

Supplementary Material

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1 Supplementary Results

1.1 Description of explanatory variables used in the models other than the four approaches

1.1.1 Boyce index

The SDM models showed variability between taxa in their performance, as indicated by Boyce index (ANOVA, $\chi^2_2 = 11.984$, $P = 0.0074$). Birds had relatively low Boyce index values, while mammals had relatively high values (see Supplementary Figure 5A).

1.1.2 Indices that describe the area of suitable habitat and its changes

Only area of suitable habitat (ASH) showed variability across different taxa (ANOVA, $\chi^2_2 = 23.79$, P < 0.0001), among the constructed GLMM models. ASH of birds (57189.66 ± 49386.30 ; mean \pm standard deviation) and mammals (59052.24 ± 69395.95) was significantly larger than that of amphibians (26223.06 ± 31106.35) and reptiles (40486.53 ± 58155.62) (see Supplementary Figure 5B).

Change of range index (CRI) and loss of range index (LRI) showed the most influence of the different SSP scenarios (Supplementary Table 4A). Specifically, this factor was significant for CRI (ANOVA: $\chi^2_2 = 31.85$, P < 0.0001) and LRI (ANOVA: $\chi^2_2 = 1226.52$, P < 0.0001), but was not for ASH. In the more severe future scenarios, the animals will face more serious threats; for example, comparing SSP5 to others, a larger area will become unsuitable, and the amount of suitable area will shrink (for SSP5: CRI: 0.060 ± 2.247 ; LRI: 0.288 ± 0.253 ; for SSP1: CRI: -0.066 ± 0.607 ; LRI: 0.212 ± 0.199).

All the different models exhibited significant impacts of the different time periods (Supplementary Table 4B) (ANOVA, ASH: $\chi^2_2 = 109.17$, P < 0.0001; CRI: $\chi^2_2 = 19.75$, P = 0.00019; LRI: $\chi^2_2 = 10.3863.91$, P < 0.0001). The further future scenario of 2080-2100 shows significantly more serious threats, that is, less area that is suitable, more area becoming unsuitable, and the overall distribution shrinking (ASH: 41284.38 ± 53644.68 ; CRI: 0.073 ± 2.380 ; LRI: 0.308 ± 0.257), than earlier time periods, for example, 2021-2040 (ASH: 41726.94 ± 50693.01 ; CRI: -0.073 ± 0.366 ; LRI: 0.185 ± 0.180).

2 Supplementary Figures and Tables

2.1 Supplementary Tables

Supplementary Table 1. Variables included in the analyses and their sources, as well as whether they were retained in the analysis after the pairwise correlation analysis (- = removed from the analysis, + = retained).

Variables	Description	Data type	Correlation	Data source
Bioclimatic				
Bio1	Annual Mean Temperature	continuous	-	(1)
Bio2	Mean Diurnal Range (Mean of monthly (max temp - min temp))	continuous	-	(1)
Bio3	Isothermality (Bio2/Bio7) (* 100)	continuous	+	(1)
Bio4	Temperature Seasonality (standard deviation *100)	continuous	+	(1)
Bio5	Max Temperature of Warmest Month	continuous	+	(1)
Bio6	Min Temperature of Coldest Month	continuous	+	(1)
Bio7	Temperature Annual Range (Bio5-Bio6)	continuous	-	(1)
Bio8	Mean Temperature of Wettest Quarter	continuous	-	(1)
Bio9	Mean Temperature of Driest Quarter	continuous	-	(1)
Bio10	Mean Temperature of Warmest Quarter	continuous	-	(1)
Bio11	Mean Temperature of Coldest Quarter	continuous	-	(1)
Bio12	Annual Precipitation	continuous	-	(1)
Bio13	Precipitation of Wettest Month	continuous	-	(1)
Bio14	Precipitation of Driest Month	continuous	+	(1)
Bio15	Precipitation Seasonality (Coefficient of Variation)	continuous	+	(1)
Bio16	Precipitation of Wettest Quarter	continuous	-	(1)
Bio17	Precipitation of Driest Quarter	continuous	-	(1)
Bio18	Precipitation of Warmest Quarter	continuous	+	(1)
Bio19	Precipitation of Coldest Quarter	continuous	-	(1)
Geographic				
Aspect	Aspect eastness	continuous	+	(2)
Slope	Slope	continuous	+	(2)
Distance from water	Euclidean distance from above ground water (m)	continuous	+	(3)
Anthropogenic				
Human population	Human population density	continuous	+	(4)
Land-use	Land-use based on the downscaling of GCAM	categorical	+	(5)

Data sources: (1) WorldClim, <http://www.worldclim.org>; (2) the United States Geological Survey (<https://www.usgs.gov/>, <http://www.earthenv.org/topography>); (3) OpenStreetMap Data Extracts (<https://download.geofabrik.de/>); (4)(M. Chen et al., 2020); (5)(Gao, 2017) (<https://data.pnnl.gov/dataset/13192>).

Supplementary Table 2. Retained and removed climatic variables. Variables in orange text were not highly correlated (Pearson $|r| < 0.7$) with any other variable. Variables in red text were highly correlated, but were retained, because they have been often selected in previous studies with these taxa (as shown in the rest of the table). Crossed out variables represent those that were removed.

Climatic variables	Reason for retention	amphibians	birds	Reference Article		
				mammals	reptiles	other
Bio1						
Bio2						
Bio3	No correlation	(Mokhatla, M.M. et al., 2015)	(Distler, T. et al., 2015; Li, X. et al., 2015; Magory Cohen, T. and Dor, R., 2019)	(Ribeiro, B.R. et al., 2018)		
Bio4	Use in previous literature	(Lemes, P. et al., 2014; Munguía, M. et al., 2012; Peñalver-Alcázar, M. et al., 2021; Sangermano, F. et al., 2015)	(Ko, C.Y. et al., 2016; Li, X. et al., 2015; Magory Cohen, T. and Dor, R., 2019)	(Bogoni, J.A. and Tagliari, M.M., 2021; Deb, J.C. et al., 2020; Guevara, L. et al., 2018; Leach, K. et al., 2017; Morueta-Holme, N. et al., 2010; Stevens, R.D., 2013)	(Barnagaud, J.Y. et al., 2021; Clusella-trullas, A.S. et al., 2011; Ribeiro, B.R. et al., 2018; Sheldon, K.S. et al., 2015)	(Mouchet, M. et al., 2015)
Bio5	Use in previous literature	(Biber, M.F. et al., 2020; Gherghel, I. et al., 2020; Groff, L.A. et al., 2014; Munguía, M. et al., 2012)	(Cohen, J.M. et al., 2020; Distler, T. et al., 2015; Huang, Q. et al., 2017; Li, X. et al., 2015)	(Guevara, L. et al., 2018; Leach, K. et al., 2017; Morán-Ordóñez, A. et al., 2017; Ribeiro, B.R. et al., 2018)	(Console, G. et al., 2020; Farashi, A. and Alizadeh-Noughani, M., 2021; Li, X. et al., 2016; Muñoz, A. et al., 2016)	(Germain, S.J. and Lutz, J.A., 2020; Li, Y. et al., 2016; Zhang, Z. et al., 2021)
Bio6	Use in previous literature	(Gherghel, I. et al., 2020; Groff, L.A. et al., 2014; Li, X. et al., 2016; Munguía, M. et al., 2012; Muñoz, A. et al., 2016; Vasconcelos, T.S. et al., 2012)	(Cohen, J.M. et al., 2020; Distler, T. et al., 2015; Huang, Q. et al., 2017; Li, X. et al., 2015)	(Guevara, L. et al., 2018; Jurestovsky, D. and Andrew Joyner, T., 2018; Morán-Ordóñez, A. et al., 2017)	(Console, G. et al., 2020; Ihlow, F. et al., 2012; Li, X. et al., 2016; Muñoz, A. et al., 2016)	(Germain, S.J. and Lutz, J.A., 2020; Li, Y. et al., 2016; Zhang, Z. et al., 2021)
Bio7						
Bio8		(Mokhatla, M.M. et al., 2015)	(Salas, E.A.L.. et al., 2017)			
Bio9		(Mokhatla, M.M. et al., 2015)	(Salas, E.A.L.. et al., 2017)			
Bio10						
Bio11						
Bio12			(Schooler, S.L. et al., 2020)			
Bio13				(Ribeiro, B.R. et al., 2018) (Deb, J.C. et al., 2019)	(Farashi, A. and Alizadeh-Noughani, M., 2021)	

Bio14	Use in previous literature	(Gherghel, I. et al., 2020; Sangermano, F. et al., 2015; Vol, H., 2021)	(Distler, T. et al., 2015; Dreitz, V.J. et al., 2012; Li, X. et al., 2015; Magory Cohen, T. and Dor, R., 2019; Salas, E.A.L.. et al., 2017; Schooler, S.L. et al., 2020)	(Guevara, L. et al., 2018; Hill, S.E. and Winder, I.C., 2019; Morueta-Holme, N. et al., 2010; Ribeiro, B.R. et al., 2018)	(Console, G. et al., 2020)	(Xu, Z., 2015)
Bio15	No correlation	(Barrett, K. et al., 2014; Gherghel, I. et al., 2020; Goudarzi, F. et al., 2021; Lemes, P. et al., 2014; Mokhatla, M.M. et al., 2015; Peñalver-Alcázar, M. et al., 2021; Vasconcelos, T.S. et al., 2012)	(Biber, M.F. et al., 2020; Ko, C.Y. et al., 2016; Li, X. et al., 2015; Salas, E.A.L.. et al., 2017)	(Biber, M.F. et al., 2020; Deb, J.C. et al., 2020; Dormann, C.F. et al., 2008; Lamelas-López, L. et al., 2020; Leach, K. et al., 2017; Morueta-Holme, N. et al., 2010)	(Barnagaud, J.Y. et al., 2021; Ihlow, F. et al., 2012; Li, X. et al., 2016; Rodrigues, J.F.M. and Lima-Ribeiro, M.S., 2018)	
Bio16					(Farashi, A. and Alizadeh-Noughani, M., 2021)	
Bio17					(Farashi, A. and Alizadeh-Noughani, M., 2021)	
Bio18	Not correlated with the remaining variables	(Araújo, M.B. et al., 2006; Barrett, K. et al., 2014; Biber, M.F. et al., 2020; Gherghel, I. et al., 2020; Vasconcelos, T.S. et al., 2012)	(Matthews, S.N. et al., 2011)(Distler, T. et al., 2015)	(Dormann, C.F. et al., 2008; Maiorano, L. et al., 2011; Mestre, F. et al., 2017; Morueta-Holme, N. et al., 2010; Ribeiro, B.R. et al., 2018)	(Araújo, M.B. et al., 2006; Farashi, A. and Alizadeh-Noughani, M., 2021; Ihlow, F. et al., 2012; Thompson, D.M. et al., 2017)	
Bio19					(Farashi, A. and Alizadeh-Noughani, M., 2021)	

Supplementary Table 3. Studied species and SDM performance, as measured by the Boyce index, in all four approaches, and models' maxTSS threshold values. Species are ordered by taxa and then scientific name.

