

Supplemental Figure 1. Mucicarmine staining of *V. victoriae* and *C. neoformans* cells in murine lungs. *C. neoformans* cells stain magenta (A) while *V. victoriae* cells stain dark purple (B), indicating the presence of a polysaccharide capsule surrounding *C. neoformans* cells but not *V. victoriae* cells. Magnification=100X (oil immersion).



**Supplemental Figure 2. Growth of** *V. victoriae* and *C. neoformans* at optimal temperature versus **37°C.** *C. neoformans* grew well at both 25°C (A, blue) and 37°C (A, red), while V. victoriae grew optimally at 15°C (B, blue) but not at all at 37°C (B, red). N=2 per growth condition, error bars represent standard deviation.



**Supplemental Figure 3: Deposition of yeast cells in lung following a single oropharyngeal aspiration exposure**. Representative micrographs of GMS-stained sections of mouse lungs approximately one hour following one exposure to PBS (left), 10<sup>4</sup> *C. neoformans* cells (middle), or 10<sup>6</sup> *V. victoriae* cells (right). Yeast cells are indicated by arrows. 400x magnification.



**Supplemental Figure 4. Mucicarmine staining of** *C. neoformans* cells in murine lungs following repeated oropharyngeal aspiration exposure. *C. neoformans* cells stained magenta in lung tissue 1-day (A) and 21-days (B) following repeated exposure to 10<sup>4</sup> cells. Magnification=100X (oil immersion).



**Supplemental Figure 5A: Flow cytometry gating strategy (myeloid panel)**. Representative dot plots showing gating strategy for myeloid cell populations of interest. Live cells were gated based on FSC-A and SSC-A, then single cells were gated from FSC-A and FSC-H. CD45<sup>+</sup> Cells were selected and gated on Siglec F and CD11b to isolate eosinophils and macrophages. Non-eosinophils and non-macrophages were gated on Ly6G and Cd11b to isolate neutrophils. The non-neutrophil population was then gated on CD11b and Ly6C to isolate Ly6C<sup>hi/med</sup> monocytes. Non-monocytes were gated on CD11c to isolate dendritic cells that further gated on CD11b and CD103 for specific CD11b<sup>+</sup> and CD103<sup>+</sup> DC populations.



**Supplemental Figure 5B: Flow cytometry gating strategy (lymphoid panel)**. Representative dot plots showing gating strategy for lymphoid cell populations of interest. Live cells were gated based on FSC-A and SSC-A, then single cells were gated from FSC-A and FSC-H. CD45<sup>+</sup> Cells were selected then lymphocytes were gated out based on size. Lymphocytes were then gated on CD49b and CD3 to isolate NK and T Cells, respectively. T cells were further gated on CD8a and CD4 to isolate CD8<sup>+</sup> and CD4<sup>+</sup> T cell subpopulations. Non-NK or non-T Cells were gated against B220 to isolate B cells.



**Supplemental Figure 6: Cell populations in BALF following repeated yeast exposure**. Cell populations in BALF following repeated exposure to PBS (black), 10<sup>4</sup> *C. neoformans* cells (yellow) or 10<sup>6</sup> *V. victoriae* cells (blue) at 1 day post final exposure (circles, crossed bars, left) or 21 days post final exposure (squares, solid bars, right). A) Eosinophils, B) Macrophages. n=3-7 per group, \*P<0.05, \*\*P < 0.01, \*\*\*P < 0.001. P values were determined by ordinary one-way ANOVA with Tukey's multiple comparisons test comparing each exposure together for the same timepoint.



