

Supplementary materials

No.	Species	Strain	5mmol/L	10mmol/L	15mmol/L	20mmol/L	25mmol/L	30mmol/L
1	<i>Ligilactobacillus agilis</i>	X62	0.23	0.23	0.23	0.24	0.32	0.32
2	<i>Bacillus safensis</i>	F60	0.28	0.28	0.28	0.28	0.37	0.38
3	<i>Bacillus subtilis</i>	W4	0.29	0.29	0.30	0.28	0.37	0.36
4	<i>Bacillus safensis</i>	W42	0.29	0.29	0.33	0.32	0.38	0.41
5	<i>Bacillus safensis</i>	W56	0.28	0.28	0.29	0.35	0.40	0.36
6	<i>Limosilactobacillus fermentum</i>	B32	0.29	0.29	0.31	0.34	0.45	0.43
7	<i>Limosilactobacillus fermentum</i>	M21	0.26	0.26	0.26	0.27	0.25	0.29
8	<i>Ligilactobacillus agilis</i>	X19	0.35	0.35	0.30	0.33	0.31	0.24
9	<i>Ligilactobacillus agilis</i>	X64	0.27	0.27	0.28	0.29	0.41	0.44
10	<i>Bacillus safensis</i>	F79	0.31	0.31	0.32	0.35	0.50	0.48
11	<i>Ligilactobacillus agilis</i>	W43	0.29	0.29	0.31	0.41	0.47	0.46
12	<i>Ligilactobacillus agilis</i>	W64	0.28	0.28	0.29	0.42	0.45	0.46
13	<i>Bacillus safensis</i>	B52	0.30	0.30	0.30	0.40	0.49	0.50
14	<i>Bacillus safensis</i>	M42	0.31	0.31	0.30	0.27	0.23	0.44
15	<i>Ligilactobacillus agilis</i>	X44	0.27	0.27	0.24	0.27	0.24	0.22
16	<i>Enterococcus camelliae</i>	S35	0.26	0.26	0.27	0.28	0.42	0.43
17	<i>Weissella confusa</i>	W26	0.32	0.32	0.31	0.38	0.47	0.48
18	<i>Limosilactobacillus fermentum</i>	W45	0.29	0.29	0.30	0.40	0.46	0.47
19	<i>Ligilactobacillus agilis</i>	W70	0.30	0.30	0.31	0.45	0.48	0.50
20	<i>Bacillus safensis</i>	B55	0.30	0.30	0.29	0.31	0.47	0.51
21	<i>Enterococcus lactis</i>	M45	0.25	0.25	0.26	0.27	0.26	0.40
22	<i>Ligilactobacillus agilis</i>	X46	0.24	0.24	0.24	0.24	0.22	0.19
23	<i>Bacillus safensis</i>	F19	0.25	0.25	0.25	0.27	0.24	0.24
24	<i>Ligilactobacillus agilis</i>	W40	0.29	0.29	0.29	0.32	0.29	0.33
25	<i>Ligilactobacillus agilis</i>	W52	0.30	0.30	0.29	0.32	0.26	0.33
26	<i>Limosilactobacillus fermentum</i>	W72	0.29	0.29	0.28	0.33	0.26	0.33

Figure S1. Heat map of 26 strains growth capacity (OD_{600nm}) under facultative anaerobic gas condition with ammonium sulfate (5, 10, 15, 20, 25, and 30 mmol/L) as the sole nitrogen source.