Species information			Boyce index		Approach 4	
Latin name	Family	IUCN	Approach 1	Approach 2&3	Approach 4	threshold
Amphibians						
<i>Amolops aniqiaoensis</i>	Ranidae	LC	0.915	0.972	0.925	0.170
<i>Amolops bellulus</i>	Ranidae	VU	0.636	0.715	1	1.000
<i>Amolops daiyunensis</i>	Ranidae	VU	0.931	0.936	0.959	0.436
<i>Amolops granulosus</i>	Ranidae	NT	0.997	0.993	0.997	0.133
<i>Amolops hainanensis</i>	Ranidae	EN	0.947	0.986	0.976	0.181
<i>Amolops hongkongensis</i>	Ranidae	EN	0.889	0.919	0.901	0.123
<i>Amolops lifanensis</i>	Ranidae	LC	0.99	0.999	0.957	0.249
<i>Amolops loloensis</i>	Ranidae	VU	0.991	0.994	0.995	0.156
<i>Amolops torrentis</i>	Ranidae	LC	0.743	0.91	0.956	0.298
<i>Amolops wuyiensis</i>	Ranidae	LC	0.991	0.995	0.993	0.082
<i>Andrias davidianus</i>	Cryptobranchidae	CR	0.994	0.996	0.997	0.173
<i>Aquixalus idiootocus</i>	Rhacophoridae	LC	0.942	0.907	0.895	0.163
<i>Bamburana exiliversabilis</i>	Ranidae	NT	0.884	0.908	0.949	0.304
<i>Bamburana nasuta</i>	Ranidae	VU	0.994	0.994	0.995	0.332
<i>Bamburana versabilis</i>	Ranidae	NT	0.88	0.88	0.908	0.432
<i>Batrachuperus karlschmidti</i>	Hynobiidae	VU	0.953	0.921	0.702	0.427
<i>Batrachuperus londongensis</i>	Hynobiidae	VU	0.923	0.923	0.899	0.064
<i>Batrachuperus pinchonii</i>	Hynobiidae	VU	0.952	0.952	0.965	0.190
<i>Batrachuperus tibetanus</i>	Hynobiidae	VU	0.993	0.993	0.989	0.075
<i>Batrachuperus yenyuanensis</i>	Hynobiidae	VU	0.997	0.997	0.806	0.264
<i>Bombina fortinuptialis</i>	Bombinatoridae	VU	0.993	0.993	0.964	0.111
<i>Bombina maxima</i>	Bombinatoridae	LC	0.967	0.967	0.962	0.299
<i>Brachytarsophrys chuannanensis</i>	Megophryidae	NT	0.921	0.921	0.757	0.717
<i>Buergeria robusta</i>	Rhacophoridae	LC	0.95	0.95	0.952	0.244
<i>Bufo bankorensis</i>	Bufonidae	LC	0.915	0.915	0.873	0.199
<i>Bufo tibetanus</i>	Bufonidae	LC	0.886	0.84	0.877	0.107
<i>Bufo tuberculatus</i>	Bufonidae	NT	0.998	0.998	0.995	0.206
<i>Cynops chenggongensis</i>	Salamandridae	CR	0.865	0.769	0.922	0.067
<i>Cynops cyanurus</i>	Salamandridae	NT	0.995	0.995	0.988	0.316
<i>Cynops orientalis</i>	Salamandridae	NT	0.868	0.868	0.997	0.238
<i>Dianrana pleuraden</i>	Ranidae	LC	0.965	0.965	0.994	0.357
<i>Duttaphrynus cyphosus</i>	Bufonidae	LC	1	1	1	0.457
<i>Feirana quadranus</i>	Dic平glossidae	NT	0.991	0.991	0.996	0.373
<i>Feirana taihangnica</i>	Dic平glossidae	VU	0.942	0.942	0.938	0.316
<i>Glandirana minima</i>	Ranidae	CR	0.975	0.975	0.985	0.299
<i>Gynandropaa liui</i>	Dic平glossidae	VU	0.874	0.874	0.939	0.114
<i>Huangixalus translineatus</i>	Rhacophoridae	VU	0.969	0.969	0.968	0.444
<i>Hyla sanchiangensis</i>	Hylidae	LC	0.978	0.978	0.994	0.274
<i>Hyla tsinlingensis</i>	Hylidae	LC	0.872	0.872	0.913	0.064
<i>Hynobius arisanensis</i>	Hynobiidae	EN	0.992	0.999	0.99	0.218

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<i>Hynobius yiwuensis</i>	Hynobiidae	VU	0.751	0.875	0.851	0.331
<i>Kaloula nonggangensis</i>	Microhylidae	DD	0.422	0.858	0.991	0.905
<i>Kaloula rugifera</i>	Microhylidae	LC	0.99	0.976	0.988	0.226
<i>Kaloula verrucosa</i>	Microhylidae	LC	0.464	0.951	0.874	0.510
<i>Leptobrachium hainanense</i>	Megophryidae	VU	0.99	0.992	0.969	0.245
<i>Leptobrachium huashen</i>	Megophryidae	NT	0.802	0.901	0.922	0.429
<i>Liangshantriton taliangensis</i>	Salamandridae	VU	0.98	0.959	0.949	0.279
<i>Limnonectes fragilis</i>	Dic平glossidae	EN	0.877	0.941	0.969	0.293
<i>Limnonectes fujianensis</i>	Dic平glossidae	NT	0.971	0.95	0.957	0.219
<i>Liua shihi</i>	Hynobiidae	NT	0.976	0.98	0.99	0.154
<i>Liuhurstana shuchinae</i>	Ranidae	NT	0.988	0.982	0.938	0.232
<i>Liuixalus hainanus</i>	Rhacophoridae	DD	0.993	0.989	0.995	0.272
<i>Liuixalus kempii</i>	Rhacophoridae	DD	0.784	0.923	0.948	0.260
<i>Liuixalus romeri</i>	Rhacophoridae	VU	0.858	0.923	0.873	0.485
<i>Liurana xizangensis</i>	Occidozygidae	DD	0.978	0.996	0.997	0.133
<i>Maculopaa maculosa</i>	Dic平glossidae	EN	0.976	0.96	0.992	0.318
<i>Megophrys binchuanensis</i>	Megophryidae	VU	0.941	0.941	0.959	0.308
<i>Megophrys boettgeri</i>	Megophryidae	LC	0.897	0.97	0.999	0.203
<i>Megophrys daweimontis</i>	Megophryidae	VU	0.97	0.987	0.993	0.296
<i>Megophrys gigantica</i>	Megophryidae	VU	0.877	0.971	0.975	0.415
<i>Megophrys huangshanensis</i>	Megophryidae	VU	0.996	0.991	0.917	0.203
<i>Megophrys jingdongensis</i>	Megophryidae	NT	0.939	0.999	0.984	0.224
<i>Megophrys jinggangensis</i>	Megophryidae	DD	0.951	0.913	0.961	0.221
<i>Megophrys lini</i>	Megophryidae	DD	0.995	0.988	0.928	0.388
<i>Megophrys mangshanensis</i>	Megophryidae	NT	0.768	0.858	0.848	0.456
<i>Megophrys medogensis</i>	Megophryidae	EN	0.957	0.921	0.917	0.178
<i>Megophrys omeimontis</i>	Megophryidae	VU	0.998	0.995	0.997	0.297
<i>Megophrys shapingensis</i>	Megophryidae	LC	0.98	0.996	0.959	0.103
<i>Megophrys spinata</i>	Megophryidae	LC	0.999	0.996	0.995	0.195
<i>Microhyla mixtura</i>	Microhylidae	LC	0.983	0.994	0.9	0.269
<i>Micryletta steinegeri</i>	Microhylidae	LC	0.971	0.973	0.987	0.041
<i>Nanorana pleskei</i>	Dic平glossidae	LC	0.999	0.954	0.997	0.267
<i>Nanorana ventripunctata</i>	Dic平glossidae	LC	0.992	0.981	0.816	0.235
<i>Nidirana daunchina</i>	Ranidae	LC	0.989	0.978	0.963	0.269
<i>Odorrana anlungensis</i>	Ranidae	VU	0.885	0.911	1	0.280
<i>Odorrana hainanensis</i>	Ranidae	VU	0.906	0.954	0.952	0.400
<i>Odorrana hejiangensis</i>	Ranidae	VU	0.999	0.999	0.998	0.286
<i>Odorrana huanggangensis</i>	Ranidae	LC	0.966	0.529	0.94	0.639
<i>Odorrana jingdongensis</i>	Ranidae	VU	0.856	0.914	0.99	0.208
<i>Odorrana junlianensis</i>	Ranidae	NT	0.997	0.919	0.923	0.217
<i>Odorrana lungshengensis</i>	Ranidae	NT	0.904	0.994	0.955	0.200
<i>Odorrana nanjiangensis</i>	Ranidae	NT	0.999	0.995	0.995	0.399
<i>Odorrana swinhoana</i>	Ranidae	LC	0.851	0.844	0.857	0.236
<i>Odorrana tiannanensis</i>	Ranidae	VU	0.978	0.975	0.967	0.180
<i>Odorrana tormota</i>	Ranidae	VU	0.96	0.974	0.972	0.408
<i>Odorrana yizhangensis</i>	Ranidae	VU	0.921	0.851	0.923	0.078
<i>Onychodactylus zhangyapingi</i>	Hynobiidae	VU	0.902	0.966	0.999	0.231

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<i>Oreolalax granulosus</i>	Megophryidae	VU	0.982	0.974	0.992	0.296
<i>Oreolalax jingdongensis</i>	Megophryidae	VU	0.943	0.984	0.991	0.294
<i>Oreolalax lichuanensis</i>	Megophryidae	NT	0.973	0.997	0.77	0.081
<i>Oreolalax major</i>	Megophryidae	VU	0.994	0.989	0.798	0.408
<i>Oreolalax pingii</i>	Megophryidae	VU	0.99	0.975	0.99	0.321
<i>Oreolalax popei</i>	Megophryidae	VU	1	1	0.994	0.297
<i>Oreolalax puxiongensis</i>	Megophryidae	EN	0.742	0.849	0.997	0.869
<i>Oreolalax rhodostigmatus</i>	Megophryidae	VU	0.995	0.994	0.967	0.196
<i>Oreolalax rugosus</i>	Megophryidae	NT	0.998	0.997	0.822	0.291
<i>Oreolalax schmidti</i>	Megophryidae	NT	0.99	0.999	1	0.240
<i>Oreolalax xiangchengensis</i>	Megophryidae	LC	0.83	0.86	0.788	0.346
<i>Pachytriton archospotus</i>	Salamandridae	LC	0.998	0.932	0.967	0.322
<i>Pachytriton brevipes</i>	Salamandridae	LC	0.902	1	0.997	0.270
<i>Pachytriton changi</i>	Salamandridae	DD	0.901	0.923	0.899	0.747
<i>Pachytriton granulosus</i>	Salamandridae	DD	0.995	0.996	0.981	0.147
<i>Pachytriton inexpectatus</i>	Salamandridae	VU	0.942	0.996	0.907	0.427
<i>Pachytriton moi</i>	Salamandridae	DD	0.963	0.982	0.982	0.265
<i>Paramegophrys alpinus</i>	Megophryidae	EN	0.963	0.958	0.833	0.241
<i>Paramegophrys liui</i>	Megophryidae	LC	0.828	0.857	0.833	0.415
<i>Paramegophrys oshanensis</i>	Megophryidae	LC	1	0.999	0.956	0.291
<i>Paramesotriton caudopunctatus</i>	Salamandridae	VU	0.993	0.954	0.96	0.398
<i>Paramesotriton chinensis</i>	Salamandridae	NT	0.984	0.984	0.998	0.142
<i>Paramesotriton fuzhongensis</i>	Salamandridae	VU	0.984	0.99	0.994	0.270
<i>Paramesotriton hongkongensis</i>	Salamandridae	NT	0.941	0.992	0.971	0.327
<i>Paramesotriton maolanensis</i>	Salamandridae	DD	-0.093	0.786	0.909	0.577
<i>Paramesotriton yunwuensis</i>	Salamandridae	EN	0.967	0.948	0.762	0.290
<i>Paramesotriton zhijinensis</i>	Salamandridae	EN	0.975	0.723	0.863	0.478
<i>Parapelophryne scalpta</i>	Bufoidae	VU	0.786	0.992	0.976	0.221
<i>Pelophylax fukienensis</i>	Ranidae	NT	0.972	0.961	0.992	0.243
<i>Pelophylax hubeiensis</i>	Ranidae	LC	0.985	0.997	0.984	0.180
<i>Pelophylax plancyi</i>	Ranidae	LC	0.909	0.995	0.903	0.294
<i>Pseudoamolops sauteri</i>	Ranidae	VU	0.998	0.999	0.972	0.186
<i>Pseudohynobius flavomaculatus</i>	Hynobiidae	VU	0.579	0.881	0.821	0.945
<i>Pseudorana sangzhiensis</i>	Ranidae	LC	0.967	0.97	0.955	0.091
<i>Pseudorana weiningensis</i>	Ranidae	NT	0.998	1	0.971	0.279
<i>Qiongbufo ledongensis</i>	Bufoidae	EN	0.952	0.846	0.74	0.059
<i>Quasipaa exilispinosa</i>	Dicroidiidae	VU	0.614	0.792	0.898	0.171
<i>Quasipaa jiulongensis</i>	Dicroidiidae	VU	0.999	0.994	0.998	0.131
<i>Quasipaa shini</i>	Dicroidiidae	VU	1	0.996	0.99	0.311
<i>Rana chaochiaoensis</i>	Ranidae	LC	0.653	0.984	0.998	0.225
<i>Rana chensinensis</i>	Ranidae	LC	0.994	0.992	0.999	0.421
<i>Rana kukunoris</i>	Ranidae	LC	0.993	0.993	0.944	0.153
<i>Rana longicrus</i>	Ranidae	LC	0.933	0.969	0.954	0.190
<i>Rana omeimontis</i>	Ranidae	LC	0.99	0.999	1	0.234
<i>Rana zhenhaiensis</i>	Ranidae	LC	0.996	0.985	0.998	0.222
<i>Rhacophorus arvalis</i>	Rhacophoridae	VU	0.939	0.907	0.962	0.150

