

Generation of lysolipin derivatives by genetic engineering

Helene Robertsen^{1,2}, Sabrina Rohrer¹, Andreas Kulik¹, Wolfgang Wohlleben^{1,2,3*},
Yvonne Mast^{1,2,4,5,6*}

¹Department of Microbiology/Biotechnology, Interfaculty Institute of Microbiology and Infection Medicine, Faculty of Science, University of Tübingen, Auf der Morgenstelle 28, D-72076 Tübingen, Germany

²German Center for Infection Research (DZIF), Partner Site Tübingen, Tübingen, Germany

³Cluster of Excellence 'Controlling Microbes to Fight Infections' (CMFI), University of Tübingen, Tübingen, Germany

⁴Department Bioresources for Bioeconomy and Health Research, Leibniz Institute DSMZ -German Collection of Microorganisms and Cell Cultures, Inhoffenstraße 7B, 38124 Braunschweig, Germany

⁵Braunschweig Integrated Centre of Systems Biology (BRICS), Rebenring 56, 38106 Braunschweig, Germany

⁶Technische Universität Braunschweig, Institut für Mikrobiologie, Rebenring 56, 38106 Braunschweig, Germany

*Correspondence:
Prof. Dr. Yvonne Mast
yvonne.mast@dsmz.de

Prof. Dr. Wolfgang Wohlleben
Wolfgang.wohlleben@biotech.uni-tuebingen.de

Running title: Novel lysolipin derivatives

Key words: actinomycetes, *Streptomyces*, antibiotic, polyketides, lysolipin, genetic engineering

Table S1: Media receipt for lysolipin production media NL800 and E1. All data refer to 1 LH₂O_{deion.}

Medium	Ingredients	pH
NL800	5 g glucose 10 g glycerin 10 g soluble starch 5 g soy flour full fat 2 g yeast extract 1 g NaCl 1 g CaCO ₃	7.2
E1	20 g glucose 20 g soluble starch 5 g yeast extract 2.5 g pharmamedia 1 g MgSO ₄ x 7H ₂ O 1.3 g KH ₂ PO ₄ x 3H ₂ O 5 g NaCl 5 g CaCO ₃	7.5

Table S2: Oligonucleotides used in this study. MunI restriction site is highlighted by bold letters.

Primer designation	Primer sequence (5'-3')
IlpRI-CmR.MunI-F	TTGATTAGTCACGCCGCACCTAGTAGCCTGGCAACTATG CAATTG GACGTCTA AGAAACCATTAT
IlpRI-CmR.MunI-R	GCCGGTTCCATCCCCGTCGGCTGGCCGGCGCGGGTCTC ACAATTG TACGCC CCGCCCTGCCAC

Table S3: Bioactivity profile of lysolipin I and its derivatives as reported in patent WO/2007/079715. Improved bioactivities are highlighted in bold.

Lysolipin (derivative)	MIC ($\mu\text{g/mL}$) (<i>E. coli</i> ATCC10536)	Cytotoxicity (IC ₅₀ , [M]) (THP-1 (human lymphoblast))
Lysolipin I (CBS42)	128	9.39E-09
CBS40	3	2.16E-08
CBS44	128	1.16E-07
CBS48	128	2.08E-07
CBS49	30	3.44E-07
CBS68	128	1.58E-07
CBS70	128	2.99E-07
CBS72	128	4.27E-08

