

Methane-derived microbial biostimulant reduces greenhouse gas emission and improves rice yield

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Fig. S1A

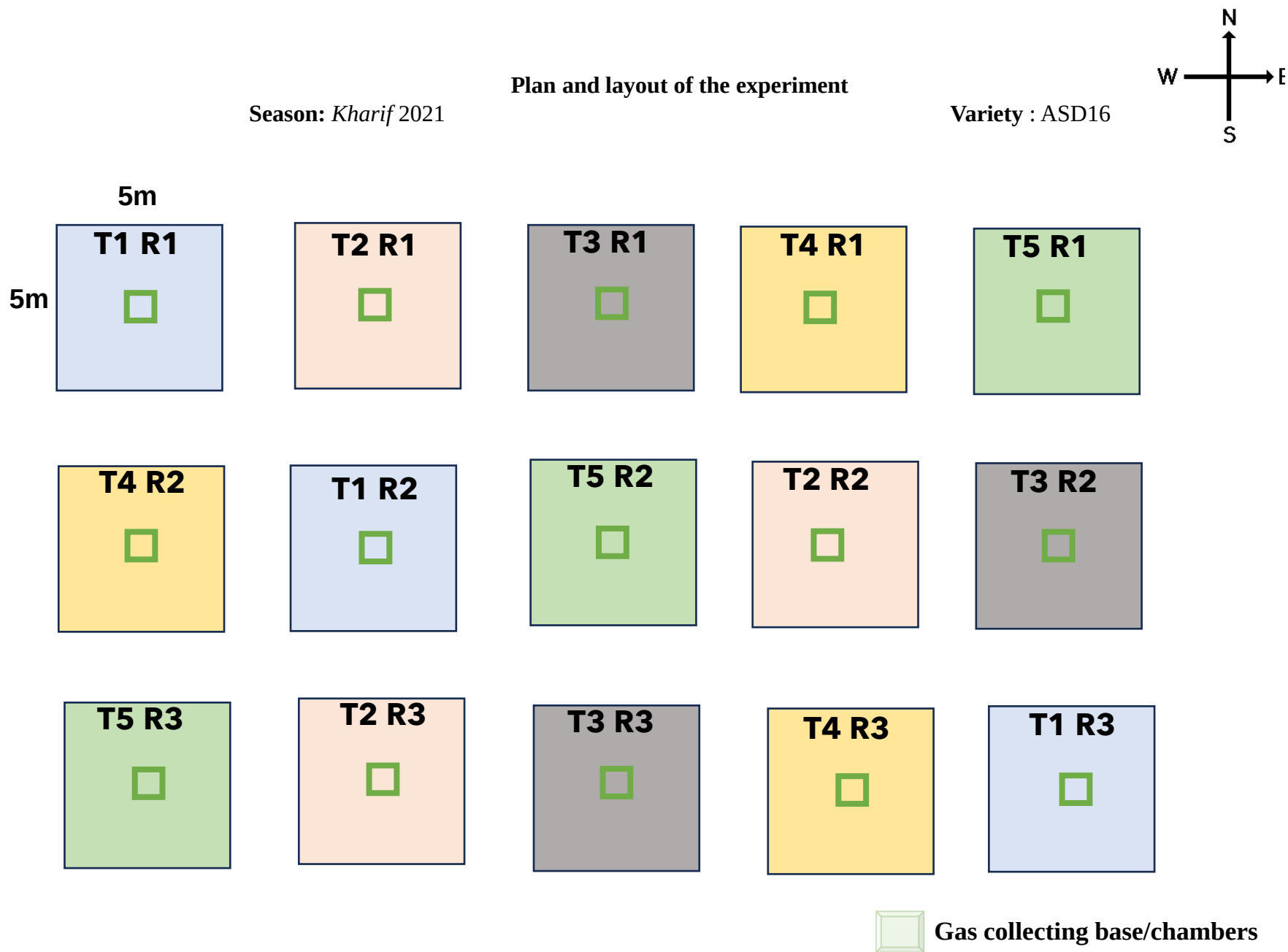


Fig S1A-Experimental field layout for season I testing- Experimental field layout for season I testing- Treatment details are below: Control (T1); 10 mL/L dose of methane- derived microbial biostimulant under two different conditions (T2 & T3), T4 is 75% N control and T5 is 75%N+ 10 mL/L dose of methane- derived microbial biostimulant. First application was seedling root dipping for 20 min for T2, T3 and T5. For T3, second application was given as soil spray whereas in T3 & T5 it was foliar spray. T4 is 75% N control and T5 is 75%N+ 10 mL/L dose of methane derived microbial biostimulant. R1, R2 and R3 respectively corresponds to replication 1, 2 and 3. Small green box indicate position of gas collection base & chambers.