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<i>Rhacophorus aurantiventralis</i>	Rhacophoridae	VU	0.934	0.868	0.943	0.204
<i>Rhacophorus chenfui</i>	Rhacophoridae	LC	0.999	0.992	0.999	0.277
<i>Rhacophorus dennysi</i>	Rhacophoridae	LC	0.828	0.836	0.811	0.386
<i>Rhacophorus hungfuensis</i>	Rhacophoridae	EN	0.997	0.981	0.993	0.238
<i>Rhacophorus leucofasciatus</i>	Rhacophoridae	VU	0.989	0.965	0.995	0.341
<i>Rhacophorus minimus</i>	Rhacophoridae	NT	0.707	1	0.855	0.634
<i>Rhacophorus nigropunctatus</i>	Rhacophoridae	NT	0.99	0.999	0.996	0.372
<i>Rhacophorus prasinatus</i>	Rhacophoridae	NT	0.957	0.955	0.706	0.252
<i>Rhacophorus rhodopus</i>	Rhacophoridae	LC	0.939	0.96	0.999	0.395
<i>Rhacophorus taipeianus</i>	Rhacophoridae	NT	0.9	0.973	0.922	0.162
<i>Rhacophorus verrucopus</i>	Rhacophoridae	DD	0.912	0.989	0.927	0.384
<i>Rhacophorus yaoshanensis</i>	Rhacophoridae	EN	0.984	0.994	0.997	0.205
<i>Rugosa tientaiensis</i>	Ranidae	NT	0.962	0.977	0.981	0.279
<i>Scutiger glandulatus</i>	Megophryidae	LC	0.99	0.983	0.991	0.116
<i>Scutiger jiulongensis</i>	Megophryidae	VU	0.982	0.989	0.959	0.186
<i>Scutiger maculatus</i>	Megophryidae	CR	0.994	0.989	0.975	0.169
<i>Scutiger mammatus</i>	Megophryidae	LC	0.929	0.894	0.949	0.288
<i>Scutiger muliensis</i>	Megophryidae	EN	0.956	0.949	0.965	0.145
<i>Scutiger ningshanensis</i>	Megophryidae	EN	0.996	1	0.999	0.424
<i>Scutiger pingwuensis</i>	Megophryidae	EN	0.99	0.984	0.978	0.160
<i>Scutiger tuberculatus</i>	Megophryidae	VU	0.999	0.997	0.989	0.320
<i>Sylvirana bannanica</i>	Ranidae	VU	0.972	0.962	0.983	0.098
<i>Sylvirana hekouensis</i>	Ranidae	VU	0.956	0.987	0.983	0.501
<i>Sylvirana latouchii</i>	Ranidae	LC	0.822	0.79	0.829	0.477
<i>Sylvirana menglaensis</i>	Ranidae	LC	0.932	0.988	0.991	0.476
<i>Sylvirana nigrotympanica</i>	Ranidae	DD	0.944	0.982	0.98	0.512
<i>Sylvirana spinulosa</i>	Ranidae	NT	0.95	0.958	0.982	0.293
<i>Taylorana liui</i>	Occidozygidae	VU	0.459	0.898	0.857	0.471
<i>Theloderma moloch</i>	Rhacophoridae	DD	0.987	0.993	0.986	0.157
<i>Torrentophryne ailaoanus</i>	Bufonidae	LC	0.845	0.949	0.945	0.314
<i>Torrentophryne aspinia</i>	Bufonidae	VU	0.693	0.979	0.98	0.632
<i>Tylototriton kweichowensis</i>	Salamandridae	VU	0.991	0.996	0.973	0.169
<i>Tylototriton pseudoverrucosus</i>	Salamandridae	NT	0.848	0.863	0.754	0.349
<i>Tylototriton yangi</i>	Salamandridae	NT	0.998	0.999	0.997	0.234
<i>Unculuana unculuanus</i>	Dicroidlossidae	EN	0.941	0.997	0.995	0.263
<i>Vibrissaphora boringii</i>	Megophryidae	EN	0.998	0.998	0.999	0.251
<i>Vibrissaphora leishanensis</i>	Megophryidae	VU	0.957	0.903	0.948	0.068
<i>Vibrissaphora liui</i>	Megophryidae	NT	0.877	0.922	0.891	0.394
<i>Yaotriton hainanensis</i>	Salamandridae	EN	0.8	0.765	0.757	0.371
<i>Yaotriton liuyangensis</i>	Salamandridae	DD	0.999	0.96	0.98	0.313
<i>Yaotriton wenxianensis</i>	Salamandridae	VU	0.742	0.964	0.971	0.255
<i>Yerana yei</i>	Dicroidlossidae	VU	1	1	1	0.402
Birds						
<i>Actinodura morrisoniana</i>	Timaliidae	LC	0.96	0.863	0.863	0.071
<i>Aegithalos fuliginosus</i>	Aegithalidae	LC	0.987	0.98	0.98	0.139
<i>Alcippe striaticollis</i>	Timaliidae	LC	0.943	0.925	0.925	0.205

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<i>Alcippe variegaticeps</i>	Timaliidae	VU	0.843	0.882	0.882	0.298
<i>Alectoris magna</i>	Phasianidae	NT	0.967	0.967	0.967	0.223
<i>Arborophila ardens</i>	Phasianidae	EN	0.942	0.939	0.939	0.320
<i>Arborophila crudigularis</i>	Phasianidae	NT	0.937	0.87	0.87	0.175
<i>Arborophila gingica</i>	Phasianidae	VU	0.983	0.953	0.953	0.137
<i>Arborophila rufipectus</i>	Phasianidae	EN	0.957	0.915	0.915	0.028
<i>Babax koslowi</i>	Timaliidae	NT	0.532	0.747	0.747	0.389
<i>Bambusicola thoracicus</i>	Phasianidae	LC	1	0.999	0.999	0.319
<i>Certhia tianquanensis</i>	Certhiidae	VU	0.905	0.939	0.939	0.029
<i>Chrysolophus pictus</i>	Phasianidae	NT	0.999	0.999	0.999	0.209
<i>Crossoptilon auritum</i>	Phasianidae	NT	0.99	0.99	0.99	0.121
<i>Crossoptilon crossoptilon</i>	Phasianidae	NT	0.984	0.99	0.99	0.178
<i>Crossoptilon harmani</i>	Phasianidae	NT	0.939	0.99	0.99	0.141
<i>Crossoptilon mantchuricum</i>	Phasianidae	VU	0.976	0.994	0.994	0.257
<i>Emberiza koslowi</i>	Emberizidae	VU	0.977	0.837	0.837	0.229
<i>Garrulax courtoisi</i>	Timaliidae	CR	0.926	0.94	0.94	0.490
<i>Garrulax davidi</i>	Timaliidae	LC	0.995	0.997	0.997	0.377
<i>Garrulax elliotii</i>	Timaliidae	LC	0.998	0.997	0.997	0.333
<i>Garrulax henrici</i>	Timaliidae	LC	0.901	0.81	0.81	0.076
<i>Garrulax lunulatus</i>	Timaliidae	LC	0.85	0.917	0.917	0.165
<i>Garrulax maximus</i>	Timaliidae	LC	0.985	0.98	0.98	0.234
<i>Garrulax morrisonianus</i>	Timaliidae	LC	-0.292	0.597	0.597	0.312
<i>Garrulax poecilorhynchus</i>	Timaliidae	LC	0.998	1	1	0.131
<i>Garrulax sukatschewi</i>	Timaliidae	VU	0.815	0.633	0.633	0.397
<i>Garrulax taewanus</i>	Timaliidae	NT	0.93	0.989	0.989	0.246
<i>Heterophasia auricularis</i>	Timaliidae	LC	0.864	0.87	0.87	0.186
<i>Kozlowia roborowskii</i>	Fringillidae	VU	0.297	0.324	0.324	0.611
<i>Latoucheornis siemsseni</i>	Emberizidae	LC	0.973	0.935	0.935	0.068
<i>Leptopoecile elegans</i>	Sylviidae	NT	0.963	0.958	0.958	0.188
<i>Liocichla omeiensis</i>	Timaliidae	VU	0.892	0.776	0.776	0.302
<i>Liocichla steerii</i>	Timaliidae	LC	0.874	0.863	0.863	0.114
<i>Lophophorus lhuysii</i>	Phasianidae	EN	0.977	0.991	0.991	0.139
<i>Lophura swinhoii</i>	Phasianidae	NT	0.961	0.949	0.949	0.176
<i>Megalima nuchalis</i>	Capitonidae	LC	0.884	0.823	0.823	0.076
<i>Montifringilla henrici</i>	Passeridae	NT	0.885	0.862	0.862	0.147
<i>Moupinia poecilotis</i>	Timaliidae	LC	0.75	0.803	0.803	0.274
<i>Myophonus insularis</i>	Turdidae	LC	0.903	0.941	0.941	0.258
<i>Paradoxornis conspicillatus</i>	Paradoxornithidae	NT	0.952	0.949	0.949	0.185
<i>Paradoxornis paradoxus</i>	Paradoxornithidae	NT	0.959	0.837	0.837	0.365
<i>Parus davidi</i>	Paridae	LC	0.851	0.917	0.917	0.023
<i>Parus holsti</i>	Paridae	LC	0.824	0.253	0.253	0.402
<i>Parus superciliosus</i>	Paridae	NT	0.907	0.938	0.938	0.207
<i>Parus venustulus</i>	Paridae	LC	0.972	0.993	0.993	0.219
<i>Perisoreus internigrans</i>	Corvidae	VU	0.927	0.925	0.925	0.278
<i>Phoenicurus alaschanicus</i>	Turdidae	EN	0.976	0.885	0.885	0.357
<i>Phylloscopus emeiensis</i>	Sylviidae	LC	0.918	0.921	0.921	0.189

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<i>Phylloscopus hainanus</i>	Sylviidae	VU	0.804	0.76	0.76	0.342
<i>Phylloscopus kansuensis</i>	Sylviidae	LC	0.97	0.986	0.986	0.231
<i>Podoces biddulphi</i>	Corvidae	VU	0.947	0.933	0.933	0.060
<i>Polyplectron katsumatae</i>	Phasianidae	CR	0.798	0.925	0.925	0.590
<i>Pseudopodoces humilis</i>	Paridae	LC	0.986	0.993	0.993	0.210
<i>Pycnonotus taivanus</i>	Pycnonotidae	VU	0.948	0.937	0.937	0.170
<i>Regulus goodfellowi</i>	Regulidae	LC	0.975	0.949	0.949	0.130
<i>Sitta yunnanensis</i>	Sittidae	VU	0.862	0.985	0.985	0.131
<i>Stachyris nonggangensis</i>	Timaliidae	EN	0.877	0.922	0.922	0.122
<i>Strix davidi</i>	Strigidae	VU	0.811	0.827	0.827	0.211
<i>Syrmaticus ellioti</i>	Phasianidae	VU	0.956	0.987	0.987	0.263
<i>Syrmaticus mikado</i>	Phasianidae	NT	0.989	0.983	0.983	0.175
<i>Syrmaticus reevesii</i>	Phasianidae	EN	0.95	0.988	0.988	0.178
<i>Tarsiger johnstoniae</i>	Turdidae	LC	0.916	0.905	0.905	0.049
<i>Tetraophasis obscurus</i>	Phasianidae	VU	0.974	0.986	0.986	0.166
<i>Tetraophasis szechenyii</i>	Phasianidae	VU	0.956	0.868	0.868	0.138
<i>Tetrastes sewerzowi</i>	Tetraonidae	NT	0.967	0.972	0.972	0.293
<i>Tragopan caboti</i>	Phasianidae	EN	0.985	0.973	0.973	0.168
<i>Turdus mupinensis</i>	Turdidae	LC	0.975	0.985	0.985	0.223
<i>Urocissa caerulea</i>	Corvidae	LC	0.984	0.99	0.99	0.189
<i>Urocynchramus pylzowi</i>	Emberizidae	NT	0.956	0.906	0.906	0.201
<i>Yuhina brunneiceps</i>	Timaliidae	LC	0.895	0.877	0.877	0.164

Mammals

<i>Aeretes melanopterus</i>	Sciuridae	NT	0.999	0.999	0.988	0.239
<i>Ailuropoda melanoleuca</i>	Ailuropodidae	VU	0.998	0.999	0.995	0.266
<i>Anourosorex yamashinai</i>	Soricidae	DD	0.95	0.984	0.901	0.243
<i>Apodemus chevrieri</i>	Muridae	LC	0.918	0.86	0.993	0.277
<i>Apodemus semotus</i>	Muridae	LC	0.913	0.937	0.47	0.461
<i>Arielulus torquatus</i>	Vespertilionidae	VU	0.945	0.993	0.928	0.320
<i>Barbastella beijingensis</i>	Vespertilionidae	DD	0.941	0.842	0.707	0.074
<i>Blarinella quadraticauda</i>	Soricidae	LC	0.994	0.997	0.997	0.198
<i>Bos mutus</i>	Bovidae	VU	0.999	0.989	0.998	0.157
<i>Brachionus przewalskii</i>	Muridae	LC	0.943	0.996	0.991	0.399
<i>Cansumys canus</i>	Cricetidae	LC	0.965	0.959	0.979	0.118
<i>Capricornis swinhoei</i>	Bovidae	LC	0.951	0.972	0.972	0.169
<i>Caryomys eva</i>	Cricetidae	LC	1	1	0.998	0.292
<i>Caryomys inez</i>	Cricetidae	LC	1	0.979	0.994	0.307
<i>Chaetocauda sichuanensis</i>	Gliridae	EN	0.988	0.9	0.954	0.204
<i>Chodsigoa hypsibia</i>	Soricidae	LC	0.884	0.9	0.986	0.264
<i>Chodsigoa lamula</i>	Soricidae	NT	0.977	0.96	0.964	0.283
<i>Chodsigoa smithii</i>	Soricidae	LC	0.995	0.999	0.995	0.151
<i>Cricetulus kamensis</i>	Cricetidae	NT	0.95	0.951	0.866	0.131
<i>Crocidura rapax</i>	Soricidae	LC	0.91	0.953	0.893	0.137
<i>Crocidura tanakae</i>	Soricidae	LC	0.966	0.977	0.943	0.065
<i>Elaphurus davidianus</i>	Cervidae	CR	0.802	0.76	-0.445	0.433
<i>Eospalax fontanieri</i>	Spalacidae	LC	0.641	0.543	0.959	0.410