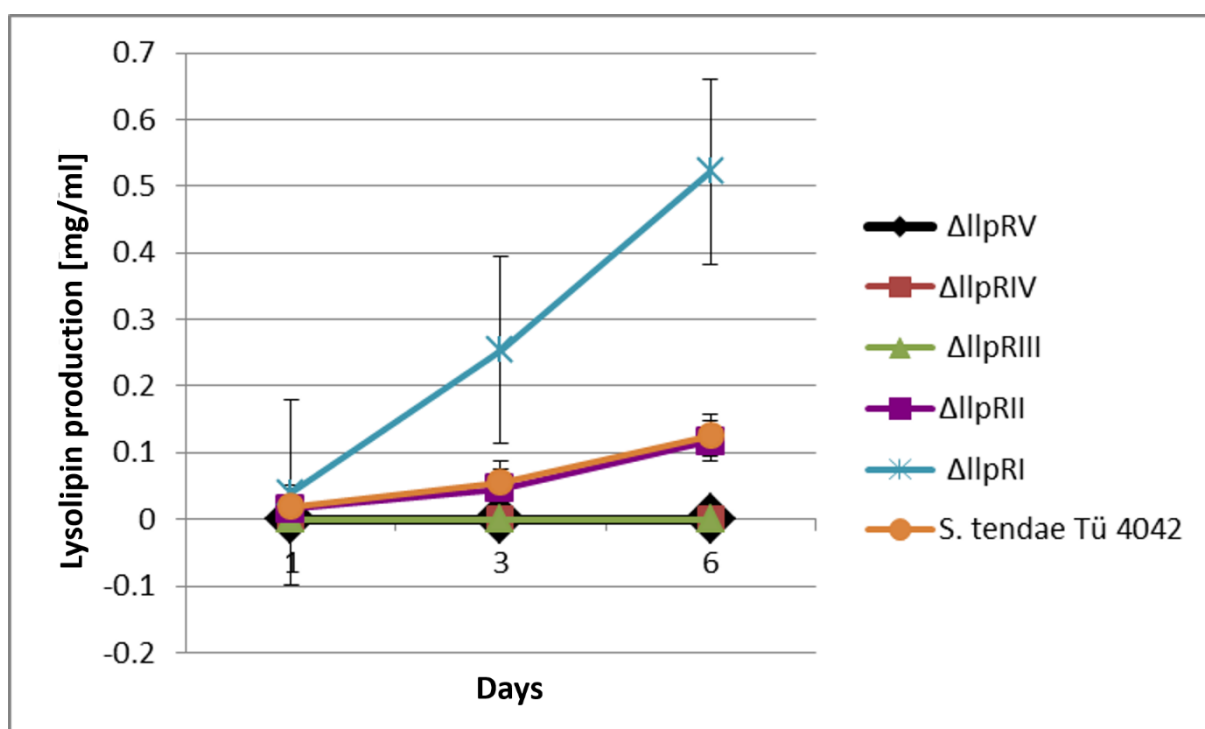


Figure S1: Lysolipin I production of *Streptomyces tendae* Tü4042 and transcriptional regulatory mutants *S. tendae* $\Delta IlpRI$, $\Delta IlpRII$, $\Delta IlpRIII$, $\Delta IlpRIV$, and $\Delta IlpRV$, respectively, according to Rohrer, 2017. Cell cultures were harvested after 1, 2 and 6 days of cultivation in E1 medium. Lysolipin production was analysed by HPLC-MS by using a

lysolipin calibration curve. Results are shown from three independent biological replicates.

