

Enhancing Chemical and Biological Diversity by Co-cultivation

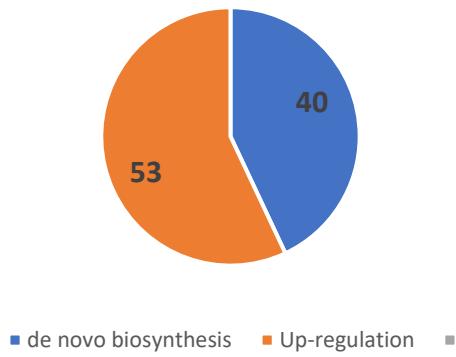
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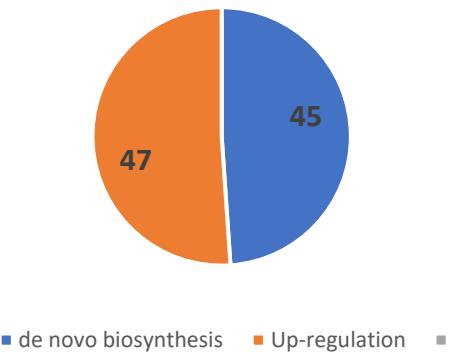
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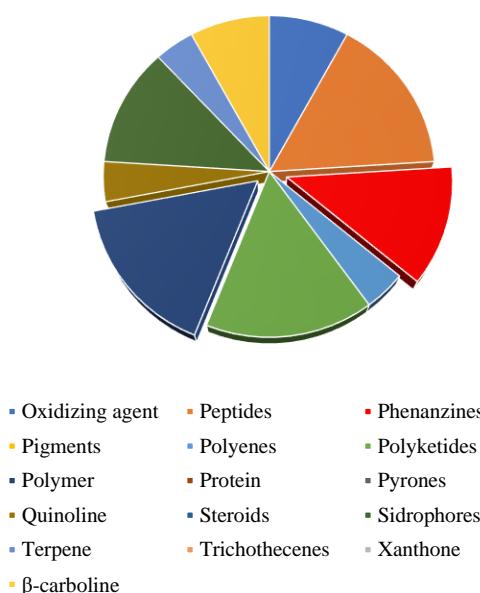
Bacteria induced metabolite in co-culture



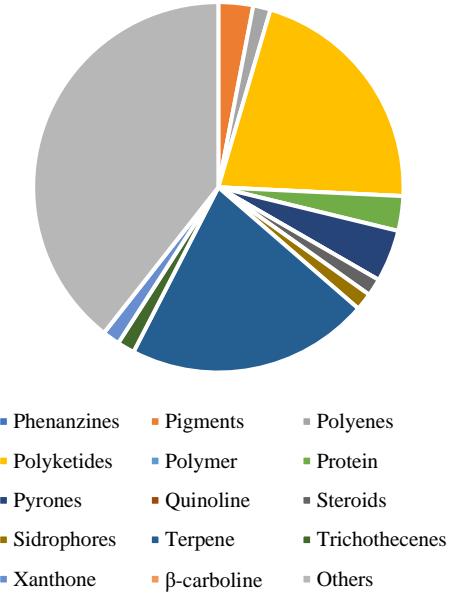
Fungi induced metabolite in co-culture



Bacteria induced metabolite class

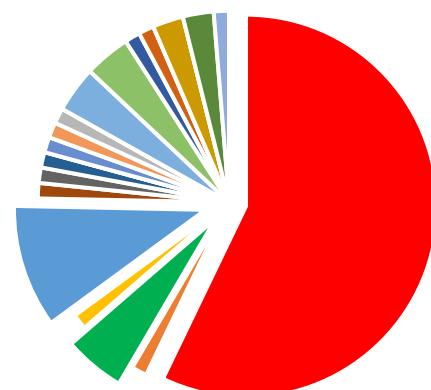


Fungi induced metabolite class



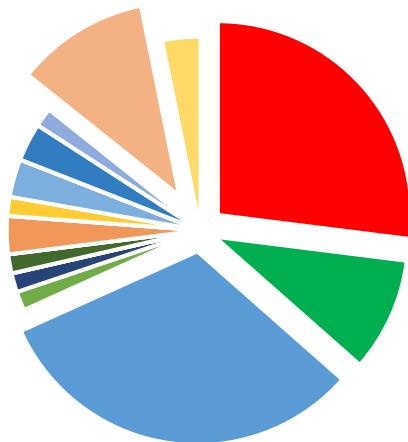
Outcome of biological activity for bacterial producing cocultures

- Antibacterial activity
- Anticancer activity
- Antifungal activity
- Antiosteoporosis activity
- Antiprotozoal activity
- Attracts Enteromorpha zoospores
- Biosurfactant activity
- Germination stimulation
- Hyphae and sporulation inhibition
- Increases antibiotic tolerance
- Inhibits swarming motility
- Phytotoxic activity
- Regulates virulence
- Suppression of virulence
- Antibiotic tolerance
- Antifouling activity
- Antioomycete activity
- Antioxidant activity
- Antiviral activity
- Biocide activity
- Enzyme Inhibition
- Herbicide activity
- Increase metabolism
- Increases survival
- Modulate gene expression
- Quorum-sensing inhibition
- Stimulate wood decay
- Transcription induction



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- Acids
- Aminoglycosides
- Bacteriocin
- Chlorinated benzophenones
- Diketopiperazines
- Diphenyl ethers
- Exopolysaccharide
- Indoles
- Iturin
- Lipopeptides
- Naphthoquinones
- Oxidizing agents
- Phenazines
- Polyenes
- Polymer
- Proteins

- Alkaloids
- Aromatic heptaenes
- Chojalactones
- Depsipeptides
- Diimides
- Enzyme
- Fatty Acids
- Indolocarbazoles
- Lactones
- Macrolides
- Not reported
- Peptides
- Pigments
- Polyketides
- Polysubstituted benzaldehyds
- Pyrones

Antifungal activity

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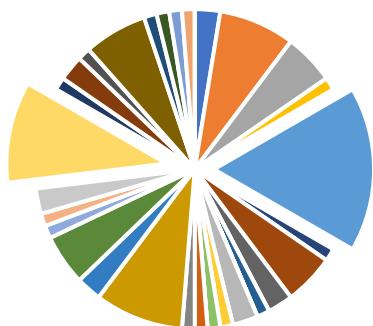
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Anticancer activity



- | | |
|---|---|
| <ul style="list-style-type: none"> ■ Acids ■ Aminoglycosides ■ Bacteriocin ■ Chlorinated benzophenones ■ Diketopiperazines ■ Diphenyl ethers ■ Exopolysaccharide ■ Indoles ■ Iturin ■ Lipopeptides ■ Naphthoquinones ■ Oxidizing agents ■ Phenazines ■ Polyenes ■ Polymer ■ Proteins ■ Quinolines ■ Tetracyclic citrinins ■ Lipopeptides | <ul style="list-style-type: none"> ■ Alkaloids ■ Aromatic heptaenes ■ Chojalactones ■ Depsipeptides ■ Diimides ■ Enzyme ■ Fatty Acids ■ Indolocarbazoles ■ Lactones ■ Macrolides ■ Not reported ■ Peptides ■ Pigments ■ Polyketides ■ Polysubstituted benzaldehyds ■ Pyrones ■ Terpenes ■ Trichothecene ■ Coumarin |
|---|---|

Antibacterial activity



| Type | Inducer microbe | Culture Medium | Challenge microbe | Compound | Compound class | Induction type | Biological activity | Reference |
|-------------|--|------------------|--|--------------------------------|-------------------|--|--|--------------------------------|
| F/F | <i>Serpula lucrymans</i> | Solid | <i>Trichoderma</i> sp (3 isolates) | laccase | Enzymes | <i>de novo</i> biosynthesis | Stimulate wood decay | (Score et al. 1997) |
| B/F and B/B | <i>Bacillus</i> sp. S3, <i>B. pumilus</i> S8, <i>B. licheniformis</i> D1 | Liquid | <i>Candida albicans</i> , <i>Yarrowia lipolytica</i> , <i>Pseudomonas aeruginosa</i> , <i>Bacillus pumilus</i> | not identified | Not identified | Up-regulation | Antifungal activity against <i>Y. lipolytica</i> | (Dusane et al. 2011) |
| B/B | <i>Streptomyces</i> sp (76 species) | Solid | <i>Bacillus subtilis</i> ATCC6633 | not identified | Not identified | Up-regulation | Antibiotic activity | (Ueda et al. 2000) |
| F/F | <i>Acremonium</i> sp. Tbp-5 | Solid | <i>Mycogone rosea</i> DSM 12973 | acremostatins A–C | Lipoaminopeptides | <i>de novo</i> biosynthesis | No reports | (Degenkolb et al. 2002) |
| B/F | <i>Actinomyces levoris</i> | Solid | <i>Candida tropicalis</i> | levorin | Aromatic heptaene | <i>de novo</i> biosynthesis | Antifungal and antibiotic activity | (Yakovleva and Bulgakova 1978) |
| F/F | <i>Alternaria tenuissima</i> | Liquid and Solid | <i>Nigrospora sphaerica</i> | stemphyperylene, alterperylene | Polyketides | <i>de novo</i> biosynthesis | Stemphyperylene shows antifungal activity | (Chagas et al. 2013) |
| F/F | <i>Antrodiella citrinella</i> , <i>Flammulina velutipes</i> , <i>Fomitopsis pinicola</i> , <i>Heterobasidion annosum</i> , <i>Junghuhnia collabens</i> , <i>Phlebia centrifuga</i> , <i>Phlebiopsis gigantea</i> , <i>Resinicium bicolor</i> | Solid | <i>Antrodiella citrinella</i> , <i>Flammulina velutipes</i> , <i>Fomitopsis pinicola</i> , <i>Heterobasidion annosum</i> , <i>Junghuhnia collabens</i> , <i>Phlebia centrifuga</i> , <i>Phlebiopsis gigantea</i> , <i>Resinicium bicolor</i> | laccase | Enzymes | Up-regulation/ <i>de novo</i> biosynthesis | Stimulate wood decay | (Iakovlev and Stenlid 2000) |

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|------------|--------------------------------------|--------|--|---|----------------------------|--|--|-----------------------|
| B/B | <i>Aspergillus austroafricanus</i> | Liquid | <i>Bacillus subtilis</i> or <i>Streptomyces lividans</i> | austramide, violaceol I, violaceol II, diorcinol | Diphenyl ethers | <i>de novo</i> biosynthesis/U p-regulation | Violaceol I-II has antibacterial active against <i>Staphylococcus aureus</i> (ATCC 700699); Diorcinol has moderate antibacterial activity against <i>B. subtilis</i> . | (Ebrahim et al. 2016) |
| F/B | <i>Aspergillus fumigatus</i> | Liquid | <i>Streptomyces rapamycinicus</i> | fumicyclines A-B | Meroterpenoids | <i>de novo</i> biosynthesis | Moderate antibacterial activity against <i>S. rapamycinicus</i> . | (König et al. 2013) |
| B/F or F/B | <i>Aspergillus fumigatus</i> | Solid | <i>Pseudomonas aeruginosa</i> | 1-Hydroxyphenazine, 1-methoxyphenazine, phenazine-1-sulfate, triacetyl fusarinine C [Al3+] and triacetyl fusarinine C [Fe3+], triacetyl fusarinine C, fusarinine C and phenazine dimers | Phenazines | Biotransformation | 1-hydroxyphenazine and 1-methoxyphenazine suppressed fungal growth; siderophores triacetyl fusarinine C, triacetyl fusarinine C [Al3+] and triacetyl fusarinine C [Fe3+] stimulate the capturing of iron | (Moree et al. 2012) |
| F/B | <i>Aspergillus fumigatus</i> | Liquid | <i>Streptomyces bullii</i> | ergosterol, brevianamide F, spirotryprostatin A, 6-methoxy spirotryprostatin B, fumitremorgin C and its 12,13-dihydroxy derivative, fumitremorgin B, verruculogen, 11-O-methylpsuerotin A, 11-O-methylpsuerotin A2, emestrins A-B | Diketopiperazine alkaloids | <i>de novo</i> biosynthesis | High toxicity and antiparasitic activity against <i>Trypanosoma brucei brucei</i> and <i>Leishmania donovani</i> (except brevianamide F and 11-O-methylpsuerotin A) | (Rateb et al. 2013) |
| F/B | <i>Aspergillus fumigatus</i> | Liquid | <i>Streptomyces peucetius</i> | Fumiformamide, N,N'-(1Z,3Z)-1,4-bis(4-methoxyphenyl)β-1,3-diene-2,3-diyl)diformamide, two known N-formyl derivatives and xanthocillin analogue BU-4704 | Alkaloids | <i>de novo</i> biosynthesis | N,N'-(1Z,3Z)-1,4-bis(4-methoxyphenyl)β-1,3-diene-2,3-diyl)diformamide has cytotoxic activity | (Zuck et al. 2011) |
| F/B | <i>Aspergillus fumigatus</i> KMC-901 | Liquid | <i>Sphingomonas</i> sp KMK-001 | glionitrin A | Diketopiperazine alkaloid | <i>de novo</i> biosynthesis | Antibacterial activity against MRSA, antitumor activity against human cancer cell | (Park et al. 2009) |

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|-------------|--|--------|---|--|--|-----------------------------|---|---|
| | | | | | | | lines HCT-116, A549, AGS, and DU145 | |
| F/F | <i>Aspergillus giganteus</i> | Liquid | <i>Aspergillus niger</i> | antifungal protein (AFP) | Protein | Down-regulation | Antifungal protein (AFP) has antifungal activity against various filamentous fungi | (Meyer and Stahl 2003) |
| F/F | <i>Aspergillus giganteus</i> | Liquid | <i>Fusarium oxysporum</i> | antifungal protein (AFP) | Protein | Up-regulation | Antifungal protein (AFP) has antifungal activity against various filamentous fungi | (Meyer and Stahl 2003) |
| F/F | <i>Aspergillus nidulans</i> | Liquid | Actinomycetes (58 soil-dwelling bacteria) and <i>Escherichia coli</i> | orsellinic acid, lecanoric acid, F-9775A and F-9775B | Archetypal polyketides and pentacyclic triterpenes | <i>de novo</i> biosynthesis | F-9775A and F-9775B has antiosteoporosis activity; orsellinic acid has antimicrobial activity; lecanoric acid inhibits ATP synthesis and electron transfer and has antimicrobial activity | (Schroeckh et al. 2009) |
| B/F and B/B | <i>Bacillus pumilus</i> | Liquid | <i>Serratia marcescens</i> , <i>Bacillus pumilus</i> or <i>Bacillus licheniformis</i> | not identified | Not identified | Up-regulation | Increase in biosurfactant activity | (Dusane et al. 2011) |
| B/B | <i>Bacillus amyloliquefaciens</i> LBM 5006 | Liquid | <i>Escherichia coli</i> ATCC 25922 | peptides | Bacteriocin/peptide | <i>de novo</i> biosynthesis | Antibacterial activity | (Benitez et al. 2011; Chanos and Mygind 2016) |
| B/F | <i>Bacillus amyloliquefaciens</i> | Solid | <i>Aspergillus fumigatus</i> and <i>Aspergillus niger</i> | Iturin | Lipopetides | <i>de novo</i> biosynthesis | Antifungal activity | (Moree et al. 2013) |
| B/B | <i>Bacillus cereus</i> UW85 | Liquid | <i>Cytophaga-Flavobacterium</i> (CF) | peptidoglycan from vegetative cells | Polymer | Up-regulation | Increase the growth of CF bacteria | (Peterson et al. 2006) |
| B/B | <i>Bacillus</i> isolate strain UA-094 | Liquid | <i>Bacillus</i> isolate strain UA-086 | indole and cyclo(Phe-Pro) diketopiperazine | Indole and diketopiperazine | Up-regulation | Antibacterial activity | (Trischman et al. 2004) |
| B/B | <i>Bacillus subtilis</i> | Solid | <i>Streptomyces griseus</i> | lactonase-homologous Protein | Protein | <i>de novo</i> biosynthesis | Antibacterial activity: inhibits development and | (Schneider et al. 2012) |