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<i>Eospalax rothschildi</i>	Spalacidae	LC	1	0.992	0.984	0.240
<i>Eospalax smithii</i>	Spalacidae	NT	0.984	0.964	0.996	0.178
<i>Eothenomys chinensis</i>	Cricetidae	NT	0.993	0.991	0.998	0.115
<i>Eothenomys custos</i>	Cricetidae	LC	0.951	0.974	0.988	0.133
<i>Eothenomys miletus</i>	Cricetidae	LC	1	1	0.97	0.083
<i>Eothenomys olitor</i>	Cricetidae	NT	0.888	0.988	0.973	0.101
<i>Eothenomys proditor</i>	Cricetidae	NT	0.981	0.98	0.472	0.231
<i>Eothenomys wardi</i>	Cricetidae	NT	0.956	0.985	0.987	0.240
<i>Eozapus setchuanus</i>	Dipodidae	LC	0.782	0.966	0.999	0.289
<i>Episoriculus fumidus</i>	Soricidae	NT	0.928	0.935	0.945	0.253
<i>Euroscaptor grandis</i>	Talpidae	VU	0.88	0.985	0.979	0.324
<i>Felis bieti</i>	Felidae	CR	0.975	0.986	0.975	0.170
<i>Lepus yarkandensis</i>	Leporidae	NT	0.939	0.996	0.997	0.260
<i>Lipotes vexillifer</i>	Lipotidae	CR	0.995	0.996	0.974	0.282
<i>Macaca thibetana</i>	Cercopithecidae	VU	0.76	0.97	0.994	0.374
<i>Mesechinus hughii</i>	Erinaceidae	NT	0.986	0.971	0.872	0.266
<i>Microtus kikuchii</i>	Cricetidae	NT	0.915	0.923	0.648	0.579
<i>Mogera kanoana</i>	Talpidae	DD	0.811	0.656	0.844	0.180
<i>Moschus anhuiensis</i>	Moschidae	CR	0.988	0.994	0.989	0.430
<i>Moschus fuscus</i>	Moschidae	CR	0.992	0.962	0.998	0.192
<i>Muntiacus crinifrons</i>	Cervidae	EN	0.922	0.982	0.963	0.164
<i>Muntiacus gongshanensis</i>	Cervidae	CR	0.97	0.966	0.963	0.188
<i>Muntiacus reevesi</i>	Cervidae	VU	0.918	0.993	0.994	0.302
<i>Murina bicolor</i>	Vespertilionidae	DD	0.895	0.87	0.924	0.176
<i>Murina chrysochaetes</i>	Vespertilionidae	DD	0.98	0.964	0.904	0.139
<i>Murina gracilis</i>	Vespertilionidae	DD	0.969	0.818	0.945	0.163
<i>Murina lorelieae</i>	Vespertilionidae	DD	0.879	0.898	0.92	0.546
<i>Murina puta</i>	Vespertilionidae	NT	0.95	0.973	0.914	0.319
<i>Murina recondita</i>	Vespertilionidae	DD	0.918	0.869	0.855	0.187
<i>Myodes shanseius</i>	Cricetidae	DD	0.815	0.947	0.996	0.229
<i>Myotis davidii</i>	Vespertilionidae	LC	0.978	0.999	0.998	0.391
<i>Myotis fimbriatus</i>	Vespertilionidae	NT	0.746	1	0.782	0.731
<i>Myotis pequinius</i>	Vespertilionidae	LC	0.995	0.977	0.992	0.119
<i>Neodon irene</i>	Cricetidae	LC	0.94	0.944	0.997	0.155
<i>Neodon linzhiensis</i>	Cricetidae	NT	0.941	0.981	0.925	0.874
<i>Neohylomys hainanensis</i>	Erinaceidae	VU	0.991	0.997	0.999	0.371
<i>Niviventer andersoni</i>	Muridae	LC	0.974	0.975	0.984	0.115
<i>Niviventer coninga</i>	Muridae	LC	0.867	0.93	0.961	0.201
<i>Niviventer culturatus</i>	Muridae	NT	0.809	0.941	0.759	0.308
<i>Niviventer excelsior</i>	Muridae	LC	0.853	0.958	0.999	0.162
<i>Nyctalus plancyi</i>	Vespertilionidae	LC	1	0.991	1	0.309
<i>Ochotona cansus</i>	Ochotonidae	LC	0.995	0.883	0.96	0.170
<i>Ochotona erythrotis</i>	Ochotonidae	LC	0.707	0.778	0.922	0.499
<i>Ochotona gloveri</i>	Ochotonidae	LC	0.797	0.858	0.816	0.123
<i>Ochotona iliensis</i>	Ochotonidae	EN	0.983	0.996	0.974	0.176
<i>Ochotona thomasi</i>	Ochotonidae	NT	0.725	0.822	0.982	0.321

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<i>Pantholops hodgsonii</i>	Bovidae	NT	0.794	0.881	0.824	0.211
<i>Petaurista alborufus</i>	Sciuridae	LC	0.963	0.972	0.996	0.298
<i>Petaurista xanthotis</i>	Sciuridae	LC	0.741	0.893	0.996	0.159
<i>Plecotus taivanus</i>	Vespertilionidae	NT	0.964	0.98	0.658	0.288
<i>Procapra picticaudata</i>	Bovidae	NT	0.924	1	0.994	0.145
<i>Proedromys bedfordi</i>	Cricetidae	VU	0.991	0.996	1	0.469
<i>Proedromys liangshanensis</i>	Cricetidae	NT	0.964	0.939	0.983	0.233
<i>Rhinolophus formosae</i>	Rhinolophidae	NT	0.981	0.975	0.995	0.160
<i>Rhinolophus huananus</i>	Rhinolophidae	NT	0.643	0.881	0.988	0.163
<i>Rhinolophus monoceros</i>	Rhinolophidae	VU	0.912	0.945	0.91	0.100
<i>Rhinolophus rex</i>	Rhinolophidae	NT	0.483	0.83	0.993	0.333
<i>Rhinolophus xinanzhongguoensis</i>	Rhinolophidae	DD	0.64	0.885	0.659	0.221
<i>Rhinopithecus bieti</i>	Cercopithecidae	EN	0.997	0.998	0.973	0.269
<i>Rhinopithecus roxellana</i>	Cercopithecidae	VU	0.999	0.998	0.997	0.265
<i>Scapanulus oweni</i>	Talpidae	NT	1	0.999	0.999	0.175
<i>Scaptochirus moschatus</i>	Talpidae	NT	0.969	0.957	0.963	0.431
<i>Sciurotamias davidianus</i>	Sciuridae	LC	0.877	0.887	0.864	0.542
<i>Sorex cylindricauda</i>	Soricidae	NT	0.97	0.923	0.952	0.196
<i>Sorex sinalis</i>	Soricidae	NT	0.992	0.997	0.995	0.038
<i>Sorex thibetanus</i>	Soricidae	NT	0.984	0.993	0.966	0.322
<i>Trogopterus xanthipes</i>	Sciuridae	VU	0.998	0.916	0.989	0.177
<i>Uropsilus investigator</i>	Talpidae	NT	0.888	0.99	0.971	0.193
<i>Uropsilus soricipes</i>	Talpidae	LC	0.999	1	0.999	0.192
<i>Volemys millicens</i>	Cricetidae	NT	0.999	0.972	0.983	0.172
<i>Volemys musseri</i>	Cricetidae	NT	0.963	0.959	0.989	0.250

Reptiles

<i>Achalinus hainanus</i>	Xenodermatidae	VU	0.561	0.836	0.904	0.435
<i>Achalinus meiguensis</i>	Xenodermatidae	LC	1	1	0.817	0.172
<i>Achalinus niger</i>	Xenodermatidae	LC	0.961	0.945	0.993	0.152
<i>Alligator sinensis</i>	Alligatoridae	CR	0.981	0.96	0.827	0.633
<i>Amphiesma miyajimae</i>	Colubridae	EN	-0.183	0.668	0.998	0.305
<i>Calamaria yunnanensis</i>	Colubridae	VU	0.921	0.923	0.974	0.239
<i>Cyrtodactylus zhaoermii</i>	Gekkonidae	DD	0.524	0.849	0.821	0.735
<i>Dopasia hainanensis</i>	Anguidae	EN	0.922	0.92	0.976	0.319
<i>Elaphe anomala</i>	Colubridae	VU	0.847	0.903	0.809	0.451
<i>Elaphe bimaculata</i>	Colubridae	LC	0.959	0.992	0.995	0.413
<i>Euprepiophis perlacea</i>	Colubridae	EN	0.959	0.991	0.999	0.286
<i>Gekko chinensis</i>	Gekkonidae	LC	0.799	0.875	0.995	0.317
<i>Gekko kikuchii</i>	Gekkonidae	VU	0.792	0.125	0.72	0.624
<i>Gekko liboensis</i>	Gekkonidae	LC	0.996	0.998	0.995	0.166
<i>Gekko melli</i>	Gekkonidae	VU	0.393	0.997	0.99	0.196
<i>Gekko scabridus</i>	Gekkonidae	LC	0.993	0.997	0.976	0.229
<i>Gekko similignum</i>	Gekkonidae	DD	0.908	0.895	0.985	0.316
<i>Gekko subpalmatus</i>	Gekkonidae	LC	0.854	0.843	0.985	0.339
<i>Gekko swinhonis</i>	Gekkonidae	VU	0.988	0.999	0.995	0.299
<i>Goniurosaurus hainanensis</i>	Eublepharidae	VU	0.982	0.922	0.911	0.234

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<i>Goniurosaurus liboensis</i>	Eublepharidae	EN	0.763	0.135	0.833	0.246
<i>Japalura brevipes</i>	Agamidae	NT	0.802	0.734	0.997	0.095
<i>Japalura makii</i>	Agamidae	VU	0.844	0.158	0.894	0.280
<i>Japalura swinhonis</i>	Agamidae	LC	0.991	0.987	0.974	0.365
<i>Laudakia wui</i>	Agamidae	NT	0.962	0.993	0.994	0.143
<i>Lycodon gongshan</i>	Colubridae	NT	0.407	0.885	0.95	0.425
<i>Lycodon multizonatus</i>	Colubridae	NT	0.986	0.987	0.996	0.181
<i>Lycodon synaptor</i>	Colubridae	DD	0.729	0.936	0.935	0.273
<i>Macropisthodon rufus</i>	Colubridae	LC	0.937	0.907	0.98	0.549
<i>Oligodon ornatus</i>	Colubridae	NT	0.856	0.99	0.984	0.061
<i>Opisthotropis andersonii</i>	Colubridae	NT	0.982	0.972	0.986	0.336
<i>Opisthotropis cheni</i>	Colubridae	NT	0.967	0.963	0.931	0.058
<i>Opisthotropis guangxiensis</i>	Colubridae	NT	0.644	0.985	0.85	0.263
<i>Opisthotropis jacobi</i>	Colubridae	NT	0.986	0.996	0.979	0.374
<i>Opisthotropis kuatunensis</i>	Colubridae	LC	0.963	0.837	0.937	0.312
<i>Opisthotropis latouchii</i>	Colubridae	LC	0.515	0.996	0.948	0.451
<i>Opisthotropis maxwelli</i>	Colubridae	NT	0.914	0.965	0.942	0.281
<i>Pareas boulengeri</i>	Pareatidae	LC	0.943	0.976	0.961	0.266
<i>Pareas chinensis</i>	Pareatidae	LC	0.881	0.85	0.902	0.430
<i>Pareas formosensis</i>	Pareatidae	NT	0.968	0.948	0.964	0.300
<i>Pareas nigriceps</i>	Pareatidae	DD	0.908	0.942	0.891	0.292
<i>Pareas stanleyi</i>	Pareatidae	LC	0.804	0.962	0.87	0.268
<i>Phrynocephalus erythrus</i>	Agamidae	LC	0.942	0.961	0.997	0.154
<i>Phrynocephalus forsythii</i>	Agamidae	LC	0.864	0.956	0.882	0.388
<i>Phrynocephalus przewalskii</i>	Agamidae	LC	0.973	0.966	0.97	0.155
<i>Phrynocephalus putjatai</i>	Agamidae	NT	0.988	0.967	0.97	0.202
<i>Plestiodon capito</i>	Scincidae	LC	1	0.991	0.997	0.343
<i>Rhabdophis adleri</i>	Colubridae	NT	0.844	0.907	0.911	0.288
<i>Rhabdophis swinhonis</i>	Colubridae	NT	0.914	0.954	0.937	0.292
<i>Scincella modesta</i>	Scincidae	LC	0.954	0.97	0.933	0.184
<i>Scincella tsinlingensis</i>	Scincidae	LC	1	1	0.995	0.289
<i>Scincella formosensis</i>	Scincidae	LC	0.996	0.99	0.997	0.301
<i>Scincella potanini</i>	Scincidae	LC	0.977	0.999	0.999	0.506
<i>Sinomicrurus hatori</i>	Elapidae	VU	0.898	0.595	0.83	0.090
<i>Sinomicrurus sauteri</i>	Elapidae	VU	0.993	0.993	0.898	0.205
<i>Sinonatrix annularis</i>	Colubridae	VU	0.877	0.926	0.819	0.409
<i>Sphenomorphus taiwanensis</i>	Scincidae	LC	0.913	0.949	0.877	0.107
<i>Takydromus formosanus</i>	Lacertidae	LC	0.973	0.96	0.882	0.166
<i>Takydromus hsuehshanensis</i>	Lacertidae	LC	0.941	0.96	0.953	0.330
<i>Takydromus intermedius</i>	Lacertidae	NT	0.651	0.794	0.857	0.532
<i>Takydromus luyeanus</i>	Lacertidae	LC	0.938	0.92	0.84	0.387
<i>Takydromus sauteri</i>	Lacertidae	NT	0.99	0.965	0.995	0.278
<i>Takydromus septentrionalis</i>	Lacertidae	LC	0.897	0.75	0.989	0.439
<i>Takydromus stejnegeri</i>	Lacertidae	LC	0.987	0.980	0.624	0.409
<i>Takydromus sylvaticus</i>	Lacertidae	EN	0.993	0.991	0.962	0.265
<i>Takydromus viridipunctatus</i>	Lacertidae	LC	0.913	0.886	0.845	0.371