No.	Species	Strain	5mmol/L	10mmol/L	15mmol/L	20mmol/L	25mmol/L	30mmol/L
1	<i>Enterococcus lactis</i>	X1	0.24	0.26	0.28	0.55	0.52	0.57
2	<i>Enterococcus faecium</i>	Y34	0.23	0.24	0.28	0.27	0.28	0.27
3	<i>Ligilactobacillus salivarius</i>	P17	0.24	0.24	0.28	0.27	0.28	0.27
4	<i>Ligilactobacillus agilis</i>	P53	0.26	0.26	0.30	0.39	0.28	0.43
5	<i>Ligilactobacillus agilis</i>	S14	0.32	0.26	0.29	0.28	0.31	0.29
6	<i>Bacillus pascis</i>	B11	0.26	0.24	0.30	0.46	0.47	0.45
7	<i>Ligilactobacillus salivarius</i>	W14	0.32	0.35	0.37	0.36	0.37	0.36
8	<i>Ligilactobacillus salivarius</i>	F15	0.20	0.22	0.24	0.25	0.25	0.24
9	<i>Bacillus licheniformis</i>	F37	0.27	0.35	0.34	0.35	0.44	0.44
10	<i>Limosilactobacillus fermentum</i>	M18	0.25	0.35	0.34	0.36	0.31	0.34
11	<i>Limosilactobacillus fermentum</i>	X18	0.27	0.34	0.35	0.56	0.51	0.51
12	<i>Enterococcus lactis</i>	Y35	0.32	0.24	0.27	0.28	0.28	0.28
13	<i>Ligilactobacillus agilis</i>	P18	0.23	0.21	0.18	0.27	0.20	0.20
14	<i>Limosilactobacillus fermentum</i>	P74	0.26	0.27	0.37	0.38	0.37	0.36
15	<i>Limosilactobacillus fermentum</i>	S15	0.27	0.27	0.32	0.63	0.63	0.60
16	<i>Ligilactobacillus salivarius</i>	B12	0.28	0.28	0.30	0.50	0.47	0.46
17	<i>Limosilactobacillus fermentum</i>	W16	0.31	0.24	0.26	0.26	0.27	0.27
18	<i>Limosilactobacillus mucosae</i>	F18	0.24	0.24	0.27	0.33	0.30	0.29
19	<i>Ligilactobacillus agilis</i>	F41	0.26	0.39	0.37	0.38	0.38	0.40
20	<i>Enterococcus lactis</i>	M21	0.26	0.30	0.30	0.65	0.65	0.65
21	<i>Ligilactobacillus agilis</i>	X20	0.27	0.27	0.32	0.62	0.54	0.56
22	<i>Limosilactobacillus fermentum</i>	Y36	0.28	0.24	0.24	0.24	0.24	0.25
23	<i>Ligilactobacillus salivarius</i>	P19	0.34	0.32	0.31	0.28	0.30	0.28
24	<i>Ligilactobacillus agilis</i>	P77	0.27	0.27	0.32	0.44	0.44	0.45
25	<i>Enterococcus faecium</i>	S16	0.26	0.30	0.37	0.39	0.40	0.39
26	<i>Enterococcus faecium</i>	B13	0.26	0.30	0.33	0.59	0.54	0.53
27	<i>Ligilactobacillus agilis</i>	W18	0.27	0.27	0.31	0.32	0.33	0.31
28	<i>Limosilactobacillus fermentum</i>	F20	0.25	0.26	0.28	0.29	0.29	0.31
29	<i>Enterococcus faecium</i>	F42	0.27	0.26	0.32	0.67	0.67	0.66
30	<i>Limosilactobacillus fermentum</i>	M27	0.29	0.30	0.31	0.31	0.32	0.30
