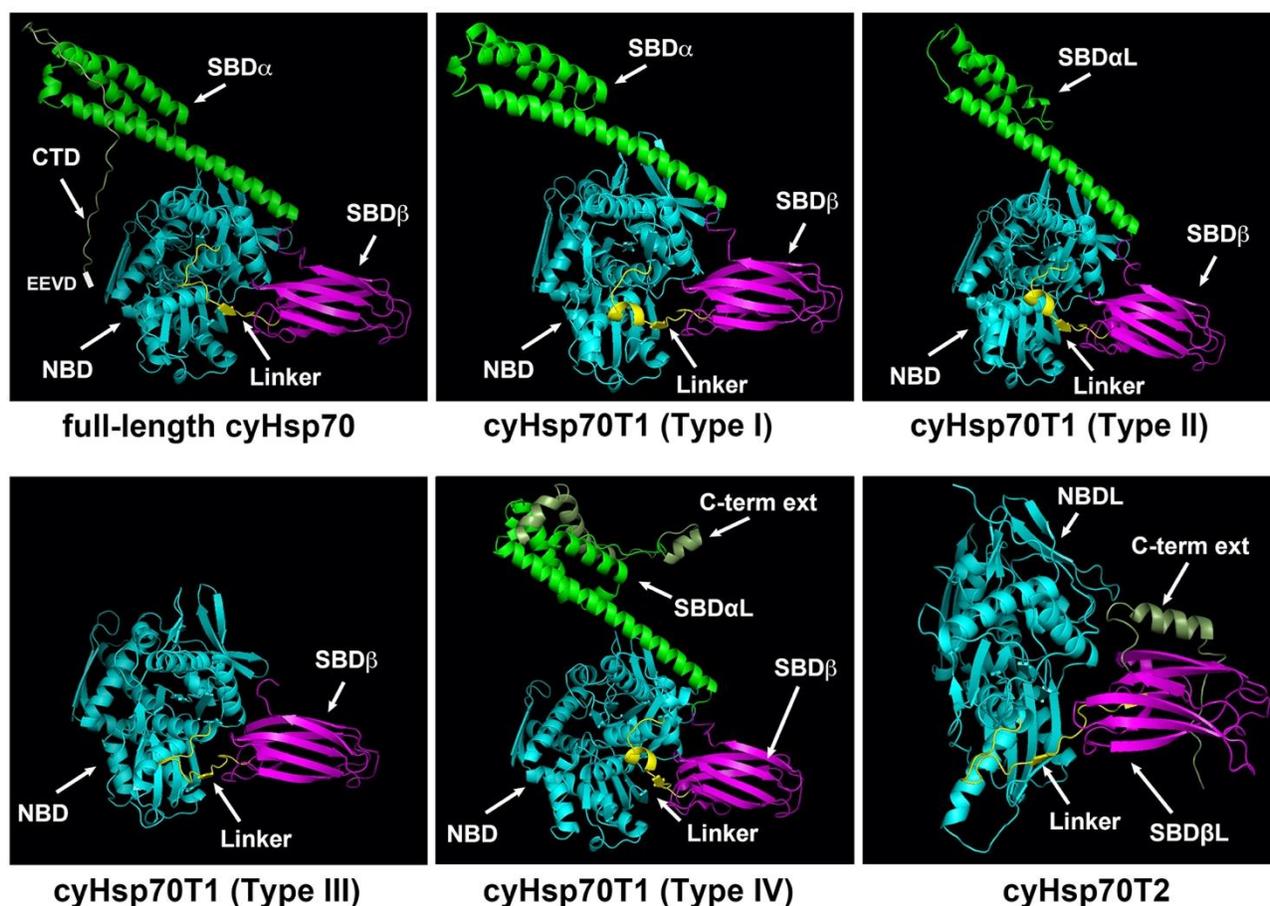
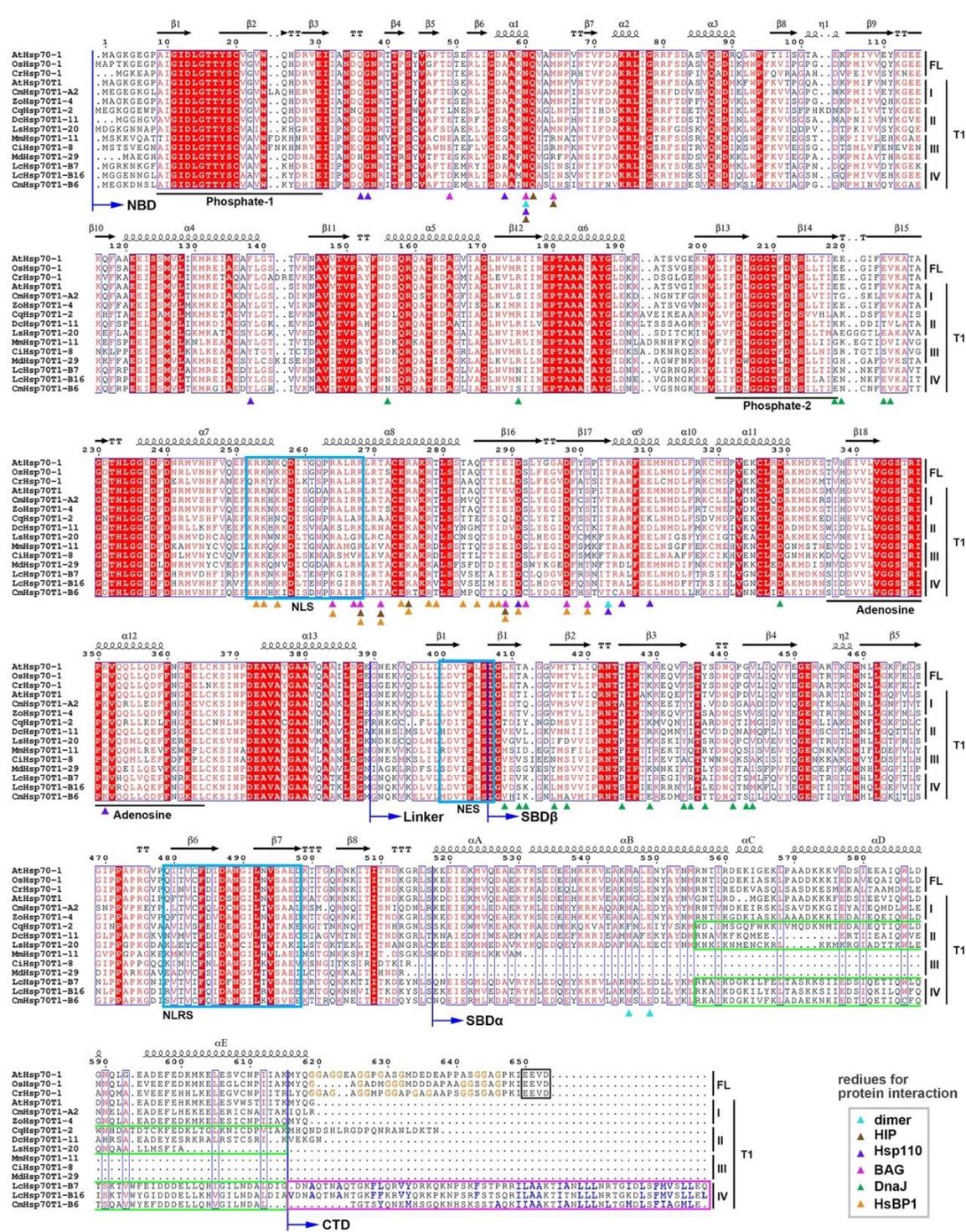


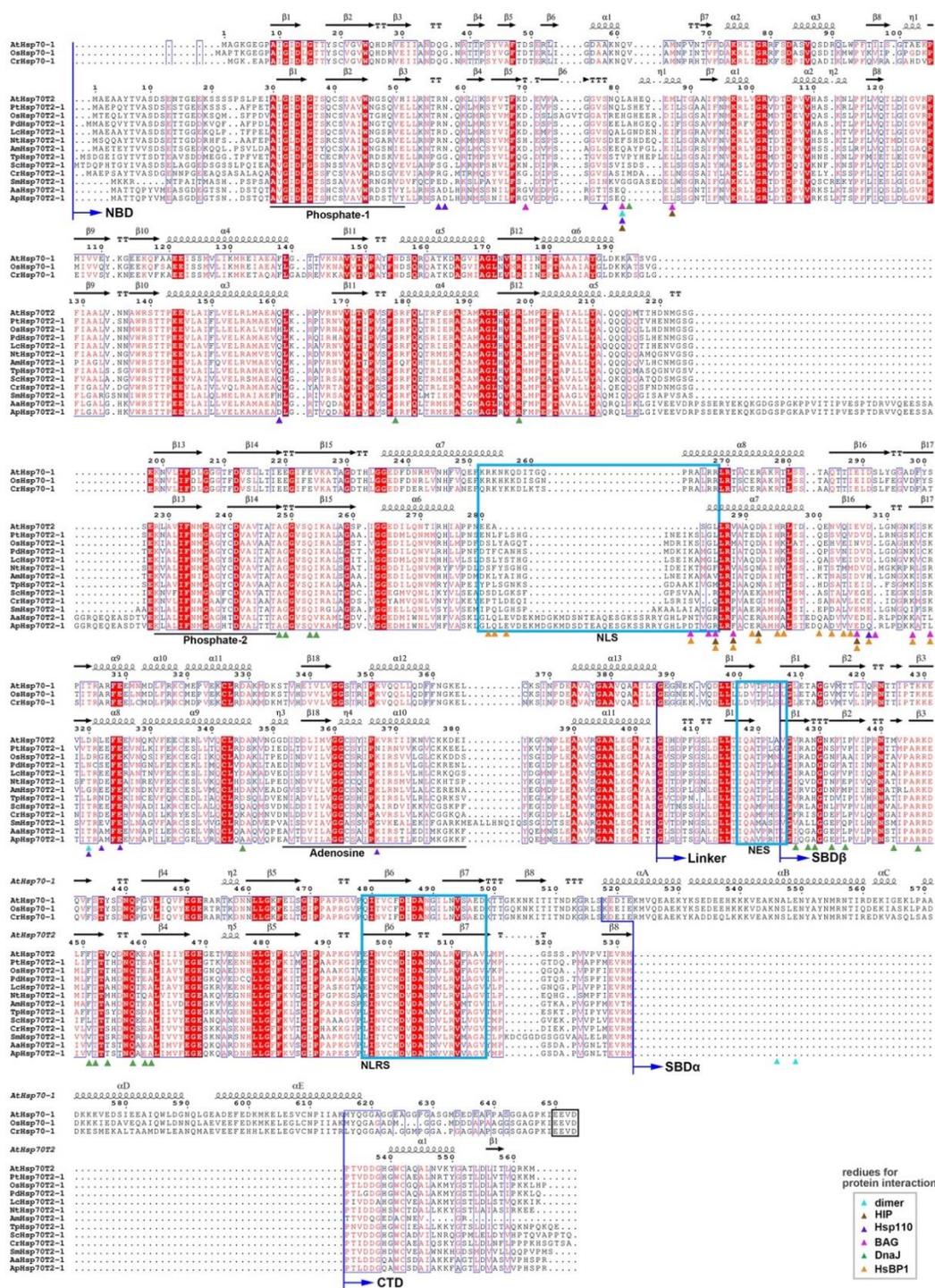
**Supplementary Figure 1. Conserved C-terminal motifs of Hsp70s.** The conserved C-terminal sequence of Hsp70s were determined by aligning the final 20 amino acids residues from the C-terminus using the MUSCLE program. The resulting sequence alignment was visualized as residue frequency at each position, employing the online tool WebLogo (<https://weblogo.berkeley.edu/logo.cgi>). This analysis utilized the protein sequences of Hsp70 obtained from 172 plants and 20 green algae.