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<i>Teratoscincus roborowskii</i>	Sphaerodactyli dae	NT	0.977	0.974	0.998	0.126
<i>Thermophis baileyi</i>	Colubridae	CR	0.989	0.969	0.826	0.237
<i>Thermophis zhaoermii</i>	Colubridae	CR	0.955	0.981	0.996	0.227
<i>Trimeresurus gracilis</i>	Viperidae	NT	0.809	0.756	0.862	0.085
<i>Xenopeltis hainanensis</i>	Xenopeltidae	NT	0.878	0.945	1.000	0.370

Supplementary Table 4A. The influence of SSPs in models testing the suitable area and its change (P-values obtained using Tukey multiple comparisons). Results that are significantly different from each other are represented using different letters, and those that are smallest have the smallest values alphabetically (i.e., a is smallest).

	ASH	CRI	LRI	ASH	CRI	LRI
SSP2 - SSP1	1	0.99942	< 0.0001	a-a	a-a	b-a
SSP3 - SSP1	1	0.00341	< 0.0001	a-a	b-a	c-a
SSP5 - SSP1	1	< 0.0001	< 0.0001	a-a	b-a	d-a
SSP3 - SSP2	1	0.00472	< 0.0001	a-a	b-a	c-b
SSP5 - SSP2	1	< 0.0001	< 0.0001	a-a	b-a	d-b
SSP5 - SSP3	1	0.71412	< 0.0001	a-a	b-b	d-c

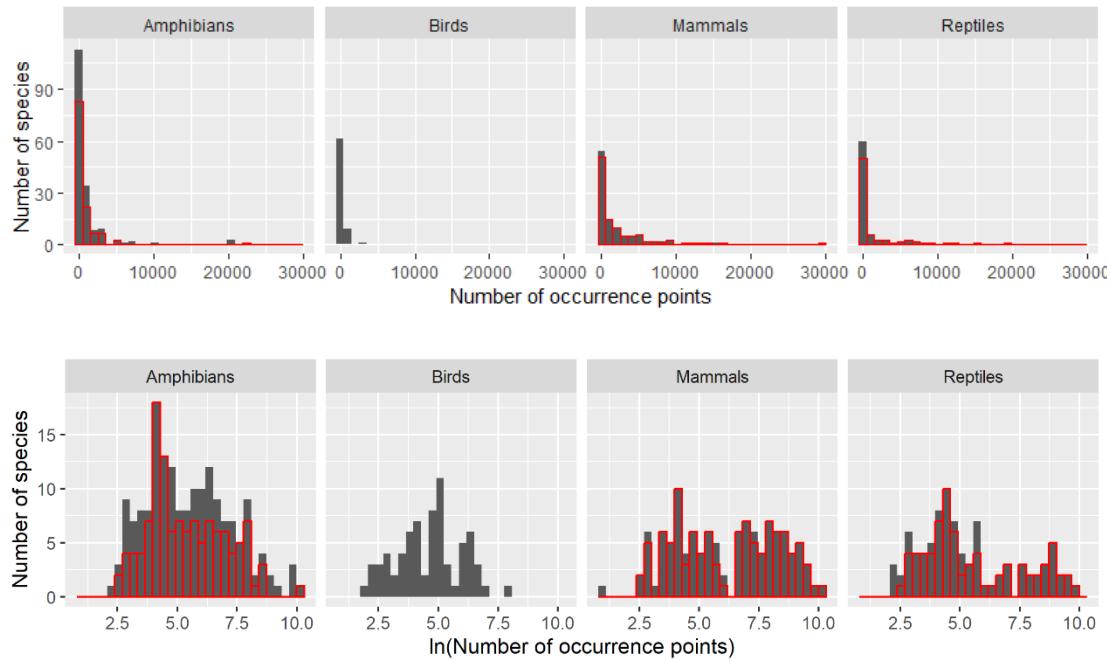
Supplementary Table 4B. The influence of Period in models testing the suitable area and its change (P-values obtained using Tukey multiple comparisons). Results that are significantly different from each other are represented using different letters, and those that are smallest have the smallest values alphabetically (i.e., a is smallest).

	ASH	CRI	LRI	ASH	CRI	LRI
2041-2060 - 2021-2040	0.0185	1.0	< 0.0001	b-c	a-a	b-a
2061-2080 - 2021-2040	< 0.0001	0.05981	< 0.0001	b-c	ab-a	c-a
2081-2100 - 2021-2040	< 0.0001	0.00196	< 0.0001	a-c	b-a	d-a
2061-2080 - 2041-2060	0.1713	0.05757	< 0.0001	b-b	ab-a	c-b
2081-2100 - 2041-2060	< 0.0001	0.00171	< 0.0001	a-b	b-a	d-b
2081-2100 - 2061-2080	< 0.0001	0.70113	< 0.0001	a-b	b-ab	d-c

Supplementary Table 5. The number of endemic vertebrates that will face a threat of extinction in the period 2080-2100 under the different future scenarios, organized by IUCN levels. Species will be threatened by extinction if their future distribution is predicted to be less than 20% of their current distribution (i.e., CRI < -0.8), or if 80% of the currently suitable area becomes unsuitable in the future (i.e., LRI > 0.8).

SSP	CRI < -0.8	LRI > 0.8	Total species	Threatened	LC	NT	Threatened %	LC %	NT %
Amphibians									
SSP1	8	10	11	8	1	2	9.88	2.13	6.45
SSP2	13	11	16	14	0	2	17.72	0.00	6.25
SSP3	13	17	17	14	1	2	17.72	2.13	6.25
SSP5	25	33	33	23	6	4	28.75	13.04	12.50
Birds									
SSP1	0	0	0	0	0	0	0.00	0.00	0.00
SSP2	1	0	1	1	0	0	3.85	0.00	0.00
SSP3	1	2	2	2	0	0	7.69	0.00	0.00
SSP5	2	3	3	3	0	0	11.54	0.00	0.00
Mammals									
SSP1	0	1	1	0	1	0	0.00	3.23	0.00
SSP2	1	0	1	1	0	0	4.76	0.00	0.00
SSP3	1	2	2	1	0	1	4.76	0.00	3.03
SSP5	6	10	10	3	3	4	14.29	10.00	12.12
Reptiles									
SSP1	2	1	2	2	0	0	10.53%	0.00%	0.00%
SSP2	4	2	4	3	0	1	15.79%	0.00%	5.26%
SSP3	4	5	5	4	0	1	21.05%	0.00%	5.26%
SSP5	5	5	5	3	0	2	15.79%	0.00%	10.53%

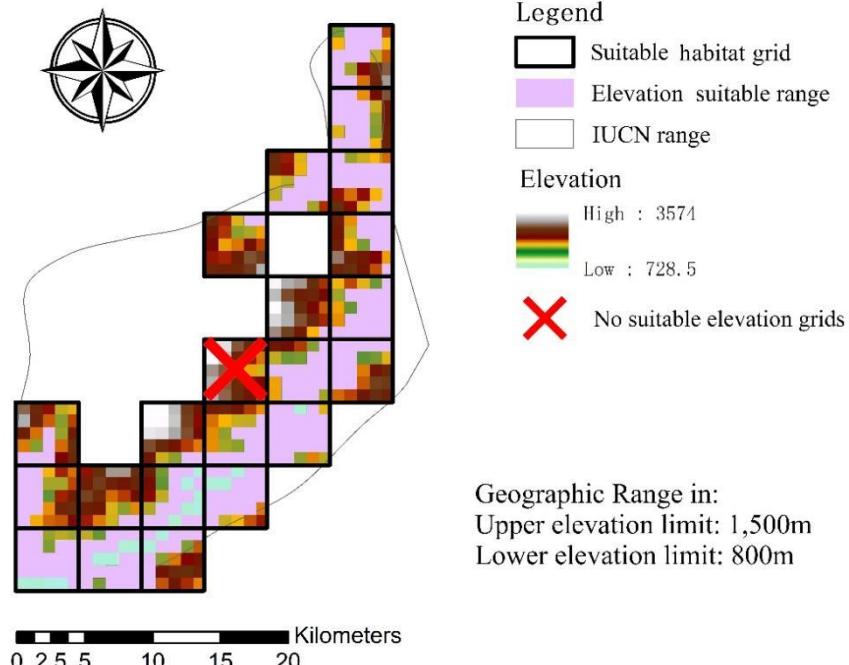
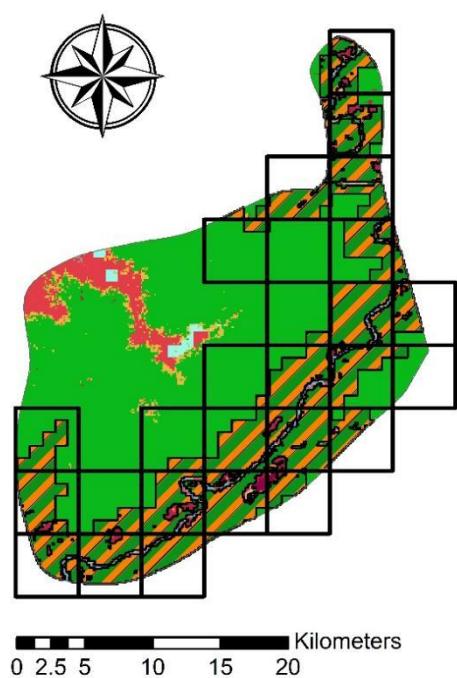
2.2 Supplementary Figures



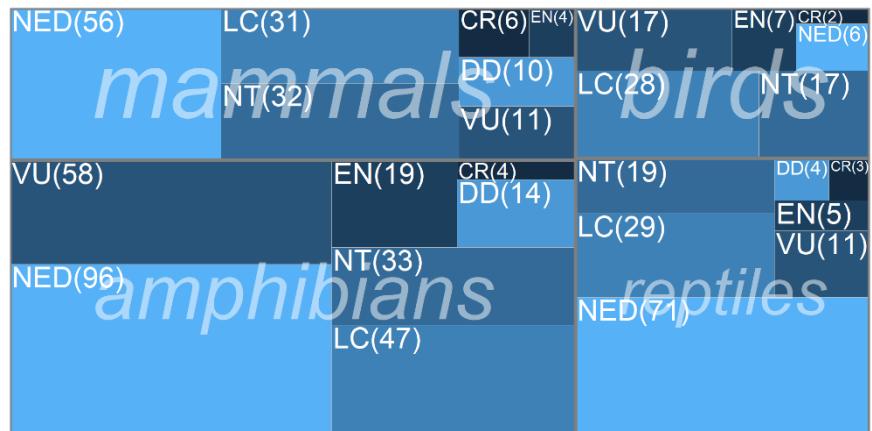
Supplementary Figure 1 . Histograms of the number of occurrence records for the different species in each taxon. Upper panels are based on the raw data, and the lower panels show a log transformation. Red color marks out the IUCN data; other data are from Global Biodiversity Information Facility (GBIF: www.gbif.org) or the Chinese Bird Report (www.birdreport.cn; for birds specifically).