31	<i>Ligilactobacillus salivarius</i>	X36	0.26	0.27	0.32	0.74	0.63	0.64
32	<i>Limosilactobacillus fermentum</i>	Y45	0.33	0.30	0.31	0.29	0.28	0.28
33	<i>Ligilactobacillus agilis</i>	P32	0.34	0.32	0.33	0.29	0.29	0.27
34	<i>Enterococcus faecium</i>	P80	0.32	0.27	0.25	0.26	0.26	0.25
35	<i>Ligilactobacillus salivarius</i>	S17	0.26	0.27	0.39	0.43	0.44	0.39
36	<i>Ligilactobacillus salivarius</i>	B14	0.27	0.26	0.32	0.61	0.59	0.61
37	<i>Ligilactobacillus agilis</i>	W23	0.20	0.35	0.31	0.33	0.33	0.32
38	<i>Enterococcus faecium</i>	F21	0.27	0.28	0.30	0.31	0.31	0.32
39	<i>Ligilactobacillus salivarius</i>	F43	0.27	0.35	0.35	0.34	0.35	0.37
40	<i>Limosilactobacillus fermentum</i>	M43	0.29	0.30	0.33	0.31	0.32	0.33
41	<i>Enterococcus lactis</i>	X55	0.32	0.26	0.18	0.20	0.19	0.27
42	<i>Enterococcus lactis</i>	Y48	0.32	0.28	0.29	0.30	0.27	0.26
43	<i>Ligilactobacillus agilis</i>	P33	0.27	0.27	0.32	0.54	0.56	0.53
44	<i>Enterococcus lactis</i>	R13	0.32	0.30	0.30	0.29	0.28	0.27
45	<i>Lactocaseibacillus paracasei</i>	S18	0.27	0.27	0.30	0.66	0.63	0.62
46	<i>Limosilactobacillus fermentum</i>	B21	0.36	0.33	0.31	0.33	0.32	0.30
47	<i>Ligilactobacillus agilis</i>	F24	0.26	0.27	0.31	0.62	0.59	0.59
48	<i>Limosilactobacillus fermentum</i>	F44	0.27	0.29	0.31	0.39	0.45	0.45
49	<i>Limosilactobacillus fermentum</i>	M50	0.27	0.30	0.34	0.72	0.70	0.66
50	<i>Enterococcus lactis</i>	Y7	0.22	0.22	0.26	0.25	0.31	0.25
51	<i>Enterococcus lactis</i>	Y50	0.25	0.22	0.21	0.21	0.21	0.21
52	<i>Ligilactobacillus agilis</i>	P38	0.27	0.30	0.34	0.53	0.51	0.51
53	<i>Ligilactobacillus salivarius</i>	S3	0.27	0.27	0.36	0.38	0.37	0.35
54	<i>Enterococcus faecium</i>	B1	0.26	0.30	0.38	0.38	0.36	0.34
55	<i>Ligilactobacillus agilis</i>	B23	0.26	0.29	0.31	0.32	0.32	0.30
56	<i>Enterococcus faecium</i>	W37	0.27	0.30	0.33	0.33	0.34	0.34
57	<i>Limosilactobacillus mucosae</i>	F27	0.27	0.30	0.31	0.33	0.32	0.34
58	<i>Limosilactobacillus fermentum</i>	F48	0.37	0.27	0.23	0.23	0.56	0.24
59	<i>Limosilactobacillus fermentum</i>	M52	0.33	0.33	0.32	0.34	0.45	0.36
60	<i>Ligilactobacillus salivarius</i>	Y24	0.14	0.17	0.12	0.15	0.20	0.11