**Supplementary Figure 2. Protein 3D modeling of all cyHSP70 types.** The three-dimensional structures of both full-length and truncated cyHSP70 proteins were predicted using the online tool SWISS-MODEL (<https://swissmodel.expasy.org/>). The respective protein IDs from the genomic database and the corresponding SMTL IDs for the modeled proteins in SWISS-MODEL for each cyHSP70 type are provided as follows: full-length cyHsp70: AT5G02500 (*Arabidopsis thaliana*) / A0A0R4J626 (*Glycine max*); cyHsp70T1 (Type I): AT1G56410 (*A. thaliana*) / A0A3Q7Y9C5 (*Cicer arietinum*); cyHsp70T1 (Type II): PaHsp70T1-B14 (*Persea americana*) / A0A1S3TT80 (*Vigna radiata* var *radiata*); cyHsp70T1 (Type III): MD14G0022900 (*Medicago truncatula*) / A0A6P5R6Z9 (*Prunus avium*); cyHsp70T1 (Type IV): LiHsp70T1-B14 (*Litsea cubeba*) / A0A3S4NG45 (*Cinnamomum micranthum* f *kanehirae*); cyHsp70T2: AT2G32120 (*A. thaliana*) / I1JAU2 (*G. max*). Distinct color codes represent the five major regions of Hsp70 (NBD, linker, SBDβ, SBDα and CTD). The term “C-term ext” designates the C-terminal extension. The atypical NBD, SBDβ and SBDα regions are labeled as NBDL, SBDβL and SBDαL, respectively. The conserved C-terminal motif “EEVD” of full-length cyHsp70 is highlighted in white.

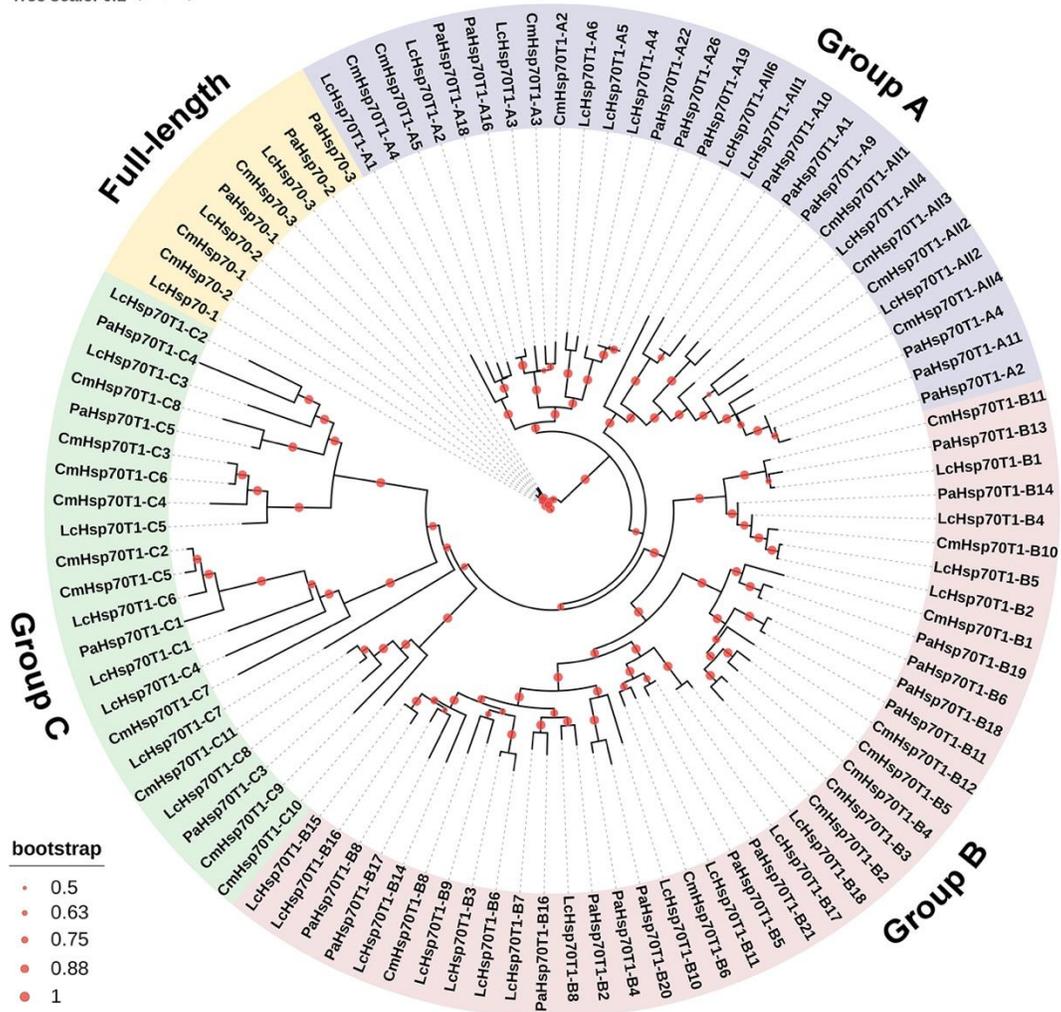


**Supplementary Figure 3. Multiple sequence alignment, molecule binding positions, and structural characteristics of full-length cyHSP70s and the T1 variant of truncated cyHsp70s.** The sequences employed for the alignment correspond to those presented in Figure 2. The secondary structure positions, as predicted in the 3D structure of AtHsp70-1 (refer to Supplementary Figure 2), are indicated. Nucleotide-interacting positions within the Nucleotide-Binding Domain (NBD) are labeled as Phosphate-1, Phosphate-2, and