vibrio cholerae O395	Nutritional/Metabolic factor	Chen et al., 2005
<i>purF</i>	Amidophosphoribosyltransferase	Victors	147673131	Vibrio cholerae O395	Nutritional/Metabolic factor	Merrell et al., 2002
<i>pykF</i>	Pyruvate kinase	Victors	147674848	Vibrio cholerae O395	Energy metabolism	Merrell et al., 2002
<i>rpmF</i>	LSU ribosomal protein L32p @ LSU ribosomal protein L32p, zinc-independent	Victors	126208850	Actinobacillus pleuropneumoniae L20		
<i>rpmJ</i>	LSU ribosomal protein L36p @ LSU ribosomal protein L36p, zinc-dependent	Victors	56707499	Francisella tularensis subsp. tularensis SCHU S4		
<i>rpoE</i>	RNA polymerase sigma factor RpoE	Victors	147675349	Vibrio cholerae O395	Regulation	Chen et al., 2005
<i>rpoS</i>	RNA polymerase sigma factor RpoS	PATRIC_VF	SEN2763	Salmonella enterica subsp. enterica serovar Enteritidis str. P125109	Regulation	Chen et al., 2005
<i>sspA</i>	Stringent starvation protein A	Victors	147675668	Vibrio cholerae O395	Cellular process	Merrell et al., 2002
<i>trpB</i>	Tryptophan synthase beta chain	PATRIC_VF	Z2550	Escherichia coli O157:H7 str. EDL933		
<i>tufA</i>	Translation elongation factor Tu	Victors	15676067	Neisseria meningitidis MC58	Adherence	Chen et al., 2005
<i>VC0032</i>	AAA+ ATPase superfamily protein YifB/ComM, associated with DNA recombination	Victors	15640064	Vibrio cholerae O1 biovar El Tor str. N16961	Unknown	
<i>VC0260</i>	Nucleoside-diphosphate sugar epimerase/dehydratase	Victors	15640289	Vibrio cholerae O1 biovar El Tor str. N16961	Immune modulation	Chen et al., 2005
<i>VC0468</i>	Glutathione synthetase	Victors	15640495	Vibrio cholerae O1 biovar El Tor str. N16961	Biosynthesis of co-factors, prosthetic groups and carriers	Merrell et al., 2002
<i>VC0727</i>	Phosphate transport system regulatory protein PhoU	Victors	15640746	Vibrio cholerae O1 biovar El Tor str. N16961	Regulation	Merrell et al., 2002

VC0965	Phosphoenolpyruvate-protein phosphotransferase of PTS system	Victors	15640981	Vibrio cholerae O1 biovar El Tor str. N16961	Transport and binding proteins	Merrell et al., 2002
VC1012	Electron transport complex protein RnfE	Victors	15641027	Vibrio cholerae O1 biovar El Tor str. N16961	Energy metabolism	Merrell et al., 2002
VC1098	Acetate kinase	Victors	15641111	Vibrio cholerae O1 biovar El Tor str. N16961	Energy metabolism	Merrell et al., 2002
VC1715	Chromosome partition protein MukE	Victors	15641719	Vibrio cholerae O1 biovar El Tor str. N16961	Cellular process	Merrell et al., 2002
VC1716	Chromosome partition protein MukF	Victors	15641720	Vibrio cholerae O1 biovar El Tor str. N16961	Cellular process	Merrell et al., 2002
VC2067	Flagellar synthesis regulator FleN	Victors	15642067	Vibrio cholerae O1 biovar El Tor str. N16961	Motility	Chen et al., 2005
VC2206	Flagellar protein FlgP	Victors	15642205	Vibrio cholerae O1 biovar El Tor str. N16961	Motility	Chen et al., 2005
VC2295	Na(+)-translocating NADH-quinone reductase subunit A	Victors	15642293	Vibrio cholerae O1 biovar El Tor str. N16961	Immune modulation	Merrell et al., 2002
VC2647	Transcriptional regulator, PadR family	Victors	15642642	Vibrio cholerae O1 biovar El Tor str. N16961	Hypothetical conserved	Merrell et al., 2002
VC2660	Translation elongation factor P	Victors	15642655	Vibrio cholerae O1 biovar El Tor str. N16961	Elongation factor P	Merrell et al., 2002
VCA0006	FIG007491: hypothetical protein YeeN	Victors	15600777	Vibrio cholerae O1 biovar El Tor str. N16961	Hypothetical conserved	Merrell et al., 2002
<i>vipA/mglA</i>	T6SS component TssB (ImpB/VipA)	VFDB	VFG002092	Vibrio cholerae O1 biovar El Tor str. N16961	Effector delivery system (Type VI secretion system)	Chen et al., 2005
<i>vipB/mglB</i>	T6SS component TssC (ImpC/VipB)	VFDB	VFG002093	Vibrio cholerae O1 biovar El Tor str. N16961	Effector delivery system (Type VI secretion system)	Chen et al., 2005
<i>zwf</i>	Glucose-6-phosphate 1-dehydrogenase	Victors	147671692	Vibrio cholerae O395	Energy metabolism	Merrell et al., 2002