61	<i>Limosilactobacillus fermentum</i>	Y67	0.30	0.27	0.26	0.25	0.25	0.27
62	<i>Limosilactobacillus fermentum</i>	P39	0.27	0.27	0.33	0.57	0.56	0.55
63	<i>Limosilactobacillus fermentum</i>	S5	0.28	0.30	0.31	0.28	0.38	0.27
64	<i>Enterococcus faecium</i>	B2	0.27	0.29	0.33	0.70	0.62	0.64
65	<i>Ligilactobacillus agilis</i>	B24	0.26	0.33	0.34	0.31	0.30	0.28
66	<i>Limosilactobacillus fermentum</i>	F3	0.28	0.29	0.32	0.54	0.35	0.51
67	<i>Limosilactobacillus mucosae</i>	F30	0.27	0.29	0.32	0.56	0.56	0.55
68	<i>Enterococcus faecium</i>	F63	0.26	0.27	0.31	0.38	0.45	0.38
69	<i>Limosilactobacillus fermentum</i>	M55	0.27	0.29	0.30	0.43	0.37	0.41
70	<i>Limosilactobacillus fermentum</i>	Y26	0.18	0.19	0.20	0.20	0.23	0.21
71	<i>Limosilactobacillus fermentum</i>	P1	0.32	0.29	0.29	0.29	0.29	0.27
72	<i>Ligilactobacillus agilis</i>	P43	0.27	0.29	0.30	0.60	0.56	0.56
73	<i>Limosilactobacillus fermentum</i>	B8	0.33	0.30	0.28	0.29	0.27	0.26
74	<i>Limosilactobacillus fermentum</i>	W6	0.31	0.29	0.26	0.27	0.26	0.26
75	<i>Limosilactobacillus fermentum</i>	F9	0.37	0.33	0.30	0.32	0.32	0.32
76	<i>Limosilactobacillus fermentum</i>	F33	0.29	0.30	0.32	0.33	0.31	0.36
77	<i>Ligilactobacillus salivarius</i>	F74	0.31	0.27	0.31	0.29	0.27	0.27
78	<i>Limosilactobacillus fermentum</i>	M61	0.27	0.29	0.21	0.20	0.27	0.28
79	<i>Limosilactobacillus fermentum</i>	Y27	0.26	0.27	0.30	0.48	0.49	0.47
80	<i>Limosilactobacillus fermentum</i>	P2	0.26	0.20	0.27	0.27	0.27	0.29
81	<i>Limosilactobacillus fermentum</i>	P52	0.28	0.29	0.29	0.52	0.53	0.49
82	<i>Limosilactobacillus fermentum</i>	S13	0.26	0.29	0.27	0.53	0.54	0.46
83	<i>Limosilactobacillus fermentum</i>	B10	0.27	0.30	0.29	0.55	0.52	0.52
84	<i>Limosilactobacillus fermentum</i>	W12	0.26	0.29	0.30	0.46	0.45	0.40
85	<i>Limosilactobacillus fermentum</i>	F14	0.27	0.30	0.27	0.30	0.27	0.30
86	<i>Bacillus licheniformis</i>	F36	0.30	0.33	0.33	0.36	0.31	0.35
87	<i>Limosilactobacillus fermentum</i>	M15	0.36	0.34	0.34	0.35	0.33	0.34
88	<i>Limosilactobacillus fermentum</i>	M69	0.28	0.29	0.33	0.27	0.28	0.29
89	<i>Pediococcus acidilactici</i>	M78	0.21	0.29	0.22	0.21	0.24	0.22

