

Supplementary Material

Experimental exposure assessment for *in vitro* cell-based bioassays in 96- and 384-well plates

Julia Huchthausen,[†] Maria König,[†] Beate I. Escher,^{†,‡} Luise Henneberger^{†,*}

[†] Helmholtz Centre for Environmental Research – UFZ, Department of Cell Toxicology, Permoserstr. 15, 04318 Leipzig, Germany

[‡] Eberhard Karls University Tübingen, Environmental Toxicology, Department of Geosciences, 72076 Tübingen, Germany

***Correspondence:**

Luise Henneberger,
luise.henneberger@ufz.de.

S1: Test chemicals

Text S1: pK_a measurement.

The acidity constants (pK_a) of six of the test chemicals were measured using a Sirius T3 titrator (Pion). A detailed description of the experimental procedure can be found in the literature (Niu et al, 2022). Briefly, 5 μ L of a 10 mM DMSO stock solution of each test chemical and 25 μ L of a phosphate buffer (14.4 mM K_2HPO_4 and 0.15 M KCl) were added to a Sirius T3 test vial. A reference vial containing 5 μ L DMSO (Roth, A994-100 ML) and 25 μ L phosphate buffer was measured with each sample. The pK_a measurement was performed using the automated UV-metric pK_a protocol of the Sirius T3 Control software (version 2.0.0.0.). Three sequential titrations were performed with the same sample and a constant ionic strength of 0.15 M KCl ranging from pH 2 to 12 by adding 0.5 M HCl or 0.5 M KOH while UV absorbance was measured. Data analysis was performed using Sirius T3 Refine software (version 2.0.0.0.). All pK_a values were reported as the mean of the three titrations (or six for 2,4-D). The pK_a of ibuprofen was measured in the presence of 25 – 52 % methanol with 0.15 M KCl. The measured pK_a was extrapolated to the pK_a at 0 % methanol with Yasuda-Shedlovsky extrapolation (Shedlovsky, 1962; Yasuda, 1959).

Table S1: Test chemicals, their chemical class, CAS number, purity, supplier and SPME desorption solution.

<i>Chemical</i>	<i>Chemical class</i>	<i>CAS</i>	<i>Purity</i>	<i>Supplier</i>	<i>SPME desorption solution</i>
caffeine	neutral	58-08-2	$\geq 99\%$	Sigma-Aldrich	10/90 ACN/H ₂ O
lamotrigine	base	84057-84-1	$\geq 98\%$	Cayman Chemical Company	50/50 ACN/H ₂ O
diclofenac sodium	acid	15307-79-6	$\geq 99\%$	Cayman Chemical Company	90/10 ACN/H ₂ O
2,4-dichloro-phenoxyacetic acid (2,4-D)	acid	94-75-7	$\geq 97\%$	Cayman Chemical Company	50/50 MeOH/H ₂ O
(S)-naproxen	acid	22204-53-1	$\geq 99\%$	Cayman Chemical Company	50/50 MeOH/H ₂ O
ibuprofen	acid	15687-27-1	$\geq 99.6\%$	Euro OTC Pharma	50/50 MeOH/H ₂ O
torasemide	acid	56211-40-6	$\geq 98\%$	Sigma-Aldrich	50/50 MeOH/H ₂ O
warfarin	acid	81-81-2	$\geq 99\%$	Sigma-Aldrich	50/50 MeOH/H ₂ O
telmisartan	multifunctional	144701-48-4	$\geq 98\%$	Cayman Chemical Company	90/10 ACN/H ₂ O

S2: Instrumental analysis

Table S2: LC parameters for the test chemicals used in this study. The eluents used are composed as follows: A) 5 % acetonitrile and 95 % water, B) 100 % acetonitrile, C) 5 % acetonitrile and 95 % water with 0.1 % formic acid, D) 95 % acetonitrile and 5 % water with 0.1 % formic acid.

