

Supplementary Material - Supplementary Figures



Supplementary Figure 1: Negative control assays demonstrating the specificity of primary and secondary antibodies used for immunostaining of podoplanin, AQP5, pro-SPC, ENaC, and Na⁺/K⁺-ATPase. Representative immunofluorescence images of lung sections (5 μ m, Scale: 100 μ m) (from WT mice), embedded with paraffin/cryomatrix (A, B, C) and cytocentrifuged ATII cells (D, E) showing an absence of non-specific signal, after staining with anti-podoplanin (A), anti-AQP5 (B), anti-pro-SPC (C), anti- α -ENaC (D) and anti-Na⁺/K⁺-ATPase (E), in absence of secondary antibodies. An absence of background is also confirmed in immunostaining assays of the corresponding secondary antibodies (anti-rabbit Alexa Fluor TM-568 or anti-mouse Alexa Fluor TM-568) alone (in absence of the primary antibody). Nuclei were stained by DAPI.



Supplementary Figure 2: Lung function parameters in naïve adult WT and KvLQT1-KO mice. Parameters are presented according to the measurement maneuvers/mathematical model from which they are derived: The single-compartment model (**A**) yields the total respiratory resistance (R_{rs}), elastance (E_{rs}) and compliance (C_{rs}). Partial step-wise PV loops (**B**) give rise to an estimate of the subject's inspiratory capacity (A), a shape parameter describing the form of deflating PV-loop (K), the area between the PV inflation and deflation limbs (Area), and the inspiratory work-of-breathing normalized to maximal pressure (WOBn). The inspiration capacity (IC) can also be extracted from the Deep Inflation maneuver as the volume at 30 cmH₂O. The constant-phase model (**C**) outputs the Newtonian (airway) resistance (R_N), tissue damping (G), and tissue elastance (H) parameters. All measurements were made with a flexiVent system in anesthetized and mechanically ventilated naïve adult WT and KvLQT1 KO mice (n=11-13). Results are reported by means ± SEM. Unpaired t-test (Agostino/Pearson normality test: positive) was practiced. *p < 0.05 vs WT mice.



Supplementary Figure 3: Effect of R-L3 on lung function in WT mice after a thiourea challenge. Parameters are presented according to the maneuvers/mathematical model from which they are derived: The single-compartment model (A) yields the total respiratory resistance (R_{rs}), elastance (E_{rs}), and compliance (C_{rs}). Partial step-wise PV loops (B) give rise to an estimate of the inspiratory capacity (A), a shape parameter describing the form of deflating PV-loop (K), the area between the PV inflation and deflation limbs (Area), and the inspiratory work-of-breathing normalized to maximal pressure (WOBn). The inspiration capacity (IC) can also be extracted from the Deep Inflation maneuver as the volume at 30 cmH₂O. The constant-phase model (C) outputs the Newtonian (airway) resistance (R_N), tissue damping (G), and tissue elastance (H) parameters. All measurements were made 4 hours after the thiourea challenge with a flexiVent in anesthetized and mechanically ventilated adult WT mice under control conditions (PBS/PBS) or challenged with thiourea (i.p.: TU, 5 mg/kg, PBS/TU) and treated or not with the KvLQT1 activator R-L3 (R-L3/TU) (n=5-9). Results are reported by means ± SEM. One-way ANOVA and Bonferroni's multiple comparisons test were practiced (normality Agostino/Pearson test: positive) was practiced. *p < 0.05 or *** p < 0.0001 vs PBS/PBS.



Supplementary Figure 4: Lung function parameters in WT and KO mice after a thiourea challenge. Parameters are presented according to the maneuvers/mathematical model from which they are derived: The single-compartment model (A) yields the total respiratory resistance (R_{rs}), elastance (E_{rs}) and compliance (C_{rs}). Partial step-wise PV loops (B) give rise to an estimate of the inspiratory capacity (A), a shape parameter describing the form of deflating PV-loop (K), the area between the PV inflation and deflation limbs (Area), and the inspiratory work-of-breathing normalized to maximal pressure (WOBn). The inspiration capacity (IC) can also be extracted from the Deep Inflation maneuver as the volume at 30 cmH₂O. The constant-phase model (C) outputs the Newtonian (airway) resistance (R_N), tissue damping (G) and tissue elastance (H) parameters. All measurements were measured made 4 hours after the thiourea challenge with a flexiVent in WT and KO mice (i.p.: TU, 5 mg/kg, WT/TU vs KO/TU, n=9-10). Results are reported by means ± SEM. All parameters were comparable between WT and KO animals.



Supplementary Figure 5: Effect of R-L3 treatment on the expression of alveolar markers and ion/liquid channels/transporters in absence of thiourea challenge. Representative immunofluorescence images (Scale: 100 μm) of podoplanin (A), AQP5 (B), α-ENaC subunit (C), and Na^+/K^+ -ATPase (**D**) stainings of slides with cytocentrifuged ATII cells isolated from WT control mice (PBS), treated with the KvLQT1 activator (R-L3, 4µM) for 24 hours before lung collection (n=2 experiments, each including a pool of 4-9 mice). Nuclei were stained by DAPI. Quantification (right panels) of all staining intensities was made with a protocol exploited by ICY Software. Values are presented as means \pm SEM. Unpaired t-test (Agostino/Pearson normality test: positive). *p < 0.05 vs PBS.