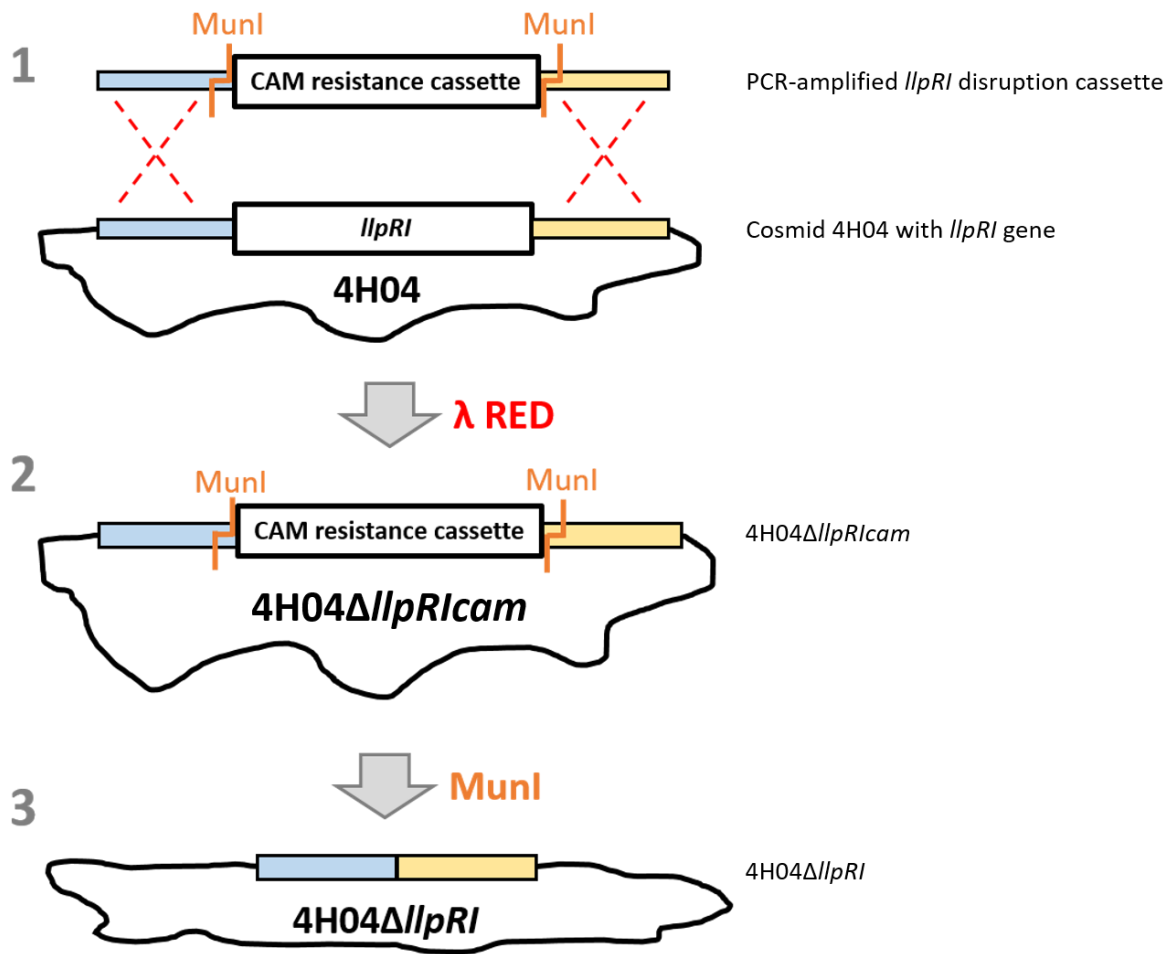


Figure S2: Schematic presentation of the cloning procedure to generate construct *4H04ΔllpRI* in *E. coli*. Up- and downstream homologous regions of *llpRI* are indicated in blue and yellow, respectively. *MunI* restriction sites are shown in orange. Homologous recombination events are highlighted as red dashed crosses 1: λ -RED-driven recombination of homologous gene regions by a double crossover event to integrate the PCR-amplified *llpRI* disruption cassette into 4H04; 2: *4H04ΔllpRIcam* with chloramphenicol resistance marker (CAM) and deleted *llpRI* gene; 3: *4H04ΔllpRI* without CAM cassette after restriction with *MunI*, resulting in *llpRI* deletion.