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|-------------|---|--------|--|--|---------------------------------|-----------------------------|---|---|
| | | | | | | | streptomycin production in <i>S. griseus</i> . | |
| B/B | <i>Bacillus subtilis</i> | Solid | <i>Streptomyces coelicolor</i> and <i>Streptomyces avermitilis</i> | bacillaene | Polyene | Up-regulation | Delays the production of pigmented antibiotics in <i>S. coelicolor</i> ; Antibiotic activity against <i>S. avermitilis</i> (inhibits the growth). | (Straight et al. 2006; Butcher et al. 2007) |
| B/B | <i>Bacillus subtilis</i> | Liquid | <i>Streptomyces coelicolor</i> | surfactin | Lipopeptide surfactant | Up-regulation | Inhibition of aerial hyphae and sporulation of <i>S. coelicolor</i> | (Straight et al. 2006) |
| F or B/B | <i>Bacillus subtilis</i> , <i>Escherichia coli</i> , <i>Saccharomyces cerevisiae</i> | Liquid | <i>Myxococcus xanthus</i> (proteobacterium) | peptidoglycan | Polymer | Up-regulation | Induction of rippling (ondulatory movement) in <i>M. xanthus</i> | (Berleman et al. 2006) |
| B/F and B/B | <i>Bacillus</i> sp. S3, <i>B. pumilus</i> S8, <i>B. licheniformis</i> D1, and <i>Serratia marcescens</i> V1 | Liquid | <i>Candida albicans</i> , <i>Yarrowia lipolytica</i> , <i>Pseudomonas aeruginosa</i> and <i>Bacillus pumilus</i> | not identified | Not identified | Up-regulation | Antibacterial activity against <i>P. aeruginosa</i> and <i>B. pumilus</i> | (Dusane et al. 2011) |
| B/B | <i>Bacillus</i> sp. S3 | Liquid | <i>Pseudomonas aeruginosa</i> | not identified | Not identified | Up-regulation | Quorum-sensing inhibition | (Dusane et al. 2011) |
| B/P | bacterial biofilms | Liquid | <i>Enteromorpha</i> (green seaweed) | N-acylhomoserine lactone (AHL) | Quorum sensing fatty acid (DSF) | Up-regulation | AHL attracts <i>Enteromorpha</i> zoospores | (Joint et al. 2002) |
| F/F | <i>Bionectria ochroleuca</i> | Solid | <i>Trichophyton rubrum</i> | 4"-hydroxysulfoxy-2,2"-dimethylthielavin P | Polyketide | <i>de novo</i> biosynthesis | No reports | (Bertrand et al. 2013) |

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|-----|-------------------------|-------|---|-------------------------------------|--|--------------------------------------|-----------------------|
| F/F | <i>Botrytis cinerea</i> | Solid | <p><i>Candida albicans</i>, <i>Aspergillus fumigatus</i>, <i>Colletotrichum acutatum</i>, <i>Fusarium proliferatum</i>, and <i>Magnaporthe grisea</i></p> <p>11-hydroxy-dehydro-botrydienol, botrytisic acid A, botrytisic acid B, dehydro-botrydienol, 3-hydroxy-4-oxocyclofarnesa-2,5,7,9-tetraen-11,8-oxide, norbotrydialone acetate, botrydienal phytotoxin, 10-oxo-dehydro-dihydro-botrydial, hydratedbotryenalol, and dehydro-botrydienal, 2-(hydroxy-5-methoxyphenoxy)acrylic acid, integrastin B, palmarumycin C15, australifungin, botryendial phytotoxin, cissetin, 3-O-acetyl-botcineric acid, equisetin, enniitantин I, ennitantин G, 11,12-hydroxy-eudesm-4-en-3-one, cordyol C, illudin C3, illudinic acid, integrastin B, 7-chrolo-6-methoxymellein, cis-4-hydroxy-6-deoxy-scylalone or xylarine, citreoviridin licicolinic acid A, 4-hydroxy-5-methylmellein, chaetoxanthone C, Cordyol C, mycorrhizin A, palmarumycins C7 and C15, chaetoxanthone C, diorcinol, destruxin A1/A4, antibiotic FR-49175, bis-dethio-bis-methyl-thio-gliotoxin, 3-hydroxymellein, 3,4-dihydro-3,8-dihydroxy-3-methyl-1H2-benzopyran-1-one, pleuromutilin, cyclopiazonic acid</p> | Fatty acids, enniантинs, and others | Up-regulation/ <i>de novo</i> biosynthesis | Antifungal activity and cytotoxicity | (Serrano et al. 2017) |
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|-------------|--|--------|--|---|---|--|---|---|
| F/F | <i>Serpula lacrymans</i> and <i>Coniophora puteana</i> | Solid | <i>Trichoderma</i> spp and <i>Scytalidium</i> FY | peroxidase | Enzymes | Up-regulation | Stimulate wood decay | (Score et al. 1997) |
| F/F | <i>Coniophora puteana</i> | Solid | <i>Scytalidium</i> FY | laccase | Enzymes | Up-regulation | Stimulate wood decay | (Score et al. 1997) |
| B/F | <i>Burkholderia cenocepacia</i> | Liquid | <i>Candida albicans</i> and <i>Xanthomonas campestris</i> pv. <i>campestris</i> | cis-2-dodecanoic acid (BDSF) | α,β Unsaturated fatty acid (DSF) | Up-regulation | BDSF restores the biofilm and extracellular polysaccharide production phenotypes of <i>X. campestris</i> and inhibits <i>C. albicans</i> germ tube formation. | (Boon et al. 2008) |
| B/B and B/P | <i>Burkholderia gladioli</i> | Liquid | <i>Burkholderia seminalis</i> and orchid | extracellular polysaccharides | Polysaccharides | Up-regulation | Polysaccharide alters hormone metabolism | (Araújo et al. 2016) |
| B/F | <i>Burkholderia gladioli</i> | Solid | <i>Rhizopus microsporus</i> | enacyloxin Iia, iso-enacyloxin Iia, enacyloxin IIIa and iso-enacyloxin IIIa | Polyketides | <i>de novo</i> biosynthesis | Antifungal and antibacterial activity | (Ross et al. 2014) |
| F/F | <i>Candida albicans</i> | Solid | <i>Trichophyton rubrum</i> , <i>Trichophyton mentagrophytes</i> , <i>Trichophyton terrestris</i> and <i>Microsporum gypseum</i> . (other common dermatophytes) | (3R,6E)-2,3-dihydrofarnesol (R-DHF) and (2E,6E)-farnesol (F-ol) | Acyclic sesquiterpenes | <i>de novo</i> biosynthesis/U p-regulation | F-ol and R-DHF has inhibitory effects on <i>Trichophyton rubrum</i> , <i>Trichophyto mentagrophytes</i> , <i>Microsporum canis</i> and <i>Epidermophyton floccosu</i> . | (Brasch et al. 2014) |
| F/B | <i>Candida albicans</i> | Liquid | <i>Pseudomonas aeruginosa</i> | farnesol | Sesquiterpene | Up-regulation | Reduces levels of quinolone and pyocyanin from <i>Pseudomonas</i> and inhibits swarming motility in <i>P. aeruginosa</i> | (Cugini et al. 2007; McAlester et al. 2008) |
| F/B | <i>Candida albicans</i> | Liquid | <i>Pseudomonas aeruginosa</i> | not identified | Not identified | Up-regulation | Inhibits virulence in <i>Pseudomonas aeruginosa</i> | (Lopez-Medina et al. 2015) |
| F/F | <i>Ceriporiopsis subvermispora</i> | Solid | <i>Pleurotus ostreatus</i> | laccase, manganese peroxidase and lignin-degrading enzymes | Enzymes | Up-regulation | Stimulate wood decay | (Chi et al. 2007) |

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|-----|--|--------|--|--|---|--|---|-----------------------|
| B/B | Diaminopimelic acid-peptidoglycan producing bacteria | Liquid | <i>Bacillus subtilis</i> | Peptidoglycan and disaccharide tripeptides (muropeptide fragments of the cell wall) | Polymers | Up-regulation | Germination stimulation of dormant <i>Bacillus subtilis</i> spores | (Shah et al. 2008) |
| F/B | <i>Emericella</i> sp strain CNL-878 | Liquid | <i>Salinisporea arenicola</i> strain CNH-665 | emericellamides A and B | Cyclic depsipeptide | Up-regulation | Emericellamide A has moderate antibacterial activity against MRSA and weak cytotoxicity against the human colon carcinoma cell line HCT-116; Emericellamide B was slightly weaker than 1. | (Oh et al. 2007) |
| F/B | <i>Chaetomium</i> sp. (endophytic fungus) | Solid | <i>Bacillus subtilis</i> | 3- and 4-hydroxybenzoic acid methyl esters, acremonisol A, SB236050, SB238569, isosulochrin, protocatechuic acid methyl ester, shikimeran A, bipherin A, chorismeron, quinomeron and serkydayn | Polyketides, dibenzoic acid, α pyrone and quinolinic alkaloids | Up-regulation/ <i>de novo</i> biosynthesis | Increase in antibacterial activity and cytotoxicity. | (Akone et al. 2016) |
| B/B | <i>Escherichia coli</i> | Liquid | <i>Salmonella typhimurium</i> | indole | Alkaloid | Up-regulation | Increase antibiotic tolerance (carbenicillin and ciprofloxacin) in <i>Salmonella typhimurium</i> | (Vega et al. 2013) |
| F/F | <i>Eutypa lata</i> | Solid | <i>Botryosphaeria obtusa</i> | O-methylmellein, 4-hydroxy-8-O-methylmellein and 5-hydroxy-8-O-methylmellein | Polyketides (mycoalexins) | Up-regulation/ <i>de novo</i> biosynthesis | Non-hydroxylated derivative have antifungal and phytotoxic activity | (Glauser et al. 2009) |
| F/F | <i>Alternaria tenuissima</i> | Solid | <i>Fusarium culmorum</i> and <i>Fusarium graminearum</i> | alternariol, alternariol monomethyl ether, altenuen | Mycotoxins | Down-regulation | Not reported | (Müller et al. 2012) |

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|-----|--|--------|--|---|--|--|---|-----------------------|
| F/F | <i>Fusarium culmorum</i> and <i>Fusarium graminearum</i> | Solid | <i>Alternaria tenuissima</i> | Deoxynivalenol and zearalenone | Trichothecene and Polyketide | Up-regulation | Deoxynivalenol has antibacterial activity; zearalenone shows cytochrome P450 3A4 inhibition | (Müller et al. 2012) |
| F/B | <i>Fusarium oxysporum</i> MSA35 (wild-type strain) | Liquid | <i>Serratia</i> sp and <i>Achromobacter</i> sp | isocaryophyllene, α -humulene, cyclocaryophyllan-4-ol | Sesquiterpenes | Up-Regulation | Antimicrobial activity of isocaryophyllene. | (Minardi et al. 2009) |
| F/F | <i>Fusarium oxysporum</i> MSA35 (wild-type strain) | Liquid | <i>F. oxysporum</i> f. sp. <i>lactucae</i> (pathogenic strain) | isocaryophyllene, α -humulene, cyclocaryophyllan-4-ol | Sesquiterpenes | Up-Regulation | Suppression of virulence in pathogenic <i>F. oxysporum</i> | (Minardi et al. 2009) |
| F/F | <i>Fusarium oxysporum</i> SIN2 | Solid | <i>Sarocladium</i> cf. <i>strictum</i> SIN29 | fusaric acid | Picolinic acid analogue | Up-regulation | Increase <i>Fusarium oxysporum</i> virulence | (Bohni et al. 2016) |
| F/B | <i>Fusarium pallidoroseum</i> | Liquid | <i>Saccharopolyspora erythraea</i> | N-demethylphloiosetin, pallidorosetin A, pallidorosetin B, equisetin, ophiosetin | Decalin-type tetramic acid analogues | <i>de novo</i> biosynthesis | No reports | (Whitt et al. 2014) |
| F/B | <i>Fusarium tricinctum</i> | Solid | <i>Bacillus subtilis</i> | ateropyrone, enniантин B, B1 and A1; fusaristatin A; macrocarpon C; (-)-citreoisocoumarin; 2-(carboxymethylamino)benzoic acid and (-)-citreoisocoumarinol | Pyrone/Depsipeptides/lipopeptide/Co umarin | Up-regulation/ <i>de novo</i> biosynthesis | Enniантин B1 and A1 have antibacterial activity against <i>Bacillus subtilis</i> , <i>Staphylococcus aureus</i> (MRSA and MSSA), <i>Streptococcus pneumoniae</i> , and <i>Enterococcus faecali</i> ; lateropyrone displayed antibacterial activity against <i>B. subtilis</i> , <i>S.aureus</i> , <i>S. pneumoniae</i> and <i>E. faecalis</i> | (Ola et al. 2013) |
| F/F | <i>Fusarium tricinctum</i> | Liquid | <i>Fusarium begoniae</i> | subenniatin A-B | Depsipeptides | <i>de novo</i> biosynthesis | Inactive in cytotoxic and antibacterial bioassays | (Wang et al. 2013b) |

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| F/F | <i>Gloeophyllum abietinum</i> | Liquid and solid | <i>Heterobasidion annosum</i> | oosponol, oospoglycol, melledonal A, melledonal C | Polyketides and Sesquiterpenes | Up-regulated | Antibacterial activity | (Sonnenbichler et al. 1994) |
| B/F | <i>Gluconobacter sp.W-315</i> | Liquid | <i>Neurospora crassa</i> or <i>Aspergillus oryzae</i> | enacyloxin IIa | Polyketide | <i>de novo</i> biosynthesis | Antibacterial activity | (Watanabe et al. 1982) |
| B/B | group A streptococcus (GAS) | Solid | group B streptococcus (GBS) | short hydrophobic peptides (SHP2/3) | Peptides | Up-regulation | Stimulate RovS-mediated gene regulation in GBS | (Cook et al. 2013) |
| B/B | group B streptococcus (GBS) | Solid | group A streptococcus (GAS) | short hydrophobic peptides (SHP1520) | Peptides | Up-regulation | Modulate Rgg2/3-regulated gene expression in GAS. | (Cook et al. 2013) |
| B/F | Gut microbiota | Liquid | <i>Candida albicans</i> | peptidoglycans | Polymer | Up-regulation | Increases hyphal development of the polymorphic fungal pathogen <i>C. albicans</i> | (Xu et al. 2008) |
| B/B | <i>Lactobacillus helveticus</i> M92, <i>Lactobacillus plantarum</i> L4, <i>Enterococcus faecium</i> L3 | Liquid | <i>Lactococcus lactis</i> subsp. <i>Lactis</i> LMG 9450 | not identified | Bacteriocin | Up-regulation | Antibiotic activity | (Kos et al. 2011; Chanos and Mygind 2016) |
| B/F | <i>Lactobacillus kefiranofaciens</i> | Liquid | <i>Saccharomyces cerevisiae</i> | kefiran | Bacteriocin (exopolysaccharide) | Up-regulation | Used as a thickener, stabilizer, emulsifier, fat substitute or gelling agent; also has antitumor activity | (Cheirsilp et al. 2003) |
| B/F | <i>Lactobacillus kefiranofaciens</i> | Liquid | <i>Saccharomyces cerevisiae</i> | kefiran | Bacteriocin (exopolysaccharide) | Up-regulation | Used as a thickener, stabilizer, emulsifier, fat substitute or gelling agent; also has antitumor activity | (Tada et al. 2007) |
| B/B | <i>Lactobacillus paracasei</i> | Liquid | <i>Bacillus subtilis</i> ATCC 11774 | paracin 1.7 | Bacteriocin | Up-regulation | Broad-spectrum antibacterial activity | (Ge et al. 2014; Chanos and Mygind 2016) |
| B/B | <i>Lactobacillus plantarum</i> DC400 | Liquid | <i>Lactobacillus sanfranciscensis</i> DPPMA174 and <i>Pediococcus pentosaceus</i> 2XA3 | plantaricin A | Bacteriocin (pheromone) | <i>de novo</i> biosynthesis | Antibacterial activity | (Di Cagno et al. 2009; Chanos and Mygind 2016) |
| B/B | <i>Lactobacillus plantarum</i> KLDS1.0391 | Liquid | <i>L. helveticus</i> KLDS1.0737, <i>Enterococcus faecium</i> KLDS4.0352, <i>Lactobacillus reuteri</i> KLDS1.0737, | plantaricin MG | Bacteriocin | Up-regulation | Antibacterial activity | (Man et al. 2012; Chanos and Mygind 2016) |

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| | | | <i>Enterococcus faecalis</i> KLDS4.0313 | | | | | |
| B/B | <i>Lactobacillus acidophilus La-5</i> | Liquid | <i>Streptococcus thermophilus</i> STY-3; <i>Lactobacillus delbrueckii</i> subsp. <i>bulgaricus</i> LBY-2; <i>L. delbrueckii</i> subsp. <i>bulgaricus</i> CECT 400; <i>L. delbrueckii</i> subsp. <i>lactis</i> CECT 282 and <i>Lactobacillus sakei</i> subsp. <i>sakei</i> CECT 906 | lactacin B | Bacteriocin | <i>de novo</i> biosynthesis/U p-regulation | Antibacterial activity | (Tabasco et al. 2009; Chanos and Mygind 2016) |
| B/B | <i>Lactobacillus acidophilus N2</i> (NCFM) | Liquid | <i>L. delbrueckii</i> , <i>L. helveticus</i> , <i>L. casei</i> , <i>L. fennatum</i> , <i>L. plantarum</i> , <i>L. viridescens</i> , <i>Streptococcus pyogenes</i> , <i>Staphylococcus aureus</i> , <i>Staphylococcus epidermidis</i> , <i>Bacillus cereus</i> , <i>Listeria monocytogenes</i> | lactacin B | Bacteriocin | <i>de novo</i> biosynthesis/U p-regulation | Antibacterial activity | (Barefoot et al. 1994; Chanos and Mygind 2016) |
| B/B | <i>Lactobacillus plantarum J23</i> | Liquid | <i>L. lactis</i> , <i>L. hilgardii</i> , <i>P. pentosaceus</i> , <i>L. lactis</i> , <i>L. brevis</i> , <i>L. fermentum</i> , <i>L. hilgardii</i> , <i>L. paracasei</i> , <i>L. pentosus</i> , <i>L. plantarum</i> , <i>L. sakei</i> , <i>L. mesenteroides</i> , <i>P. acidilacti</i> , <i>P. Parvulus</i> , <i>P. pentosaceus</i> | Not identified | Bacteriocin | <i>de novo</i> biosynthesis | Antibacterial activity against <i>Oenococcus oeni</i> , and a range of <i>Lactobacillus</i> and <i>Pediococcus</i> species | (Rojo-Bezares et al. 2007; Chanos and Mygind 2016) |