Suitable habitat for *Amolops aniqiaoensis* according to its IUCN range vs. AOH.

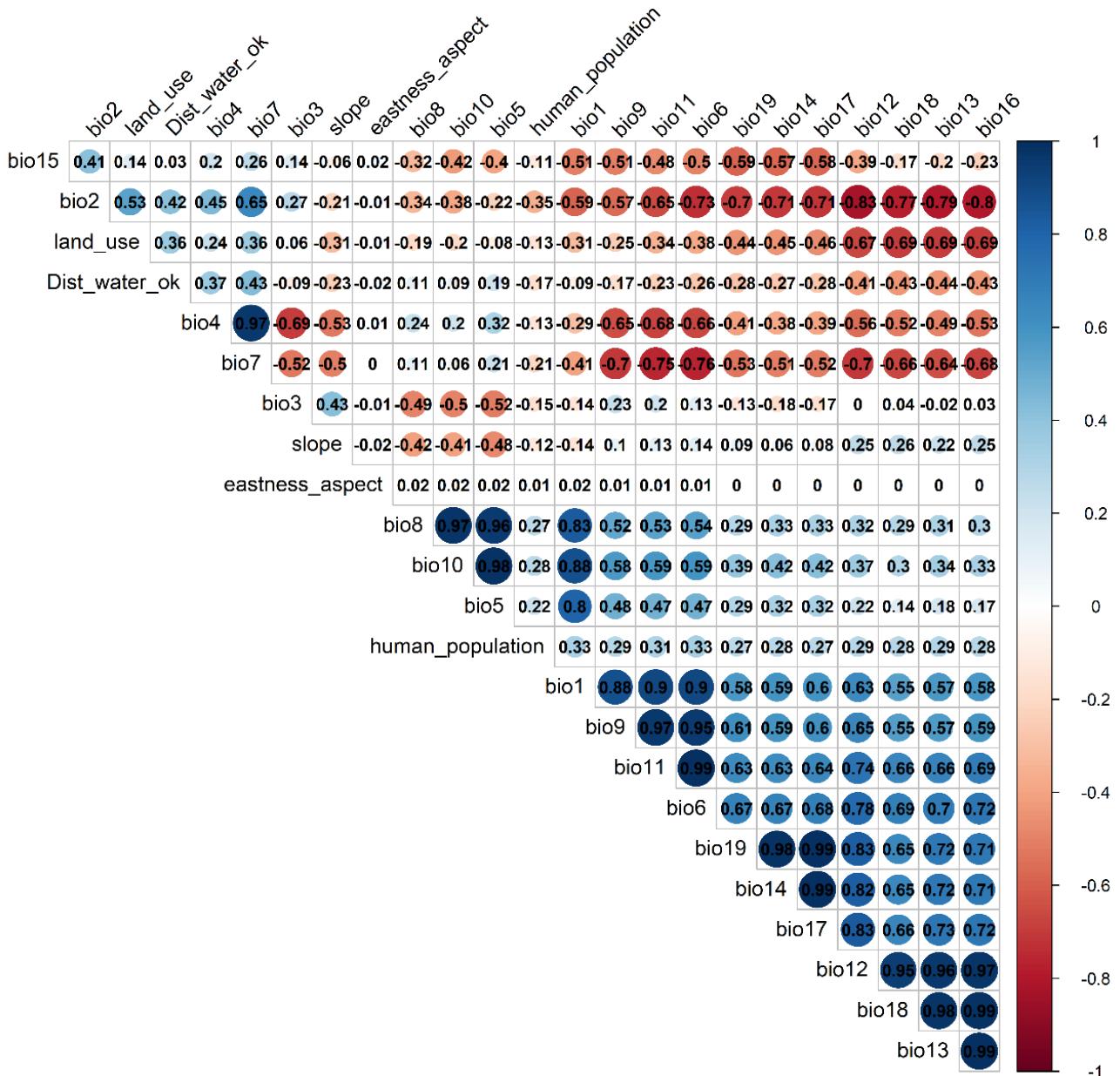
Level 1	Level 2
1. Forest	1.6. Forest - Subtropical/Tropical Moist Lowland 1.9. Forest - Subtropical/Tropical Moist Montane
5. Wetlands (inland)	5.1. Wetlands (inland) - Permanent Rivers/Streams/Creeks (includes waterfalls)



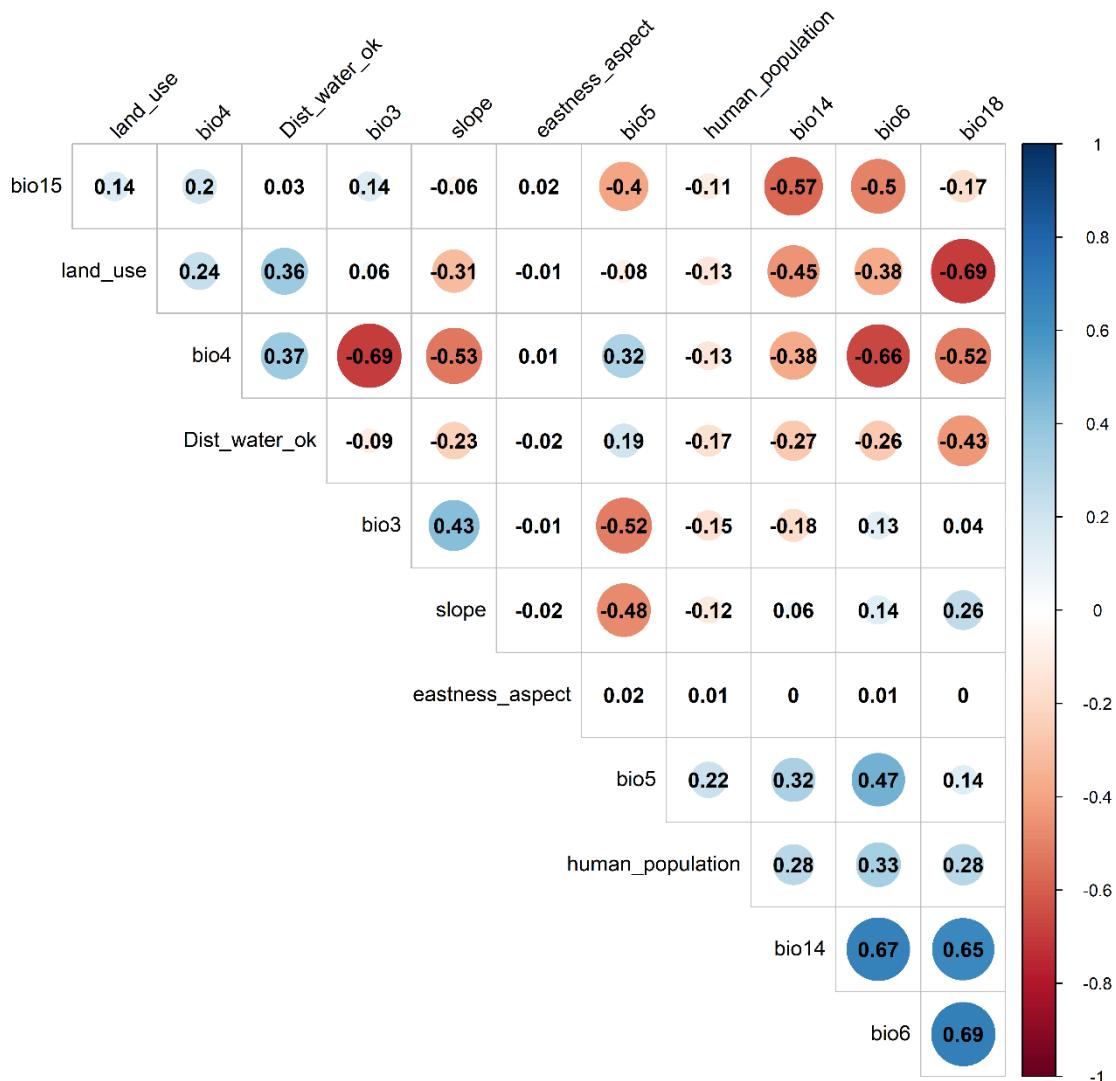
Supplementary Figure 2 . Example of *Amolops aniqiaoensis* illustrating Approach 4. The gridcell is shown if any area within it was suitable. In this example, one gridcell was removed because it did not contain suitable elevation.



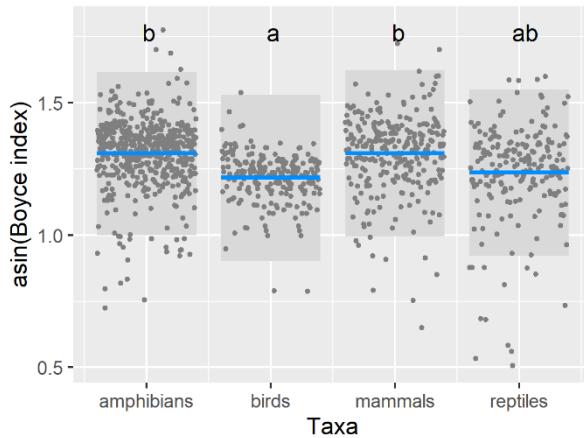
Supplementary Figure 3. A treemap showing the sample size for each taxon, and subgroups at different category levels (higher category levels in dark blue). Species that did not have enough data for the analysis, that is, species that had less than 10 occurrence points after unsuitable habitat was removed, are represented by Not Enough Data (NED). Other abbreviations represent the species threat status: LC = Least Concern, NT = Near Threatened, VU = Vulnerable, EN = Endangered, CR = Critically Endangered, DD = Data Deficient.



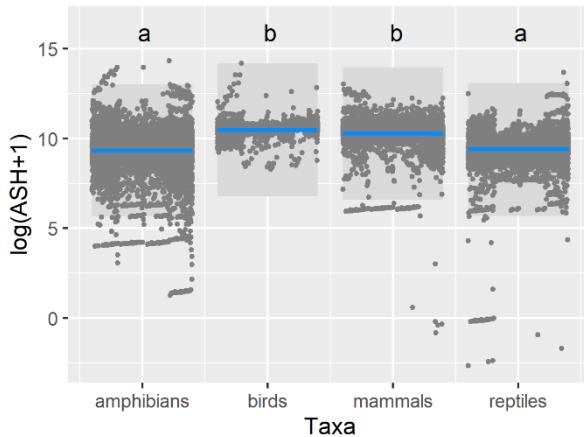
Supplementary Figure 4A. Matrix of pairwise correlations as assessed by the Pearson correlation coefficient for the original 24 variables.



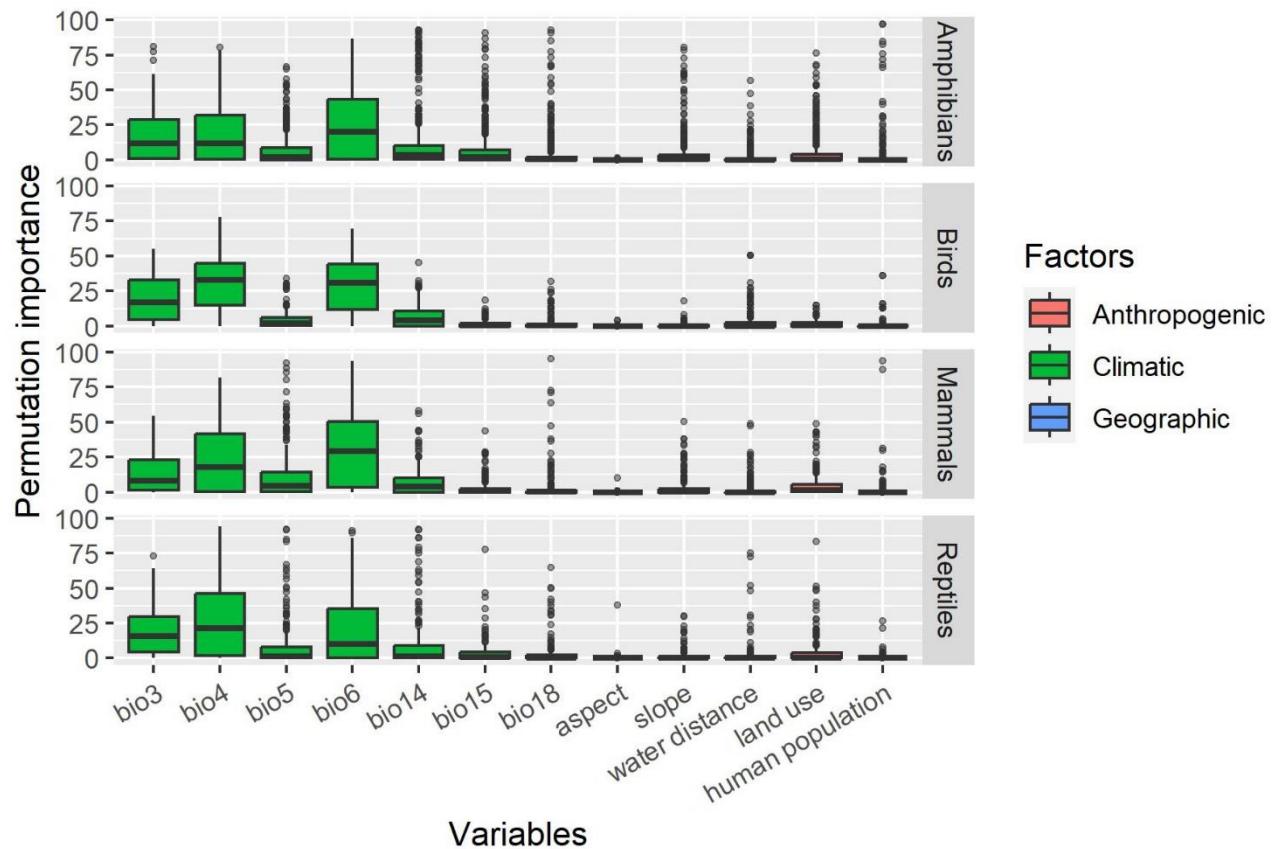
Supplementary Figure 4B. The same matrix for the remaining 12 variables after correlated variables ($|r| > 0.7$) were removed.