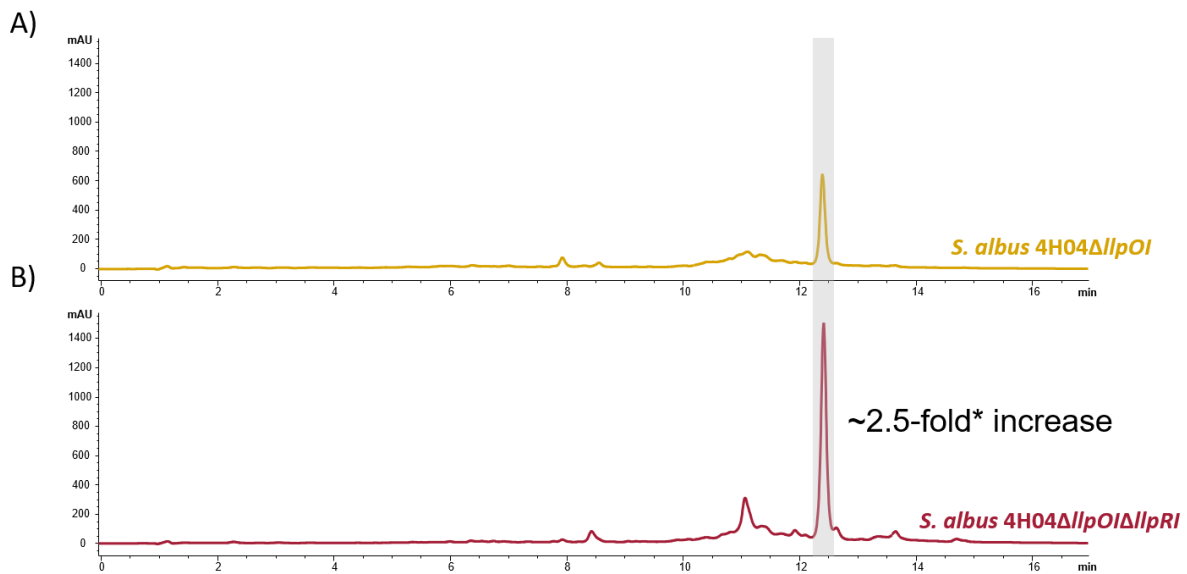


Figure S3: HPLC chromatograms of extract samples from A) *S. albus* 4H04ΔllpOI (yellow) and B) *S. albus* 4H04ΔllpOIΔllpRI (red). Peak at RT 12.4 correlates to lysolipin derivative (CBS40) production and is highlighted in grey. Samples were obtained from cultures grown for seven days at 28°C.

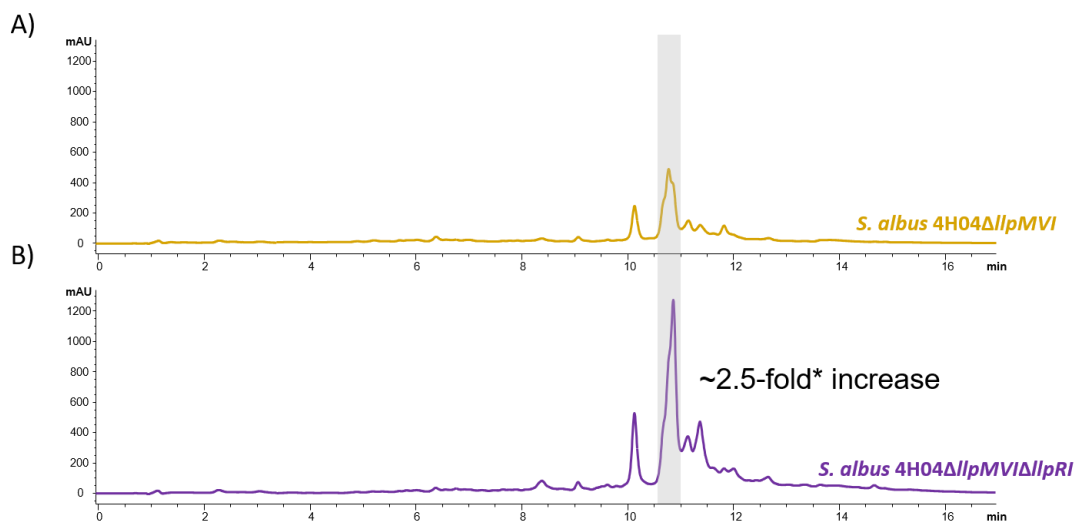


Figure S4: HPLC chromatograms of extract samples from A) *S. albus* 4H04ΔllpMVI (yellow) and B) *S. albus* 4H04ΔllpMVIΔllpRI (purple). Peak at RT 10.6 correlates to lysolipin derivatives (CBS 70+72) production and is highlighted in grey. Samples were obtained from cultures grown for seven days at 28°C.