Fig. S1B

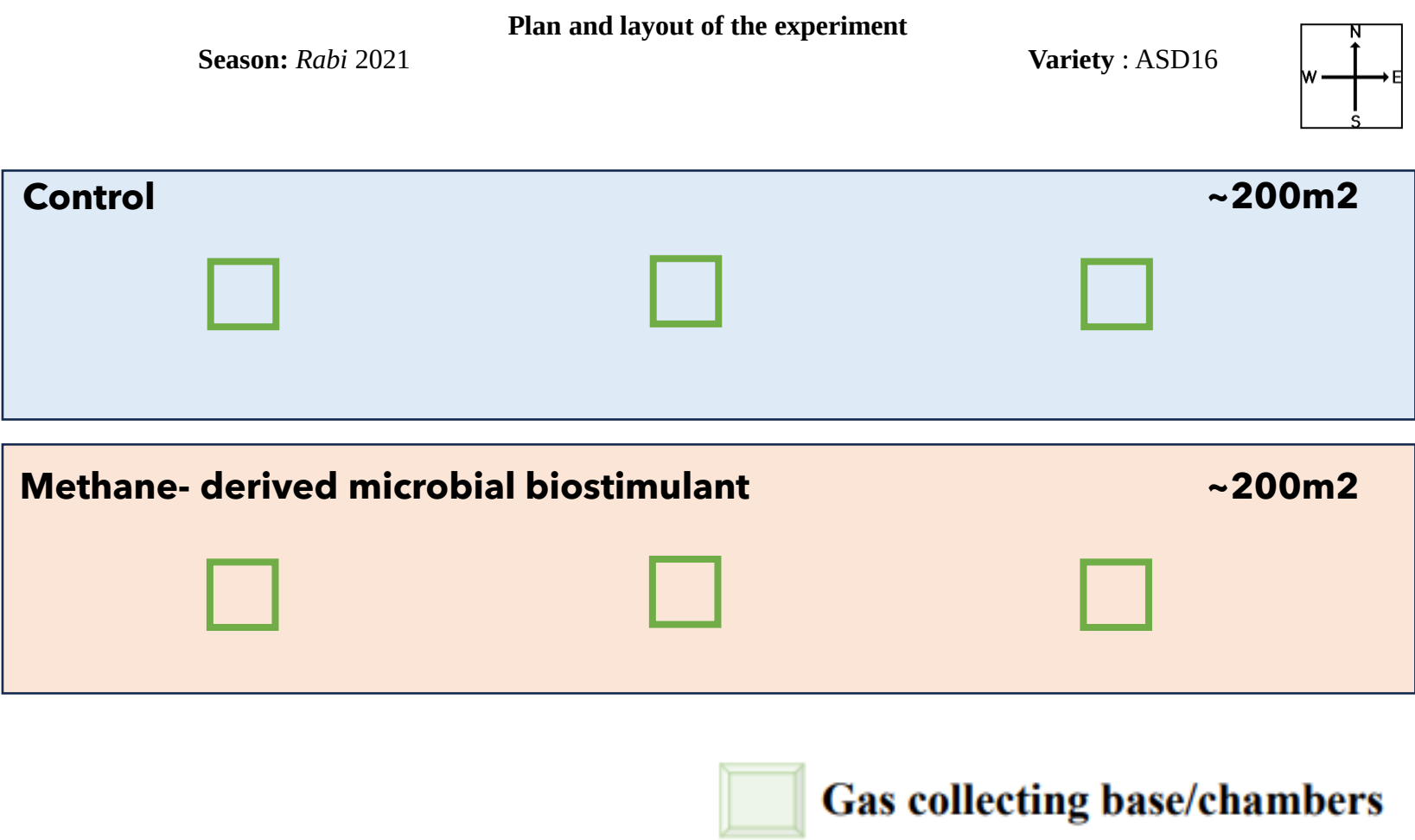


Fig S1B- Experimental field layout for season II testing- Treatment details are below: Control (T1) and methane- derived microbial biostimulant- 10mL/L (T2). Seedling root dipping during transplantation and foliar application during tillering stage and panicle development stage was followed for the application of microbial biostimulant. Control plants received water spray at the same time. Small square box indicates position of gas collection base & chambers.

Fig. S1C

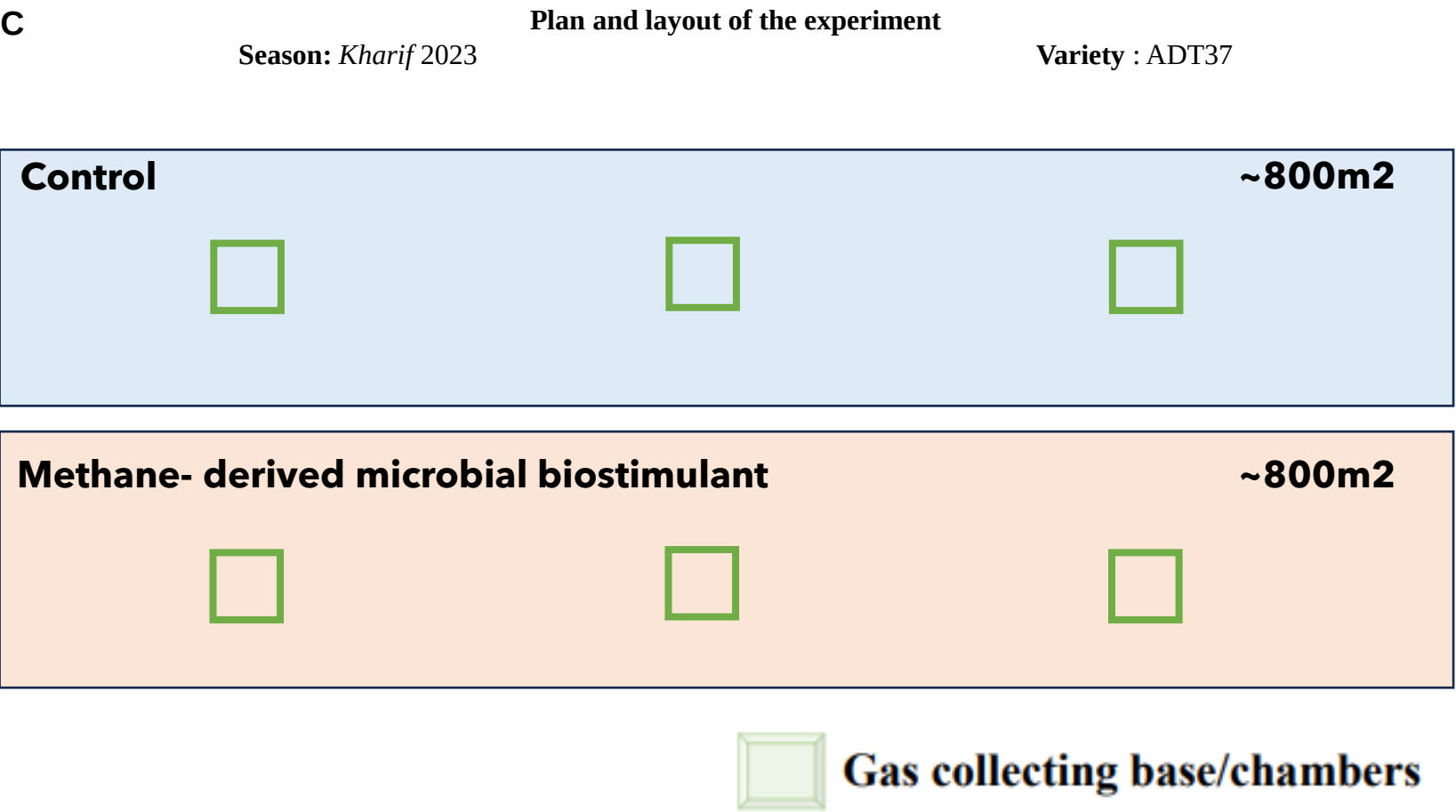


Fig S1C- Experimental field layout for season III testing - Treatment details are below: Control (T1) and methane- derived microbial biostimulant- 10mL/L (T2). Seedling root dipping during transplantation and foliar application during tillering stage and panicle development stage was followed for the application of microbial biostimulant. Control plants received water spray at the same time. Small square box indicates position of gas collection base & chambers.

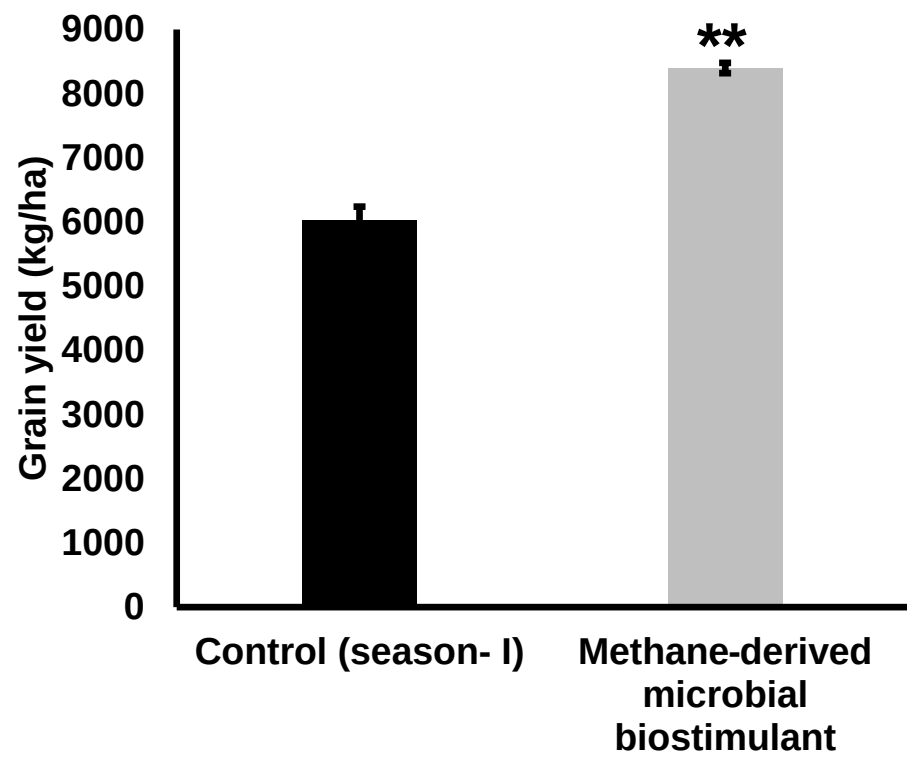


Figure S2A- Influence of methane- derived microbial biostimulant on grain yield- Second application was given as soil spray instead of foliar spray. Differences were evaluated using the two-tailed Student's *t* test and significant differences at *P* < 0.01 is represented by “**”.