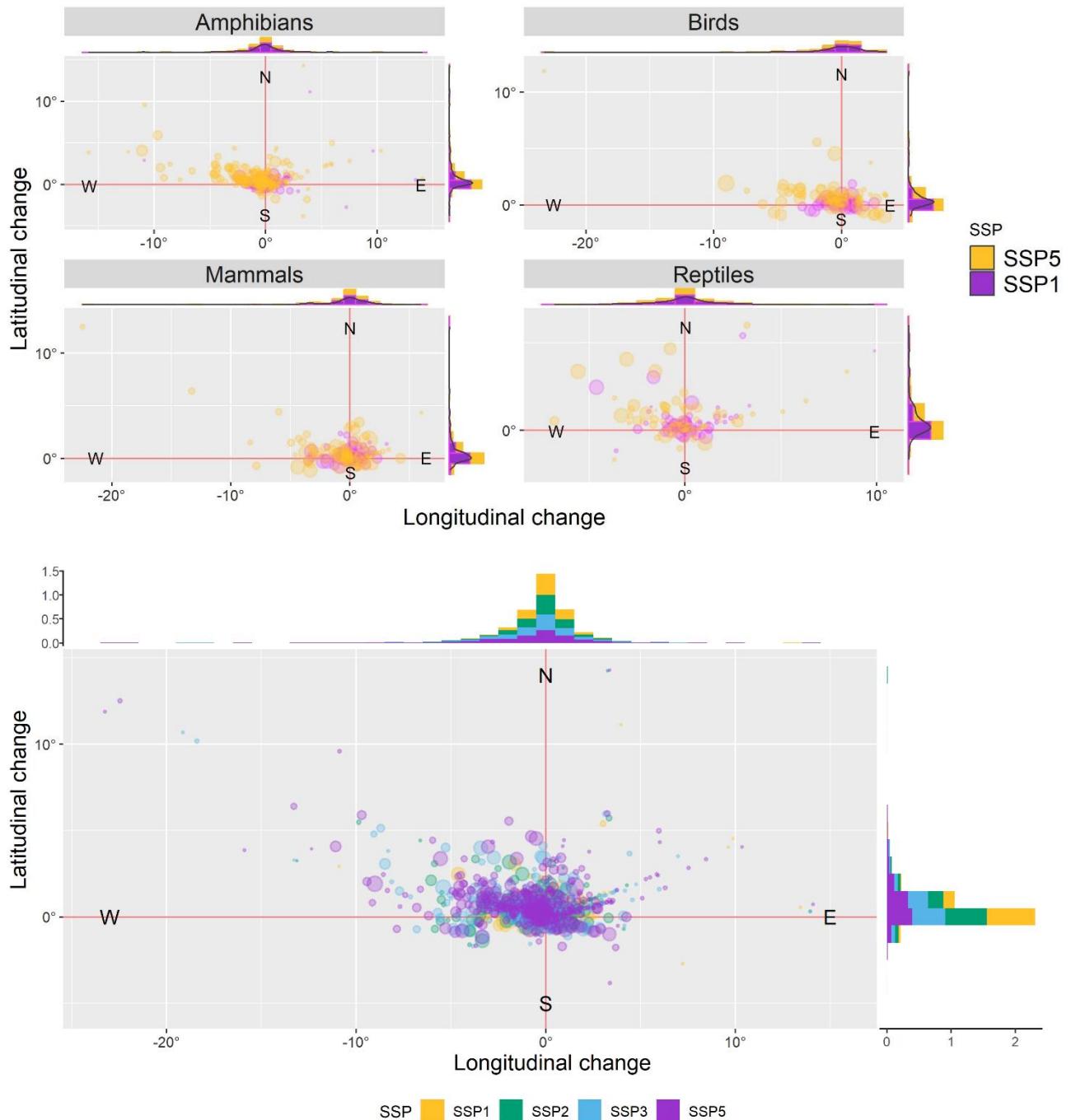
Supplementary Figure 5A. The models analyzing Boyce index varied by Taxa. Blue is the predicted mean value, and the gray area represents the 95% confidence interval. Columns with the same letter are not significantly different.

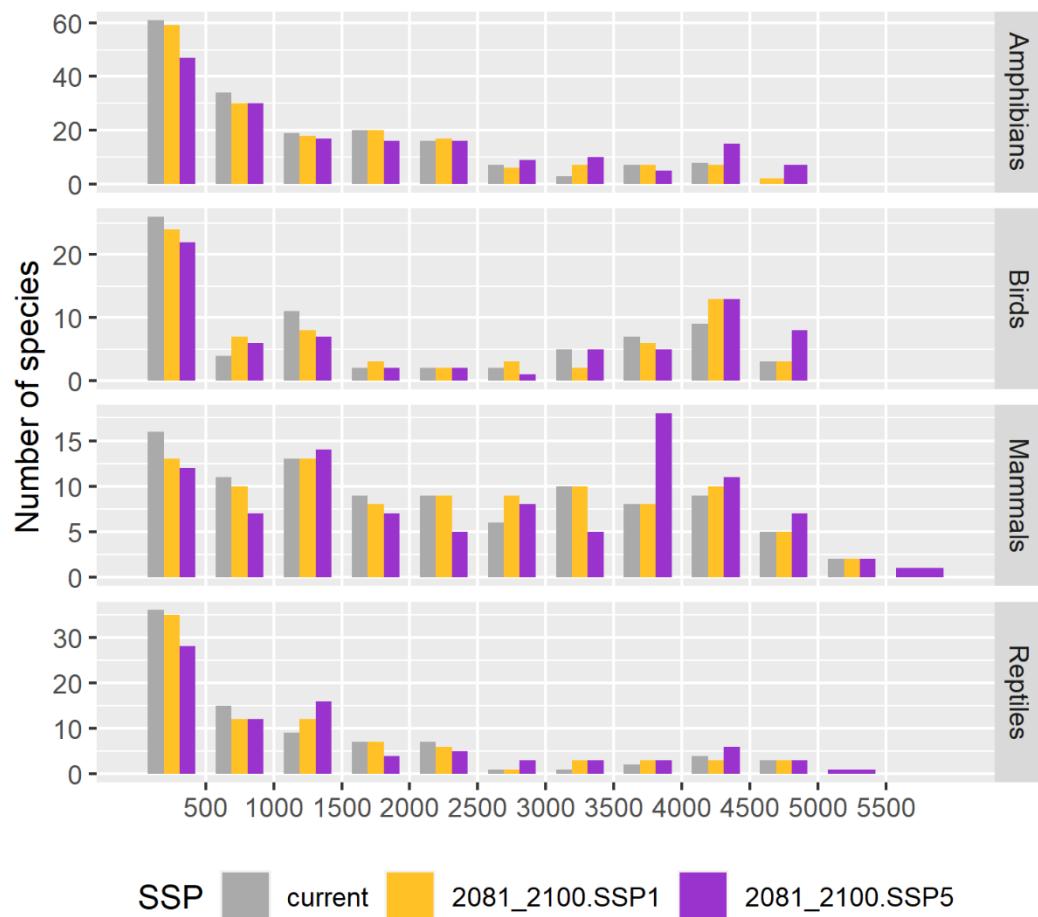
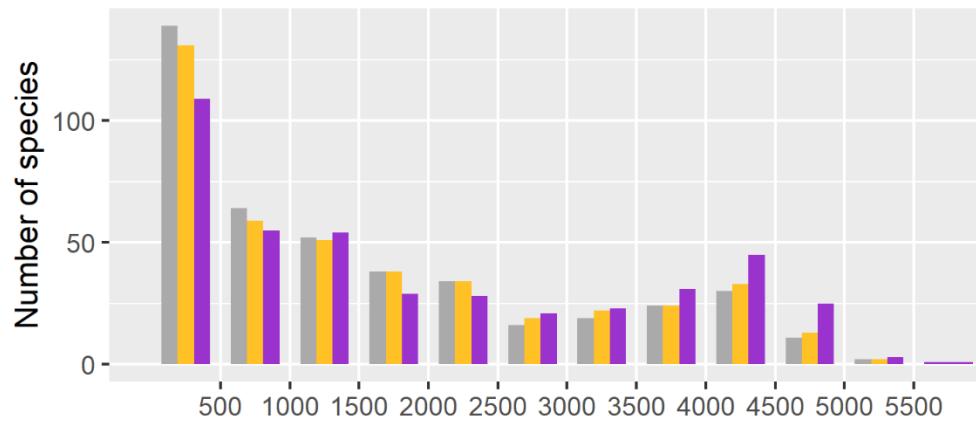


Supplementary Figure 5B. The models analyzing ASH varied by Taxa. Blue is the predicted mean value, and the gray area represents the 95% confidence interval. Columns with the same letter are not significantly different.



Supplementary Figure 6. A boxplot illustrating the permutation importance of each of the twelve explanatory variables in the best models based on Approach 4 for each taxon. The variables are classified into three different classes: climatic, geographic and anthropogenic.





Supplementary Figure 8. The mean elevation (in m a.s.l.) for each species in Approach 4, currently, and under the different climate scenarios in above, with different taxa in bottom.

3 Supplementary References

- Araújo, M. B., Thuiller, W., and Pearson, R. G. (2006). Climate warming and the decline of amphibians and reptiles in Europe. *Journal of Biogeography*, 33(10), 1712–1728. <https://doi.org/10.1111/j.1365-2699.2006.01482.x>
- Barnagaud, J. Y., Geniez, P., Cheylan, M., and Crochet, P. A. (2021). Climate overrides the effects of land use on the functional composition and diversity of Mediterranean reptile assemblages. *Diversity and Distributions*, 27(1), 50–64. <https://doi.org/10.1111/ddi.13176>
- Barrett, K., Nibbelink, N. P., and Maerz, J. C. (2014). Identifying priority species and conservation opportunities under future climate scenarios: Amphibians in a biodiversity hotspot. *Journal of Fish and Wildlife Management*, 5(2), 282–297. <https://doi.org/10.3996/022014-JFWM-015>
- Biber, M. F., Voskamp, A., Niamir, A., Hickler, T., and Hof, C. (2020). A comparison of macroecological and stacked species distribution models to predict future global terrestrial vertebrate richness. *Journal of Biogeography*, 47(1), 114–129. <https://doi.org/10.1111/jbi.13696>
- Bogoni, J. A., and Tagliari, M. M. (2021). Potential distribution of piscivores across the Atlantic Forest: From bats and marsupials to large-bodied mammals under a trophic-guild viewpoint. *Ecological Informatics*, 64(April), 101357. <https://doi.org/10.1016/j.ecoinf.2021.101357>
- Clusella-trullas, A. S., Blackburn, T. M., Chown, S. L., Clusella-trullas, S., Blackburn, T. M., and Chown, S. L. (2011). *Climatic Predictors of Temperature Performance Curve Parameters in Ectotherms Imply Complex Responses to Climate Change*. 177(6), 738–751. <https://doi.org/10.1086/660021>
- Cohen, J. M., Fink, D., and Zuckerberg, B. (2020). *Avian responses to extreme weather across functional traits and temporal scales*. March, 1–11. <https://doi.org/10.1111/gcb.15133>
- Console, G., Iannella, M., Cerasoli, F., D'Alessandro, P., and Biondi, M. (2020). A European perspective of the conservation status of the threatened meadow viper *Vipera ursinii* (BONAPARTE, 1835) (Reptilia, Viperidae). *Wildlife Biology*, 2020(2). <https://doi.org/10.2981/wlb.00604>
- Cunningham, H. R., Rissler, L. J., Buckley, L. B., and Urban, M. C. (2016). Abiotic and biotic constraints across reptile and amphibian ranges. *Ecography*, 39(1), 1–8. <https://doi.org/10.1111/ecog.01369>
- Deb, J. C., Forbes, G., and MacLean, D. A. (2020). Modelling the spatial distribution of selected North American woodland mammals under future climate scenarios. *Mammal Review*, 50(4), 440–452. <https://doi.org/10.1111/mam.12210>
- Deb, J. C., Phinn, S., Butt, N., and McAlpine, C. A. (2019). Adaptive management and planning for the conservation of four threatened large Asian mammals in a changing climate. *Mitigation and Adaptation Strategies for Global Change*, 24(2), 259–280. <https://doi.org/10.1007/s11027-018-9810-3>