**Table S3:** Antibiotic resistance genes identified in *V. gazogenes* PB1 using PATRIC

AMR Mechanism	Family/Genes
Antibiotic activation enzyme	<i>katG</i>
Antibiotic inactivation enzyme	CatB family
Antibiotic target in susceptible species	Alr, Ddl, dxr, EF-G, EF-Tu, folA, Dfr, folP, gyrA, gyrB, Iso-tRNA, kasA, MurA, rho, rpoB, rpoC, S10p, S12p
Antibiotic target protection protein	QnrB family
Antibiotic target replacement protein	<i>fabV</i>
Efflux pump conferring antibiotic resistance	EmrAB-OMF, EmrAB-TolC, MacA, MacB, Tet(35), TolC/OpmH
Gene conferring resistance via absence	<i>gidB</i>
Protein altering cell wall charge conferring antibiotic resistance	GdpD, PgsA
Regulator modulating expression of antibiotic resistance genes	H-NS, OxyR

**Table S4:** Prophage regions identified by PHASTER in the *V. gazogenes* PB1 genome.

Region	Region Length	Completeness	Score	Total Proteins	Region Position	Most Common Phage (Number of similar proteins)	NCBI Accession	GC %
<b>Chromosome 1 (CP092587.1)</b>								
1	9Kb	incomplete	30	9	887974-897006	PHAGE_Escher_500465_1 (7)	NC_049342	43.63%
2	21.1Kb	intact	140	27	2061980-2083082	PHAGE_Vibrio_VP882 (14)	NC_009016	45.64%
<b>Chromosome 2 (CP092588.1)</b>								
1	25.8Kb	questionable	80	32	526331-552169	PHAGE_Vibrio_8 (14)	NC_022747	46.34%
2	39.2Kb	intact	123	55	701354-740613	PHAGE_Vibrio_12B12 (42)	NC_021070	46.34%

**Table S5:** Genomic islands predicted by IslandPath-DIMOB in the *V. gazogenes* PB1 genome

Chromosome 1 (CP092587.1)		
Genome Island #	Start	End
GI_1	116587	131071
GI_2	767987	781104
GI_3	880889	892019
GI_4	1128508	1173292
GI_5	2330710	2340786
GI_6	2889764	2907705

Chromosome 2 (CP092588.1)		
Genome Island #	Start	End
GI_1	306912	323990
GI_2	697776	735882
GI_3	918370	940627
GI_4	1162062	1189180

**Table S6:** Comparison of the genomes between *V. gazogenes* PB1 and other members of the *Vibrio* genus

Species	<i>Vibrio gazogenes</i>	<i>Vibrio gazogenes</i>	<i>Vibrio spartinae</i>	<i>Vibrio cholerae</i>	<i>Vibrio parahaemolyticus</i>	<i>Vibrio natriegens</i>	<i>Vibrio alginolyticus</i>	<i>Vibrio harveyi</i>	<i>Vibrio campbellii</i>
Strain	PB1	ATCC 43942	3.6	RFB16	O3:K6 substr. RIMD 2210633	ATCC 14048	E110	ATCC 33843	BoB-53
NCBI Genome ID	14723	14723	56603	505	691	14718	2331	685	11077
Genome size (Mb)	4.71	4.79	5.01	4.14	5.17	5.18	5.15	5.88	5.43
Chromosome 1 (NCBI RefSeq Accession)	NZ_CP092587.1	NZ_CP018835.1	NZ_CP046268.1	NZ_CP043554.1	NC_004603.1	NZ_CP009977.1	NZ_CP098033.1	NZ_CP009467.1	NZ_CP026321.1
Chromosome 1 (Mb)	3.52	3.47	3.82	2.95	3.29	3.25	3.29	3.62	3.47
Chromosome 2 (NCBI RefSeq Accession)	NZ_CP092588.1	NZ_CP018836.1	NZ_CP046269.1	NZ_CP043556.1	NC_004605.1	NZ_CP009978.1	NZ_CP098034.1	NZ_CP009468.1	NZ_CP026322.1
Chromosome 2 (Mb)	1.2	1.3	1.19	1.14	1.88	1.93	1.82	2.26	1.96
Total number of CDS	3988	4084	4167	3639	4519	4488	4589	5205	4768
Average Nucleotide Identity (ANI) (%)	-	94.22	91.92	71.61	71.4	71.4	71.15	71.15	71.26
Average Amino Acid Identity (AAI) (%)	-	96.58	94.77	73.21	72.4	72.02	72.12	71.87	71.84
Percentage of Conserved Proteins (%)	-	85.46	84.4	59.36	54.81	55.92	55.56	52.7	53.75