Figure S2. Heat map of 89 strains growth capacity (OD_{600nm}) under anaerobic gas condition with ammonium sulfate (5, 10, 15, 20, 25, and 30 mmol/L) as the sole nitrogen source.

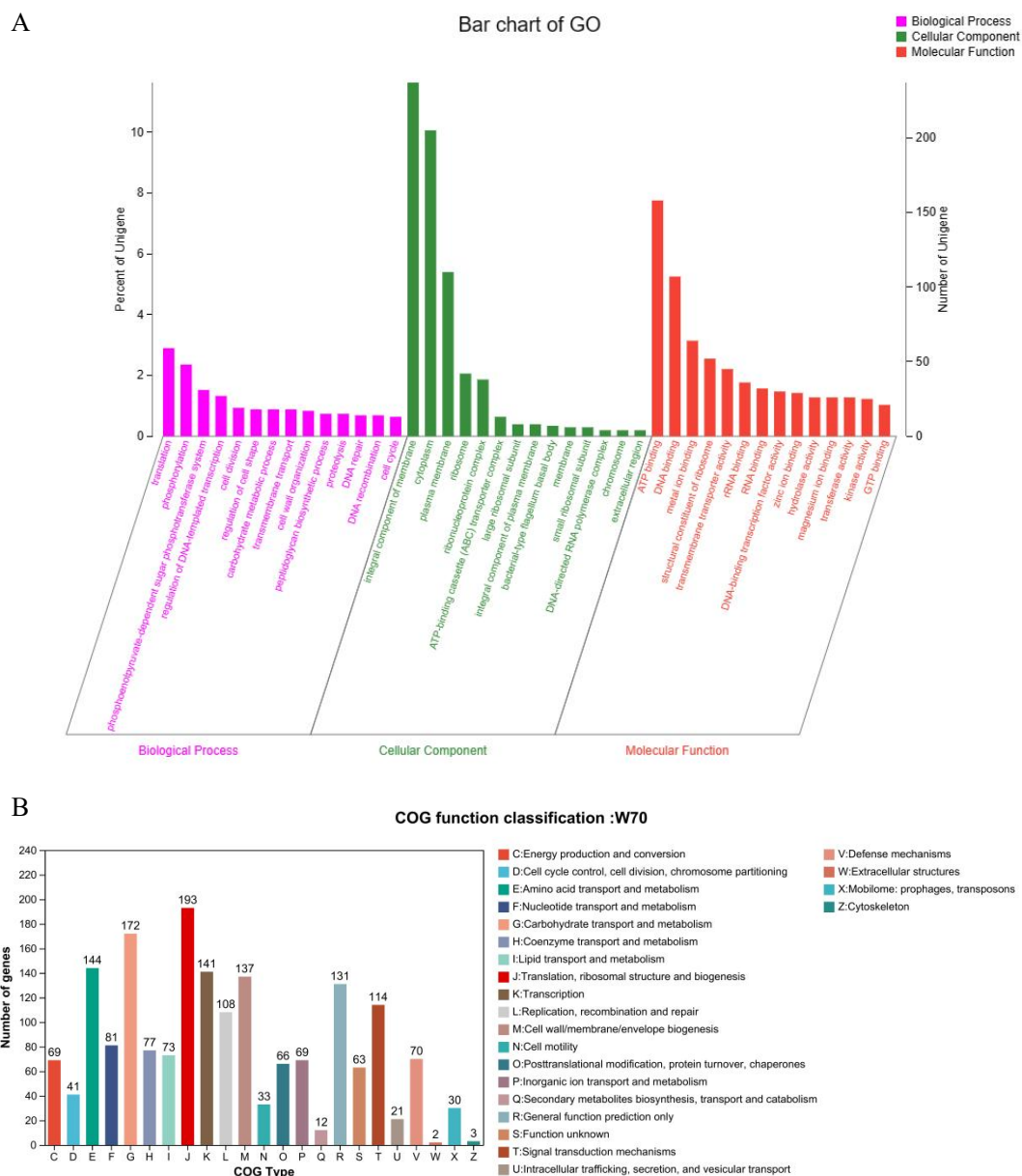


Figure S3. GO and COG annotation analysis of *Ligilactobacillus agilis* W70. **(A)** GO classification of putative proteins. **(B)** COG classification of putative proteins.