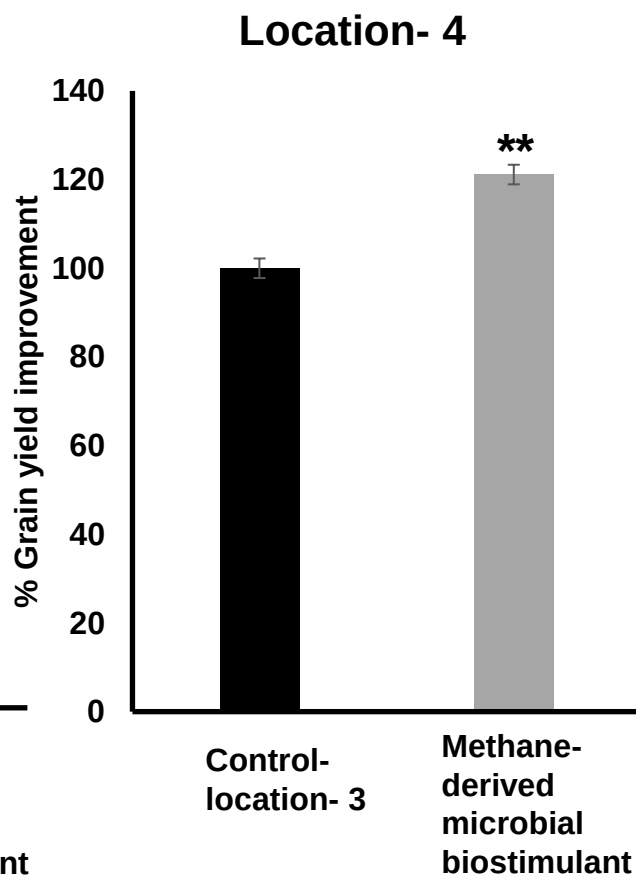
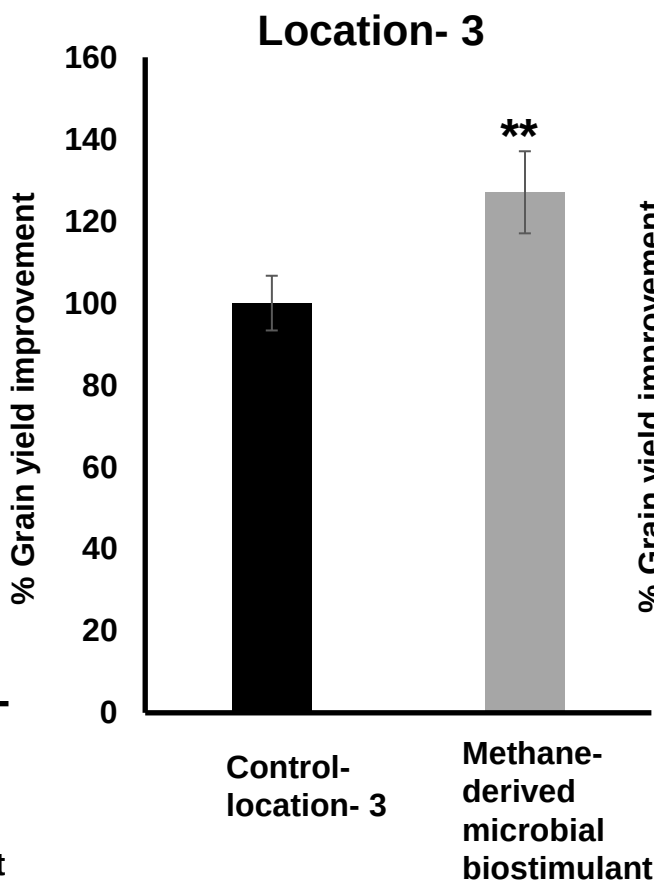
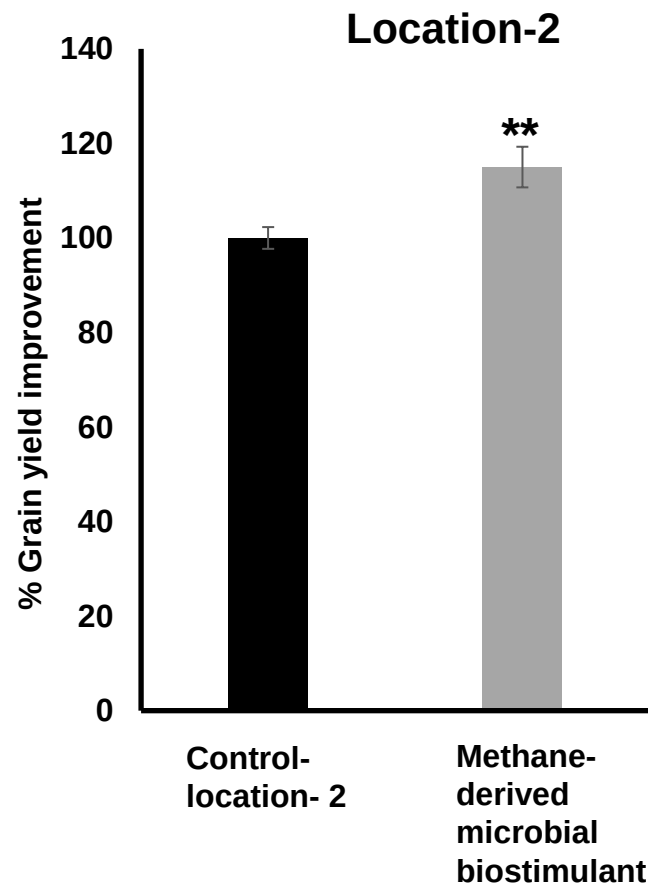


Figure S2B-D- Multilocation microbial biostimulant validation data- Grain yield improvement mediated by methane- derived microbial biostimulant under different agro-ecological locations in India. Differences were evaluated using the two-tailed Student's *t* test and significant differences *P* < 0.01 is represented by “***”.