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| B/B | <i>Lactobacillus plantarum</i> NC8 | Liquid | <i>Bacillus cereus</i> , <i>Enterococcus faecalis</i> , <i>E. Faecium</i> , <i>L. acidophilus</i> , <i>L. brevis</i> , <i>L. bulgaricus</i> , <i>L. casei</i> , <i>L. fermentum</i> , <i>L. helveticus</i> , <i>L. hilgardii</i> , <i>L. reuteri</i> , <i>L. sake</i> , <i>L. salivarius</i> , <i>L. lactis</i> , <i>L. mesenteroides</i> , <i>Listeria innocua</i> , <i>P. pentosaceous</i> , <i>Staphylococcus carnosus</i> , <i>Streptococcus thermophilus</i> | plantaricin NC8 | Bacteriocin (Class Iib) | Up-regulated | Antibacterial activity | (Maldonado et al. 2004; Chanos and Mygind 2016) |
| B/B | <i>Lactococcus lactis</i> sub sp. <i>Lactis</i> | Liquid | <i>Yarrowia lipolytica</i> | nisin | Bacteriocin (Polycyclic peptide) | Up-regulation | Antibacterial activity | (Ariana and Hamed 2017) |
| B/F | <i>Lactococcus lactis</i> subsp. <i>lactis</i> ATCC 11454 | Liquid | <i>Saccharomyces cerevisiae</i> | nisin | Bacteriocin (Polycyclic peptide) | Up-regulation | Antibacterial activity | (Liu et al. 2006) |
| B/F | <i>Lactococcus lactis</i> subsp. <i>lactis</i> ATCC11454 | Liquid | <i>Kluyveromyces marxianus</i> | nisin | Bacteriocin (Polycyclic peptide) | Up-regulation | Antibacterial activity | (Shimizu et al. 1999) |
| B/B | <i>Leuconostoc citreum</i> GJ7 | Liquid | <i>Lactobacillus plantarum</i> KFRI 464, <i>Lactobacillus delbrueckii</i> KFRI 347 and <i>Leuconostoc mesenteroides</i> KCTC 1628 | kimchicin G7 | Bacteriocin | Up-regulation | Antibacterial activity | (Chang et al. 2007; Chanos and Mygind 2016) |
| F/B | <i>Libertella</i> sp | Liquid | Antibiotic-resistant marine a-proteobacterium | libertellenones A-D | Diterpenoids | <i>de novo</i> biosynthesis | Antitumoral activity against HCT-116 human adenocarcinoma cell line (libertellenone D is the most potent). | (Oh et al. 2005) |
| F/F | <i>Aspergillus</i> sp. (mangrove epiphytic) | Liquid | <i>Aspergillus</i> sp. (mangrove epiphytic) | asperginic, neoaspergillic acid, ergosterol | Alkaloids and Steroid | <i>de novo</i> biosynthesis | Asperginic, neoaspergillic acid show antibacterial activity against selected Gram bacteria. | (Zhu et al. 2011) |
| F/F | Mangrove fungi strain No. K38 | Liquid | Mangrove fungi strain No. E33 | 8-hydroxy-3-methyl-9-oxo-9H-xanthene-1-carboxylic acid methyl ether | Xanthone derivative | <i>de novo</i> biosynthesis | Antifungal activity against <i>Gloeaspisporum musae</i> and <i>Peronophthora cichoraelearum</i> | (Li et al. 2011) |

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| F/F | <i>Phomopsis</i> sp K38 (mangrove fungi) | Liquid | <i>Alternaria</i> sp E33 (mangrove fungi) | cyclo-(L-leucyl-trans-4-hydroxy-L-prolyl-D-leucyl-trans-4-hydroxy-L-proline) | Cyclopeptide (cyclic tetrapeptide) | <i>de novo</i> biosynthesis | Antifungal activity against <i>Gaeumannomyces graminis</i> , <i>Rhizoctonia cerealis</i> , <i>Helminthosporium sativum</i> and <i>Fusarium graminearum</i> | (Li et al. 2014) |
| F/F | <i>Phomopsis</i> sp K38 (mangrove fungi) | Liquid | <i>Alternaria</i> sp E33 (mangrove fungi) | cyclo (D-Pro-L-Tyr-L-Pro-L-Tyr) and cyclo (Gly-L-Phe-L-Pro-L-Tyr) | Cyclopeptide (cyclic tetrapeptide) | <i>de novo</i> biosynthesis | Moderate antifungal activity against <i>C. albicans</i> , <i>G. graminis</i> , <i>R. cerealis</i> , <i>H. sativum</i> and <i>F. graminearum</i> , Cyclo (Gly-L-Phe-L-Pro-L-Tyr) was more active than cyclo (D-Pro-L-Tyr-L-Pro-L-Tyr). | (Huang et al. 2014) |
| F/F | <i>Phomopsis</i> sp K38 (mangrove fungi) | Liquid | <i>Alternaria</i> sp E33 (mangrove fungi) | (-)-byssochlamic acid bisdiimid | Diimide derivative | <i>de novo</i> biosynthesis | (-)-byssochlamic has weak cytotoxic activity against Hep-2 and HepG2 cells. | (Li et al. 2010) |
| F/F | <i>Phomopsis</i> sp K38 (mangrove fungi) | Liquid | <i>Alternaria</i> sp E33 (mangrove fungi) | ethyl 5-ethoxy-2-formyl-3-hydroxy-4-methylbenzoate | Polysubstituted benzaldehyde derivative | <i>de novo</i> biosynthesis | Antifungal activity against <i>Fusarium graminearum</i> , <i>Gloeosporium musae</i> , <i>Rhizoctonia solani Kuhn</i> and <i>Phytophthora soja</i> . | (Wang et al. 2013a) |
| F/F | <i>Marasmius pallescens</i> | Solid and Liquid | <i>Marasmiellus troyanus</i> | laccase and manganese peroxidase activity | Enzymes | Up-regulation | Stimulate wood decay | (Ferreira Gregorio et al. 2006) |
| B/B | Marine bacteria | Solid | 85 other marine isolates | not identified | Not identified | Up-regulation | Antibacterial activity | (Long and Azam 2001) |
| B/B | <i>Prochlorococcus</i> strain MIT9313 and MED4 (marine cyanobacterium) | Liquid | heterotrophic marine bacteria | not identified | Not identified | Up-regulation/ <i>de novo</i> biosynthesis | Not reported. | (Sher et al. 2011) |
| B/B | <i>Micromonospora</i> spp., <i>Verrucosispora</i> spp., and <i>Solwaraspore</i> spp. (marine bacteria) | Liquid | <i>Mycobacterium</i> sp. WMMA-183 and <i>Rhodococcus</i> sp. WMMA-185 (marine mycolic acid-containing bacteria) | not identified | Not identified | Up-regulation/ <i>de novo</i> biosynthesis | Antibacterial activity | (Adnani et al. 2015) |
| F/F | Marine-derived mangrove endohytic fungi Strain n. 1924 | Liquid | Marine-derived mangrove endohytic fungi Strain n. 3893 | cyclo(phe-phe)dipeptide and 6-methylsalicylic acid | Peptide and beta-hydroxy acid | <i>de novo</i> biosynthesis | 6-methylsalicylic acid has biocide activity against pests <i>Heliothis armigera</i> Huhner and <i>Sinergasilus</i> spp. | (Zhu et al. 2007) |

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| F/F | <i>Monascus</i> sp | Solid and Liquid | <i>Saccharomyces cerevisiae</i> and <i>Aspergillus oryzae</i> | not identified | Pigment | Up-regulation | Overproduction of hydrophobic substances to block enzyme attack (defense mechanism) | (Shin et al. 1998) |
| F/F/F/F/F | <i>Ovadendron sulphureoochraceum</i> , <i>Ascochyta pisi</i> , <i>Emericellopsis minima</i> , <i>Cylindrocarpon destructans</i> and <i>Fusarium oxysporum</i> | Liquid | <i>Ovadendron sulphureoochraceum</i> , <i>Ascochyta pisi</i> , <i>Emericellopsis minima</i> , <i>Cylindrocarpon destructans</i> and <i>Fusarium oxysporum</i> | lateritin | N-methylated peptide (depsipeptide) | <i>de novo</i> biosynthesis | Antitumoral activity against human cancer cell lines pancreas BXPC-3, breast MCF-7, CNS SF268, lung NSC H460, colon KM20L2 and prostate DU145); antibacterial activity against <i>Micrococcus luteus</i> , <i>S. aureus</i> , <i>E. faecalis</i> and <i>S. pneumoniae</i> ; antifungal activity against <i>Candida albicans</i> | (Pettit et al. 2010) |
| F/F | <i>Paraconiothyrium SSM001</i> (Taxus endophyte) | Liquid | <i>Alternaria</i> and <i>Phomopsis</i> (bark fungi) | taxol | Diterpenoid | Up-regulation | Anticancer activity | (Soliman and Raizada 2013) |
| F/F | <i>Penicillium citrinum</i> | Liquid | <i>Beauveria feline</i> | citrifelins A–B | Citrinin adducts with a unique tetracyclic framework | <i>de novo</i> biosynthesis | Citrifelins A and B have antibacterial activity against <i>E. coli</i> , <i>S. aureus</i> , <i>A. hydrophilia</i> , <i>V. parahemolyticus</i> , <i>V. harveyi</i> , <i>E. tarda</i> , <i>V. alginolyticus</i> , and <i>V. anguillarums</i> . | (Meng et al. 2015) |
| F/F | <i>Penicillium fuscum</i> | Liquid | <i>Penicillium camembertii/clavigerum</i> | berkeleylactones A–H, macrolides A26771B, patulin, and citrinin | 16-membered-ring macrolides | <i>de novo</i> biosynthesis | Berkeleylactone A has potent antibacterial and antifungal activities against four MRSA, <i>Bacillus anthracis</i> , <i>Streptococcus pyogenes</i> , <i>Candida albicans</i> , and <i>Candida glabrata</i> | (Stierle et al. 2017) |
| B/F | <i>Penicillium pinophilum</i> FKI-5653 | Liquid | <i>Trichoderma harzianum</i> FKI-5655 | secopenicillide C, penicillide, MC-141, pestalasin A and stromemycin | Lactones | Up-regulation | Penicillide acts as antibiotic, plant growth inhibitor, mycotoxin, acyl-CoA-cholesterol acyltransferase inhibitor; anticancer and cytotoxic, non-peptidic oxytocin receptor antagonist and calpain inhibitor. | (Nonaka et al. 2011) |

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| F/B | <i>Penicillium</i> sp DT-F29 | Solid | <i>Bacillus</i> sp B31 | verruculogen TR-2; 12-hydroxyverruculogen TR-2; 12b-hydroxy-13a-methoxyverruculogen TR-2; 12b-hydroxy-13a-ethoxyverruculogen TR-2; 12b-hydroxy-13a-butoxyethoxyverruculogen TR-2; 12a-hydroxy-13a-prenylverruculogen TR-2; spirotryprostatin C; cycloprostatin C; 12,13-dihydroxyfumitremorgin C; cyclotryprostatin B; hydrocycloprostatin A; hydrocycloprostatin B; neofipiperazine C; fumitremorgin B; prenylcycloprostatin B; 25-hydroxyfumitremorgin B; 13-prenyl fumitremorgin B; 12b-hydroxy-13a-butoxyethoxyfumitremorgin B; 12b-hydroxy-13a-methoxyverruculogen B; fumitremorgin A; 26a-hydroxyfumitremorgin A; 25-hydroxyfumitremorgin A, and diprostatin A | 2,5-diketopiperazines | <i>de novo</i> biosynthesis/U p-regulation | 2,5-DKPs show bromodomain-containing protein 4 (BRD4) inhibitory activities; fumitremorgin B is a potential BRD4 inhibitor | (Yu et al. 2017) |
| F/B | <i>Pestalotia</i> sp | Liquid | Antibiotic-resistant marine α-proteobacterium | pestalone | Chlorinated benzophenone | <i>de novo</i> biosynthesis | Moderate antitumoral activity against human tumor cell lines; antibacterial activity against MRSA and vancomycin-resistant <i>Enterococcus faecium</i> | (Cueto et al. 2001) |
| F/F | <i>Phlebia radiata</i> , <i>Phlebia rufa</i> , <i>Coriolus versicolor</i> , <i>Stereum hirsutum</i> , <i>Phanerochaete velutina</i> and <i>Hypholoma fasciculare</i> | Solid | <i>Phlebia radiata</i> , <i>Phlebia rufa</i> , <i>Coriolus versicolor</i> , <i>Stereum hirsutum</i> , <i>Phanerochaete velutina</i> and <i>Hypholoma fasciculare</i> | phenoloxidase | Enzymes | Up-regulation/ <i>de novo</i> biosynthesis | Stimulate wood decay | (White and Boddy 1992) |
| O/Oomycetes | <i>Phytophthora nicotianae</i> | Liquid | <i>Phytophthora nicotianae</i> | mating hormones α1 and α2 | Hormones | <i>de novo</i> biosynthesis | Induction of sexual reproduction in the opposite mating type | (Ojika et al. 2011) |
| B/F | <i>Pseudomonas aeruginosa</i> | Solid | <i>Candida albicans</i> | 5-methyl-phenazinium-1-carboxylate (5MPCA) | Phenazines derivative (red pigment) | <i>de novo</i> biosynthesis | Redox-active antifungal activity | (Gibson et al. 2009) |
| B/B | <i>Pseudomonas aeruginosa</i> | Liquid | <i>Enterobacter</i> sp | pyocyanin | Phenazine derivative | <i>de novo</i> biosynthesis | Antibacterial activity | (Angell et al. 2006) |

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| B/F | <i>Pseudomonas aeruginosa</i> | Liquid | <i>Candida albicans</i> | N-3-oxo-dodecanoyl-L-Homoserine lactone (3-oxo-C12HSL) | Fatty acid (DSF) | Up-regulation | Prevents <i>Candida</i> morphology switch from yeast to hyphal (filamentous) growth. | (Cugini et al. 2007; McAlester et al. 2008) |
| B/B or F | <i>Pseudomonas aeruginosa</i> | Liquid | <i>Escherichia coli</i> , <i>Klebsiella pneumoniae</i> , <i>Proteus mirabilis</i> , <i>Streptococcus pyogenes</i> , <i>Bacillus subtilis</i> , <i>Staphylococcus aureus</i> and <i>Candida albicans</i> | cis-2-decenoic acid | a,b Unsaturated fatty acid (DSF) | Up-regulation | Induces the dispersion of biofilm microcolonies on the challenged microbes | (Davies and Marques 2009) |
| B/B | <i>Pseudomonas aeruginosa</i> | Solid | <i>Staphylococcus aureus</i> | 4-hydroxy-2-heptylquinoline-N-Oxide (HQNO) | Quinoline | <i>de novo</i> biosynthesis | Suppression of <i>S. aureus</i> respiration, protecting <i>S. aureus</i> from death by commonly used aminoglycoside antibiotics; Selection of typical <i>S. aureus</i> small-colony variants (SCVs), which are stable aminoglycoside resistance and persistence in chronic infections | (Hoffman et al. 2006) |
| B/B | <i>Rhodococcus fascians</i> (multi-antibiotic resistant mutant) | Liquid | <i>Streptomyces padanus</i> | rhodostreptomycin A-B | Aminoglycosides | Gene transfer | Antibacterial activity. | (Kurosawa et al. 2008) |
| F/F | <i>Schizophyllum commune</i> | Solid | <i>Hypholoma fasciculare</i> | indigo and indirubin | Pigments | Up-regulation | Indirubin is a potent inhibitors of cyclin dependent kinases and has antiviral, antibacterial and antifungal activities | (Menezes et al. 2015) |
| B/B | <i>Actinokineospora</i> sp. EG49 (sponge-associated Actinomycetes) | Liquid | <i>Nocardiopsis</i> sp. RV163 (sponge-associated Actinomycetes) | N-(2-hydroxyphenyl)-acetamide, 1,6-dihydroxyphenazine and 5a,6,11a,12-tetrahydro-5a,11a-dimethyl[1,4]benzoxazin o[3,2-b][1,4]benzoxazine | Angucycline, diketopiperazine and β-carboline | Up-regulation/ <i>de novo</i> biosynthesis | 1,6-dihydroxyphenazine has antibacterial activity against <i>Bacillus</i> sp. P25 and <i>Actinokineospora</i> sp. EG49 and antiprotozoal activity against <i>Trypanosoma brucei</i> | (Dashti et al. 2014) |
| B/B | <i>Stenotrophomonas maltophilia</i> | Liquid | <i>Pseudomonas aeruginosa</i> | cis-11-methyl-2-dodecanoic acid | a,b Unsaturated fatty acid (DSF) | Up-regulation | Stimulates biofilm filamentation, increases levels of a number of proteins with roles in bacterial stress tolerance and increased tolerance to | (Fouhy et al. 2007; Ryan et al. 2008) |