**Supplemental Figure 7: Lymphoid cell populations in BAL-depleted Lung following repeated yeast exposure**. Cell populations in BAL-depleted lung following repeated exposure to PBS (black), 10<sup>4</sup> *C. neoformans* cells (yellow) or 10<sup>6</sup> *V. victoriae* cells (blue) at 1 day post final exposure (circles, crossed bars, left) or 21 days post final exposure (squares, solid bars, right). A) Lymphocytes, B) NK Cells, C) B Cells, D) T Cells, E) CD4<sup>+</sup> T Cells, F) CD8<sup>+</sup> T Cells. n=3-7 per group, \*P<0.05, \*\*P < 0.01. P values were determined by ordinary one-way ANOVA with Tukey's multiple comparisons test comparing each exposure together for the same timepoint.



**Supplemental Figure 8: Myeloid cell populations in BAL-depleted Lung following repeated yeast exposure**. Cell populations in BAL-depleted lung following repeated exposure to PBS (black), 10<sup>4</sup> *C. neoformans* cells (yellow) or 10<sup>6</sup> *V. victoriae* cells (blue) at 1 day post final exposure (circles, crossed bars, left) or 21 days post final exposure (squares, filled bars, right). A) CD45<sup>+</sup>, B) Eosinophils, C) Macrophages, D) Neutrophils, E) Ly6C<sup>hi/med</sup> Monocytes, F) CD11b<sup>+</sup> Dendritic Cells, G) CD103<sup>+</sup> Dendritic Cells. n=3-7 per group, \*P<0.05, \*\*P < 0.01. P values were determined by ordinary one-way ANOVA with Tukey's multiple comparisons test comparing each exposure together for the same timepoint.



Supplemental Figure 9: Changes in BAL-depleted Lung cell population quantifications from 1 day to 21 days post final exposure. Mean cell quantifications in lung following repeated exposure to PBS,  $10^4$  *C. neoformans* cells or  $10^6$  *V. victoriae* cells at 1 day post final exposure compared to 21 days post final exposure. n=3-7 per group. Values used for this heatmap were obtained by calculating the average quantities at 21 days post final exposure and subtracting the average quantities at 1 day post final exposure. A positive Z score (pink) indicates that the quantification of cells increased at 21 days post final exposure compared to 1 day post final exposure. Negative Z scores (blue) indicate that the cell quantifications decreased by 21 days post final exposure compared to 1 day post final exposure final exposure. Asterisks overlaying the heatmap indicate P values comparing the quantifications of each time point for the same exposure. \*P<0.05, \*\*P < 0.01, \*\*\*P < 0.001, \*\*\*P < 0.0001. P values were determined by ordinary one-way ANOVA with Sidak's multiple comparisons test comparing the same exposure at 1 day post final exposure and 21 days post final exposure.

Lymphoid Panel:										
Marker	Species	Fluorochrome	Clone Company		Catalog Number					
CD45	Rat anti-mouse	PE Cy 7	13:2.3	Southern Biotech	166017					
CD49b	Rat anti-mouse	EF 506	DX5	Invitrogen	69-5971-82					
CD3	Rat anti-mouse	APC	17A2	Invitrogen	17-0032-82					
CD4	Rat anti-mouse	BV 650	RM4-5	Biolegend	100545					
CD8a	Rat anti-mouse	APC Cy 7	53-6.7	BD	561967					
B220	Rat anti-mouse	PE	RA3-6B2	BD	553090					
Myeloid Panel:										
Marker	Species	Fluorochrome	Clone	Company	Catalog Number					
CD45	anti-mouse	BV 711	30-F11	Biolegend	103147					
CD11b	Rat anti-mouse/human	APC Cy 7	M1/70	BD	561039					
Siglec F	Rat anti-mouse	BV 605	E50-2440	BD	740388					
Ly6G	Rat anti-mouse	BV 510	1A8	BD	740157					
Ly6C	Rat anti-mouse	Per CP Cy 5.5	HK1.4	Invitrogen	45-5932-80					
CD11c	anti-mouse	APC	N418	Biolegend	117310					
CD103	Rat anti-mouse	BV 421	M290	BD	562771					
Reagents:										
Name	Company	Catalog Number								
DMEM, high glucose, pyruvate	Gibco	11995-040								
Fetal Bovine Serum	Gibco 1600-044									
Collagenase/Dispase	Sigma Aldrich 11097113001									
Deoxyribonuclease I	Alfa Aesar	J61061								
10x RBC Lysis Buffer	Invitrogen	00-4300-54								
PBS 1x	HyClone	SH30256.02								
EDTA 0.5 M, pH 8.0	Affymetrix	15684								
Rat Serum	Sigma Aldrich	rich S7648								
CD16/CD32 Rat anti-mouse unlabeled Clone 2.4G2	BD	553141								
Cytofix Fixation Buffer	BD	554655								
Sphero AccuCount Blank Particles	Spherotech Inc	ACBP-50-10								
Ultra Comp eBeads	Invitrogen	01-2222-42								
Rabbit anti-Mouse IgM Secondary Antibody, AP	Invitrogen	31333								
IgG Fc Goat anti-Mouse, AP	Invitrogen	31325								
Goat Anti-Mouse IgE-AP	Southern Biotech	1110-04	1							
Thermo Sceintific Pierce PNPP Substrate Kit	Invitrogen	37620								