S3A

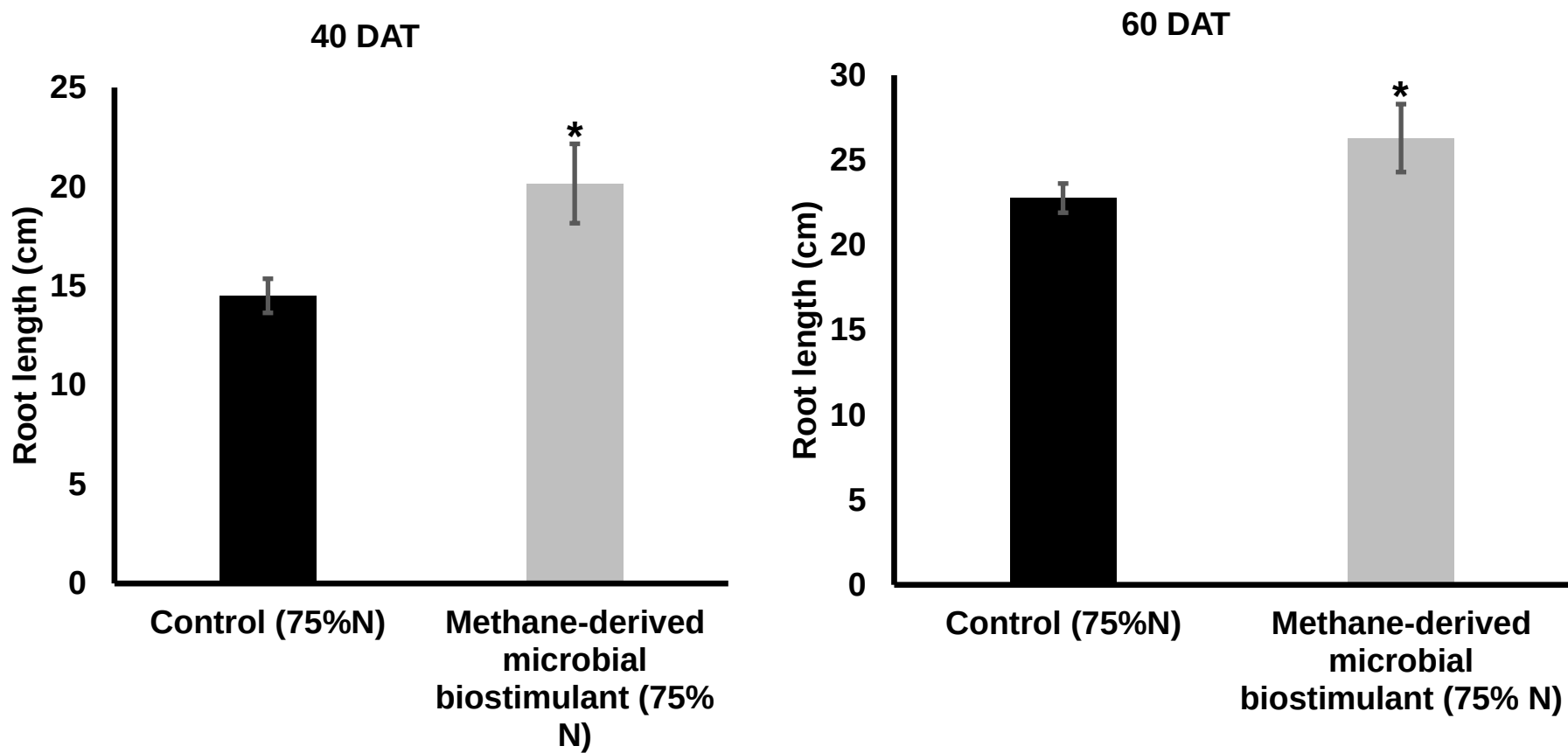


Figure S3A- Effect of microbial biostimulant on root length at 75% N fertilizer- Seedling root dipping was performed in paddy roots with microbial biostimulant. Five independent plants were uprooted from each treatment and root length was measured at 40 DAT and 60 DAT. Student's t-test Significant differences at $P < 0.05$ is represented by “*”.

S3B

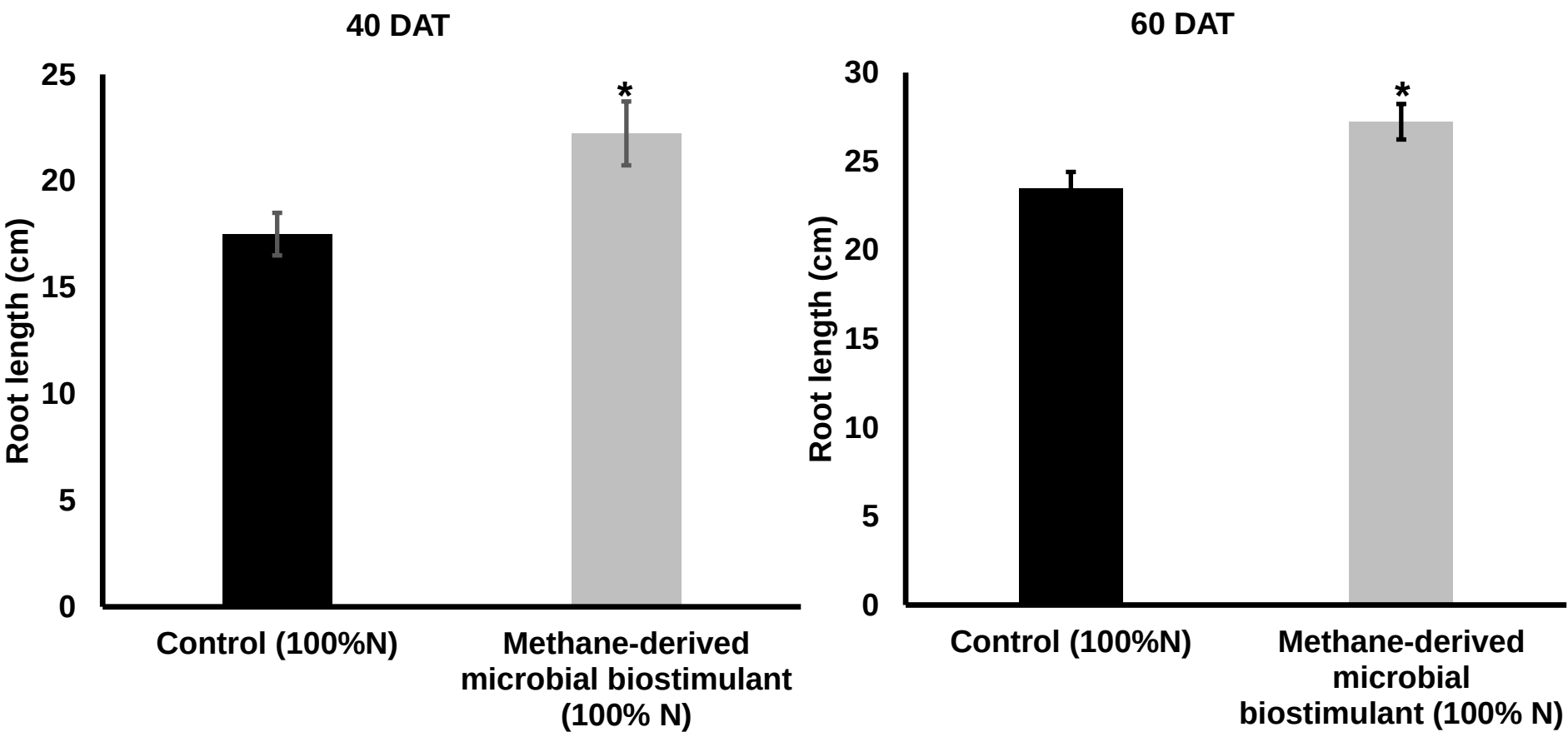
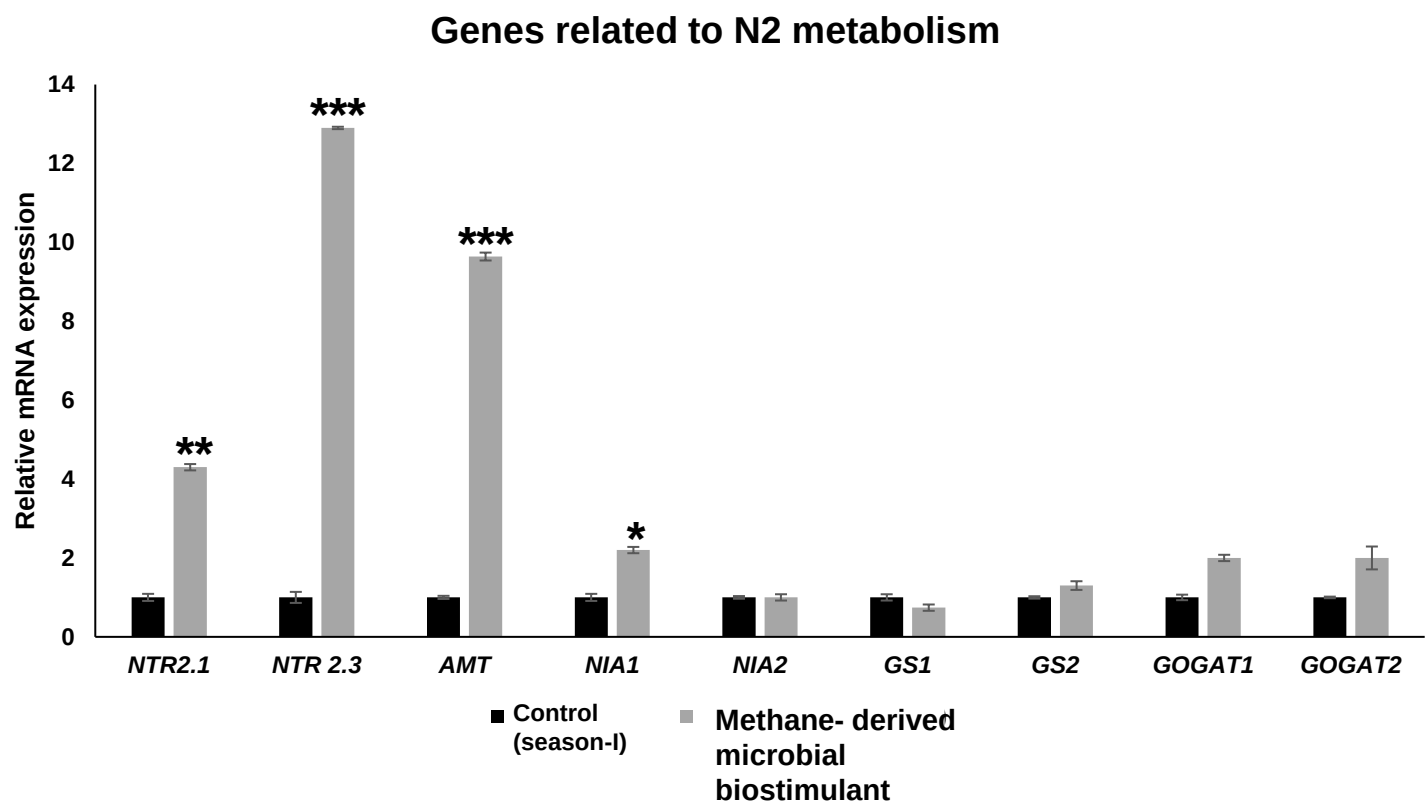