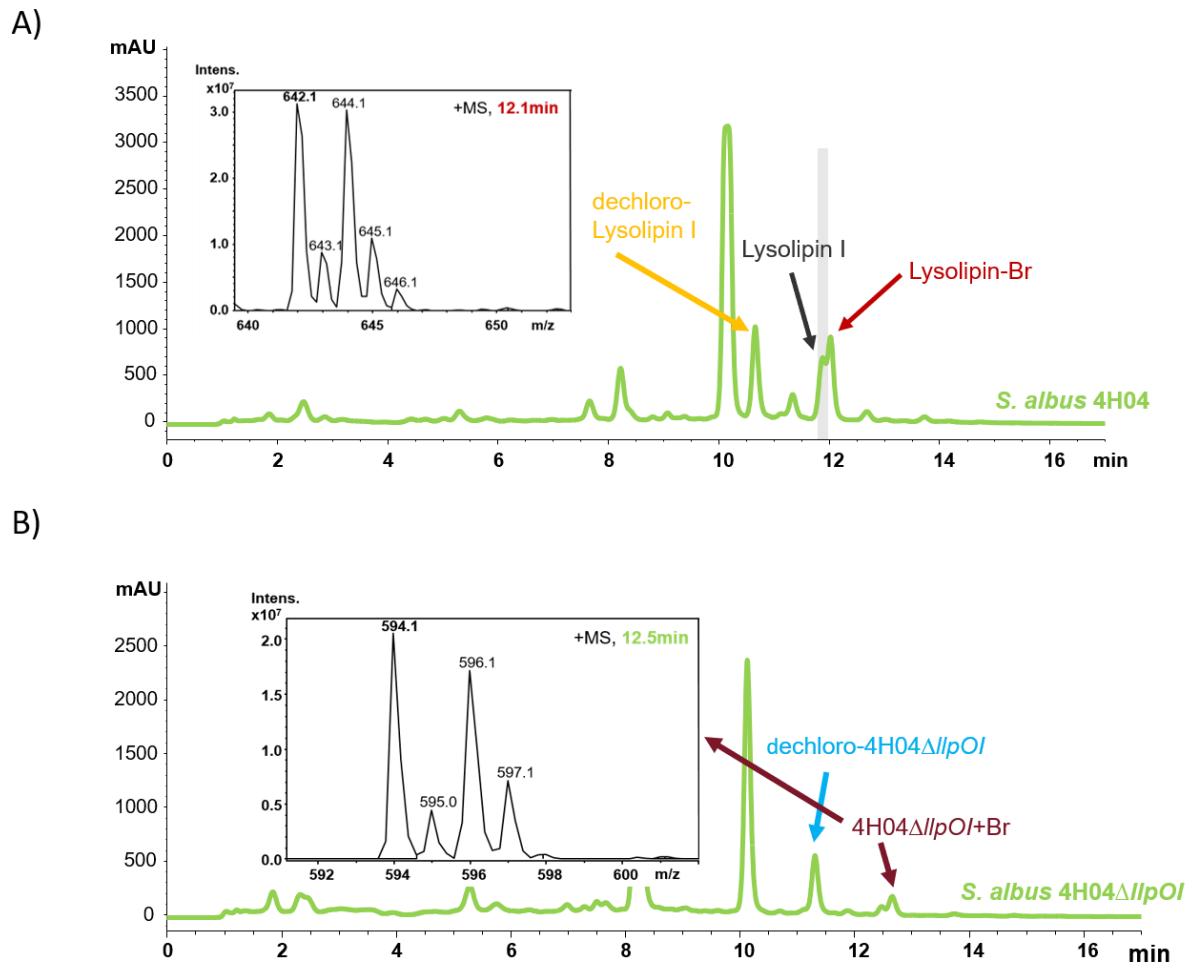


Figure S5: HPLC chromatograms of extract samples from A) *S. albus* 4H04 (green) and B) *S. albus* 4H04 Δ lpOI (green). Peaks correlating to lysolipin I and lysolipin derivatives are highlighted by arrows. MS spectra are shown in black. Samples were obtained from cultures grown for seven days at 28°C.