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| | | | | | | | polymyxins of <i>P. aeruginosa</i> | |
| B/B | <i>Streptococcus mutans</i> | Solid/Liquid | <i>Streptococcus sanguinis</i> | mutacins | Bacteriocin | Up-regulation | Antibacterial activity against <i>Streptococcus sanguinis</i> . | (Kreth et al. 2005) |
| B/B | <i>Streptococcus oligofermentas</i> | Solid/Liquid | <i>Streptococcus mutans</i> (major caries-causing pathogen) | hydrogen peroxide (H ₂ O ₂) | Oxidizing agent | Up-regulation | Antibacterial activity against <i>Streptococcus mutans</i> | (Tong et al. 2007) |
| B/F | <i>Streptococcus oralis</i> | Liquid | <i>Actinomyces naeslundii</i> T14V | autoinducer 2 | Furanosyl borate diester | Up-regulation | Induces biofilm growth in commensal bacteria | (Rickard et al. 2006) |
| B/B | <i>Streptococcus sanguinis</i> | Solid/Liquid | <i>Streptococcus mutans</i> (major caries-causing pathogen) | hydrogen peroxide (H ₂ O ₂) | Oxidizing agent | Up-regulation | Antibacterial activity against <i>Streptococcus mutans</i> . | (Kreth et al. 2005) |
| B/B | <i>Streptomyces bikiniensis</i> | Solid | <i>Salmonella dusseldorf</i> | streptomycin | Aminoglycoside | Up-regulation | Antibacterial activity against <i>Salmonella dusseldorf</i> . | (Turpin et al. 1992) |
| B/B | <i>Streptomyces cinnabarinus</i> PK209 | Liquid | <i>Alteromonas</i> sp. KNS-16 | lobocompactol | Diterpene | <i>de novo</i> biosynthesis | Antifouling activity against macroalgae <i>Ulva</i> and <i>Navicula</i> ; antioxidant and anticancer activity. | (Cho and Kim 2012) |
| B/B | <i>Streptomyces cinnamoneus</i> NBRC 13823 | Liquid | <i>Tsukamurella pulmonis</i> (mycolic acid-containing bacteria) | arcyriaflavin E | Indolocarbazole alkaloid | <i>de novo</i> biosynthesis | Citotoxicity | (Hoshino et al. 2015c) |
| B/B | <i>Streptomyces coelicolor</i> | Solid | <i>Streptomyces coelicolor</i> , <i>Amycolatopsis</i> sp., <i>Streptomyces</i> sp., <i>Streptomyces viridochromogenes</i> | four new acyl-desferrioxamines | Hydroxamic acid | Up-regulated/ <i>de novo</i> biosynthesis | Antibacterial activity | (Traxler et al. 2013) |
| B/B | <i>Streptomyces endus</i> S-522 | Liquid | <i>Tsukamurella pulmonis</i> S-522 or <i>Corynebacterium glutamicum</i> (mycolic acid-containing bacteria) | alchivemycin A | Polycyclic polyketide | <i>de novo</i> biosynthesis | Antibacterial activity against <i>Micrococcus luteus</i> | (Onaka et al. 2011) |

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| B/B | <i>Streptomyces lividans</i> TK23 | Liquid | <i>Tsukamurella pulmonis</i> , <i>T. pseudospumae</i> , <i>T. spumae</i> , <i>T. strandjordii</i> , <i>Corynebacterium glutamicum</i> , <i>C. efficiens</i> , <i>Rhodococcus erythropolis</i> , <i>R. coprophilus</i> , <i>R. wratislaviensis</i> , <i>R. zoppii</i> , <i>Gordonia rubripertincta</i> , <i>G. bronchialis</i> , <i>Nocardia farcinica</i> , <i>N. asteroides</i> , <i>Mycobacterium smegmatis</i> , <i>M. chlorophenolicum</i> , <i>Williamsia muralis</i> and <i>Dietzia cinnamea</i> | actinorhodin and undecylprodigiosin | Benzoisochromanequinone polyketide (red pigment) | <i>de novo</i> biosynthesis | Indicator of secondary metabolism | (Onaka et al. 2011) |
| B/B | <i>Streptomyces lividans</i> TK24 | Solid | <i>Salmonella dusseldorf</i> | not identified | Not identified | Up-regulation | <i>Streptomyces lividans</i> increased the survival of <i>Salmonella dusseldorf</i> | (Turpin et al. 1992) |
| F/F | <i>Streptomyces nodosus</i> strain 258 | Liquid | <i>C. brumptii</i> , <i>C. parapsilosis</i> , <i>P. lactosa</i> , <i>R. anrantica</i> , <i>R. rubra</i> , <i>Cryptococcus neoformans</i> , <i>C. albicans</i> , <i>C. krusei</i> , <i>C. tropicalis</i> , <i>C. puleherimma</i> and <i>C. utilis</i> | amphotericin B | Polyene | Up-regulation | Antifungal activity | (Yakovleva and Bulgakova 1978) |
| B/B | <i>Streptomyces</i> sp CJ-5 | Liquid | <i>Tsukamurella pulmonis</i> TP-B0596 (mycolic acid-containing bacteria) | chojalactones A–C | butanolide chojalactones with unusual γ -butyrolactone scaffolds | <i>de novo</i> biosynthesis | Chojalactones A and B showed moderate cytotoxicity against P388 cells. | (Hoshino et al. 2015b) |
| B/B | <i>Streptomyces</i> sp NZ-6 | Liquid | <i>Tsukamurella pulmonis</i> TP-B0596 (mycolic acid-containing bacteria) | niizalactams A–C | Di- and tricyclic macrolactams | <i>de novo</i> biosynthesis | Did not show antimicrobial activity or cytotoxicity. | (Hoshino et al. 2015a) |

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| B/B | <i>Streptomyces</i> sp. strain B033 | Liquid | <i>Burkholderia vietnamensis</i> ATCC BAA-248, <i>Brucella neotomae</i> ATCC 23459, <i>Yersinia pestis</i> A1122 and <i>Xanthomonas axonopodis</i> ATCC 8718 (pathogenic strains of the phylum <i>Proteobacteria</i>). | resistomycin | Aminoglycosides | <i>de novo</i> biosynthesis | Increase in antibacterial activity. | (Carlson et al. 2015) |
| B/B | <i>Streptomyces</i> sp. Mg1 | Solid | <i>Bacillus subtilis</i> | surfactin hydrolase | Enzymes | Biotransformation | Increase in surfactin hydrolase activity (detoxification). | (Hoefler et al. 2012) |
| B/B | <i>Streptomyces</i> sp. strain PTY087I2 | Liquid | <i>Bacillus subtilis</i> , MSSA, MRSA and <i>Pseudomonas aeruginosa</i> (human pathogens) | granaticin, granatomycin D, and dihydrogranaticin B | Naphthoquinone | Up-regulation | Antibacterial activity against the all gram positive human pathogens used in these experiments | (Sung et al. 2017) |
| F/B | <i>Streptomyces</i> spp. | Liquid | <i>Bacillus subtilis</i> | staurosporine | Indolocarbazole alkaloid | Up-regulation | Antibacterial activity and protein kinase inhibition; Germination stimulation of dormant <i>Bacillus subtilis</i> spores | (Shah et al. 2008) |
| B/B | <i>Streptomyces tenimariensis</i> | Liquid | 23 different marine bacteria | istamycins A–B | Aminoglycosides | Up-regulation | Antibacterial activity against competitor colonies | (Slattery et al. 2001) |
| B/B | <i>Symbiobacterium thermophilum</i> | Liquid | <i>Bacillus</i> sp | 2,2-bis(3-indolyl)indoxyl and 1,1-bis(3-indolyl)ethane | Indole | <i>de novo</i> biosynthesis | Antibacterial activities against <i>Staphylococcus aureus</i> and <i>Bacillus subtilis</i> | (Watsuji et al. 2007) |
| F/F | <i>Trichoderma harzianum</i> SIWT 25 | Solid | <i>Serpula lacrymans</i> and <i>Coniophora puteana</i> | tyrosinase | Enzymes | Up-regulation | Stimulate wood decay | (Score et al. 1997) |
| F/F | <i>Trametes versicolor</i> | Liquid | <i>Pleurotus ostreatus</i> | laccase | Enzymes | Up-regulation | Stimulate wood decay | (Baldrian 2004) |

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| F/F | <i>Trametes versicolor</i> | Solid | <i>Stereum gausapatum</i> , <i>Daldinia concentrica</i> , <i>Bjerkandera adusta</i> , <i>Fomes fomentarius</i> and <i>Hypholoma fasciculare</i> (five other wood decay fungi) | laccase and manganese-repressed peroxidase (MRP) | Enzymes | Up-regulation | Stimulate wood decay. Laccase acts to oxidise a variety of phenolic compounds and aromatic diamines; peroxidases utilise hydrogen peroxide (H_2O_2) to catalyse the oxidation of a range of organic and inorganic compounds. | (Hiscox et al. 2010) |
| F/F | <i>Trametes versicolor</i> | Solid | <i>Ganoderma applanatum</i> | (N-(4-methoxyphenyl)formamide 2-O- β -D-xyloside and N-(4-methoxyphenyl)formamide 2-O-beta-D-xylobioside), 3-phenyllactic acid, gluconic acid and orsellinic acid | Xylosides and Carboxylic acids | Up-regulation/ <i>de novo</i> biosynthesis | N-(4-methoxyphenyl)formamide 2-O-beta-D-xylobioside) has potential to enhance the cell viability of human immortalized bronchial epithelial cell line of Beas-2B | (Yao et al. 2016) |
| F/F | <i>Trametes versicolor</i> and <i>Pleurotus ostreatus</i> | Liquid | <i>Trichoderma harzianum</i> , bacteria and yeasts (soil microbes) | laccase | Enzymes | Up-regulation | Stimulate wood decay | (Baldrian 2004) |
| B/B | <i>Escherichia coli</i> P5v (transformed bacteria engineered to provide DHS precursor) | Liquid | <i>Escherichia coli</i> (transformed strains engineered with 3AB synthase and the DHS transporter to provide 3AB) | 3-amino-benzoic acid (3AB) | Organic Acid | Up-regulation | 3AB is an important building block molecule for biosynthesis of several natural products | (Zhang and Stephanopoulos 2016) |
| F/P | <i>Trichoderma atroviride</i> | Liquid | <i>Arabidopsis thaliana</i> seedlings | Indole-acetic acid-related indoles | Indoles | Up-regulation | Stimulatory effect on plant growth | (Salas-Marina et al. 2011) |
| F/F | <i>Trichoderma harzianum</i> | Solid | <i>Moniliophthora roreri</i> | T39 butenolide, harzianolide, sorbicillinol | Lactones | <i>de novo</i> biosynthesis | T39 butenolide and harzianolide have antifungal activity against phytopathogens. | (Tata et al. 2015) |
| F/F | <i>Trichoderma</i> sp | Liquid | <i>Lentinula edodes</i> | β -mannosidase and laccase | Enzymes | Up-regulation | Stimulate wood decay. Lowering of Mn-dependent peroxidase activity | (Savoie and Mata 1999) |
| F/F | Unidentified Fungi (strain Nos. 1924) | Liquid | Unidentified Fungi (strain Nos. 3893) | marinamide (4-(2-pyrrolyl)-1-isoquinolone-3-carboxylic acid) and methyl ester marinamide (methyl 4-(2-pyrrolyl)-1-isoquinolone-3-carboxylate) | Alkaloids | <i>de novo</i> biosynthesis | Antibacterial activity against <i>E. coli</i> , <i>Pseudomonas pyocyanne</i> , and <i>S. aureus</i> ; Antitumoral activity against tumor cell lines | (Zhu et al. 2007; Zhu et al. 2013) |

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| B/B | Unidentified surface-associated marine bacteria | Liquid | <i>Staphylococcus aureus</i> , <i>Pseudomonas aeruginosa</i> or <i>Escherichia coli</i> (human pathogens) | not identified | Not identified | Up-regulation | Antibacterial activity | (Mearns-Spragg et al. 1998) |
| F/F | <i>Trametes versicolor</i> | Liquid | <i>Trichoderma harzianum</i> (soil fungus and biocontrol organism) | laccase and cellobiase | Enzymes | Up-regulation | Stimulate wood decay | (Freitag and Morrell 1992) |
| B/F | <i>Xanthomonas campestris</i> pv. <i>campestris</i> | Liquid | <i>Candida albicans</i> and <i>Xanthomonas campestris</i> pv. <i>Campestris</i> | cis -11-methyl-2-dodecenoic acid | a,b Unsaturated fatty acid (DSF) | Up-regulation | Regulates virulence in <i>X. Campestris</i> ; Regulates morphological transition and virulence by <i>C. albicans</i> | (Wang et al. 2004) |
| B/B | <i>Xylella fastidiosa</i> | Liquid | <i>Methylobacterium mesophilicum</i> | not identified | Not identified | Up-regulation | <i>X. fastidiosa</i> down-regulate genes related to growth and up-regulate genes related to energy production, stress, transport, and motility, | (Dourado et al. 2015) |
| F/B | <i>Fusarium oxysporum</i> f. sp. <i>radicis-lycopersici</i> (soil-borne fungus) | Liquid | <i>Pseudomonas chlororaphis</i> strain PCL1391 | fusaric acid | Polyketide | Up-regulation | Represses the production of signal N-hexanoyl-L-homoserine lactone (C6-HSL) and phenazine-1-carboxamide | (van Rij et al. 2005) |
| F/B | <i>Fusarium oxysporum</i> | Liquid | <i>Pseudomonas fluorescens</i> CHA0 (biocontrol strain) | fusaric acid | Polyketide | Up-regulation | Suppress 2,4-diacetylphloroglucinol (2,4-DAPG) production in strain CHA0 | (Notz et al. 2002) |
| F/B | <i>Fusarium oxysporum</i> | Liquid | <i>Pseudomonas fluorescens</i> | Enzyme | Enzymes | Up-regulation | Degradate 2,4-DAPG into less toxic derivatives monoacetyl-phloroglucinol and phloroglucinol | (Schouten et al. 2004) |
| F/F | biocontrol agent <i>Trichoderma harzianum</i> M10 | Liquid | endophyte <i>Talaromyces pinophilus</i> F36CF | harziaphilic acid | tetramic acid derivative | <i>de novo</i> biosynthesis | Harziaphilic acids did not affect viability of colorectal cancer and healthy colonic epithelial cells, but selectively reduced cancer cell proliferation. | (Vinale et al. 2017) |
| F/F | biocontrol agent <i>Trichoderma harzianum</i> M10 | Liquid | endophyte <i>Talaromyces pinophilus</i> F36CF | ferricrocin, coprogen B and dimerumic acid | siderophores | Up-regulation | - | (Vinale et al. 2017) |

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| F/F | biocontrol agent <i>Trichoderma harzianum</i> M10 | Liquid | endophyte <i>Talaromyces pinophilus</i> F36CF | 3-O-methylfunicone and herquline B | γ -pyrone | Down-regulation | - | (Vinale et al. 2017) |
| F/B | <i>Aspergillus niger</i> | Liquid | <i>Streptomyces coelicolor</i> | cyclo(Phe-Phe) and 2-hydroxyphenylacetic acid | cyclic dipeptide | Up-regulation | - | (Wu et al. 2015) |
| F/B | <i>Aspergillus fumigatus</i> | Liquid | <i>Streptomyces rapamycinicus</i> | fumigermin | polyketide | <i>de novo</i> biosynthesis | Fumigermin inhibits germination of spores of the inducing <i>S. rapamycinicus</i> | (Stroe et al. 2020) |
| B/F | marine <i>Streptomyces</i> | Liquid | <i>Bacillus</i> strains | dentigerumycin E | cyclic hexapeptide incorporating three piperazic acids | <i>de novo</i> biosynthesis | Dentigerumycin E exhibited antiproliferative and antimetastatic activities against human cancer cells | (Shin et al. 2018) |
| F/B | <i>Fusarium tricinctum</i> | Solid | <i>Streptomyces lividans</i> | fusatricinones A–D (1), dihydrolateropyrone (2), zearalenone, (−)-citreoisocoumarin, macrocarpon C and 7-hydroxy-2-(2-hydroxypropyl)-5-methylchromone | naphthoquinone dimers (1), lateropyrone derivative (2) | <i>de novo</i> biosynthesis | - | (Moussa et al. 2019) |
| F/B | <i>Fusarium tricinctum</i> | Solid | <i>Streptomyces lividans</i> | lateropyrone, the enniatins B, B1 and A1, and fusaristatin A | Depsipeptides and lipopeptide | Up-regulation | - | (Moussa et al. 2019) |
| B/F | <i>Pseudomonas aeruginosa</i> | Solid | <i>Fusarium tricinctum</i> | 2-heptyl- 4-hydroxy-quinolone (HHQ), phenazine-1-carboxylic acid (PCA) and phenazine-1-carboxamide (PCN) | quorum sensing molecule and phenazine alkaloids | <i>de novo</i> biosynthesis | - | (Moussa et al. 2020) |
| F/F | <i>Ustilago maydis</i> | solid | <i>Fusarium verticillioides</i> | Over 20 chromatographic peaks | - | <i>de novo</i> biosynthesis | <i>F. verticillioides</i> is a strong antagonist of <i>U. maydis</i> as its presence leads to large reductions in <i>U. maydis</i> biomass. | (Estrada et al. 2011) |