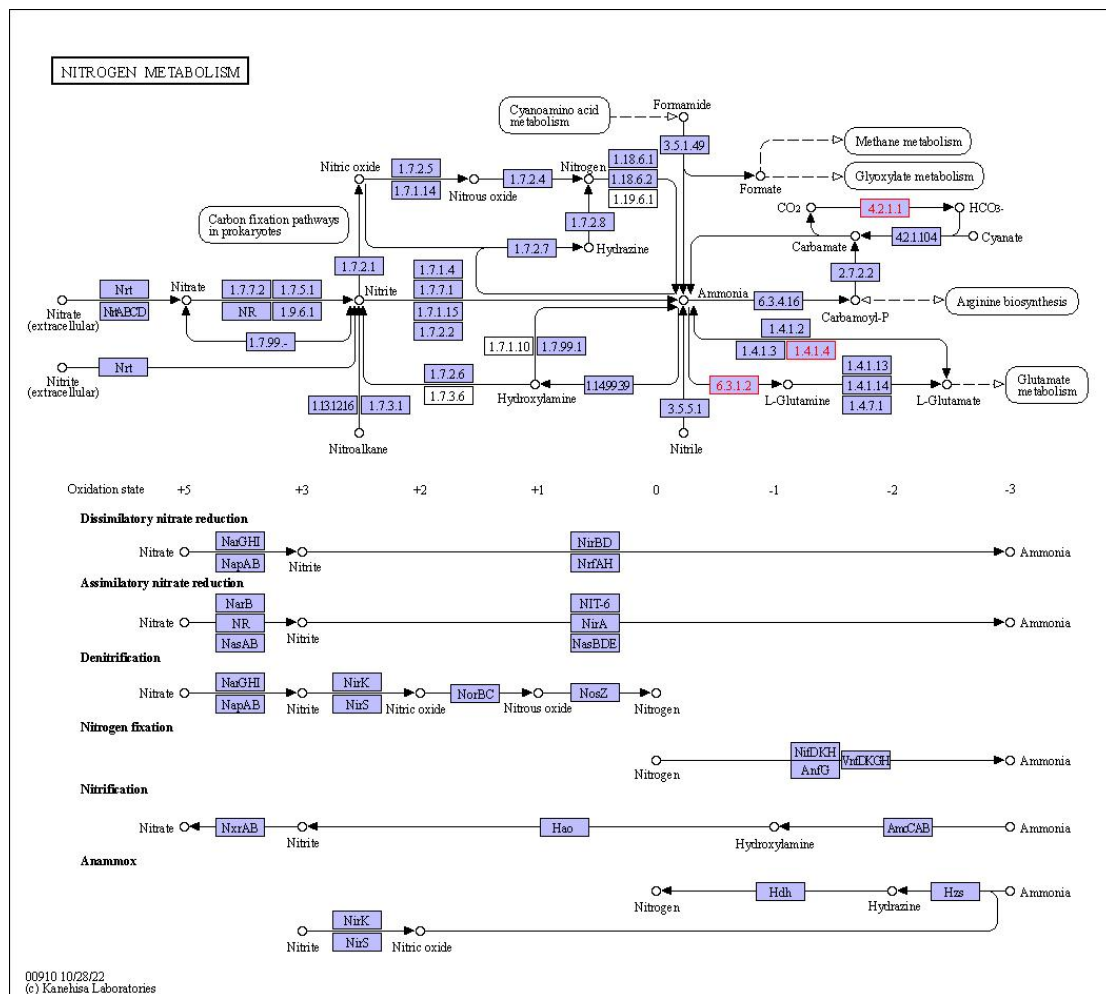


Figure S4. Nitrogen metabolism in *Ligilactobacillus agilis* W70 using the KEGG database. The red boxes represent the enzymes involved in nitrogen metabolism in strain W70. EC:6.3.1.2, glutamine synthetase; EC:1.4.1.4, glutamate dehydrogenase (NADP⁺).