**Table S7:** Annotated genes in the 100 kbp region of chromosome 2 of *V. gazogenes* ATCC 43942 that has been deleted in *V. gazogenes* PB1.

Annotation	Start	End	Length	Strand
<b>Region 1: 569,917-590,569</b>				
toxin HipA CDS	570,310	571,549	1,240	forward
transcriptional regulator CDS	571,534	572,016	483	forward
hypothetical protein CDS	572,805	573,683	879	forward
allulose-6-phosphate 3-epimerase CDS	573,858	574,550	693	reverse
PTS fructose transporter subunit IIC CDS	574,738	575,838	1,101	reverse
PTS fructose transporter subunit IIB CDS	575,895	576,218	324	reverse
PTS fructose transporter subunit IIA CDS	576,287	576,754	468	reverse
PTS fructose transporter subunit IIA CDS	577,149	577,604	456	reverse
hypothetical protein CDS	577,628	579,268	1,641	reverse
DNA helicase CDS	579,722	583,243	3,522	forward
zeta toxin family protein CDS	583,779	584,492	714	reverse
metal-dependent hydrolase CDS	584,551	584,819	269	reverse
hypothetical protein CDS	584,941	587,802	2,862	reverse
hypothetical protein CDS	587,846	590,104	2,259	reverse
<b>Region 2: 753,693-819,888</b>				
hypothetical protein CDS	753,681	754,148	468	forward
hypothetical protein CDS	754,418	755,185	768	reverse
threonine-tRNA ligase CDS	755,245	757,200	1,956	reverse
hypothetical protein CDS	757,671	758,885	1,215	reverse
hypothetical protein CDS	759,021	763,778	4,758	reverse
hypothetical protein CDS	763,775	767,893	4,119	reverse
hypothetical protein CDS	767,893	773,442	5,550	reverse
hypothetical protein CDS	773,439	781,196	7,758	reverse
hypothetical protein CDS	781,228	787,713	6,486	reverse
hypothetical protein CDS	787,809	793,187	5,379	reverse
hypothetical protein CDS	793,216	799,848	6,633	reverse
hypothetical protein CDS	799,845	807,335	7,491	reverse
transposase CDS	808,357	808,623	267	forward
transposase CDS	808,665	809,468	804	forward
hypothetical protein CDS	809,717	810,127	411	reverse
hypothetical protein CDS	810,142	811,065	924	reverse
hypothetical protein CDS	811,995	812,870	876	forward
endonuclease V CDS	812,963	813,604	642	forward
hypothetical protein CDS	813,596	814,486	891	reverse
hypothetical protein CDS	814,488	816,266	1,779	reverse
hypothetical protein CDS	816,786	817,661	876	forward