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| F/F | <i>Stereum hirsutum</i> | Solid | <i>Coprinus micaceus</i> and <i>Coprinus disseminatus</i> | - | - | Up-regulation | <i>S. hirsutum</i> mycelial fronts always overgrew those of <i>C. micaceus</i> , there were down-regulations of metabolites in the interaction zone, compared to monocultures of both <i>S. hirsutum</i> and <i>C. micaceus</i> . | (Peiris et al. 2008) |
| F/F | <i>Hypholoma fasciculare</i> | Solid | <i>Resinicium bicolor</i> | benzoic acid methyl ester, a benzyl alcohol, a quinolinium type compound and other volatiles | sesquiterpenes | <i>de novo</i> biosynthesis | two wood decay basidiomycete fungi | (Hynes et al. 2007) |
| F/F | <i>Rhodotorula glutinis</i> DBVPG 3853 | Liquid | <i>Debaryomyces castellii</i> DBVPG 3503 | β-carotene, torulene, torularhodin | carotenoid pigments | Up-regulation | - | (Buzzini 2001) |
| F/B | <i>Penicillium notatum</i> | Liquid | <i>Cyanobacterium Scytonema ocelatum</i> | <i>tolytoxin</i> | Phytoalexin | <i>de novo</i> biosynthesis | Antifungal | (Patterson and Bolis 1997) |
| B/F | <i>Streptomyces sp. US80</i> | Liquid | heat-killed fungi | irumamycin, X-14952 B and 17-hydroxy-venturicidin A | - | Up-regulation | Antifungal | (Fourati-Ben Fguira et al. 2008) |
| B/B | <i>Streptomyces sp. RKBH-B178</i> | Liquid | autoclaved <i>Mycobacterium smegmatis</i> | hydrazidomycin D | hydrazide | <i>de novo</i> biosynthesis | - | (Liang et al. 2019) |
| B/B | <i>Streptomyces sp. RKBH-B178</i> | Liquid | autoclaved <i>Pseudomonas aeruginosa</i> | Glucuronidated-PQS | glucuronidated analog of the pseudomonas quinolone signal (PQS) | <i>de novo</i> biosynthesis | - | (Liang et al. 2019) |
| B/B | <i>Bacillus cereus</i> , <i>Flavobacterium johnsoniae</i> , and <i>Pseudomonas koreensis</i> | Liquid | <i>Bacillus cereus</i> , <i>Flavobacterium johnsoniae</i> , and <i>Pseudomonas koreensis</i> | zwittermicin, petrobactin, derivatives of lokisin | sidephores | Up-regulation | Antibiotic | (Chevrette et al. 2022) |

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| B/B | <i>Streptomyces hygroscopicus</i> HOK021 | Liquid | <i>Tsukamurella pulmonis</i> TP-B0596 | harundomycin A | conjugate between FabF inhibitor and catechol-type siderophore | <i>de novo</i> biosynthesis | Bifunctional Antibacterial Conjugate | (Asamizu et al. 2022) |
| F/F | <i>Aspergillus carneus</i> KMM 4638 | Liquid | <i>Beauveria felina</i> (= <i>Isaria felina</i>) KMM 4639 | asperflavinoids B, D and E | drimane-Type Sesquiterpenes | <i>de novo</i> biosynthesis | Cytotoxic | (Zhuravleva et al. 2022) |
| F/B | <i>Aspergillus nidulans</i> | Liquid | <i>Streptomycetes</i> | anhydrosepedonin (1) and antibiotic C (2) | iron-chelating tropolones and tripyridone | <i>de novo</i> biosynthesis and up-regulation | antibacterial | (Gerke et al. 2022) |
| F/F | <i>Alternaria</i> | Liquid | <i>Trichoderma atroviride</i> | alternariol | benzochromenone | Up-regulation | mycotoxin | (Tian et al. 2023) |
| F/F | <i>Trichoderma atr ovinide</i> | Liquid | <i>Alternaria</i> | Hydroxylation of alternariol (AOH-OH) | benzochromenone | Biotransformation | - | (Tian et al. 2023) |
| B/F | <i>Streptomyces</i> sp 13F051 | Liquid | <i>Leohumicola minima</i> 15S071 | ulleungdolin | polyketide-peptide hybrid | <i>de novo</i> biosynthesis | - | (Hwang et al. 2022) |
| F/F | <i>Botrytis cinerea</i> | Liquid | <i>Pichia fermentans</i> (two strains), <i>Issatchenkia terricola</i> and <i>Wickerhamomyces anomalous</i> . | 46 differentially secreted metabolites, including trans-cinnamic acid and the indole-3-carboxaldehyde | Phenylpropanoid and alkaloid | Up-regulation | Antifungal for biocontrol | (Fernandez-San Millan et al. 2022) |
| F/F | <i>Aspergillus oryzae</i> | Solid | <i>Zygosaccharomyces rouxii</i> | l-Arginine and l-Glutamate increase and l-Isoleucine decreased | Aminoacids | Regulation | <ul style="list-style-type: none"> - fungal breakage rate increased - enzyme activity decreased | (Liu et al. 2022b) |
| F/F | <i>Trichoderma</i> strains | Liquid | <i>Trichoderma</i> strains | Unkown | Unkown | Regulation | <ul style="list-style-type: none"> - synergistically promote plant growth - antagonistic activity against pathogens | (Liu et al. 2022a) |
| F/F | <i>Aspergillus nidulans</i> | Liquid | <i>Aspergillus fumigatus</i> | diphenyl ethers | polyketides | Up-regulation | Antibacterial | (Ninomiya et al. 2022) |

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| F/F | <i>Aspergillus fumigatus</i> | | <i>Aspergillus nidulans</i> | Unkown | siderophore | Up-regulation | - | (Ninomiya et al. 2022) |
| F/F | <i>Fusarium oxysporum</i> AB2 | Liquid | <i>Epicoccum nigrum</i> TORT | Beauvericin | Depsipeptides | Up-regulation | insecticidal and antibacterial activities | (Vásquez-Bonilla et al. 2022) |
| F/F | <i>Azospirillum oryzae</i> NBT506 | Liquid | <i>Bacillus velezensis</i> UTB96 | Surfactin and indole-3-acetic acid (AA) | surfactin production, indole-3-acetic acid (IAA) production, phosphate solubilization and enzyme activities | Up-regulation | plant growth-promoting bacteria (PGPB) | (Bagheri et al. 2022) |
| F/F | <i>Aspergillus oryzae</i> | Liquid | <i>Zygosaccharomyces rouxii</i> | Unkown | Unkown | Up-regulation | Antibacterial | (Liu et al. 2022c) |
| F/F | deep-sea-derived fungi <i>Penicillium crustosum</i> PRB-2 | Solid | <i>Penicillium fellutatum</i> HDN14-323 | penifellutins A (1), B (2), penifellutins C (3) and D (4), | Polyketides | <i>de novo</i> biosynthesis | - | (Yu et al. 2022) |
| F/B | <i>Aspergillus flavipes</i> | Solid | <i>Bacillus subtilis</i> | Taxol | - | Up-regulation | Chemotherapy | (El-Sayed et al. 2021) |
| F/F | <i>Penicillium fuscum</i> | Solid | <i>P. camembertii/clavigerum</i> | berkeleypenostatins A–G (1–7), berkeleylactones A–H, the known macrolide A26771B, citrinin, and patulin | macrolide | <i>de novo</i> biosynthesis and Up-regulation | Anticancer | (Stierle et al. 2021) |
| B/F | <i>Streptomyces lunalinharesii</i> | Solid | Phytopathogen <i>Rhizoctonia solani</i> | desferrioxamine E and anisomycin | siderophores | Up-regulation | - | (Maimone et al. 2021) |
| B/F | wild-type lactic acid bacteria | Liquid | auxotrophic <i>Saccharomyces cerevisiae</i> | riboflavin and folate | B-group vitamins | Up-regulation | - | (Konstantinidis et al. 2021) |
| F/F | Marine <i>Aspergillus terreus</i> C23-3 | Liquid | Marine <i>Aspergillus unguis</i> DLEP200800 1 | unguisin A, lovastatin, and nidulin and others | Multiple | Up-regulation | Antimicrobial | (Wang et al. 2022) |

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| F/F | <i>Irpex lacteus</i> | Solid | <i>Nigrospora oryzae</i> | irperide | butenolide | <i>de novo</i> biosynthesis | Antifungal | (Wu et al. 2022) |
| F/F | <i>Alternaria, Botryosphaeria, Phoma and Talaromyces</i> | Solid | <i>Fusarium pseudograminearum</i> (Fp) | Unkown | Unkown | Unkown | Antifungal | (Zhao et al. 2022) |
| F/B | <i>Aspergillus sydowii</i> | Liquid | <i>Bacillus subtilis</i> | s (-)- (7S)- 10-hydroxysydonic acid, serine sydonate and macrolactin U' | - | Up-regulation | Antibacterial against <i>Staphylococcus aureus</i> | (Sun et al. 2022) |
| B/B | <i>Enterococcus faecium</i> 135 | Liquid | <i>Ligilactobacillus salivarius</i> and <i>Limosilactobacillus reuteri</i> | bacteriocin-like inhibitory substances (BLIS) | Bacteriocin | Up-regulation | Antibacterial | (Piazzentini et al. 2022) |
| B/B | <i>Streptomyces sp</i> | Liquid | <i>Achromobacter sp</i> | Ligiamycins A and B | Decalin-Amino-Maleimides | Up-regulation | - | (Lim et al. 2022) |
| F/F | <i>Cosmospora sp.</i> | Solid | Phytopathogen <i>Magnaporthe oryzae</i> | soudanones A, E, D (1-3) and their two new derivatives, soudanones H-I (4-5), the known isochromans, pseudoangillosporins A and B (6, 7), naphtho-γ-pyrone, cephalochromin and ustilaginoidin G (8, 9), and ergosterol (10). | Isochromanones | <i>de novo</i> biosynthesis and Up-regulation | - | (Oppong-Danquah et al. 2022) |
| B/B | <i>Streptomyces sp</i> | Solid | <i>Pandoraea sp.</i> | Gwanakosides A and B | 6-Deoxy-α-l-talopyranose-Bearing Aromatic Metabolites | <i>de novo</i> biosynthesis | Antibacterial | (Huynh et al. 2022) |
| B/B | Soil <i>Bacillus velezensis</i> | Solid | <i>Pseudomonas</i> | surfactin | lipopeptides | Up-regulation | - enhance motility - chemical trap to reduce the toxicity | (Andrić, S et al. 2021) |

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| F/F | <i>Trametes coccinea</i> (F3) | Solid | <i>Leiotrametes lactinea</i> (F9) and <i>T. versicolor</i> (F1) | Melanin and laccase | Group of multicopper oxidases involved in detoxification of toxic compounds | Up-regulation | - | (Dullah et al. 2021) |
| F/F | <i>Talaromyces calidicanius</i> | Solid | <i>Aspergillus chevalieri</i> | Unkown | Unkown | Up-regulation | inhibiting the enzyme acetylcholinesterase | (de Oliveira et al. 2022) |
| F/B | <i>Aspergillus nidulans</i> | Liquid | <i>Streptomyces iranensis</i> | orsellinic acid, lecanoric acid, F-9775A and F-9775B | Archetypal polyketides and pentacyclic triterpenes | <i>de novo</i> biosynthesis | Arginine-derived polyketides serve as the bacterial signals | (Krespach et al. 2022) |
| B/F | <i>Paenibacillus peoriae</i> strain M48F | Solid | <i>Pleurotus ostreatus</i> HK35 and <i>Pleurotus eryngii</i> DSMZ 8264 | 5-diisopropylpyrazine | VOC | Up-regulation | enhance mycelial growth | (Orban et al. 2023) |
| F/B | <i>Pleurotus ostreatus</i> HK35 and <i>Pleurotus eryngii</i> DSMZ 8264 | Solid | <i>Paenibacillus peoriae</i> strain M48F | β -bisabolene | sesquiterpenes | Up-regulation | fungal defense reaction | (Orban et al. 2023) |
| A/F | marine dinoflagellate <i>Prorocentrum lima</i> | Liquid | <i>Aspergillus pseudoglaucus</i> | okadaic acid and dinophysistoxin 1 | Up-regulation | Up-regulation | Toxines | (Berry et al. 2022) |

References

- Adnani N, Vazquez-Rivera E, Adibhatla SN, Ellis GA, Braun DR, Bugni TS (2015) Investigation of interspecies interactions within marine micromonosporaceae using an improved co-culture approach. *Marine Drugs* 13:6082–6098. <https://doi.org/10.3390/md13106082>
- Akone SH, Mándi A, Kurtán T, Hartmann R, Lin W, Daletos G, Proksch P (2016) Inducing secondary metabolite production by the endophytic fungus *Chaetomium* sp. through fungal e bacterial co-culture and epigenetic modification. *Tetrahedron* 72:6340–6347. <https://doi.org/10.1016/j.tet.2016.08.022>
- Andrić S, Meyer T, Rigolet A, Prigent-Combaret C, Höfte M, Balleux G, Steels S, Hoff G, De Mot R, McCann A, De Pauw E, Arias AA, Ongena M (2021) Lipopeptide Interplay Mediates Molecular Interactions between Soil Bacilli and Pseudomonads. *Microbiology Spectrum* 9:e02038-21. <https://doi.org/10.1128/spectrum.02038-21>
- Angell S, Bench BJ, Williams H, Watanabe CMH (2006) Pyocyanin Isolated from a Marine Microbial Population: Synergistic Production between Two Distinct Bacterial Species and Mode of Action. *Chemistry and Biology* 13:1349–1359. <https://doi.org/10.1016/j.chembiol.2006.10.012>
- Araújo WL, Creason AL, Mano ET, Camargo-Neves AA, Minami SN, Chang JH, Loper JE (2016) Genome Sequencing and Transposon Mutagenesis of *Burkholderia seminalis* TC3.4.2R3 Identify Genes Contributing to Suppression of Orchid Necrosis Caused by *B. gladioli*. Molecular plant-microbe interactions : MPMI 29:MPMI02160047R. <https://doi.org/10.1094/MPMI-02-16-0047-R>
- Ariana M, Hamed J (2017) Enhanced production of nisin by co-culture of *Lactococcus lactis* sub sp. *lactis* and *Yarrowia lipolytica* in molasses based medium. *Journal of Biotechnology* 256:21–26. <https://doi.org/10.1016/j.jbiotec.2017.07.009>
- Asamizu S, Pramana AAC, Kawai S, Arakawa Y, Onaka H (2022) Comparative Metabolomics Reveals a Bifunctional Antibacterial Conjugate from Combined-Culture of *Streptomyces hygroscopicus* HOK021 and *Tsukamurella pulmonis* TP-B0596. *ACS Chem Biol* 17:2664–2672. <https://doi.org/10.1021/acschembio.2c00585>
- Bagheri N, Ahmadzadeh M, Mariotte P, Jouzani GS (2022) Behavior and interactions of the plant growth-promoting bacteria *Azospirillum oryzae* NBT506 and *Bacillus velezensis* UTB96 in a co-culture system. *World Journal of Microbiology and Biotechnology* 38:101. <https://doi.org/10.1007/s11274-022-03283-8>
- Baldrian P (2004) Increase of laccase activity during interspecific interactions of white-rot fungi. *FEMS Microbiology Ecology* 50:245–253. <https://doi.org/10.1016/j.femsec.2004.07.005>
- Barefoot SF, Chen Y, Hughes TA, Bodine AB, Shearer MY, Hughes MD (1994) Identification and Purification of a Protein That Induces Production of the *Lactobacillus acidophilus* Bacteriocin Lactacin Bt. *Applied and Environmental Microbiology* 60:3522–3528
- Benitez L, Correa A, Daroit D, Brandelli A (2011) Antimicrobial activity of *Bacillus amyloliquefaciens* LBM 5006 is enhanced in the presence of *Escherichia coli*. *Current Microbiology* 62:1017–1022. <https://doi.org/10.1007/s00284-010-9814-z>
- Berleman JE, Chumley T, Cheung P, Kirby JR (2006) Rippling is a predatory behavior in *Myxococcus xanthus*. *Journal of Bacteriology* 188:5888–5895. <https://doi.org/10.1128/JB.00559-06>
- Berry O, Briand E, Bagot A, Chaigné M, Meslet-Cladière L, Wang J, Grovel O, Jansen JJ, Ruiz N, Robiou du Pont T, Pouchus YF, Hess P, Bertrand S (2022) Deciphering interactions between the marine dinoflagellate *Prorocentrum lima* and the fungus *Aspergillus pseudoglaucus*. *Environmental Microbiology* n/a. <https://doi.org/10.1111/1462-2920.16271>
- Bertrand S, Schumpp O, Bohni N, Monod M, Gindro K, Wolfender J-L (2013) De novo production of metabolites by fungal co-culture of *Trichophyton rubrum* and *Bionectria ochroleuca*. *Journal of Natural Products* 76:1157–1165. <https://doi.org/10.1021/np400258f>