Figure S3B- Effect of microbial biostimulant on root length at 100% N fertilizer- Seedling root dipping was performed in paddy roots with microbial biostimulant. Five independent plants were uprooted from each treatment and root length was measured at 40 DAT and 60 DAT. Student's t-test Significant differences at $P < 0.05$ is represented by “*”.

S4A



S4B

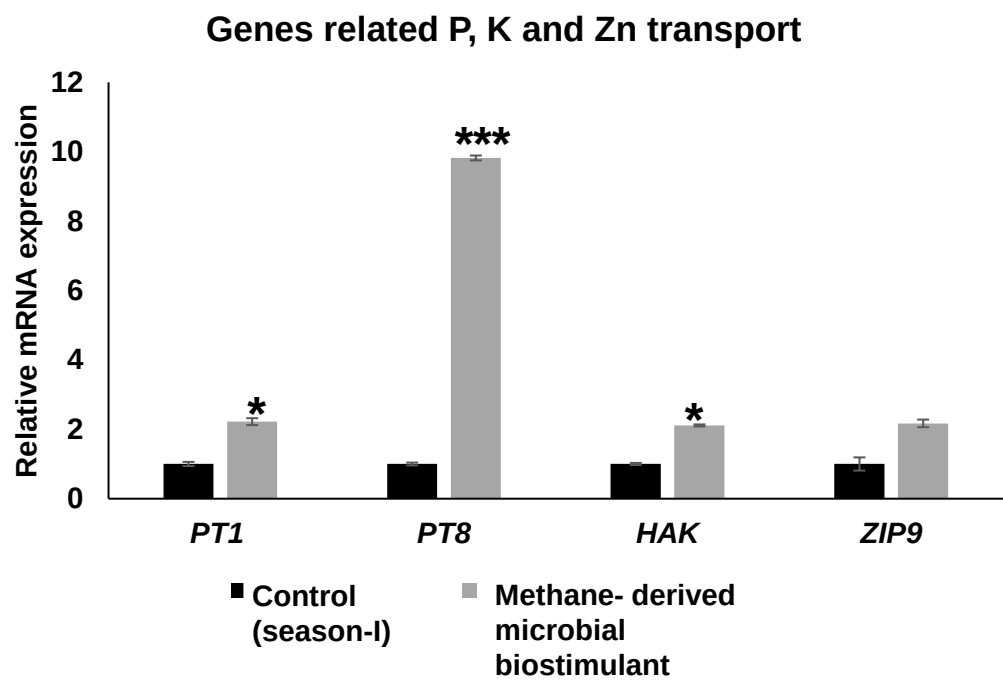


Figure S4- Influence of methane- derived microbial biostimulant on expression of root nutrient uptake and transporter genes- qRT-PCR analysis showing the expression of genes related to macronutrient transport and metabolism in roots of microbial biostimulant treated plants. Expression levels of genes were normalized to the endogenous reference gene actin and are represented relative to respective control roots, which was set to 1. Pooled root samples from control and microbial biostimulant treated roots used for RNA extraction. The results shown are from three independent experiments. Error bars indicate mean $\pm$ SE. Student's t-test: significant differences at $P < 0.05$, $P < 0.01$ and $P < 0.001$ are represented by *, ** and ***, respectively.

Nitrate transporter (*NRT*); Ammonium transporter (*AMT*); nitrate reductase (*NIA*); Glutamine synthetase (*GS*); glutamate synthase (*GOGAT*); Phosphate transporter (*PT*); High affinity potassium transporter (*HAK*); Zinc transporter (*ZIP*).