hypothetical protein CDS	817,763	818,062	300	reverse
hypothetical protein CDS	818,055	818,285	231	reverse
hypothetical protein CDS	818,433	821,108	2,676	reverse
<b>Region 3: 821,079-831,326</b>				
hypothetical protein CDS	821,535	822,413	879	forward
hypothetical protein CDS	822,556	822,837	282	reverse
hypothetical protein CDS	823,240	823,659	420	reverse
hypothetical protein CDS	823,656	823,925	270	reverse
hypothetical protein CDS	824,355	824,813	459	reverse
hypothetical protein CDS	824,820	826,034	1,215	reverse
hypothetical protein CDS	826,037	826,219	183	forward
hypothetical protein CDS	826,286	826,624	339	reverse
hypothetical protein CDS	826,627	827,766	1,140	reverse
IS256 family transposase CDS	827,940	828,178	239	forward
hypothetical protein CDS	828,252	828,701	450	reverse
hypothetical protein CDS	828,815	829,054	240	reverse
hypothetical protein CDS	829,390	829,815	426	reverse
hypothetical protein CDS	829,974	830,339	366	reverse
hypothetical protein CDS	830,649	831,032	384	reverse
hypothetical protein CDS	831,223	831,489	267	forward
<b>Region 4: 1,244,876-1,261,341</b>				
hypothetical protein CDS	1,245,021	1,245,206	186	forward
amino acid adenylation CDS	1,245,262	1,245,930	669	forward
MBL fold hydrolase CDS	1,246,119	1,247,210	1,092	forward
hypothetical protein CDS	1,247,212	1,247,853	642	forward
5'-nucleotidase CDS	1,248,154	1,249,113	960	forward
hypothetical protein CDS	1,249,214	1,250,101	888	forward
endonuclease V CDS	1,250,093	1,250,773	681	reverse
hypothetical protein CDS	1,250,827	1,251,702	876	reverse
hypothetical protein CDS	1,252,196	1,252,570	375	forward
hypothetical protein CDS	1,252,765	1,253,187	423	reverse
hypothetical protein CDS	1,253,254	1,253,982	729	forward
hypothetical protein CDS	1,253,997	1,254,401	405	forward
hypothetical protein CDS	1,254,678	1,254,929	252	reverse
hypothetical protein CDS	1,255,120	1,255,503	384	forward
hypothetical protein CDS	1,255,791	1,256,174	384	forward
hypothetical protein CDS	1,256,512	1,256,910	399	forward
hypothetical protein CDS	1,257,198	1,257,754	557	forward
hypothetical protein CDS	1,258,329	1,258,715	387	forward
hypothetical protein CDS	1,258,811	1,259,689	879	reverse

**Table S8.** CRISPR array and CRISPR-Cas operons identified in *V. gazogenes* PB1 using CRISPRCasFinder

Chrom osome	Start	End	Array length	Direct repeat sequence	Repeat Length	# of spacers	Evidence level	CRISPR- Cas type	Cas genes
1	1916871	1918302	1432	CGGTTCATCCCCGTGTACGGGAAACAC	29	23	4	-	-
1	2855893	2856160	268	TTTCTAACGCTGCCTATGCCAGTGAAC	28	4	4	-	-
1	3441994	3442426	433	ATCGGAAGACACTGGCCGAGACAGGCCGTT	30	6	3	-	-
2	495287	496047	761	GTGTTCCCCGTATGCACGGGATGAACCG	29	12	4	I-E	Cas3, Cas8, Cse2, Cas7, Cas5e, Cas6, Cas1, Cas2
2	874590	875029	440	GTCTTAACGGCCTGTTCGGCCAAGGTATTCCAC	35	6	4	III-B	Cmr6, Cmr5, Cmr4, Cmr3, Cmr2, Cas10, Csm3, Cas1, Cas2
2	1088527	1089043	517	CGGTTCATCCCCGTGCATAACGGGAAACAC	29	8	4	III-D	Cas10, Csm3, Csm3, Cas1, Cas2
2	1110778	1111161	384	GTTCAGTAGGTCTGCCGTTAGGCCGTTGAGAC	35	5	4	-	-

**Table S9:** Results of spacer BLASTN search

Chromosome	CRISPR array start	CRISPR array end	Spacer start	Spacer end	Spacer sequence	% identity	Query coverage	NCBI accession	Organism
1	1916871	1918302	1916900	1916931	GATAATGACCTGCTGGTTGCGGTCGCCGTTA	100%	100%	JX556417	<i>Vibrio</i> phage vB_VpaM_MAR
						100%	100%	FN297812	<i>Vibrio</i> phage VP58.5
						100%	100%	AY133112	<i>Vibrio</i> phage VHML
2	495287	496047	495926	495957	CGTTCATTACCGACCGGGATTGTGGGAGTAC	100%	100%	CP020037.1	<i>Vibrio parahaemolyticus</i> strain 20130629002S01 plasmid pVPGX2