- Bohni N, Hofstetter V, Gindro K, Buyck B, Schumpp O, Bertrand S, Monod M, Wolfender J-L (2016) Production of fusaric acid by *Fusarium* spp. in pure culture and in solid medium co-cultures. *Molecules* 21:370–386. <https://doi.org/10.3390/molecules21030370>
- Boon C, Deng Y, Wang LH, He Y, Xu JL, Fan Y, Pan SQ, Zhang LH (2008) A novel DSF-like signal from *Burkholderia cenocepacia* interferes with *Candida albicans* morphological transition. *ISME Journal* 2:27–36. <https://doi.org/10.1038/ismej.2007.76>
- Brasch J, Hortsch F, Fritsch D, Beck-Jendroschek V, Troger A, Francke W (2014) Acyclic sesquiterpenes released by *Candida albicans* inhibit growth of dermatophytes. *Medical Mycology* 52:48–55. <https://doi.org/10.3109/13693786.2013.814174>
- Butcher RA, Schroeder FC, Fischbach MA, Straight PD, Kolter R, Walsh CT, Clardy J (2007) The identification of bacillaene, the product of the PksX megacomplex in *Bacillus subtilis*. *Proceedings of the National Academy of Sciences* 104:1506–1509. <https://doi.org/10.1073/pnas.0610503104>
- Buzzini P (2001) Batch and fed-batch carotenoid production by *Rhodotorula glutinis* - *Debaryomyces castellii* co-cultures in corn syrup. *Journal of Applied Microbiology* 90:843–847. <https://doi.org/10.1046/j.1365-2672.2001.01319.x>
- Carlson S, Tanouye U, Omarsdottir S, Murphy BT (2015) Phylum-specific regulation of resistomycin production in a streptomyces sp. via microbial coculture. *Journal of Natural Products* 78:381–387. <https://doi.org/10.1021/np500767u>
- Chagas FO, Dias LG, Pupo MT (2013) A Mixed Culture of Endophytic Fungi Increases Production of Antifungal Polyketides. *Journal of Chemical Ecology* 39:1335–1342. <https://doi.org/10.1007/s10886-013-0351-7>
- Chang JY, Lee HJ, Chang HC (2007) Identification of the agent from *Lactobacillus plantarum* KFRI464 that enhances bacteriocin production by *Leuconostoc citreum* GJ7. *Journal of Applied Microbiology* 103:2504–2515. <https://doi.org/10.1111/j.1365-2672.2007.03543.x>
- Chanos P, Mygind T (2016) Co-culture-inducible bacteriocin production in lactic acid bacteria. *Applied Microbiology and Biotechnology* 100:4297–4308. <https://doi.org/10.1007/s00253-016-7486-8>
- Cheirsilp B, Shimizu H, Shioya S (2003) Enhanced kefiran production by mixed culture of and. *Journal of Biotechnology* 100:43–53. [https://doi.org/10.1016/S0168-1656\(02\)00228-6](https://doi.org/10.1016/S0168-1656(02)00228-6)
- Chevrette MG, Thomas CS, Hurley A, Rosario-Meléndez N, Sankaran K, Tu Y, Hall A, Magesh S, Handelsman J (2022) Microbiome composition modulates secondary metabolism in a multispecies bacterial community. *Proc Natl Acad Sci USA* 119:e2212930119. <https://doi.org/10.1073/pnas.2212930119>
- Chi Y, Hatakka A, Maijala P (2007) Can co-culturing of two white-rot fungi increase lignin degradation and the production of lignin-degrading enzymes? *International Biodeterioration and Biodegradation* 59:32–39. <https://doi.org/10.1016/j.ibiod.2006.06.025>
- Cho JY, Kim MS (2012) Induction of Antifouling Diterpene Production by *Streptomyces cinnabarinus* PK209 in Co-Culture with Marine-Derived *Alteromonas* sp. KNS-16. *Bioscience, Biotechnology, and Biochemistry* 76:1849–1854. <https://doi.org/10.1271/bbb.120221>
- Cook LC, LaSarre B, Federle MJ (2013) Interspecies communication among commensal and pathogenic streptococci. *mBio* 4:1–11. <https://doi.org/10.1128/mBio.00382-13>
- Cueto M, Jensen PR, Kauffman C, Fenical W, Lobkovsky E, Clardy J (2001) Pestalone, a New Antibiotic Produced by a Marine Fungus in Response to Bacterial Challenge. *Journal of Natural Products* 64:1444–1446
- Cugini C, Calfee MW, Farrow JM, Morales DK, Pesci EC, Hogan DA (2007) Farnesol, a common sesquiterpene, inhibits PQS production in *Pseudomonas aeruginosa*. *Molecular Microbiology* 65:896–906. <https://doi.org/10.1111/j.1365-2958.2007.05840.x>

- Dashti Y, Grkovic T, Abdelmohsen UR, Hentschel U, Quinn RJ (2014) Production of induced secondary metabolites by a co-culture of sponge-associated actinomycetes, *Actinokineospora* sp. EG49 and *Nocardiopsis* sp. RV163. *Marine Drugs* 12:3046–3059. <https://doi.org/10.3390/md12053046>
- Davies DG, Marques CNH (2009) A fatty acid messenger is responsible for inducing dispersion in microbial biofilms. *Journal of Bacteriology* 191:1393–1403. <https://doi.org/10.1128/JB.01214-08>
- de Oliveira GP, Barreto DLC, Ramalho Silva M, Augusti R, Evódio Marriel I, Gomes de Paula Lana U, Takahashi e A (2022) Biotic stress caused by in vitro co-inoculation enhances the expression of acetylcholinesterase inhibitors by fungi. *Natural Product Research* 36:4266–4270. <https://doi.org/10.1080/14786419.2021.1975701>
- Degenkolb T, Heinze S, Schlegel B, Strobel G, Gräfe U (2002) Formation of new lipoaminopeptides, acremostatins A, B, and C, by co-cultivation of *Acremonium* sp. Tbp-5 and *Mycogone rosea* DSM 12973. *Bioscience, Biotechnology, and Biochemistry* 66:883–886. <https://doi.org/10.1271/bbb.66.883>
- Di Cagno R, De Angelis M, Coda R, Minervini F, Gobbetti M (2009) Molecular adaptation of sourdough *Lactobacillus plantarum* DC400 under co-cultivation with other lactobacilli. *Research in Microbiology* 160:358–366. <https://doi.org/10.1016/j.resmic.2009.04.006>
- Dourado MN, Santos DS, Nunes LR, Costa de Oliveira RLB, de Oliveira M V, Araújo WL (2015) Differential gene expression in *Xylella fastidiosa* 9a5c during co-cultivation with the endophytic bacterium *Methylobacterium mesophilicum* SR1.6/6. *Journal of Basic Microbiology* 55:1357–1366. <https://doi.org/10.1002/jobm.201400916>
- Dullah S, Hazarika DJ, Goswami G, Borgohain T, Ghosh A, Barooah M, Bhattacharyya A, Boro RC (2021) Melanin production and laccase mediated oxidative stress alleviation during fungal-fungal interaction among basidiomycete fungi. *IMA Fungus* 12:33. <https://doi.org/10.1186/s43008-021-00082-y>
- Dusane DH, Matkar P, Venugopalan VP, Kumar AR, Zinjarde SS (2011) Cross-species induction of antimicrobial compounds, biosurfactants and quorum-sensing inhibitors in tropical marine epibiotic bacteria by pathogens and biofouling microorganisms. *Current Microbiology* 62:974–980. <https://doi.org/10.1007/s00284-010-9812-1>
- Ebrahim W, El-Neketi M, Lewald LI, Orfali RS, Lin W, Rehberg N, Kalscheuer R, Daletos G, Proksch P (2016) Metabolites from the Fungal Endophyte *Aspergillus austroafricanus* in Axenic Culture and in Fungal-Bacterial Mixed Cultures. *Journal of Natural Products* 79:914–922. <https://doi.org/10.1021/acs.jnatprod.5b00975>
- El-Sayed ASA, Shindia AA, AbouZeid A, Koura A, Hassanein SE, Ahmed RM (2021) Triggering the biosynthetic machinery of Taxol by *Aspergillus flavipes* via cocultivation with *Bacillus subtilis*: proteomic analyses emphasize the chromatin remodeling upon fungal-bacterial interaction. *Environmental Science and Pollution Research* 28:39866–39881. <https://doi.org/10.1007/s11356-021-13533-1>
- Estrada AER, Hegeman A, Kistler HC, May G (2011) In vitro interactions between *Fusarium verticillioides* and *Ustilago maydis* through real-time PCR and metabolic profiling. *Fungal Genetics and Biology* 48:874–885. <https://doi.org/10.1016/j.fgb.2011.06.006>
- Fernandez-San Millan A, Gamir J, Larraya L, Farran I, Veramendi J (2022) Towards understanding of fungal biocontrol mechanisms of different yeasts antagonistic to *Botrytis cinerea* through exometabolomic analysis. *Biological Control* 174:105033. <https://doi.org/10.1016/j.bioc.2022.105033>
- Ferreira Gregorio AP, Da Silva IR, Sedarati MR, Hedger JN (2006) Changes in production of lignin degrading enzymes during interactions between mycelia of the tropical decomposer basidiomycetes *Marasmiellus troyanus* and *Marasmius pallescens*. *Mycological Research* 110:161–168. <https://doi.org/10.1016/j.mycres.2005.10.002>

- Fouhy Y, Scanlon K, Schouest K, Spillane C, Crossman L, Avison MB, Ryan RP, Dow JM (2007) Diffusible signal factor-dependent cell-cell signaling and virulence in the nosocomial pathogen *Stenotrophomonas maltophilia*. *Journal of Bacteriology* 189:4964–4968. <https://doi.org/10.1128/JB.00310-07>
- Fourati-Ben Fguira L, Smaoui S, Karray-Rebai I, Bejar S, Mellouli L (2008) The antifungal activity of the terrestrial *Streptomyces* US80 strain is induced by heat-killed fungi. *Biotechnology journal* 3:1058–1066. <https://doi.org/10.1002/biot.200700155>
- Freitag M, Morrell JJ (1992) Changes in selected enzyme activities during growth of pure and mixed cultures of the white-rot decay fungus *Trametes versicolor* and the potential biocontrol fungus *Trichoderma harzianum*. *Canadian Journal of Microbiology* 38:317–23. <https://doi.org/10.1139/m92-053>
- Ge J, Fang B, Wang Y, Song G, Ping W (2014) *Bacillus subtilis* enhances production of Paracin1.7, a bacteriocin produced by *Lactobacillus paracasei* HD1-7, isolated from Chinese fermented cabbage. *Annals of Microbiology* 64:1735–1743. <https://doi.org/10.1007/s13213-014-0817-z>
- Gerke J, Köhler AM, Wennrich J-P, Große V, Shao L, Heinrich AK, Bode HB, Chen W, Surup F, Braus GH (2022) Biosynthesis of Antibacterial Iron-Chelating Tropolones in *Aspergillus nidulans* as Response to Glycopeptide-Producing Streptomycetes. *Frontiers in Fungal Biology* 2
- Gibson J, Sood A, Hogan DA (2009) *Pseudomonas aeruginosa*-*Candida albicans* interactions: Localization and fungal toxicity of a phenazine derivative. *Applied and Environmental Microbiology* 75:504–513. <https://doi.org/10.1128/AEM.01037-08>
- Glauser G, Gindro K, Fringeli J, De Joffrey JP, Rudaz S, Wolfender J-L (2009) Differential Analysis of Mycoalexins in Confrontation Zones of Grapevine Fungal Pathogens by Ultrahigh Pressure Liquid Chromatography / Time-of-Flight Mass Spectrometry and Capillary Nuclear Magnetic Resonance. *Journal of Agricultural and Food Chemistry* 57:1127–1134. <https://doi.org/10.1021/jf8033539>
- Hiscox J, Baldrian P, Rogers HJ, Boddy L (2010) Changes in oxidative enzyme activity during interspecific mycelial interactions involving the white-rot fungus *Trametes versicolor*. *Fungal Genetics and Biology* 47:562–571. <https://doi.org/10.1016/j.fgb.2010.03.007>
- Hoefler BC, Gorzelnik K V, Yang JY, Hendricks N, Dorrestein PC, Straight PD (2012) Enzymatic resistance to the lipopeptide surfactin as identified through imaging mass spectrometry of bacterial competition. *Proceedings of the National Academy of Sciences* 109:13082–13087. <https://doi.org/10.1073/pnas.1205586109>
- Hoffman LR, Deziel E, D'Argenio DA, Lepine F, Emerson J, McNamara S, Gibson RL, Ramsey BW, Miller SI (2006) Selection for *Staphylococcus aureus* small-colony variants due to growth in the presence of *Pseudomonas aeruginosa*. *Proceedings of the National Academy of Sciences* 103:19890–19895. <https://doi.org/10.1073/pnas.0606756104>
- Hoshino S, Okada M, Wakimoto T, Zhang H, Hayashi F, Onaka H, Abe I (2015a) Niizalactams A-C, Multicyclic Macrolactams Isolated from Combined Culture of *Streptomyces* with Mycolic Acid-Containing Bacterium. *Journal of Natural Products* 78:3011–3017. <https://doi.org/10.1021/acs.jnatprod.5b00804>
- Hoshino S, Wakimoto T, Onaka H, Abe I (2015b) Chojalactones A-C, cytotoxic butanolides isolated from *streptomyces* sp. cultivated with mycolic acid containing bacterium. *Organic Letters* 17:1501–1504. <https://doi.org/10.1021/acs.orglett.5b00385>
- Hoshino S, Zhang L, Awakawa T, Wakimoto T, Onaka H, Abe I (2015c) Arcyriaflavin E, a new cytotoxic indolocarbazole alkaloid isolated by combined-culture of mycolic acid-containing bacteria and *Streptomyces cinnamoneus* NBRC 13823. *Journal of Antibiotics* 68:342–344
- Huang S, Ding W, Li C, Cox DG (2014) Two new cyclopeptides from the co-culture broth of two marine mangrove fungi and their antifungal activity. *Pharmacogn Mag* 10:410–414. <https://doi.org/10.4103/0973-1296.141781>

Huynh T-H, Lee J, Moon DH, Nguyen TQ, Son S, Hwang S, Du YE, Cui J, Jang J-H, Nam S-J, Shin J, Jang J, Lee SK, Oh K-B, Oh D-C (2022) Gwanakosides A and B, 6-Deoxy- α -l-talopyranose-Bearing Aromatic Metabolites from a Streptomyces sp. and Coculture with Pandoraea sp. *J Nat Prod* 85:83–90. <https://doi.org/10.1021/acs.jnatprod.1c00703>

Hwang GJ, Jang M, Son S, Kim GS, Lee B, Heo KT, Kim GJ, Choi H, Hur J-S, Jang J-P, Ko S-K, Hong Y-S, Ahn JS, Jang J-H (2022) Ulleungdolin, a Polyketide–Peptide Hybrid Bearing a 2,4-Di-O-methyl- β -d-antiarose from Streptomyces sp. 13F051 Co-cultured with Leohumicola minima 15S071. *J Nat Prod* 85:2445–2453. <https://doi.org/10.1021/acs.jnatprod.2c00682>

Hynes J, Müller CT, Jones TH, Boddy L (2007) Changes in Volatile Production During the Course of Fungal Mycelial Interactions Between Hypholoma fasciculare and Resinicum bicolor. *Journal of Chemical Ecology* 33:43–57. <https://doi.org/10.1007/s10886-006-9209-6>

Iakovlev A, Stenlid J (2000) Spatiotemporal Patterns of Laccase Activity in Interacting Mycelia of Wood-Decaying Basidiomycete Fungi. *Microbial Ecology* 39:236–245. <https://doi.org/10.1007/s002480000022>

Joint I, Tait K, Callow ME, Callow JA, Milton D, Williams P, Cámarra M (2002) Cell-to-cell communication across the prokaryote-eukaryote boundary. *Science* 298:1207. <https://doi.org/10.1126/science.1077075>

König CC, Scherlach K, Schroeckh V, Horn F, Nietzsche S, Brakhage AA, Hertweck C (2013) Bacterium Induces Cryptic Meroterpenoid Pathway in the Pathogenic Fungus *Aspergillus fumigatus*. *ChemBioChem* 14:938–942. <https://doi.org/10.1002/cbic.201300070>

Konstantinidis D, Pereira F, Geissen E-M, Grkovska K, Kafkia E, Jouhten P, Kim Y, Devendran S, Zimmermann M, Patil KR (2021) Adaptive laboratory evolution of microbial co-cultures for improved metabolite secretion. *Molecular Systems Biology* 17:e10189. <https://doi.org/10.1525/msb.202010189>

Kos B, Beganović J, Jurašić L, Švađumović M (2011) Coculture-inducible bacteriocin biosynthesis of different probiotic strains by dairy starter culture *Lactococcus lactis*. *Mlječarstvo* 61:273–282