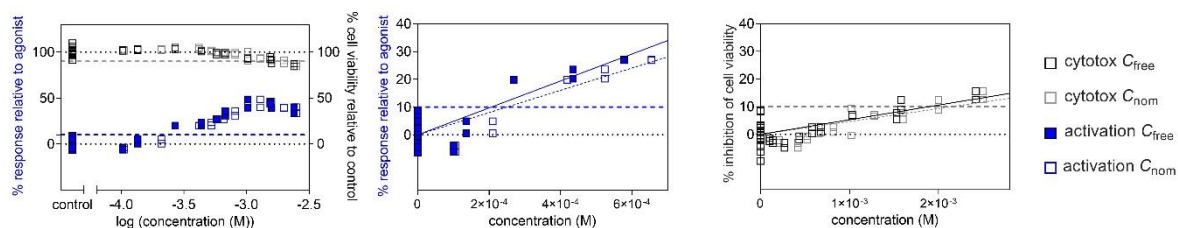
<i>Chemical</i>	<i>Column</i>	<i>Eluent</i>	<i>Retention time [min]</i>	<i>Flow rate [mL/min]</i>	<i>Injection volume [μL]</i>
caffeine	Phenomenex Luna Omega Polar C18 1.6 μ m 50 $\times$ 2.1 mm, 25°C	95 % A 5 % B	1.75	0.5	1
lamotrigine	BioZen peptide PS-C18 1.6 μ m 50 $\times$ 2.1 mm, 25°C	90 % C 10 % D	0.79	0.5	1
diclofenac	Phenomenex Kinetex C18 1.7 μ m 50 $\times$ 2.1 mm, 30°C	50 % C 50 % D	1.27	0.5	1
2,4-D	Phenomenex Kinetex C18 1.7 μ m 50 $\times$ 2.1 mm, 40°C	60 % C 40 % D	0.96	0.5	1
naproxen	Phenomenex Kinetex C18 1.7 μ m 50 $\times$ 2.1 mm, 40°C	55 % C 45 % D	0.83	0.5	PBS: 2 Desorption solution: 15
ibuprofen	Phenomenex Kinetex C18 1.7 μ m 50 $\times$ 2.1 mm, 40°C	45 % C 55 % D	0.98	0.5	5
torasemide	Phenomenex Kinetex C18 1.7 μ m 50 $\times$ 2.1 mm, 40°C	75 % C 25 % D	0.95	0.5	1
warfarin	Phenomenex Kinetex C18 1.7 μ m 50 $\times$ 2.1 mm, 30°C	55 % C 45 % D	1.2	0.5	1
telmisartan	Phenomenex Kinetex C18 1.7 μ m 50 $\times$ 2.1 mm, 40°C	90 % C 10 % D	0.82	0.5	1

Table S3: MS parameters for the test chemicals used in this study.

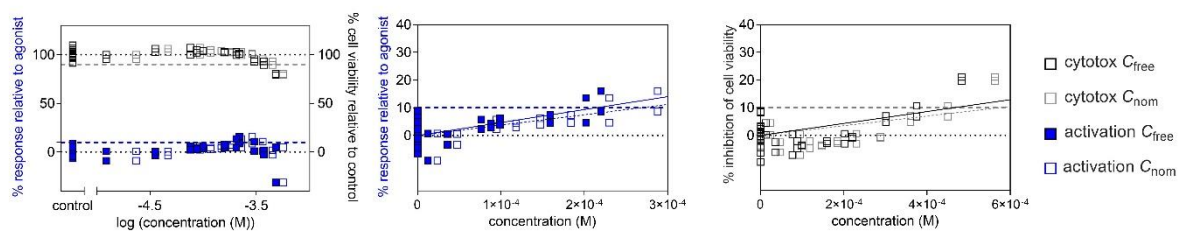
<i>Chemical</i>	<i>Ion source</i>	<i>Frag-mentor voltage [V]</i>	<i>MRM transitions (Collision energy [V])</i>	<i>Source parameters: capillary voltage [V]; gas flow [ml/min]; gas temperature [°C]; nebulizer [psi]</i>
caffeine	ESI+	110	195.2 → 138/110.1 (17/25)	5000; 13; 310; 60
lamotrigine	ESI+	150	256 → 211/145 (26/45)	3000; 13; 290; 50
diclofenac	ESI-	80	294 → 249.8/213.9 (9/21)	2000; 13; 350; 25
2,4-D	ESI-	80	218.96 → 160.9/124.9 (12/28)	2000; 13; 350; 25
naproxen	ESI-	80	229.08 → 185/168.9 (1/33)	5000; 13; 290; 25
ibuprofen	ESI-	80	205.12 → 161.1 (4)	4000; 13; 320; 25
torasemide	ESI+	120	349.14 → 263.9/183 (12/36)	2000; 13; 350; 50
warfarin	ESI+	120	309.12 → 162.9/250.9 (12/16)	4500; 13; 350; 30
telmisartan	ESI+	230	515 → 497/276 (38/40)	2000; 8; 350; 60