- Distler, T., Schuetz, J. G., and Vel, J. (2015). *Stacked species distribution models and macroecological models provide congruent projections of avian species richness under climate change*. 976–988. <https://doi.org/10.1111/jbi.12479>
- Dormann, C. F., Gruber, B., Winter, M., and Lett, D. H. B. (2008). *Detailed methods and additional results accompanying “Evolution of climate niches in European mammals” by*. 1–11.
- Dreitz, V. J., Conrey, R. Y., and Skagen, S. K. (2012). *Drought and Cooler Temperatures Are Associated with Higher Nest Survival in Mountain Plovers La sécheresse et les températures fraîches sont associées au taux de survie des nids supérieur chez le Pluvier montagnard*. 7(1).
- Farashi, A., and Alizadeh-Noughani, M. (2021). Predicting the invasion risk of non-native reptiles as pets in the Middle East. *Global Ecology and Conservation*, 31(June), e01818. <https://doi.org/10.1016/j.gecco.2021.e01818>
- Germain, S. J., and Lutz, J. A. (2020). Climate extremes may be more important than climate means when predicting species range shifts. *Climatic Change*, 163(1), 579–598. <https://doi.org/10.1007/s10584-020-02868-2>
- Gherghel, I., Brischoux, F., Nyári, Á. S., and Papeş, M. (2020). A simple framework for estimating potential distributions of amphibious marine species and implications for conservation. *Coral Reefs*, 39(4), 1081–1090. <https://doi.org/10.1007/s00338-020-01937-3>
- Goudarzi, F., Hemami, M. R., Malekian, M., Fakheran, S., and Martínez-Freiría, F. (2021). Species versus within-species niches: a multi-modelling approach to assess range size of a spring-dwelling amphibian. *Scientific Reports*, 11(1), 1–11. <https://doi.org/10.1038/s41598-020-79783-0>
- Groff, L. A., Groff, L. A., and Hayes, M. P. (2014). Using ecological niche models to direct rare amphibian surveys: A case study using the oregon spotted frog (*Rana pretiosa*). *Herpetological Conservation and Biology*, 9(2), 354–368.
- Guevara, L., Gerstner, B. E., Kass, J. M., and Anderson, R. P. (2018). Toward ecologically realistic predictions of species distributions: A cross-time example from tropical montane cloud forests. *Global Change Biology*, 24(4), 1511–1522. <https://doi.org/10.1111/gcb.13992>
- Hill, S. E., and Winder, I. C. (2019). *Predicting the impacts of climate change on Papio baboon biogeography : Are widespread , generalist primates ‘safe ’? May 2018*, 1–26. <https://doi.org/10.1111/jbi.13582>
- Huang, Q., Sauer, J. R., and Dubayah, R. O. (2017). *Multi-directional Abundance Shifts Among North American Birds and the Relative Influence of Multi- Faceted Climate Factors*. 0–2. <https://doi.org/10.1111/ijlh.12426>
- Ihlow, F., Dambach, J., Engler, J. O., Flecks, M., Hartmann, T., Nekum, S., Rajaei, H., and Rödder, D. (2012). On the brink of extinction? How climate change may affect global chelonian species richness and distribution. *Global Change Biology*, 18(5), 1520–1530. <https://doi.org/10.1111/j.1365-2486.2011.02623.x>

- Jurestovsky, D., and Andrew Joyner, T. (2018). Applications of species distribution modeling for palaeontological fossil detection: late Pleistocene models of Saiga (Artiodactyla: Bovidae, *Saiga tatarica*). *Palaeobiodiversity and Palaeoenvironments*, 98(2), 277–285. <https://doi.org/10.1007/s12549-017-0298-8>
- Ko, C. Y., Schmitz, O. J., and Jetz, W. (2016). The limits of direct community modeling approaches for broad-scale predictions of ecological assemblage structure. *Biological Conservation*, 201, 396–404. <https://doi.org/10.1016/j.biocon.2016.07.026>
- Lamelas-López, L., Pardavila, X., Borges, P. A. V., Santos-Reis, M., Amorim, I. R., and Santos, M. J. (2020). Modelling the distribution of *Mustela nivalis* and *M. putorius* in the Azores archipelago based on native and introduced ranges. *PLoS ONE*, 15(8 August), 1–19. <https://doi.org/10.1371/journal.pone.0237216>
- Leach, K., Montgomery, W. I., and Reid, N. (2017). Characterizing biotic interactions within the Order Lagomorpha using Joint Species Distribution Models at 3 different spatial scales. *Journal of Mammalogy*, 98(5), 1434–1442. <https://doi.org/10.1093/jmammal/gyx105>
- Lemes, P., Melo, A. S., and Loyola, R. D. (2014). Climate change threatens protected areas of the Atlantic Forest. *Biodiversity and Conservation*, 23(2), 357–368. <https://doi.org/10.1007/s10531-013-0605-2>
- Li, X., Clinton, N., Si, Y., Liao, J., Liang, L., and Gong, P. (2015). Projected impacts of climate change on protected birds and nature reserves in China. *Science Bulletin*, 60(19), 1644–1653. <https://doi.org/10.1007/s11434-015-0892-y>
- Li, X., Liu, X., Kraus, F., Tingley, R., and Li, Y. (2016). Risk of biological invasions is concentrated in biodiversity hotspots. *Frontiers in Ecology and the Environment*, 14(8), 411–417. <https://doi.org/10.1002/fee.1321>
- Li, Y., Li, X., Sandel, B., Blank, D., Liu, Z., Liu, X., and Yan, S. (2016). *Climate and topography explain range sizes of terrestrial vertebrates*. 6(818). <https://doi.org/10.1038/NCLIMATE2895>
- Magory Cohen, T., and Dor, R. (2019). The effect of local species composition on the distribution of an avian invader. *Scientific Reports*, 9(1), 1–9. <https://doi.org/10.1038/s41598-019-52256-9>
- Maiorano, L., Falcucci, A., Zimmermann, N. E., Psomas, A., Pottier, J., Baisero, D., Rondinini, C., Guisan, A., and Boitani, L. (2011). The future of terrestrial mammals in the Mediterranean basin under climate change. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 366(1578), 2681–2692. <https://doi.org/10.1098/rstb.2011.0121>
- Matthews, S. N., Iverson, L. R., Prasad, A. M., and Peters, M. P. (2011). Changes in potential habitat of 147 North American breeding bird species in response to redistribution of trees and climate following predicted climate change. *Ecography*, 34(6), 933–945. <https://doi.org/10.1111/j.1600-0587.2011.06803.x>
- Mestre, F., Risk, B. B., Mira, A., Beja, P., and Pita, R. (2017). A metapopulation approach to predict species range shifts under different climate change and landscape connectivity scenarios. *Ecological Modelling*, 359, 406–414. <https://doi.org/10.1016/j.ecolmodel.2017.06.013>

- Mokhatla, M. M., Dennis Rödder, and G. John Measey. (2015). Assessing the effects of climate change on distributions of Cape Floristic Region amphibians. *South African Journal of Science*, *Volume 111*(Number 11/12), 1–7. <https://doi.org/10.17159/sajs.2015/20140389>
- Morán-Ordóñez, A., Briscoe, N. J., and Wintle1, B. A. (2017). *Modelling species responses to extreme weather provides new insights into constraints on range and likely climate change impacts for Australian mammals.*
- Morueta-Holme, N., Fløjgaard, C., and Svenning, J. C. (2010). Climate change risks and conservation implications for a threatened small-range mammal species. *PLoS ONE*, *5*(4). <https://doi.org/10.1371/journal.pone.0010360>
- Mouchet, M., Levers, C., Zupan, L., and Kuemmerle, T. (2015). *Testing the Effectiveness of Environmental Variables to Explain European Terrestrial Vertebrate Species Richness across Biogeographical Scales*. 1–16. <https://doi.org/10.1371/journal.pone.0131924>
- Munguía, M., Rahbek, C., Rangel, T. F., Diniz-Filho, J. A. F., and Araújo, M. B. (2012). Equilibrium of global amphibian species distributions with climate. *PloS One*, *7*(4), 1–9. <https://doi.org/10.1371/journal.pone.0034420>
- Muñoz, A., Santos, X., and Felicísimo, Á. M. (2016). Local-scale models reveal ecological niche variability in amphibian and reptile communities from two contrasting biogeographic regions. *PeerJ*, *2016*(10), 1–20. <https://doi.org/10.7717/peerj.2405>
- Peñalver-Alcázar, M., Jiménez-Valverde, A., and Aragón, P. (2021). Niche differentiation between deeply divergent phylogenetic lineages of an endemic newt: implications for Species Distribution Models. *Zoology*, *144*(June 2020). <https://doi.org/10.1016/j.zool.2020.125852>
- Ribeiro, B. R., Sales, L. P., and Loyola, R. (2018). Strategies for mammal conservation under climate change in the Amazon. *Biodiversity and Conservation*, *27*(8), 1943–1959. <https://doi.org/10.1007/s10531-018-1518-x>
- Rodrigues, J. F. M., and Lima-Ribeiro, M. S. (2018). Predicting where species could go: climate is more important than dispersal for explaining the distribution of a South American turtle. *Hydrobiologia*, *808*(1), 343–352. <https://doi.org/10.1007/s10750-017-3436-4>
- Salas, E. A. L. , A. Seamster, V., G. Boykin, K., M. Harings, N., and W. Dixon, K. (2017). Modeling the impacts of climate change on Species of Concern (birds) in South Central U.S. based on bioclimatic variables. *AIMS Environmental Science*, *4*(2), 358–385. <https://doi.org/10.3934/environsci.2017.2.358>
- Sangermano, F., Bol, L., Galvis, P., Gullison, R. E., Hardner, J., and Ross, G. S. (2015). Habitat suitability and protection status of four species of amphibians in the Dominican Republic. *Applied Geography*, *63*, 55–65. <https://doi.org/10.1016/j.apgeog.2015.06.002>
- Schoolder, S. L., Johnson, M. D., Njoroge, P., and Bean, W. T. (2020). *Shade trees preserve avian insectivore biodiversity on coffee farms in a warming climate. October.* <https://doi.org/10.1002/ece3.6879>

- Sheldon, K. S., Leaché, A. D., and Cruz, F. B. (2015). *The influence of temperature seasonality on elevational range size across latitude : a test using Liolaemus lizards.* <https://doi.org/10.1111/geb.12284>
- Stevens, R. D. (2013). Gradients of bat diversity in atlantic forest of South America: Environmental seasonality, sampling effort and spatial autocorrelation. *Biotropica*, 45(6), 764–770. <https://doi.org/10.1111/btp.12056>
- Thompson, D. M., Ligon, D. B., Patton, J. C., and Papeş, M. (2017). Effects of life-history requirements on the distribution of a threatened reptile. *Conservation Biology*, 31(2), 427–436. <https://doi.org/10.1111/cobi.12800>
- Vasconcelos, T. S., Rodríguez, M. Á., and Hawkins, B. A. (2012). Species distribution modelling as a macroecological tool: A case study using New World amphibians. *Ecography*, 35(6), 539–548. <https://doi.org/10.1111/j.1600-0587.2011.07050.x>
- Vol, H. (2021). *Kidov A . A ., Litvinchuk S . N . Distribution and conservation status of the Hyrcanian wood frog (Rana pseudodalmatina) in Azerbaijan // Russian.* 28(May). <https://doi.org/10.30906/1026-2296-2021-28-2-97-107>
- Xu, Z. (2015). Potential distribution of invasive alien species in the upper Ili river basin: determination and mechanism of bioclimatic variables under climate change. *Environmental Earth Sciences*, 73(2), 779–786. <https://doi.org/10.1007/s12665-014-3083-2>
- Zhang, Z., Kass, J. M., Mammola, S., Koizumi, I., Li, X., Tanaka, K., Ikeda, K., Suzuki, T., Yokota, M., and Usio, N. (2021). Lineage-level distribution models lead to more realistic climate change predictions for a threatened crayfish. *Diversity and Distributions*, 27(4), 684–695. <https://doi.org/10.1111/ddi.13225>