**Table S10:** Primers used for the RT-qPCR analysis

Gene	Approximate amplicon sizes (bp)		Primer Set 1	Primer set 2
USP16020.1 ( <i>glgX</i> )	72	123	<i>glgX_set 1_F</i> : CACATCGATGGCTTCCGTTTCG <i>glgX_set 1_R</i> : CGCATTGGACTGAAGGTTT	<i>glgX_set 2_F</i> : CTGATCGCAGAACCTGGG <i>glgX_set 2_R</i> : CTTGATCACACGCCAGAAG
USP15696.1 (starch-binding protein)	96	171	<i>SBP_set 1_F</i> : CGCACCAATGCCAGTAATGA <i>SBP_set 1_R</i> : CGGATAGCCGTAAGGGTAGG	forward <i>SBP_set 2_F</i> : CCACAGCGATGATGCACCG <i>SBP_set 2_R</i> : CCGTTATCCCACCAGTGAG
USP13629.1 (alpha-glucosidase)	91	95	<i>AG_set 1_F</i> : GGCGGTTATACGACCCCTAC <i>AG_set 1_R</i> : CCTTCATGGCTACGCATAATC	forward <i>AG_set 2_F</i> : CAAGGTTATCTGGCAACGCA <i>AG_set 2_R</i> : GCATCCGGATTGGTGAAGTC
USP15540.1 (glycoside hydrolase family 9 protein)	102	126	<i>GHF9_set 1_F</i> : GAGCCGGTGCAAATTATCG <i>GHF9_set 1_R</i> : ATCAATCGCAAAGGGATGGC	<i>GHF9_set 2_F</i> : AATGGGAAAGGCAGATCCGA <i>GHF9_set 2_R</i> : GGGCCATCAACGAAGTAACC
USP14760.1 (glycoside hydrolase family 6 protein)	161	102	<i>GHF6_set 1_F</i> : GTAGCAGCGATGACCGTAAC <i>GHF6_set 1_R</i> : GCATACGTGTGTGATGTGCTG	<i>GHF6_set 2_F</i> : CGAAACTAACCGTCAGTGATGGC <i>GHF6_set 2_R</i> : GTTACGGTCATCGCTGCTAC
USP15545.1 (GH1 family beta-glucosidase)	179	127	<i>GH1BG_set 1_F</i> : GTAACAAACCCGGCAAAGTGG <i>GH1BG_set 1_R</i> : CGCTAACCTTGGGATTGAC	<i>GH1BG_set 2_F</i> : CTATACCGGAATGTGGTCG <i>GH1BG_set 2_R</i> : CGGACGAGTAAATCCGTC
16S RNA	95	154	<i>HK1_16S_set 1_F</i> : CGATGCAACGCGAAGAACCC <i>HK1_16S_set 1_R</i> : GCACCTGTCTCTCAGTTCC	<i>HK1_16S_set 2_F</i> : GGAAGTGAGAGACAGGTGC <i>HK1_16S_set 2_R</i> : CCCACCTTCCTCCGGTTTAT

**Table S11:** A one-way ANOVA was performed to compare the effect of three different carbon sources on the transcript levels of selected genes related to cellulose and starch metabolism, as determined from the PGAP annotation pipeline. A p-value of less than 0.05 was considered significant.

	Cultivation time (18 hours)			F-value	P-value
	Glucose	Cellulose	Starch		
<b>USP16020.1</b> isoamylase	1.0	0.2	0.1	44.6	0.00000049
	1.0	0.2	0.7		
	1.0	0.3	0.2		
	1.0	0.5	0.3		
	1.0	0.3	0.3		
	1.0	0.4	0.2		
<b>USP15696.1</b> $\alpha$ -amylase	1.0	0.1	16.7	11.0	0.00116875
	1.0	0.1	8.0		
	1.0	0.0	2.6		
	1.0	0.2	18.3		
	1.0	0.1	6.9		
	1.0	0.1	4.0		
<b>USP15545.1</b> $\beta$ -glucosidase	1.0	0.9	0.3	64.6	0.00000004
	1.0	0.3	0.1		
	1.0	0.4	0.1		
	1.0	0.5	0.3		
	1.0	0.5	0.2		
	1.0	0.3	0.0		
<b>USP14760.1</b> exoglucanase	1.0	30.0	7.1	15.7	0.00021070
	1.0	35.3	6.9		
	1.0	18.1	1.2		
	1.0	37.0	7.9		
	1.0	9.2	1.5		
	1.0	10.2	0.9		
<b>USP13629.1</b> $\alpha$ -glucosidase	1.0	43.5	13.2	18.8	0.00008263
	1.0	49.6	9.8		
	1.0	41.5	12.7		
	1.0	17.0	3.6		
	1.0	19.4	3.4		
	1.0	17.8	3.1		
<b>USP15540.1</b> endo-1,4- $\beta$ -glucanase	ND	ND	ND		