S3: Bioassay results

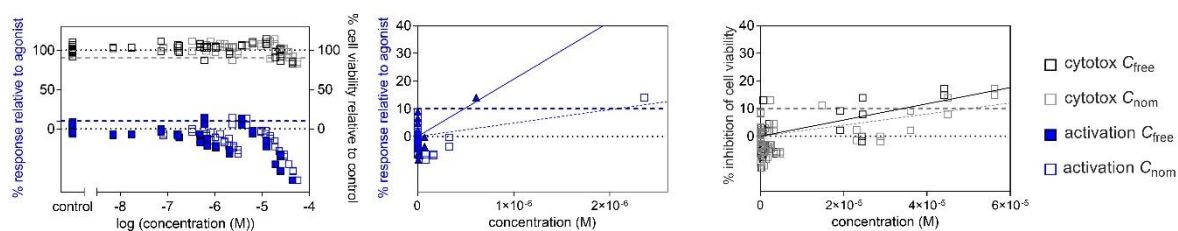
Caffeine



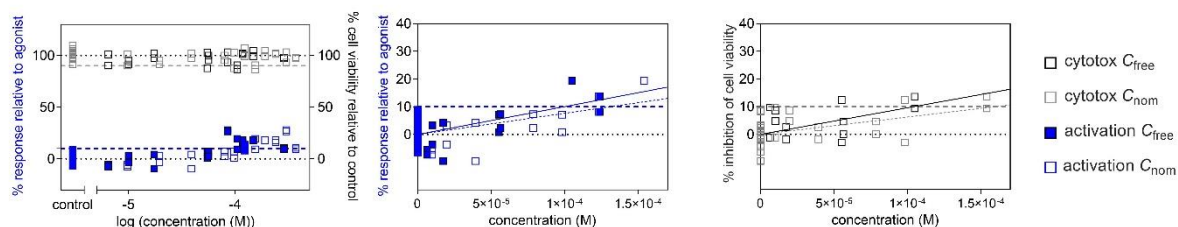
Lamotrigine



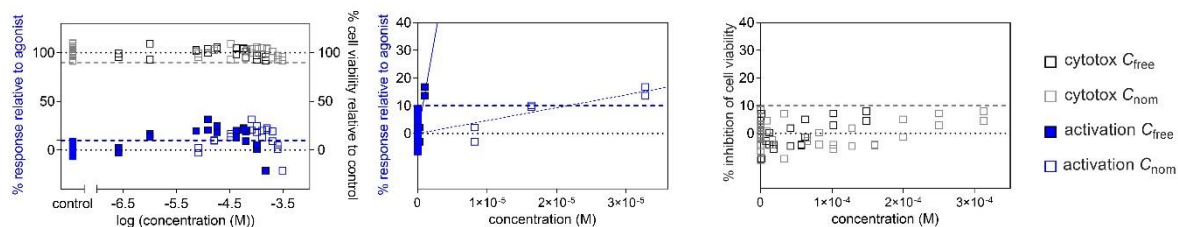
Diclofenac



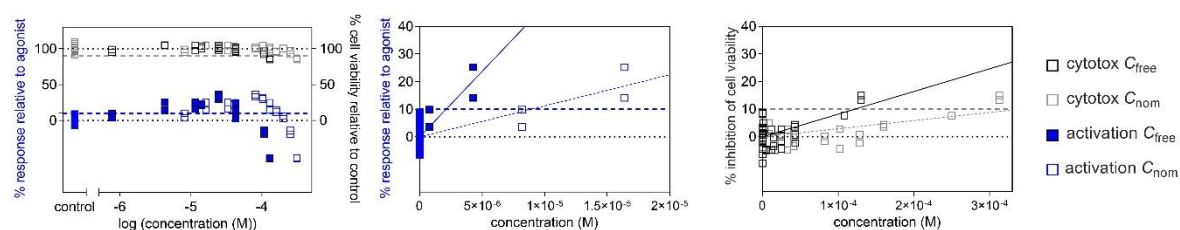
2,4-D



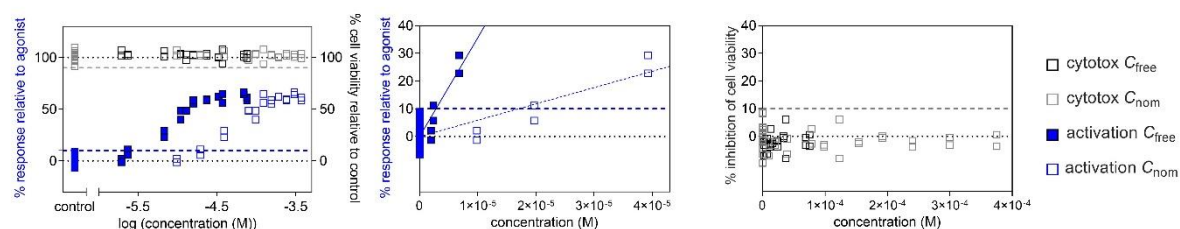
Naproxen



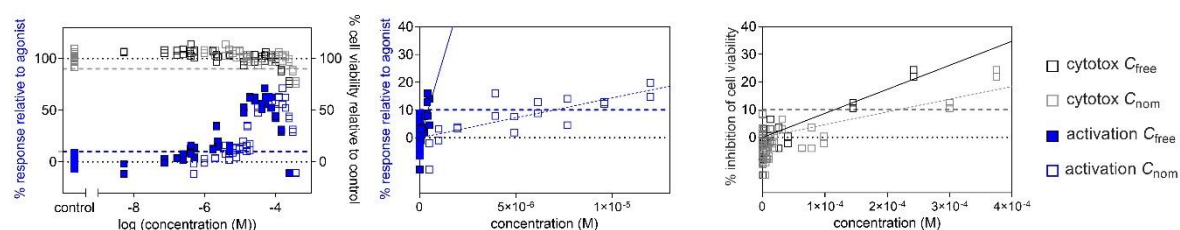
Ibuprofen



Torsemide



Warfarin



Telmisartan

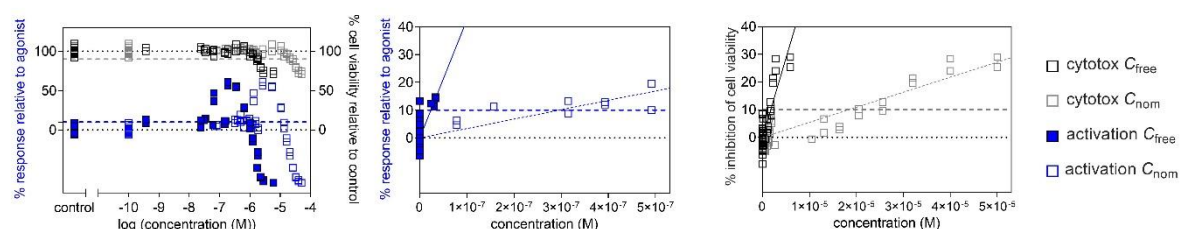


Figure S1: Nominal and freely dissolved concentration-response curves of all chemicals obtained from 384-well plates. [Left] Full concentration-response curve for cell viability and induction of the PPAR γ . [Middle] Linear range of the concentration-response curve for the induction of the PPAR γ at low concentration levels. [Right] Linear range of the concentration-response curve for cytotoxicity