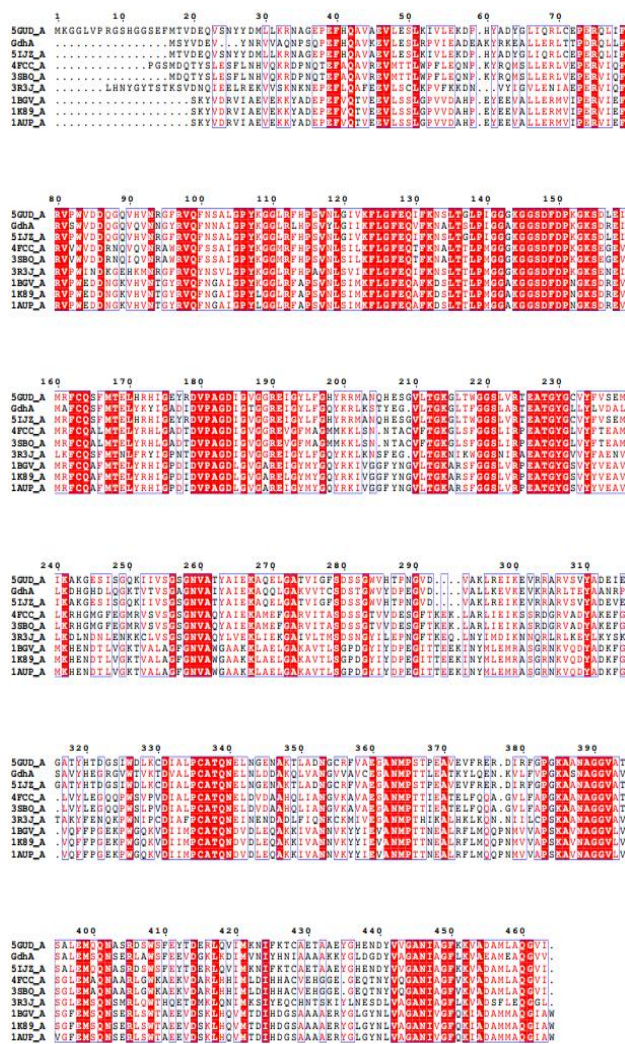


Figure S5. Multiple sequence alignment of the 5GUD, gdhA, 5IJZ, 4FCC, 3SBO, 3R3J, 1BGV, 1K89, and 1AUP proteins. (PDB templates 5GUD and 5IJZ from *Corynebacterium glutamicum*, 4FCC and 3SBO from *Escherichia coli*, 3R3J from *Plasmodium falciparum* 3D7, 1BGV, 1K89, and 1AUP from *Clostridium symbiosum*, and gdhA from *Ligilactobacillus agilis* W70)