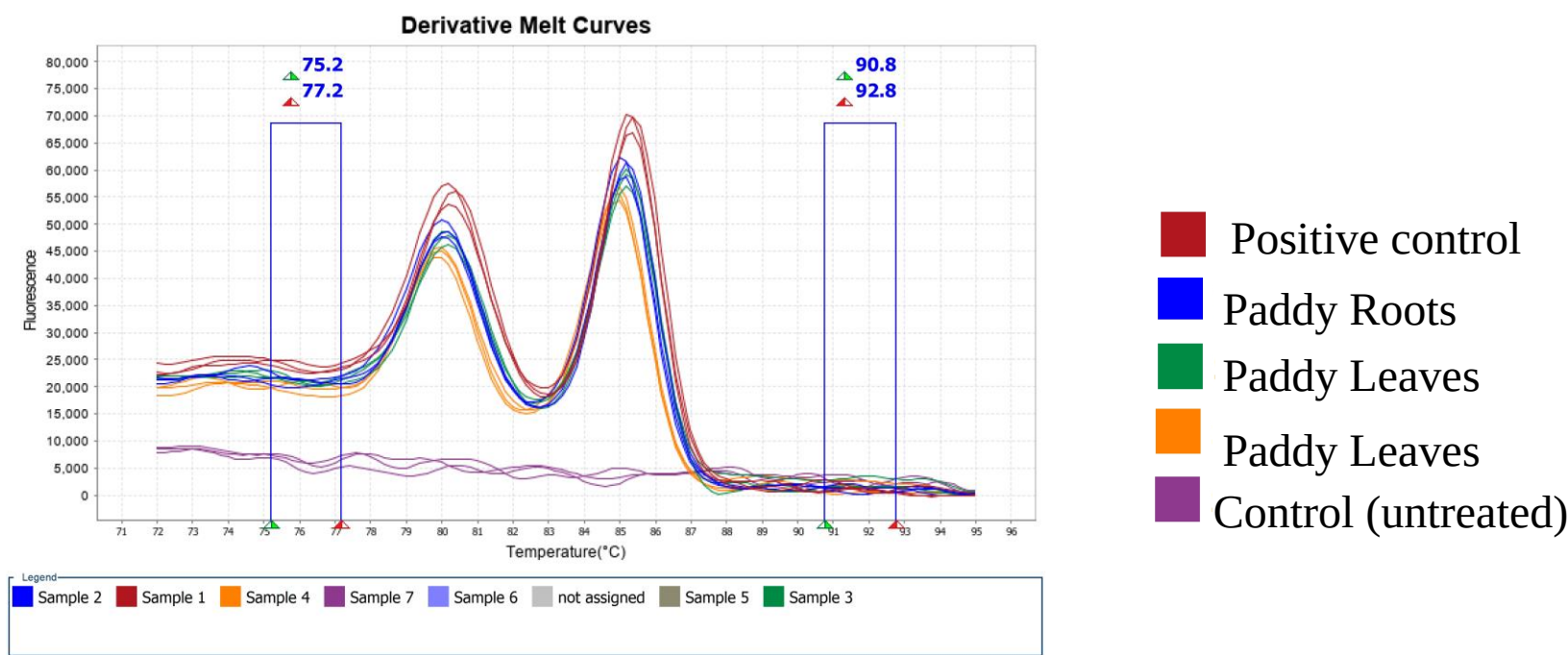


Figure S5- qRT-PCR based derivative melt curve analysis showing the presence of *M. capsulatus* in paddy roots and leaves.

Fig 6A

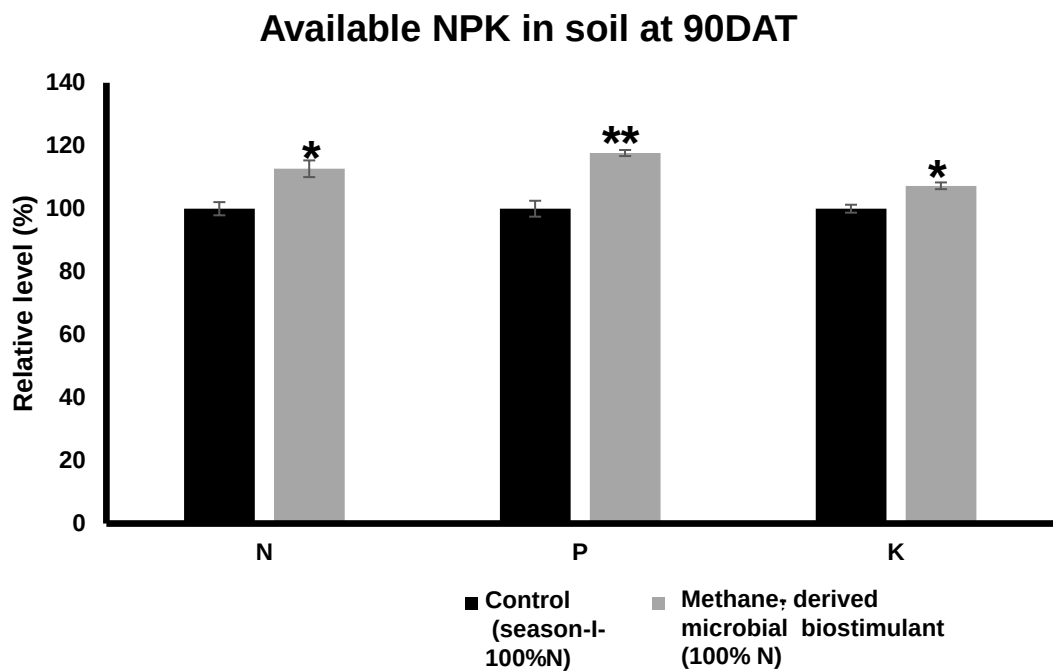


Fig 6B

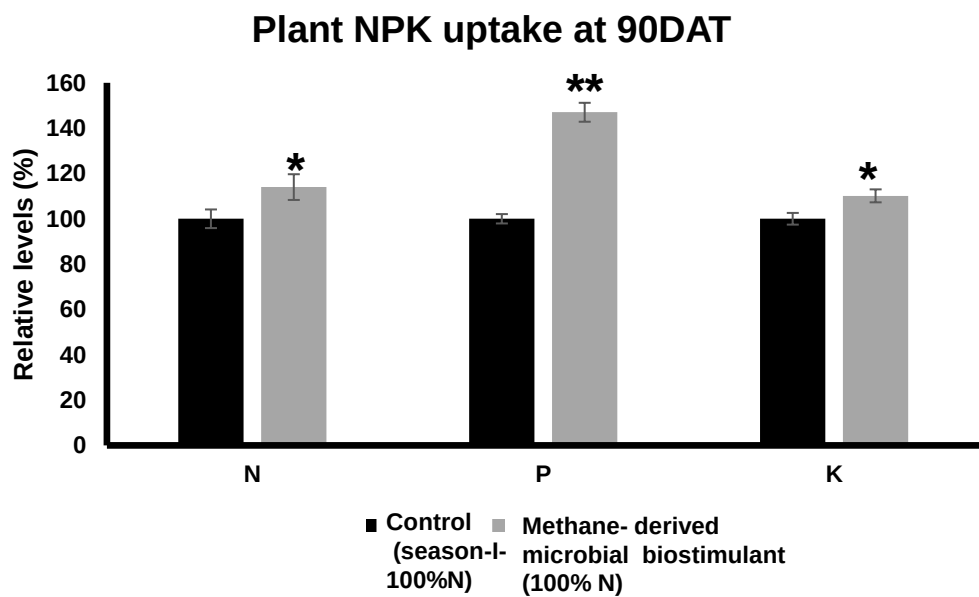


Figure S6A&B- Soil and plant nutrient analysis- Influence of microbial biostimulant on soil NPK levels (A) and plant NPK levels (B). Student's t-test: significant differences at $P < 0.05$ and $P < 0.01$ are represented by “*” and “**” respectively.