Kresbach MKC, Stroe MC, Netzker T, Rosin M, Zehner LM, Komor AJ, Beilmann JM, Krüger T, Kniemeyer O, Schroeckh V, Hertweck C, Brakhage AA (2022) Ubiquitous bacterial polyketides induce cross-kingdom microbial interactions. *bioRxiv* 2022.05.09.491136. <https://doi.org/10.1101/2022.05.09.491136>

Kreth J, Merritt J, Shi W, Qi F (2005) Competition and Coexistence between *Streptococcus mutans* and *Streptococcus sanguinis* in the Dental Biofilm Competition and Coexistence between *Streptococcus mutans* and *Streptococcus sanguinis* in the Dental Biofilm. *Journal of Bacteriology* 187:7193–7203. <https://doi.org/10.1128/JB.187.21.7193>

Kurosawa K, Ghiviriga I, Sambandan TG, Lessard PA, Barbara JE, Rha C, Sinskey AJ (2008) Rhodostreptomycins, antibiotics biosynthesized following horizontal gene transfer from *Streptomyces padanus* to *Rhodococcus fascians*. *Journal of the American Chemical Society* 130:1126–1127. <https://doi.org/10.1021/ja077821p>

Li C, Wang J, Luo C, Ding W, Cox DG (2014) A new cyclopeptide with antifungal activity from the co-culture broth of two marine mangrove fungi. *Natural Product Research* 28:616–621. <https://doi.org/10.1080/14786419.2014.887074>

Li C, Zhang J, Shao C, Ding W, She Z, Lin J (2011) A New Xanthone Derivative from the co-culture broth of two marine fungi (strain No. E33 and K38). *Chemistry of Natural Compounds* 47:342–344

Li CY, Ding WJ, Shao CL, She ZG, Lin Y-C (2010) A new diimide derivative from the co-culture broth of two mangrove fungi (strain nos. E33 and K38). *Journal of Asian Natural Products Research* 12:809–813. <https://doi.org/10.1007/s10600-013-0751-5>

Liang L, Sproule A, Haltli B, Marchbank DH, Berrué F, Overy DP, McQuillan K, Lanteigne M, Duncan N, Correa H, Kerr RG (2019) Discovery of a New Natural Product and a Deactivation of a Quorum

Sensing System by Culturing a “Producer” Bacterium With a Heat-Killed “Inducer” Culture. *Frontiers in Microbiology* 9:3351

Lim H-J, An JS, Bae ES, Cho E, Hwang S, Nam S-J, Oh K-B, Lee SK, Oh D-C (2022) Ligiamycins A and B, Decalin-Amino-Maleimides from the Co-Culture of *Streptomyces* sp. and *Achromobacter* sp. Isolated from the Marine Wharf Roach, *Ligia exotica*. *Marine Drugs* 20. <https://doi.org/10.3390/md20020083>

Liu C, Hu B, Liu Y, Chen S (2006) Stimulation of nisin production from whey by a mixed culture of *Lactococcus lactis* and *Saccharomyces cerevisiae*. *Applied Biochemistry and Biotechnology* 131:751–761. <https://doi.org/10.1385/ABAB:131:1:751>

Liu H, Hao D, Li Y, Wang X, Chen J (2022a) Approaches for the establishment of optimized co-culture system of multiple *Trichoderma* strains for culture metabolites highly effective in cucumber growth promotion. *Frontiers in Microbiology* 13

Liu Z, Fu B, Duan X, Lv W, Kang S, Zhou M, Wang C, Li D, Xu N (2022b) Effects of cell-cell interactions between *A. oryzae* and *Z. rouxii* on morphology and secondary metabolites. *LWT* 170:114035. <https://doi.org/10.1016/j.lwt.2022.114035>

Liu Z, Kang B, Duan X, Hu Y, Li W, Wang C, Li D, Xu N (2022c) Metabolomic profiles of the liquid state fermentation in co-culture of *A. oryzae* and *Z. rouxii*. *Food Microbiology* 103:103966. <https://doi.org/10.1016/j.fm.2021.103966>

Long RA, Azam F (2001) Antagonistic interactions among marine pelagic bacteria. *Applied and Environmental Microbiology* 67:4975. <https://doi.org/10.1128/AEM.67.11.4975>

Lopez-Medina E, Fan D, Coughlin LA, Ho EX, Lamont IL, Reimann C, Hooper L V, Koh AY (2015) *Candida albicans* Inhibits *Pseudomonas aeruginosa* Virulence through Suppression of Pyochelin and Pyoverdine Biosynthesis. *PLoS Pathogens* 11:1–34. <https://doi.org/10.1371/journal.ppat.1005129>

Maimone NM, de Oliveira LFP, Santos SN, de Lira SP (2021) Elicitation of *Streptomyces lunalinharesii* secondary metabolism through co-cultivation with *Rhizoctonia solani*. *Microbiological Research* 251:126836. <https://doi.org/10.1016/j.micres.2021.126836>

Maldonado A, Ruiz-Barba JL, Jiménez-Díaz R (2004) Production of plantaricin NC8 by *Lactobacillus plantarum* NC8 is induced in the presence of different types of gram-positive bacteria. *Archives of Microbiology* 181:8–16. <https://doi.org/10.1007/s00203-003-0606-8>

Man LL, Meng XC, Zhao RH (2012) Induction of plantaricin MG under co-culture with certain lactic acid bacterial strains and identification of LuxS mediated quorum sensing system in *Lactobacillus plantarum* KLDS1.0391. *Food Control* 23:462–469. <https://doi.org/10.1016/j.foodcont.2011.08.015>

McAlester G, O’Gara F, Morrissey JP (2008) Signal-mediated interactions between *Pseudomonas aeruginosa* and *Candida albicans*. *Journal of Medical Microbiology* 57:563–569. <https://doi.org/10.1099/jmm.0.47705-0>

Mearns-Spragg A, Bregu M, Boyd KG, Burgess JG (1998) Cross-species induction and enhancement of antimicrobial activity produced by epibiotic bacteria from marine algae and invertebrates, after exposure to terrestrial bacteria. *Letters in applied microbiology* 27:142–146. <https://doi.org/10.1046/j.1472-765X.1998.00416.x>

Menezes RC, Kai M, Krause K, Matthäus C, Svatoš A, Popp J, Kothe E (2015) Monitoring metabolites from *Schizophyllum commune* interacting with *Hypholoma fasciculare* combining LESA-HR mass spectrometry and Raman microscopy. *Analytical and Bioanalytical Chemistry* 407:2273–2282. <https://doi.org/10.1007/s00216-014-8383-6>

Meng LH, Liu Y, Li XM, Xu GM, Ji NY, Wang BG (2015) Citrifelins A and B, Citrinin Adducts with a Tetracyclic Framework from Cocultures of Marine-Derived Isolates of *Penicillium citrinum* and *Beauveria felina*. *Journal of Natural Products* 78:2301–2305. <https://doi.org/10.1021/acs.jnatprod.5b00450>

Meyer V, Stahl U (2003) The influence of co-cultivation on expression of the antifungal protein in *Aspergillus giganteus*. *Journal of Basic Microbiology* 43:68–74. <https://doi.org/10.1002/jobm.200390007>

Minerdi D, Bossi S, Gullino ML, Garibaldi A (2009) Volatile organic compounds: A potential direct long-distance mechanism for antagonistic action of *Fusarium oxysporum* strain MSA 35. *Environmental Microbiology* 11:844–854. <https://doi.org/10.1111/j.1462-2920.2008.01805.x>

Moree WJ, Phelan V V, Wu C-H, Bandeira N, Cornett DS, Duggan BM, Dorrestein PC (2012) Interkingdom metabolic transformations captured by microbial imaging mass spectrometry. *Proceedings of the National Academy of Sciences* 109:13811–13816. <https://doi.org/10.1073/pnas.1206855109>

Moree WJ, Yang JY, Zhao X, Liu WT, Gutiérrez M, Dorrestein PC, Aparicio M, Atencio L, Ballesteros J, Sánchez J, Gavilán RG, Gutiérrez M, Dorrestein PC (2013) IMAGING MASS SPECTROMETRY OF A CORAL MICROBE INTERACTION WITH FUNGI. *Journal of Chemical Ecology* 39:1–14. <https://doi.org/10.1007/s10886-013-0320-1.IMAGING>

Moussa M, Ebrahim W, Bonus M, Gohlke H, Márdi A, Kurtán T, Hartmann R, Kalscheuer R, Lin W, Liu Z, Proksch P (2019) Co-culture of the fungus *Fusarium tricinctum* with *Streptomyces lividans* induces production of cryptic naphthoquinone dimers. *RSC Advances* 9:1491–1500. <https://doi.org/10.1039/C8RA09067J>

Moussa M, Ebrahim W, Kalscheuer R, Liu Z, Proksch P (2020) Co-culture of the bacterium *Pseudomonas aeruginosa* with the fungus *Fusarium tricinctum* induces bacterial antifungal and quorum sensing signaling molecules. *Phytochemistry Letters* 36:37–41. <https://doi.org/10.1016/j.phytol.2020.01.013>

Müller MEH, Steier I, Köppen R, Siegel D, Proske M, Korn U, Koch M (2012) Cocultivation of phytopathogenic *Fusarium* and *Alternaria* strains affects fungal growth and mycotoxin production. *Journal of Applied Microbiology* 113:874–887. <https://doi.org/10.1111/j.1365-2672.2012.05388.x>

Ninomiya A, Urayama S, Hagiwara D (2022) Antibacterial diphenyl ether production induced by co-culture of *Aspergillus nidulans* and *Aspergillus fumigatus*. *Applied Microbiology and Biotechnology* 106:4169–4185. <https://doi.org/10.1007/s00253-022-11964-5>

Nonaka K, Abe T, Iwatsuki M, Mori M, Yamamoto T, Shiomi K, Ômura S, Masuma R (2011) Enhancement of metabolites productivity of *Penicillium pinophilum* FKI-5653, by co-culture with *Trichoderma harzianum* FKI-5655. *Journal of Antibiotics* 64:769–774. <https://doi.org/10.1038/ja.2011.91>

Notz R, Maurhofer M, Dubach H (2002) Fusaric acid-producing strains of *Fusarium oxysporum* alter 2, 4-diacetylphloroglucinol biosynthetic gene expression in *Pseudomonas fluorescens* CHA0 in vitro and. *Applied and Environmental Microbiology* 68:2229–2235. <https://doi.org/10.1128/AEM.68.5.2229>

Oh DC, Jensen PR, Kauffman CA, Fenical W (2005) Libertellenones A-D: Induction of cytotoxic diterpenoid biosynthesis by marine microbial competition. *Bioorganic and Medicinal Chemistry* 13:5267–5273. <https://doi.org/10.1016/j.bmc.2005.05.068>

Oh DC, Kauffman CA, Jensen PR, Fenical W (2007) Induced production of emericellamides A and B from the marine-derived fungus *Emericella* sp. in competing co-culture. *Journal of Natural Products* 70:515–520. <https://doi.org/10.1021/np060381f>

Ojika M, Molli SD, Kanazawa H, Yajima A, Toda K, Nukada T, Mao H, Murata R, Asano T, Qi J, Sakagami Y (2011) The second Phytophthora mating hormone defines interspecies biosynthetic crosstalk. *Nature Chemical Biology* 7:591–593. <https://doi.org/10.1038/nchembio.617>

Ola ARB, Thomy D, Lai D, Brötz-Oesterhelt H, Proksch P (2013) Inducing Secondary Metabolite Production by the Endophytic Fungus *Fusarium tricinctum* through Coculture with *Bacillus subtilis*. *Journal of Natural Products* 76:2094–2099. <https://doi.org/10.1021/np400589h>

Onaka H, Mori Y, Igarashi Y, Furumai T (2011) Mycolic acid-containing bacteria induce natural-product biosynthesis in *Streptomyces* species. *Applied and Environmental Microbiology* 77:400–406. <https://doi.org/10.1128/AEM.01337-10>

Oppong-Danquah E, Blümel M, Scarpato S, Mangoni A, Tasdemir D (2022) Induction of Isochromanones by Co-Cultivation of the Marine Fungus *Cosmospora* sp. and the Phytopathogen *Magnaporthe oryzae*. International Journal of Molecular Sciences 23. <https://doi.org/10.3390/ijms23020782>

Orban A, Jerschow JJ, Birk F, Suarez C, Schnell S, Rühl M (2023) Effect of bacterial volatiles on the mycelial growth of mushrooms. Microbiological Research 266:127250. <https://doi.org/10.1016/j.micres.2022.127250>

Park HB, Kwon HC, Lee CH, Yang HO (2009) Glionitrin A , an Antibiotic-Antitumor Metabolite Derived from Competitive Interaction. Journal of Natural Products 72:248–252. <https://doi.org/10.1021/np800606e>

Patterson GML, Bolis CM (1997) Fungal Cell-Wall Polysaccharides Elicit An Antifungal Secondary Metabolite (Phytoalexin) In The Cyanobacterium *Scytonema Ocelatum*2. Journal of Phycology 33:54–60. <https://doi.org/doi:10.1111/j.0022-3646.1997.00054.x>

Peiris D, Dunn WB, Brown M, Kell DB, Roy I, Hedger JN (2008) Metabolite profiles of interacting mycelial fronts differ for pairings of the wood decay basidiomycete fungus, *Stereum hirsutum* with its competitors *Coprinus micaceus* and *Coprinus disseminatus*. Metabolomics 4:52–62. <https://doi.org/10.1007/s11306-007-0100-4>

Peterson SB, Dunn AK, Klimowicz AK, Handelsman J (2006) Peptidoglycan from *Bacillus cereus* mediates commensalism with rhizosphere bacteria from the Cytophaga-Flavobacterium group. Applied and Environmental Microbiology 72:5421–5427. <https://doi.org/10.1128/AEM.02928-05>

Pettit RK, Pettit GR, Xu JP, Weber CA, Richert LA (2010) Isolation of human cancer cell growth inhibitory, antimicrobial lateritin from a mixed fungal culture. Planta Medica 76:500–501. <https://doi.org/10.1055/s-0029-1240617>

Piazzentini ACM, Mendonça CMN, Vallejo M, Mussatto SI, de Souza Oliveira RP (2022) Bacteriocin-like inhibitory substances production by *Enterococcus faecium* 135 in co-culture with *Ligilactobacillus salivarius* and *Limosilactobacillus reuteri*. Brazilian Journal of Microbiology 53:131–141. <https://doi.org/10.1007/s42770-021-00661-6>

Rateb ME, Hallyburton I, Houssen WE, Bull AT, Goodfellow M, Santhanam R, Jaspars M, Ebel R (2013) Induction of diverse secondary metabolites in *Aspergillus fumigatus* by microbial co-culture. RSC Advances 3:14444. <https://doi.org/10.1039/c3ra42378f>

Rickard AH, Palmer RJ, Blehert DS, Campagna SR, Semmelhack MF, Egland PG, Bassler BL, Kolenbrander PE (2006) Autoinducer 2: A concentration-dependent signal for mutualistic bacterial biofilm growth. Molecular Microbiology 60:1446–1456. <https://doi.org/10.1111/j.1365-2958.2006.05202.x>

Rojo-Bezares B, Sáenz Y, Navarro L, Zarazaga M, Ruiz-Larrea F, Torres C (2007) Coculture-inducible bacteriocin activity of *Lactobacillus plantarum* strain J23 isolated from grape must. Food Microbiology 24:482–491. <https://doi.org/10.1016/j.fm.2006.09.003>

Ross C, Opel V, Scherlach K, Hertweck C (2014) Biosynthesis of antifungal and antibacterial polyketides by *Burkholderia gladioli* in coculture with *Rhizopus microsporus*. Mycoses 57:48–55. <https://doi.org/10.1111/myc.12246>

Ryan RP, Fouhy Y, Garcia BF, Watt SA, Niehaus K, Yang L, Tolker-Nielsen T, Dow JM (2008) Interspecies signalling via the *Stenotrophomonas maltophilia* diffusible signal factor influences biofilm formation and polymyxin tolerance in *Pseudomonas aeruginosa*. Molecular Microbiology 68:75–86. <https://doi.org/10.1111/j.1365-2958.2008.06132.x>

Salas-Marina MA, Silva-Flores MA, Uresti-Rivera EE, Castro-Longoria E, Herrera-Estrella A, Casas-Flores S (2011) Colonization of *Arabidopsis* roots by *Trichoderma atroviride* promotes growth and enhances systemic disease resistance through jasmonic acid/ethylene and salicylic acid pathways. European Journal of Plant Pathology 131:15–26. <https://doi.org/10.1007/s10658-011-9782-6>

Savoie JM, Mata G (1999) The antagonistic action of *Trichoderma* sp. hyphae to *Lentinula edodes* hyphae changes lignocellulolytic activities during cultivation in wheat straw. *World Journal of Microbiology and Biotechnology* 15:369–373. <https://doi.org/10.1023/A:1008979701853>

Schneider J, Yepes A, Garcia-Betancur JC, Westedt I, Mielich B, López D (2012) Streptomycin-induced expression in *Bacillus subtilis* of YtnP, a lactonase-homologous protein that inhibits development and streptomycin production in *Streptomyces griseus*. *Applied and Environmental Microbiology* 78:599–603. <https://doi.org/10.1128/AEM.06992-11>

