

## Supplementary Code: R Script for Data Analysis

*## This code uses R to analyze experimental data, including data preprocessing, statistical analysis.*

*# rawdata to cleandata*

```
dir("./rawdata")->mmm
```

```
dir.create("cleandata")
```

```
system.time(parLapply(cl,mmm,function(i){
```

```
code <-sprintf("gunzip -c ./%s | NanoFilt -q 10 -l 1000 --headcrop 50 --tailcrop 50|
```

```
gzip > ./cleandata/%s",i,i)
```

```
system(code}}))
```

*# Assemble the reference genome using long-read and short-read data*

```
system("nohup unicycler -1 ./sa17155_1.fq.gz -2 ./sa17155_2.fq.gz -l nanopore.gz -o ref &")
```

*# Use Flye to assemble the genome with only long-read sequencing*

```
system("software/Flye/bin/flye -o ./flye/ -i 10 -t 8 -g 5m --nano-raw ./nanopore.fq --plasmids")
```

*# Polish the genome using Madaka*

```
code= ("medaka_consensus -i ./nanopore.fq -d ./assembly.fasta -o ./medaka_output -t 4 -m
```

```
r103_min_high_g360")
```

```
system(code)
```

*# Use BLAST to get the identity of the fastq file compared to the reference genome*

```
system("fastq_to_fasta -i nanopore.fastq -o nanopore.fasta")
```

```
system("makeblastdb -i sa17155_ref.fa -dbtype nucl -o ref")
```

```
system("blastn -query nanopore.fasta -db ref -out nanopore.tab -outfmt '6' ")
```

*# Use fastANI to get the identity of assemble.fasta compared to the reference genome*

```
system("fastANI -q assemble.fasta -r sa17155_ref.fa -o output.txt")
```

*# Use snippy to compare assemblefasta to a reference genome. Get the number of SNPs, INSSs, DELs*

```
system("snippy --outdir 130.fq --ctgs ./assemble.fasta --ref ./sa17155_ref.fa ")
```