Gene	Gene Accession No.	Forward Primer sequence (5' - 3')	Reverse Primer sequence (5' - 3')
<i>OsActin</i>	AB047313.1	ACCATTGGTGCTGAGCGTTT	CGCAGCTTCCATTCCCTATGAA
<i>OsPsaH</i>	Os05g0560000	GAGGACATCGGCAACACCA	GCCCTTCTTGATCGGAAGCA
<i>OsFD1</i>	LOC_Os08g01380	AGCAACAAGCTGGGAGACAG	GAGTAAGGCAGGTTCGATCCC
<i>OsET</i>	LOC_Os04g33630	GTGGGTGGCGGTGGCAAG	CACGACGGGCCTCAGCTC
<i>OsPsbD</i>	LOC_Osp1g00170	CTGCTACTGCTGTTTTCT	GATGTTATGCTCTGCCTG
<i>OsPsbP</i>	Os07g0141400	CACGGAGTTCATCGCCTACA	AAGCAAGAAGTCGACCTGGG
<i>OsPsbR3</i>	Os08g0200300	GAGGTGGCATGTTCAGTCGAT	TTTGCGCCGTACTTGTCAAC
<i>OsPsbS1</i>	LOC_Os01g64960	GCTGTTTCGGCTTCACCAAGG	ACGCCGGTCTCGATGTTGA
<i>OsCAB1R</i>	LOC_Os09g17740	GTTCTCCATGTTTCGGCTTCT	GACGAAGTTGGTGGCGTAG
<i>OsCAB2R</i>	LOC_Os01g41710	TGTTCTCCATGTTTCGGCTTCT	GCTACGGTCCCCACTTCACT
<i>OsCP24</i>	XM_015781325.2	CCTTAACCTTAACGGCCATTTC	CCCGCAGTAAATCCTGAGC
<i>OsOEP3</i>	Os07g0544800	AGCCGCTCATCGAGAAGAAG	GTACTTCTCCGCCTCTTCGG
<i>OsTLP</i>	LOC_Os08g39430	GGAGGGAGACGGGGTGGG	TCAGAAAGGGAGGAGAGCCGT
<i>OsLHC2.1</i>	LOC_Os02g52650	GGCGCGGTCTGAACGAGCT	GTTTAGCCATTAGTCAAAGCAATC
<i>OsChlI</i>	LOC_Os03g36540	CTTCACCGTCTGCAATGTAG	GATCTTAGGGTCGATGACGTT
<i>OsChlH</i>	LOC_Os03g20700	GCACGGGAACCTTGGCGTTTCATTA	ACATGTCCTGGAGCTGCTTCTCAT
<i>OsChlD</i>	LOC_Os03g59640	TAGCACAGCTGTCAGAGTGGGTTT	TTGCCAGCCACCTCAAGTATCTCA
<i>OsHEMA</i>	AB011416.1	GATGCAATCACTGCTGGAAAGCGT	CCATCTTGCCAGCACCAATCAACA
<i>OsHEMC</i>	LOC_Os02g07230	GAGCTGTCTCATTTCAGATCTGT	AGGGATTACAATAACCGTGGAA
<i>OsYGL13</i>	LOC_Os08g06630	CTCTCCCACAAGATCAAGGATG	GTTTGTATGAGGGCTCATTTC
<i>OsYGL8</i>	Loc_Os01g73450,	TGGATCTAACATGACACGCACCCA	ACTGTAACGGCATTCTTCTCCGGT
<i>OsRCA</i>	U74321.1	CTCTTCGTGCCCGTGTTTAC	TCGGAGTTAGCGTCACCAAG
<i>OsRbcS2</i>	L22155.1	AGTCTGGTGGCAACTAAGCC	GCACGGCCGGTAAATCAAA
<i>OsRbcS3</i>	AK068555.1	ACCATCTCAATGGCCTCTGC	TGTGTGCATATAGCCGGAGC
<i>OsRbcS4</i>	AK070257.1	AACGTTAGGCAGGTGCAGTT	TGCAGCTTAACACGGACACA
<i>OsRbcS5</i>	AK099574.1	GGAGTCCGGCGGAAACTAAG	GGAAACCAATGCAAGGTGGC
<i>OsHYR</i>	Os03g02650	CCGAGGGCTTGATGATGAG	CGTAAGCCCATTTCAGGAATG
<i>OsMOC1</i>	AY242058	CTGCTCCGGCTGCACTAC	CCACGCTGAGACGGAGAG
<i>OsSLR1</i>	Os03g0707600	GAGTCGCTGCACTACTACTCC	GCGCGTGTGCCAGCCCAGCGT
<i>OsLAX2</i>	AB669025	TTCTTGCCTCAGATTCCAAG	CCTCTGCATGTTATCTCCAC
<i>OsMADS57</i>	LOC_Os02g49840	CAGATTATGTTGTCTGGATGCTC	AAAGCAATAGAGAGTAAGCAGGGT
<i>OsHSF2AD</i>	LOC_Os03g06630	CAGCAGGCACCTTGGCACC	TTCTTGTACGCTTTAGCCTGT
<i>OsCKX11</i>	Os08g0460600	CAACGCAATCATTGACGCC	TTGCACCCCTCCCAAATGT
<i>OsRGN1</i>	LOC_Os01g49160	CGGCTACACCGACCAGGAG	CGCGATGATGGACCACCT
<i>OsNOG1</i>	MF687920	TCCGACTTACAATGAACAC	GGTAGCAGGACTCCACTT
<i>OsSPL9</i>	LOC_Os05g33810	AGATGGGCAGGTGATTAT	TGTGGGAGAGCTTTAGTC
<i>OsIPA1</i>	LOC_Os08g39890	CGGTCTGACTAGCTGCATCTGTTGG	CATCGTGTGCTGTTTGGTGAAG
<i>OsSPL14</i>	Os08g0509600	CAAGGGTTCCAAGCAGCGTAA	TGCACCTCATCAAGTGAGAC
<i>OsLC2</i>	AK101341	AGCATCAGCTTTGGACGAGGA	CAGTTGGTGGAAATAGAGCCAGAAT
<i>OsVIL2</i>	XM_015764905.2	GGAGTATGCTTTCCGGATCA	GTGGGAAACAACATGTGCAG
<i>OsGRF4</i>	LC333011.1	GAAAGCCTGTGGAAACGCA	CAACGCCGAGCCAAATGAG
<i>OsNRT2.1</i>	NM_001401744.1	CTTCACGTCGTCGAGGTACT	CACTCGGAGCCGTAGTAGTG
<i>OsNRT2.3A</i>	XM_015773038.2	CGCTGCTGCCGCTCATCCG	CCGTGCCCATGGCCAGAC
<i>OsNIA1</i>	XM_015767224.2	TCAAGGTGTGGTACGTGGTG	CGAGGTCATAGCCCATCTTC
<i>OsNIA2</i>	NM_001422594.1	TGTACCAGGTCATCCAGTCG	CGATGACGTACCACACCTTG
<i>OsAMT1</i>	AF001505.2	GGTTTCTCTCCCTCTCCGAT	CCACCTTCACACCACACATT
<i>OsGS1</i>	AB180688.1	TGTTTCTCCTCATCCCTGC	TCACAGTCCTCGCTTTGC
<i>OsGS2</i>	X14246.1	GGAGAGGTCATGCCTGGTCAGT	ACTACACCAGCCTGCTCCGTTA
<i>OsGOGAT1</i>	XM_015793761.2	GTGCAGCCTGTTGCAGCATAAA	CGGCATTTACCATGCAAATC
<i>OsGOGAT2</i>	NM_001402625.	CCTGTCGAAGGATGATGAAGGTGAA CC	TGCATGGCCCTACTATCTTCGCATCA
<i>OsPT1</i>	AF536961.1	CGCTTCCGTACGAGTGGTAGT	GGTTCTTTCAAATCCAGGGAAA
<i>OsPT8</i>	AF536968.1	AGAAGGCAAAAGAAATGTGTGTTAAAT	AAAATGTATTCGTGCCAAATTGCT
<i>OsHAK1</i>	NM_001402365.1	GTTGATGATGCTGATGTTGGAAG	CCAACACTTTCAGCTGAAAC
<i>OsZIP9</i>	NM_001420533.1	CATCAGTTCTTCGAAGGGATAGG	TGTGGTTAGCGAGAAGAAGATG
<i>McMopB</i>	AF031148.1	ACAGGCCGAAGAGACTTTCA	GGTGGTGTGCTTCGTAAT

Supplementary Table 1- List of oligonucleotide primers used in this study.