Adenosine, corresponding to  $\alpha$ -phosphate,  $\beta$ -phosphate, and adenosine binding sites, respectively. The regions analogous to SBD $\alpha$  (SBD $\alpha$ L) and the C-terminal extension are highlighted by unfilled green and magenta boxes, respectively.



**Supplementary Figure 4. Multiple sequence alignment, molecule binding positions, and structural characteristics of full-length cyHSP70s and the T2 variant of truncated cyHsp70s.** The sequences analyzed were collected from *A. thaliana*, *Oryza sativa*, *Populus trichocarpa*, *Nicotiana tabacum*, *Phoenix dactylifera*, *Litsea cubeba*, *Amborella trichopoda*, *Thuja plicata*, *Selaginella moellendorffii*, *Salvinia cucullata*, *Anthoceros agrestis*, *Anthoceros punctatus* and *Chlamydomonas reinhardtii*. The secondary structure positions, as predicted in the 3D structure of AtHsp70-1 and AtHsp70T2 (refer to Supplementary Figure 2), are indicated. Nucleotide-interacting positions within the Nucleotide-Binding Domain (NBD) are labeled as Phosphate-1, Phosphate-2, and Adenosine, corresponding to  $\alpha$ -phosphate,  $\beta$ -phosphate, and adenosine binding sites, respectively.

Tree scale: 0.1



**Supplementary Figure 5. Circular phylogenetic tree of cyHsp70T1 sequences in investigated Lauraceae species.** Protein sequences of full-length cyHsp70 and cyHsp70T1 obtained from three Lauraceae species (*Litsea cubeba*, *Cinnamomum micranthum*, and *Persea americana*) were aligned using the MAFFT program. The Neighbor-Joining (NJ) method was employed to generate phylogenetic tree, with support from 1,000 bootstrap resampling. Branches with bootstrap values exceeding 0.5 are indicated by red dots. The sequences of full-length cyHsp70 are labeled as full-length. The scale bar represents 0.1 amino acid substitutions per site.