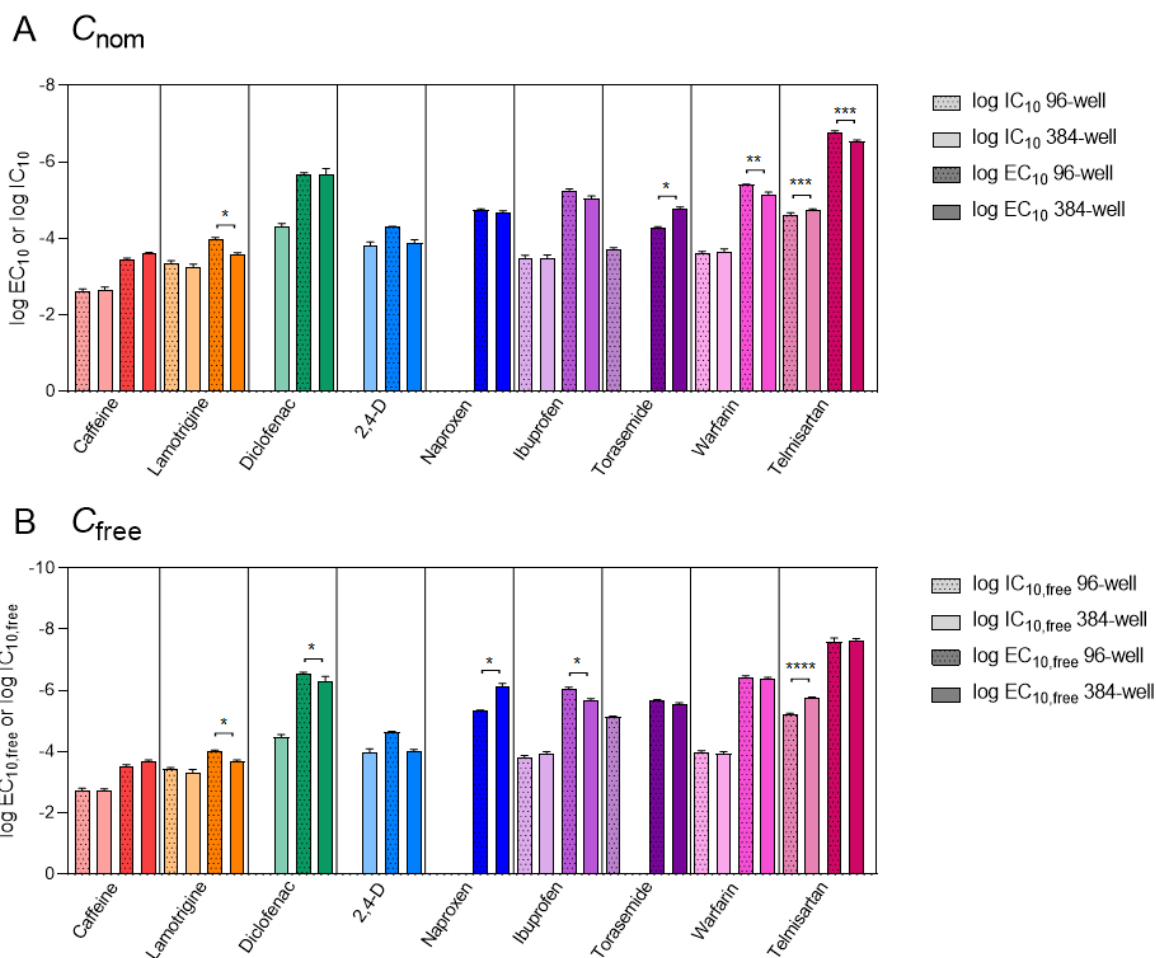


Figure S2: Unpaired t-test of $\log IC_{10}$ and $\log EC_{10}$ of the test chemicals obtained from 96-well plates (Huchthausen et al, 2020) compared to 384-well plates (A). Unpaired t-test of $\log IC_{10,free}$ and $\log EC_{10,free}$ of the test chemicals obtained from 96-well plates (Huchthausen et al, 2020) compared to 384-well plates (B).

Table S4: P-values of unpaired t-test of $\log IC_{10}$, $IC_{10,free}$ and $\log EC_{10}$ or $EC_{10,free}$ of the test chemicals obtained from 96-well plates (Huchthausen et al, 2020) compared to 384-well plates.

<i>Chemical</i>	<i>P-value</i> <i>unpaired t-test</i> <i>96 vs. 384 of</i> <i>IC_{10}</i>	<i>P-value</i> <i>unpaired t-test</i> <i>96 vs. 384 of</i> <i>EC_{10}</i>	<i>P-value</i> <i>unpaired t-test</i> <i>96 vs. 384 of</i> <i>$IC_{10,free}$</i>	<i>P-value</i> <i>unpaired t-test</i> <i>96 vs. 384 of</i> <i>$EC_{10,free}$</i>
caffeine	0.3601	0.0839	0.7278	0.0572
lamotrigine	0.2814	0.0374	0.3713	0.0492
diclofenac		0.9048		0.0380
2,4-D		0.0924		0.0517
naproxen		0.3017		0.0405
ibuprofen	0.7462	0.0910	0.3761	0.0325
torasemide		0.0272		0.1509
warfarin	0.3457	0.0016	0.3059	0.0960
telmisartan	0.0003	0.0004	0.0001	0.4752

S4: References

- Huchthausen, J., Mühlenbrink, M., König, M., Escher, B. I. & Henneberger, L. (2020) Experimental Exposure Assessment of Ionizable Organic Chemicals in In Vitro Cell-Based Bioassays. *Chemical Research in Toxicology*, 33(7), 1845-1854. doi 10.1021/acs.chemrestox.0c00067
- Niu, L., Henneberger, L., Huchthausen, J., Krauss, M., Ogefere, A. & Escher, B. I. (2022) pH-Dependent Partitioning of Ionizable Organic Chemicals between the Silicone Polymer Polydimethylsiloxane (PDMS) and Water. *ACS Environmental Au*, 2(3), 253–262. doi: 10.1021/acsenvironau.1c00056
- Shedlovsky, T. (1962) The behaviour of carboxylic acids in mixed solvents, in Pesce, B. (ed), *Electrolytes*. New York: Pergamon Press, 146–151.
- Yasuda, M. (1959) Dissociation Constants of Some Carboxylic Acids in Mixed Aqueous Solvents. *Bulletin of the Chemical Society of Japan*, 32(5), 429-432. doi: 10.1246/bcsj.32.429