## Supplemental Table 1: Flow Cytometry Antibodies and Reagents

**Supplemental Table 1: Flow Cytometry Antibodies and Reagents**. Antibody and reagent names, manufacturers and catalog numbers.

		H&E						PAS	GMS	
		Mononuclear Inflammation			Onemulamentaria	Alveolar	Inflammation	Goblet	Veed	Histiocyte
Group:	Specimen	Perivascular	Peribronchial	Parenchymal	Granulomatous	Histiocytes	Score	cells	reast	granules
PBS 1 day	F52	0	0	0	0	0	0	1	0	0
	F53	1	0	0	0	1	2	1	0	1
	F54	1	0	0	0	1	2	1	0	1
<i>C. neo</i> 10 <sup>4</sup> 1 day	F45	1	1	0	0	1	3	2	1	1
	F46	1	1	0	0	0	2	0	0	1
	F47	1 mixed	2 mixed	2	2	2	6	2	2	2
<i>V. vic</i> 10 <sup>4</sup> 1 day	F61	0	0	0	0	0	0	1	0	1
	F62	0	0	0	0	0	0	0	0	1
	F63	1	1	0	0	0	2	0	0	1
<i>V. vic</i> 10 <sup>6</sup> 1 day	F70	2	2	0	0	2	6	2	0	1
	F71	2	1	0	0	1	4	2	0	2
	F72	1	2	0	0	1	4	3	0	2
PBS 21 days	F06	0	0	0	0	0	0	1	0	0
	F07	0	0	0	0	0	0	1	0	0
	F08	1	1	0	0	0	2	0	0	1
<i>C. neo</i> 10 <sup>4</sup> 21 days	F16	2 mixed	2 mixed	0	2	2	4	2	0	1
	F17	2	2	2	3	0	9	4	2	2
	F18	2	2	1	1	2	8	3	1	2
<i>V. vic</i> 10 <sup>4</sup> 21 days	F26	0	0	0	0	0	0	0	0	1
	F27	0	0	0	0	0	0	1	0	1
	F28	0	0	0	0	0	0	1	0	0
<i>V. vic</i> 10 <sup>6</sup> 21 days	F35	1	0	0	0	1	2	0	0	2
	F36	1	1	0	0	1	3	2	0	1
	F37	1	1	0	0	0	2	2	0	2
*mixed = neutrophils also present										
1 day = tissue collected one day after final exposure										
21 days = tissue collected 21 days after final exposure										

## Supplemental Table 2 – Repeated Exposure Histology Scores

**Supplemental Table 2: Repeated Exposure Histology Scores**. Provided histology scores from blinded histopathologist examining sectioned and stained lung tissue from mice repeatedly exposed to yeast or PBS control. H&E = hematoxylin and eosin stain, PAS = Periodic acid shift stain, and GMS = Grocott's methenamine silver stain.