Sample	IAA concentration (mg/L)
Microbial biostimulant without Tryptophan	0
Microbial biostimulant with Tryptophan	1.83-3.61

Table S2- Indole acetic acid levels observed in microbial biostimulant grown in presence or absence of Tryptophan.

		CH ₄ flux (g/ha/h)									
Season	Treatment	10 DAT	20 DAT	30 DAT	40 DAT	50 DAT	60 DAT	70 DAT	80 DAT	90 DAT	
Season II	Control	31.33± 3.38	28.67 ±3.84	70 ± 1.53	109.66 ± 4.7	123.33 ± 6.33	64 ± 4.62	34.2 ± 0.9	22.43 ± 1.68	9.86 + 0.61	
	Methane - derived microbial biostimulant	23.66 ± 1.2	35.66 ± 1.42	53.66 + 1.76	71.33 ± 4.98	61.73 ± 1.92	49.46 ± 4.66	23.13 ± 1.73	15.73 ± 1. 07	5.80 ± 0.45	
		N ₂ O flux (g/ha/h)									
	Control	0.76 ± 0.02	0.83 ± 0.07	0.78 ± 0.05	2.67 ± 0.08	1.32 ± 0.05	1.82 ± 0.03	3.91 ± 0.25	3.91 ± 0.11	2.14 ± 0.15	
	Methane- derived microbial biostimulant	0.41 ± 0.07	0.55 ± 0.03	0.57 ± 0.03	0.82 ± 0.04	0.51 ±0.06	0.69 ± 0.04	2.07 ± 0.33	2.65 ± 0.13	1.47 + 0.08	
		CH ₄ flux (g/ha/h)									
Season	Treatment	10 DAT	20 DAT	30 DAT	40 DAT	50 DAT	60 DAT	70 DAT	80 DAT	90 DAT	100 DAT
Season III	Control	59.33 ± 1.86	44 ± 1.73	86.66 ± 0.88	103 ± 2.31	128 ± 2.65	149.33 ± 1.45	92.33 ± 3.28	49 ± 1.15	36.66 ± 1.45	28.33 ± 1.45
	Methane- derived microbial biostimulant	37.33 ± 1.20	26.33 ± 0.88	54.33 ± 1.45	47.66 ± 2.96	58 ± 3.46	72.66± 2.46	47± 1.53	16.33± 1.45	20.33± 0.88	12.66±0.67
		N ₂ O flux (g/ha/h)									
	Control	1.28±0 .03	0.94 ± 0.02	0.76 ± 0.02	1.05 ± 0.03	1.13 ± 0.03	1.28 ± 0.07	1.79 ± 0.04	4.72 ± 0.03	2.70 ± 0.03	3.46 ± 0.26
	Methane- derived microbial biostimulant	0.58 ± 0.03	0.68 ± 0.03	0.45 ± 0.02	0.40 ± 0.02	0.62 ± 0.03	0.68 ± 0.02	0.66 ± 0.01	2.2 ± 0.2	1.06 ± 0.11	1.25 ± 0.03

Table S3A- Temporal CH₄ and N₂O emission from control and microbial biostimulant- treated paddy fields collected during season II and season III trials

Season	Treatment	Average CH ₄ emission (kg/ha/season)	Average N ₂ O emission (kg/ha/season)
Season I Kharif 2022	Control (Season-I)	213.65	16.24
	Methane- derived microbial biostimulant	82.43	10.73
Season II Rabi 2022	Control (Season-II)	131.60	5.32
	Methane- derived microbial biostimulant	90.72	2.80
Season III Kharif 2023	Control (Season-III)	226.97	3.10
	Methane- derived microbial biostimulant	117.83	1.54

Table S3B- Average CH₄ and N₂O emission from control and microbial biostimulant- treated paddy fields.

Season	Treatment	Grain yield (kg/ha)	% improvement in grain yield over control	CH ₄ emission				N ₂ O emission			
				Average CH ₄ emission (kg/ha/season)	% reduction in CH ₄ emission over control	CO ₂ eq CH ₄ emission at 25- year time period (kg/ha/season)	Yield-scaled CO ₂ -eq emission (kg CO ₂ -eq/t)	Average N ₂ O emission (kg/ha/season)	% reduction in N ₂ O emission over control	CO ₂ eq N ₂ O emission at 25- year time period (kg/ha/season)	Yield-scaled CO ₂ -eq emission (kg CO ₂ -eq/t)
Season I Kharif 2022	Control (Season-I)	6024	-	213.65	-	17946.43	2979.16	16.24	-	4839.52	803.37
	Methane- derived microbial biostimulant	8004	32.87	82.43	-61.42	6924.29	865.10	10.73	-33.93	3197.54	399.49
Season II Rabi 2022	Control (Season-II)	5015	-	131.60	-	11054.40	2204.27	5.32	-	1585.36	316.12
	Methane- derived microbial biostimulant	6997	39.52	90.72	-31.06	7620.48	1089.11	2.80	-47.37	834.40	119.25
Season III Kharif 2023	Control (Season-III)	5081	-	226.97	-	19065.60	3752.33	3.10	-	923.80	181.81
	Methane- derived microbial biostimulant	7058	38.91	117.83	-48.09	9897.41	1402.30	1.54	-50.32	458.92	65.02

Table S4- Effect of methane-derived microbial biostimulant on yield-scaled CO₂-eq emission in rice-
Yield-scaled CO₂e-emission of CH₄ and N₂O were found to be significantly lower in methane-derived microbial biostimulant treatment compared to controls.

Conversion factor for CH₄ (1kg of CH₄= 84Kg of CO₂ equivalent) and N₂O (1Kg of N₂O= 298Kg of CO₂ equivalent) were taken from climatechangeconnection.org/emissions/co2-equivalents/

Methane emission (million metric ton)	380		
Methane emission from paddy - 10% contribution (million metric ton)	30.4		
Global land under paddy cultivation (million ha)	162		
Reduction in methane emission from methane-derived microbial biostimulant (percentage per ha)	60%		
Global targeted methane reduction per annum (2% of total - COP26 target) million metric tonnes	7.60		
Percentage of global paddy cultivation area targeted formethane-derived microbial biostimulant application	10.00%	30.00%	50.00%
Actual paddy cultivated area targeted for methane-derived microbial biostimulant application (million ha)	16.2	48.6	81
Total methane emission from target area (million metric tonnes)	3.04	9.12	15.2
Methane emission from methane-derived microbial biostimulant use in target area (million metric tonnes)	1.824	5.472	9.12
Percentage of global methane target achieved	24%	72%	120%

Table S5- CH₄ emission reduction from paddy field using methane-derived microbial biostimulant to meet COP26 target for methane reduction by 2030.