Supplementary Material

Binary Stress Induces an Increase in Indole Alkaloid Biosynthesis in *Catharanthus roseus*

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1.1. Supplementary Tables

Supplemental Tal	ole 1. List of	primers used	for aRT-PCR	experiments

Gene	Forward primer (5' to 3')	Reverse primer (5' to 3')		
tdc	TCCGAAAACAAGCCCATCGT	AAGGAGCGGTTTCGGGGGATA		
g10h	TGAATGCTTGGGCAATTGGA	GCAAATTCTTCGGCCAGCAC		
sls	GTTCCTTCTCACCGGAGTTG	CCCATTTGGTCAACATGTCA		
str	ACCATTGTGTGGGAGGACAT	ATTTGAATGGCACTCCTTGC		
sgd	TCACAAAGCTGCTGTGGAAG	CACCCGTTGTTAATGGCTCT		
orca3	CGAATTCAATGGCGGAAAGC	CCTTATCTCCGCCGCGAACT		
t16h	AGGACCTTGTTGATGTGCTAC	CATTGCCCAATCGACTGTTG		
d4h	TACCCTGCATGCCCTCAACC	TTGAAGGCCGCCAATTTGAT		
dat	GACCTAGTCCTTCCCAAACG	CCTCCATCAGCAACTTTGTG		
rps9	GAGGGCCAAAACAAACTTGA	CCCTTATGTGCCTTTGCCTA		

Supplemental Table 2. Quantitative analysis of alkaloid contents in C. roseus under different UV-B irradiation times with 72 h dark incubation

Group*	Strictosidine (mg/g)	Vindoline (mg/g)	Catharanthine (mg/g)	Ajmalicine (mg/g)
Control	0.384±0.016	3.924±0.046	3.738±0.034	0.148±0.022
1	2.411±0.050	4.713±0.058	4.448±0.030	0.624 ± 0.040
2	1.428 ± 0.050	4.849 ± 0.070	3.926±0.044	0.571±0.028
3	1.027 ± 0.054	4.174±0.022	3.103 ± 0.038	0.582 ± 0.022

* Control: no UV-B irradiation and 72 h dark incubation; 1: 1 h UV-B irradiation and 72 h dark incubation; 2: 2 h UV-B irradiation and 72 h dark incubation; 3: 4 h UV-B irradiation and 72 h dark incubation

Supplemental Table 4.	Contents of photo	synthesis pigment	between control	and treated group
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	Chla (mg/g)	Chlb (mg/g)	Car (mg/g)	Tchl (mg/g)	Rchl (mg/g)
Control	1.031±0.030	0.380±0.017	0.173±0.002	1.411±0.047	2.711±0.041
Treated	1.071 ± 0.008	0.401 ± 0.010	0.171 ± 0.001	1.472 ± 0.017	2.669 ± 0.056



1.2. Supplementary Figures



Supplementary Figure 1. Experimental design of this proteomic study. The *C. roseus* plantlets were treated with UV-B irradiation for 1 h and dark incubation for 72 h. Leaves were collected, and proteins, alkaloids, total RNA were extracted from leaves respectively. And the content of alkaloid was analyzed by HPLC. The expression of key genes in the alkaloid biosynthesis pathway were analyzed by qRT-PCR. The proteins were used for gel-free analyses. Three independent experiments were performed.



Supplementary Figure 2. HPLC chromatogram of the standards. The standards of strictosidine, vindoline, catharanthine, and ajmalicine were dissolved in methanol and diluted at six different concentrations to make calibration curve. Peaks were measured at 220 nm. (A) UV spectra with strictosidine; (B) UV spectra with vindoline; (C) UV spectra with catharanthine; (D) UV spectra with ajmalicine.



Supplementary Figure 3. Principle component analysis of the peptide profile of total proteins. The datasets of *C. roseus* proteins not exposed to UV-B irradiation plus 72 h of dark incubation, and following 1 h of UV-B irradiation plus 72 h of dark incubation were plotted using PCA. PCA was fully integrated into the SIEVE software. The blue plot was a control group which was treated only by 72 h of dark incubation. The red plot was a treated group, which was irradiated by UV-B for 1h and 72 h of dark incubation.

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Supplementary Figure 4. The biosynthetic pathway of indole alkaloids in C. roseus. The induced

alkaloids were marked by blue blocks. The alkaloid content is of alkaloids are shown in Supplemental Table 2. The abbreviations used are as follows: *tdc*, tryptophan decarboxylase; *g10h*, geraniol-10-hydroxylase; *10-hgo*, 10-hydroxygeraniol oxidoreductase; *sls*, secologanin synthase; *str*, strictosidine synthase; *sgd*, strictosidine β -D-glucosidase; *cr*, cathenamine reductase; *t16h*, tabersonine 16-hydroxylase; *16omt*, 16-hydroxytabersonine-16-O-methytransferase; *d4h*, deacetoxyvindoline 4-hydroxylase; *dat*, deacetylvindoline O-acetyltransferase.