

Supplementary Material:

Molecular characterization, localization, and physiological roles of ITP and ITP-L in the mosquito, *Aedes aegypti*

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Oligo Name	Sequence (5' – 3')	Function	Product size
<i>Itp & Itp-l</i> qPCR F	AATCGCACTGGTACCTCTGG	qPCR amplification of <i>Itp</i> & <i>Itp-l</i>	195bp (<i>Itp &</i> <i>Itp-l</i>)
<i>Itp</i> qPCR R	CCAAAACAGCCCTCTTTGCAG	qPCR amplification of <i>Itp</i>	
<i>Itp-l</i> qPCR R	GTGAAGCACGACTTTTTGCAG	qPCR amplification of Itp-l	
<i>Itp</i> dsRNA F	GACTGAATCAAATATGTGTTCCCG	Amplification of ds <i>Itp</i> target	- 201bp
<i>Itp</i> dsRNA R	AACAACGAGTAGCAGTCTTCG	Amplification of ds <i>Itp</i> target	
<i>Itp-l</i> dsRNA F	CCAAACACGTTATAAGCAGCC	Amplification of ds <i>ltp-l</i> target	- 486bp
<i>Itp-l</i> dsRNA R	ACGTCAAGATTTCCGGATGG	Amplification of ds <i>ltp-l</i> target	
<i>Egfp</i> dsRNA F	ACTCGTGACCACCCTGACCTACG	Amplification of ds <i>Egfp</i> target	- 324bp
<i>Egfp</i> dsRNA R	AGATCTTGAAGTTCACCTTGATGCC	Amplification of ds <i>Egfp</i> target	

Table S1. Gene-specific primer information used for amplification of *A. aegypti Itp* and *Itp-l* for RTqPCR analysis, FISH probe synthesis and dsRNA synthesis. T7 promoter sequence is italicized.



Figure S1. *Aedae*ITP and *Aedae*ITP-L preabsorbed controls and no primary controls. Central nervous system tissues were incubated in either (A,B) anti-*Aedae*ITP/ITP-L primary antiserum preincubated with 10 μ M antigen or (C,D) no primary control. No staining was observed in the nervous tissues including the brain (A,C) or abdominal ganglia (B,D). Scale bars: (A,C) 200 μ M; (B,D) 100 μ M.

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Figure S2. Primer sets mapped to *A. aegypti Itp* and *Itp-l* transcripts, and sequence alignment of *Aedae*ITP and *Aedae*ITP-L deduced peptides. (A) The *Itp* transcript coding sequence spans three exons, and (B) *Itp-l* spans four exons, denoted in purple. Targets for dsRNA synthesis included regions bracketed by primers shown in yellow and qPCR analysis shown in pink. Amino acid region encoding the antigen target sequence of custom *Aedae*ITP/ITP-L primary antisera is shown in red. The 3' end of common exon for *Itp* and *Itp-l* is shown in green. Top letters indicate nucleotides (positions of which are denoted by numbers above). Second panel of letter indicates the translated sequence. (C) Aligned amino acid sequences of ITP and ITP-L from *A. aegypti*, ITP (Genbank: AY950504), and ITP-L (Genbank: AY950506). Highlighting of residues indicates % identity with grey denoting 100% sequence identity.



A. aegypti. primers were designed over a common exon 2 restifting in the knockdown of both Aedaellp and AedaeItp-l transcripts, whereas dsAedaeItp^{2-l} primers were designed over a²⁰ right que exon ³ targeting only the AedaeItp-l transcript. One day old adult male (B,E,G) and female (C,F,H) mosquitoes were injected with ds *AedaeItp*, ds *AedaeItp* $\frac{3}{4}$ or $\frac{1}{6}$ trol ds *Egfp* (D) and collected four (B,C) six (E,F) and eight (G,H) days post-injection to determine knock down efficiency. Transcript evels are shown $\frac{2}{7}$ relative to control ds Egfp mosquitoes $\underline{\breve{a}}(\underline{\breve{b}})^{1}$ ds \overline{E} njected animas compared to n_infiected T animals. Data shown as mean±SEM; Ene-way DVA with Mukey's multiple partson, asterisk (*) denotes significant knockdown, $p_{\pm}^{0.05}$, plogical replicates ens denot b statistical significance). 0.0 TP.1 TP2 <? R R R R R R R

dsEGFP

ds/TP

ds/TP-L

dsEGFP

ds/TP

ds/TP-L



Figure S4. Immunolocalization of *Aedae*ITP and *Aedae*ITP-L in the central nervous system of adult *A. aegypti* mosquitoes following ds*AedaeItp* and ds*AedaeItp-l* injection. *Aedae*ITP- and *Aedae*ITP-L-like immunoreactivity is abolished in (A) ds*AedaeItp*-injected male and (B) female brain, (C) abdominal ganglia and (D) terminal ganglion. (E) ds*AedaeItp-l* treated male and (F) female brain reveal immunoreactivity in four pairs of neurosecretory cells (indicated by white arrowheads), with axonal processes emanating anteriorly towards release sites (indicated by empty arrowheads). (G) No immunostaining was observed in the abdominal ganglia following ds*AedaeItp-l* treatment whereas (H) one pair of lateral anterior cells were observed in the terminal ganglion. Scale bars: (A,B,E,F) 200 μ M; (C,D,G,H) 100 μ M.



dsEGFP-M + dsEGFP-F

Non-injected

dsEGFP-M

dsEGFP-F



Figure S6. Effect of control ds*Egfp* on blood feeding, egg laying, and larval hatching (egg viability) in comparison to non-injected adult *A. aegypti*. The effect of control ds*Egfp* knockdown was tested on (A) preference for blood feeding, (B) weight of blood fed female before and after egg-laying, (C) number of eggs laid, and (D) percentage of larval hatching. Abbreviations: non-injected (NI), male (M) and female (F). Data labeled with different letters are significantly different from non-injected adults (mean±SEM; one-way ANOVA with Bonferroni multiple comparison, p<0.05, n=6–12 mating replicates, each point represents individual replicate values), (ns denotes no statistical significance).



Figure S7. Representative images of immunofluorescent nuclei of total spermatozoa in the male seminal vesicle and testes, and female spermathecae of adult *A. aegypti* following RNAi (dsRNA)-mediated knockdown of *Aedaeltp* or *Aedaeltp-l*. Images of immunofluorescent nuclei (DAPI) from fixed spermatozoa in 1 µl droplet of DPBS with sperm from the seminal vesicle (A-G), testes (H-N) and spermatheca (O-U) of ds*Aedaeltp* or ds*Aedaeltp-l* injected animals from different mating treatments. Abbreviations: male (M) and female (F). Scale bar: 200 µm.



Figure S8. Total spermatozoa in the male testes and seminal vesicle and female spermathecae of adult *A. aegypti* following ds*Egfp* injection. Number of spermatozoa per adult four-days post ds*Egfp* injection and non-injected four-day old adults (mean \pm SEM; one-way ANOVA with Bonferroni multiple comparison, p<0.05, n=8–11 mating replicates, each point represents individual replicate values), (ns denotes no statistical significance).