Schouten A, van den Berg G, Edel-Hermann V, Steinberg C, Gautheron N, Alabouvette C, de Vos CH, Lemanceau P, Raaijmakers JM (2004) Defense responses of *Fusarium oxysporum* to 2,4-diacetylphloroglucinol, a broad-spectrum antibiotic produced by *Pseudomonas fluorescens*. *Molecular plant-microbe interactions: MPMI* 17:1201–1211. <https://doi.org/10.1094/MPMI.2004.17.11.1201>

Schroeckh V, Scherlach K, Nutzmann HW, Shelest E, Schmidt-Heck W, Schuemann J, Martin K, Hertweck C, Brakhage AA (2009) Intimate bacterial-fungal interaction triggers biosynthesis of archetypal polyketides in *Aspergillus nidulans*. *Proceedings of the National Academy of Sciences* 106:14558–14563. <https://doi.org/10.1073/pnas.0901870106>

Score AJ, Palfreyman JW, White NA (1997) Extracellular phenoloxidase and peroxidase enzyme production during interspecific fungal interactions. *International Biodegradation and Biodegradation* 39:225–233. [https://doi.org/10.1016/S0964-8305\(97\)00012-7](https://doi.org/10.1016/S0964-8305(97)00012-7)

Serrano R, González-Menéndez V, Rodríguez L, Martín J, Tormo JR, Genilloud O (2017) Co-culturing of fungal strains against *Botrytis cinerea* as a model for the induction of chemical diversity and therapeutic agents. *Frontiers in Microbiology* 8:649–664. <https://doi.org/10.3389/fmicb.2017.00649>

Shah IM, Laaberki MH, Popham DL, Dworkin J (2008) A Eukaryotic-like Ser/Thr Kinase Signals Bacteria to Exit Dormancy in Response to Peptidoglycan Fragments. *Cell* 135:486–496. <https://doi.org/10.1016/j.cell.2008.08.039>

Sher D, Thompson JW, Kashtan N, Croal L, Chisholm SW (2011) Response of *Prochlorococcus* ecotypes to co-culture with diverse marine bacteria. *The ISME Journal* 5:1125–1132. <https://doi.org/10.1038/ismej.2011.1>

Shimizu H, Mizuguchi T, Tanaka E, Shioya S (1999) Nisin production by a mixed-culture system consisting of *Lactococcus lactis* and *Kluyveromyces marxianus*. *Applied and Environmental Microbiology* 65:3134–3141

Shin CS, Kim HJ, Kim MJ, Ju JY (1998) Morphological change and enhanced pigment production of *Monascus* when cocultured with *Saccharomyces cerevisiae* or *Aspergillus oryzae*. *Biotechnology and Bioengineering* 59:576–581. [https://doi.org/10.1002/\(SICI\)1097-0290\(19980905\)59:5<576::AID-BIT7>3.0.CO;2-7](https://doi.org/10.1002/(SICI)1097-0290(19980905)59:5<576::AID-BIT7>3.0.CO;2-7)

Shin D, Byun WS, Moon K, Kwon Y, Bae M, Um S, Lee SK, Oh DC (2018) Coculture of Marine *Streptomyces* sp. With *Bacillus* sp. Produces a New Piperazic Acid-Bearing Cyclic Peptide. *Frontiers in Chemistry* 6:498. <https://doi.org/10.3389/fchem.2018.00498>

Slattery M, Rajbhandari I, Wesson K (2001) Competition-Mediated Antibiotic Induction in the Marine Bacterium *Streptomyces tenjimariensis*. *Microbial Ecology* 41:90–96. https://doi.org/10.1007/978-1-60327-140-0_4

Soliman SSM, Raizada MN (2013) Interactions between co-habitating fungi elicit synthesis of Taxol from an endophytic fungus in host *Taxus* plants. *Frontiers in Microbiology* 4:1–14. <https://doi.org/10.3389/fmicb.2013.00003>

Sonnenbichler J, Dietrich J, Peipp H (1994) Secondary Fungal Metabolites and Their Biological Activities, V. Investigations Concerning the Induction of the Biosynthesis of Toxic Secondary Metabolites in Basidiomycetes. *Biological Chemistry Hoppe-Seyler* 375:71–80. <https://doi.org/10.1515/bchm3.1994.375.1.71>

- Stierle AA, Stierle DB, Decato D, Alverson J, Apedaile L (2021) Cryptic Biosynthesis of the Berkeleypenostatins from Coculture of Extremophilic Penicillium sp. *J Nat Prod* 84:1656–1665. <https://doi.org/10.1021/acs.jnatprod.1c00248>
- Stierle AA, Stierle DB, Decato D, Priestley ND, Alverson JB, Hoody J, McGrath K, Klepacki D (2017) The Berkeleylactones, Antibiotic Macrolides from Fungal Coculture. *Journal of Natural Products* 80:1150–1160. <https://doi.org/10.1021/acs.jnatprod.7b00133>
- Straight PD, Willey JM, Kolter R (2006) Interactions between *Streptomyces coelicolor* and *Bacillus subtilis*: Role of surfactants in raising aerial structures. *Journal of Bacteriology* 188:4918–4925. <https://doi.org/10.1128/JB.00162-06>
- Stroe MC, Netzker T, Scherlach K, Krüger T, Hertweck C, Valiante V, Brakhage AA (2020) Targeted induction of a silent fungal gene cluster encoding the bacteria-specific germination inhibitor fumigermin. *eLife* 9:1–20. <https://doi.org/10.7554/eLife.52541>
- Sun Y, Shi X, Xing Y, Ren X-X, Zhang D-Y, Li X, Xiu Z-L, Dong Y-S (2022) Co-culture of *Aspergillus sydowii* and *Bacillus subtilis* induces the production of antibacterial metabolites. *Fungal Biology* 126:320–332. <https://doi.org/10.1016/j.funbio.2022.01.002>
- Sung AA, Gromek SM, Balunas MJ (2017) Upregulation and identification of antibiotic activity of a marine-derived *Streptomyces* sp. via co-cultures with human pathogens. *Marine Drugs* 15. <https://doi.org/10.3390/md15080250>
- Tabasco R, García-Cayuela T, Peláez C, Requena T (2009) *Lactobacillus acidophilus* La-5 increases lactacin B production when it senses live target bacteria. *International Journal of Food Microbiology* 132:109–116. <https://doi.org/10.1016/j.ijfoodmicro.2009.04.004>
- Tada S, Katakura Y, Ninomiya K, Shioya S (2007) Fed-batch coculture of *Lactobacillus kefiranofaciens* with *Saccharomyces cerevisiae* for effective production of kefiran. *Journal of Bioscience and Bioengineering* 103:557–562. <https://doi.org/10.1263/jbb.103.557>
- Tata A, Perez C, Campos ML, Bayfield MA, Eberlin MN, Ifa DR (2015) Imprint Desorption Electrospray Ionization Mass Spectrometry Imaging for Monitoring Secondary Metabolites Production during Antagonistic Interaction of Fungi. *Analytical Chemistry* 87:12298–12305. <https://doi.org/10.1021/acs.analchem.5b03614>
- Tian Y, Abdallah MF, De Boevre M, Audenaert K, Wang C, De Saeger S, Wu A (2023) Deciphering *Alternaria* metabolic responses in microbial confrontation via an integrated mass spectrometric targeted and non-targeted strategy. *Food Chemistry* 404:134694. <https://doi.org/10.1016/j.foodchem.2022.134694>
- Tong H, Chen W, Merritt J, Qi F, Shi W, Dong X (2007) *Streptococcus oligofermentans* inhibits *Streptococcus mutans* through conversion of lactic acid into inhibitory H₂O₂: A possible counteroffensive strategy for interspecies competition. *Molecular Microbiology* 63:872–880. <https://doi.org/10.1111/j.1365-2958.2006.05546.x>
- Traxler MF, Watrous JD, Alexandrov T, Dorrestein PC, Kolter R (2013) Interspecies interactions stimulate diversification of the *Streptomyces coelicolor* secreted metabolome. *mBio* 4:459–13. <https://doi.org/10.1128/mBio.00459-13>
- Trischman JA, Oeffner RE, De Luna MG, Kazaoka M (2004) Competitive induction and enhancement of indole and a diketopiperazine in marine bacteria. *Marine Biotechnology* 6:215–220. <https://doi.org/10.1007/s10126-003-0010-z>
- Turpin PE, Dhir VK, Maycroft KA, Rowland C, Wllington EMH (1992) The effect of *Streptomycetes* species on the survival of *Salmonella* in soil. *FEMS Microbiology Ecology* 101:271–280
- Ueda K, Kawai S, Ogawa H, Kiyama A, Kubota T, Kawanobe H, Beppu T (2000) Wide Distribution of Interspecific Stimulatory Events on Antibiotic Production and Sporulation among *Streptomyces* species. *The Journal of antibiotics* 53:979–982. <https://doi.org/11099234>

- van Rij ET, Girard G, Lugtenberg BJJ, Bloemberg G V (2005) Influence of fusaric acid on phenazine-1-carboxamide synthesis and gene expression of *Pseudomonas chlororaphis* strain PCL1391. *Microbiology* 151:2805–2814. <https://doi.org/10.1099/mic.0.28063-0>
- Vásquez-Bonilla JN, Barranco-Florido JE, Ponce-Alquicira E, Rincón-Guevara MA, Loera O (2022) Improvement of beauvericin production by *Fusarium oxysporum* AB2 under solid-state fermentation using an optimised liquid medium and co-cultures. *Mycotoxin Research* 38:175–183. <https://doi.org/10.1007/s12550-022-00458-y>
- Vega NM, Allison KR, Samuels AN, Klempner MS, Collins JJ (2013) *Salmonella typhimurium* intercepts *Escherichia coli* signaling to enhance antibiotic tolerance. *Proceedings of the National Academy of Sciences* 110:14420–14425. <https://doi.org/10.1073/pnas.1308085110>
- Vinale F, Nicoletti R, Borrelli F, Mangoni A, Parisi OA, Marra R, Lombardi N, Lacatena F, Grauso L, Finizio S, Lorito M, Woo SL (2017) Co-Culture of Plant Beneficial Microbes as Source of Bioactive Metabolites. *Scientific Reports* 7:1–12. <https://doi.org/10.1038/s41598-017-14569-5>
- Wang J, Ding W, Li C, Huang S, She Z, Lin Y (2013a) A New Polusubstituted Benzaldehyde from the co-culture broth of two marine fungi (strains Nos. E33 and K38). *Chemistry of Natural Compounds* 49:689–691
- Wang J, Lin W, Wray V, Lai D, Proksch P (2013b) Induced production of depsipeptides by co-culturing *Fusarium tricinctum* and *Fusarium begoniae*. *Tetrahedron Letters* 54:2492–2496. <https://doi.org/10.1016/j.tetlet.2013.03.005>
- Wang LH, He Y, Gao Y, Wu JE, Dong YH, He C, Wang SX, Weng LX, Xu JL, Tay L, Fang RX, Zhang LH (2004) A bacterial cell-cell communication signal with cross-kingdom structural analogues. *Molecular Microbiology* 51:903–912. <https://doi.org/10.1046/j.1365-2958.2003.03883.x>
- Wang Y, Glukhov E, He Y, Liu Y, Zhou L, Ma X, Hu X, Hong P, Gerwick WH, Zhang Y (2022) Secondary Metabolite Variation and Bioactivities of Two Marine *Aspergillus* Strains in Static Co-Culture Investigated by Molecular Network Analysis and Multiple Database Mining Based on LC-PDA-MS/MS. *Antibiotics* 11. <https://doi.org/10.3390/antibiotics11040513>
- Watanabe T, Izaki K, Takahashi H (1982) New Polyenic Antibiotics Against Gram-positive and -Negative Bacteria. *The Journal of Antibiotics* 35:1141–1147
- Watsuji T, Yamada S, Yamabe T, Watanabe Y, Kato T, Saito T, Ueda K, Beppu T (2007) Identification of indole derivatives as self-growth inhibitors of *Symbiobacterium thermophilum*, a unique bacterium whose growth depends on coculture with a *Bacillus* sp. *Applied and Environmental Microbiology* 73:6159–6165. <https://doi.org/10.1128/AEM.02835-06>
- White N, Boddy L (1992) Extracellular enzyme localization during interspecific fungal interactions. *FEMS Microbiology Letters* 98:75–80. <https://doi.org/10.1111/j.1574-6968.1992.tb05493.x>
- Whitt J, Shipley SM, Newman DJ, Zuck KM (2014) Tetramic acid analogues produced by coculture of *saccharopolyspora erythraea* with *fusarium pallidoroseum*. *Journal of Natural Products* 77:173–177. <https://doi.org/10.1021/np400761g>
- Wu C, Zacchetti B, Ram AFJ, Van Wezel GP, Claessen D, Choi YH (2015) Expanding the chemical space for natural products by *Aspergillus*-*Streptomyces* co-cultivation and biotransformation. *Scientific Reports* 5:1–10. <https://doi.org/10.1038/srep10868>
- Wu Y-M, Yang X-Q, Chen J-X, Wang T, Li T-R, Liao F-R, Liu R-T, Yang Y-B, Ding Z-T (2022) A new butenolide with antifungal activity from solid co-cultivation of *Irpex lacteus* and *Nigrospora oryzae*. *Natural Product Research* 1–5. <https://doi.org/10.1080/14786419.2022.2037589>
- Xu XL, Lee RTH, Fang HM, Wang YM, Li R, Zou H, Zhu Y, Wang Y (2008) Bacterial Peptidoglycan Triggers *Candida albicans* Hyphal Growth by Directly Activating the Adenylyl Cyclase Cyr1p. *Cell Host and Microbe* 4:28–39. <https://doi.org/10.1016/j.chom.2008.05.014>

Yakovleva EP, Bulgakova TN (1978) Formation of Amphotericin B in Mixed Cultures. Pharmaceutical Chemistry Journal 12:1483–1488

Yao L, Zhu LP, Xu XY, Tan LL, Sadilek M, Fan H, Hu B, Shen XT, Yang J, Qiao B, Yang S (2016) Discovery of novel xylosides in co-culture of basidiomycetes *Trametes versicolor* and *Ganoderma applanatum* by integrated metabolomics and bioinformatics. *Scientific Reports* 6:1–13.
<https://doi.org/10.1038/srep33237>

Yu G, Sun P, Aierken R, Sun C, Zhang Z, Che Q, Zhang G, Zhu T, Gu Q, Li M, Li D (2022) Linear polyketides produced by co-culture of *Penicillium crustosum* and *Penicillium fellutanum*. *Marine Life Science & Technology* 4:237–244. <https://doi.org/10.1007/s42995-021-00125-8>

Yu L, Ding W, Wang Q, Ma Z, Xu X, Zhao X, Chen Z (2017) Induction of cryptic bioactive 2,5-diketopiperazines in fungus *Penicillium* sp. DT-F29 by microbial co-culture. *Tetrahedron* 73:907–914. <https://doi.org/10.1016/j.tet.2016.12.077>

Zhang H, Stephanopoulos G (2016) Co-culture engineering for microbial biosynthesis of 3-amino-benzoic acid in *Escherichia coli*. *Biotechnology Journal* 11:981–987. <https://doi.org/10.1002/biot.201600013>

Zhao X, Hou D, Xu J, Wang K, Hu Z (2022) Antagonistic Activity of Fungal Strains against Fusarium Crown Rot. *Plants* 11. <https://doi.org/10.3390/plants11030255>

Zhu F, Chen G, Chen X, Huang M, Wan X (2011) Asperginin, a new antibacterial alkaloid produced by mixed fermentation of two marine-derived mangrove epiphytic fungi. *Chemistry of Natural Compounds* 47:767–769. <https://doi.org/10.1007/s10600-011-0053-8>

Zhu F, Chen GY, Wu JS, Pan JH (2013) Structure revision and cytotoxic activity of marinamide and its methyl ester, novel alkaloids produced by co-cultures of two marine-derived mangrove endophytic fungi. *Natural Product Research* 27:1960–1964. <https://doi.org/10.1080/14786419.2013.800980>

Zhu F, Lin Y, Ding J, Wang X, Huang L (2007) Secondary metabolites of two marine-derived mangrove endophytic fungi (strain nos. 1924# and 3893#) by mixed fermentation. *Chemistry and Industry of Forest Products* 27:8–20

Zhuravleva OI, Belousova EB, Oleinikova GK, Antonov AS, Khudyakova YV, Rasin AB, Popov RS, Menchinskaya ES, Trinh PTH, Yurchenko AN, Yurchenko EA (2022) Cytotoxic Drimane-Type Sesquiterpenes from Co-Culture of the Marine-Derived Fungi *Aspergillus carneus* KMM 4638 and *Beauveria felina* (=*Isaria felina*) KMM 4639. *Marine Drugs* 20:584.
<https://doi.org/10.3390/md20090584>

Zuck KM, Shipley S, Newman DJ (2011) Induced Production of N-Formyl Alkaloids from *Aspergillus fumigatus* by Co-culture with *Streptomyces peucetius*. *Journal of Natural Products* 74:1653–1657. <https://doi.org/10.1021/np200255f>