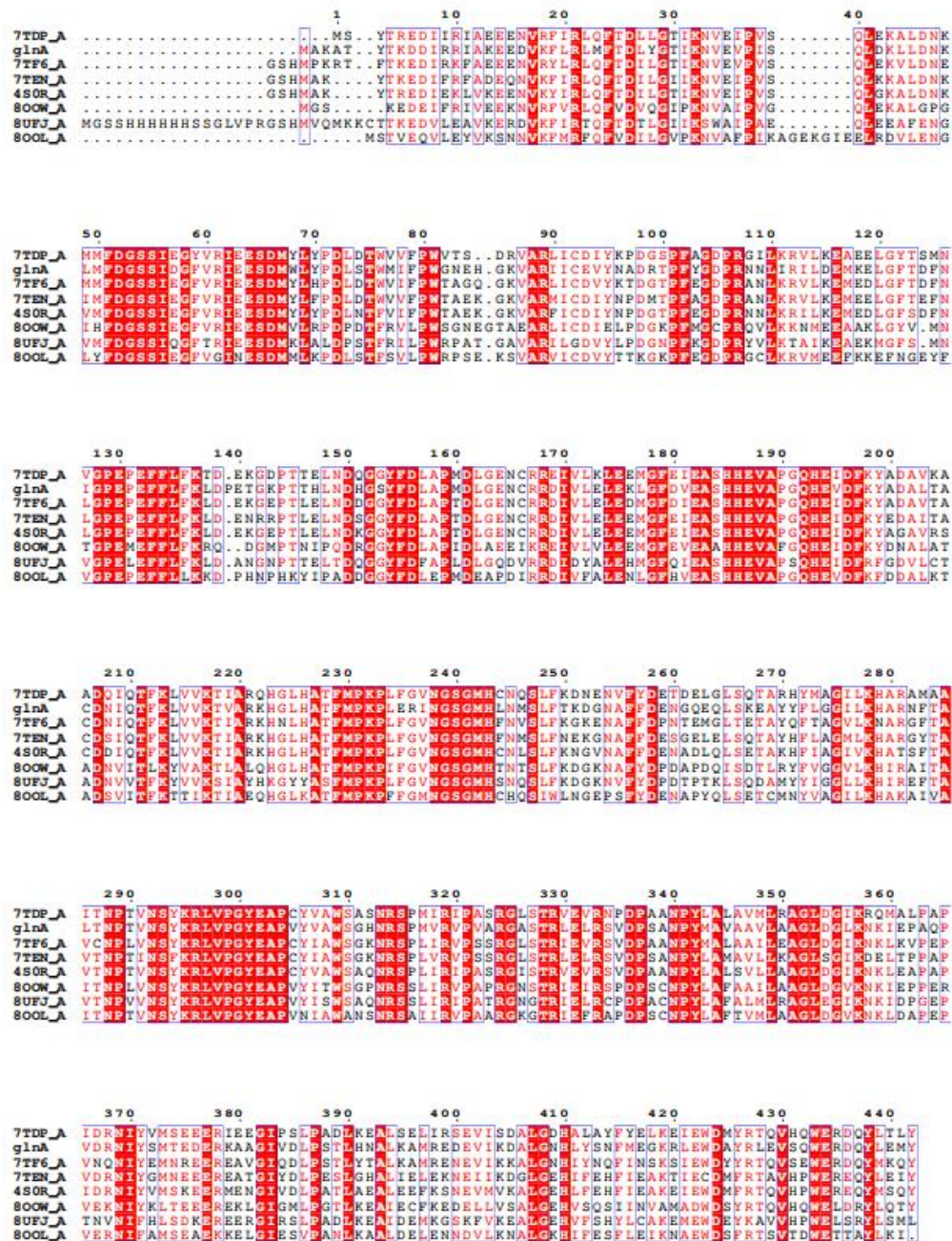


Figure S6. Multiple sequence alignment of the 7TDP, glnA, 7TF6, 7TEN, 4S0R, 8OOW, 8UFJ, and 8OOL proteins. (PDB templates 7TDP from *Paenibacillus polymyxa*, 7TF6 from *Staphylococcus aureus*, 7TEN from *Listeria monocytogenes*, 4S0R from *Bacillus subtilis*, 8OOW from *Methermicoccus shengliensis* DSM 18856, 8UFJ from *Methanosarcina mazei* Go1, and 8OOL from *Methanothermococcus thermolithotrophicus*, and glnA from *Ligilactobacillus agilis* W70)

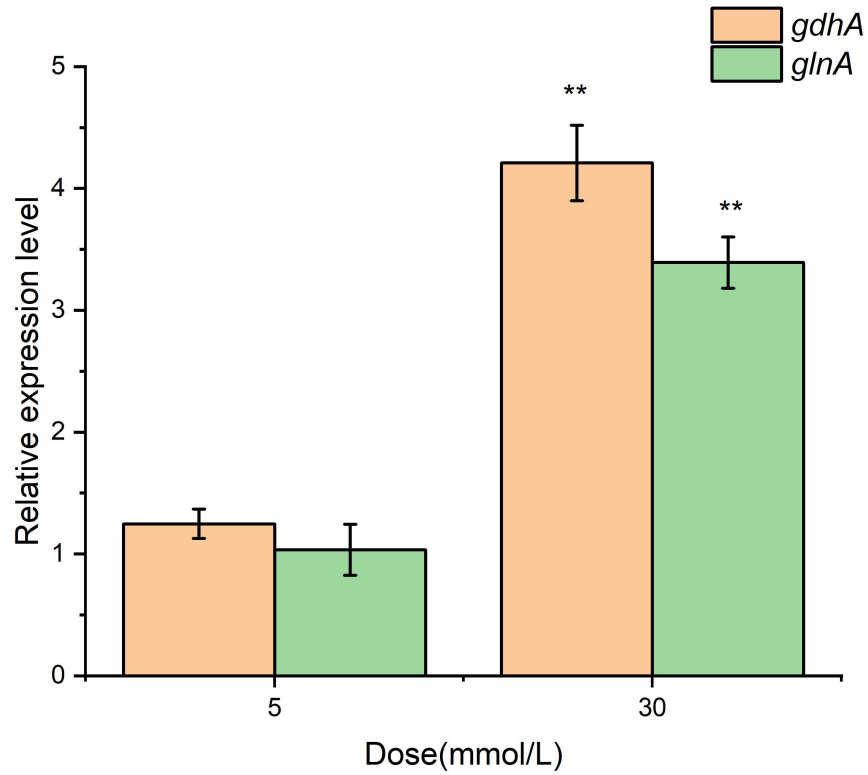


Figure S7. Relative expression of *gdhA* and *glnA* in 5 mmol/L and 30 mmol/L ammonium sulfate as the sole nitrogen source modified MRS medium. These results are means $\pm$ SD. * $P < 0.05$, ** $P < 0.01$ versus control.

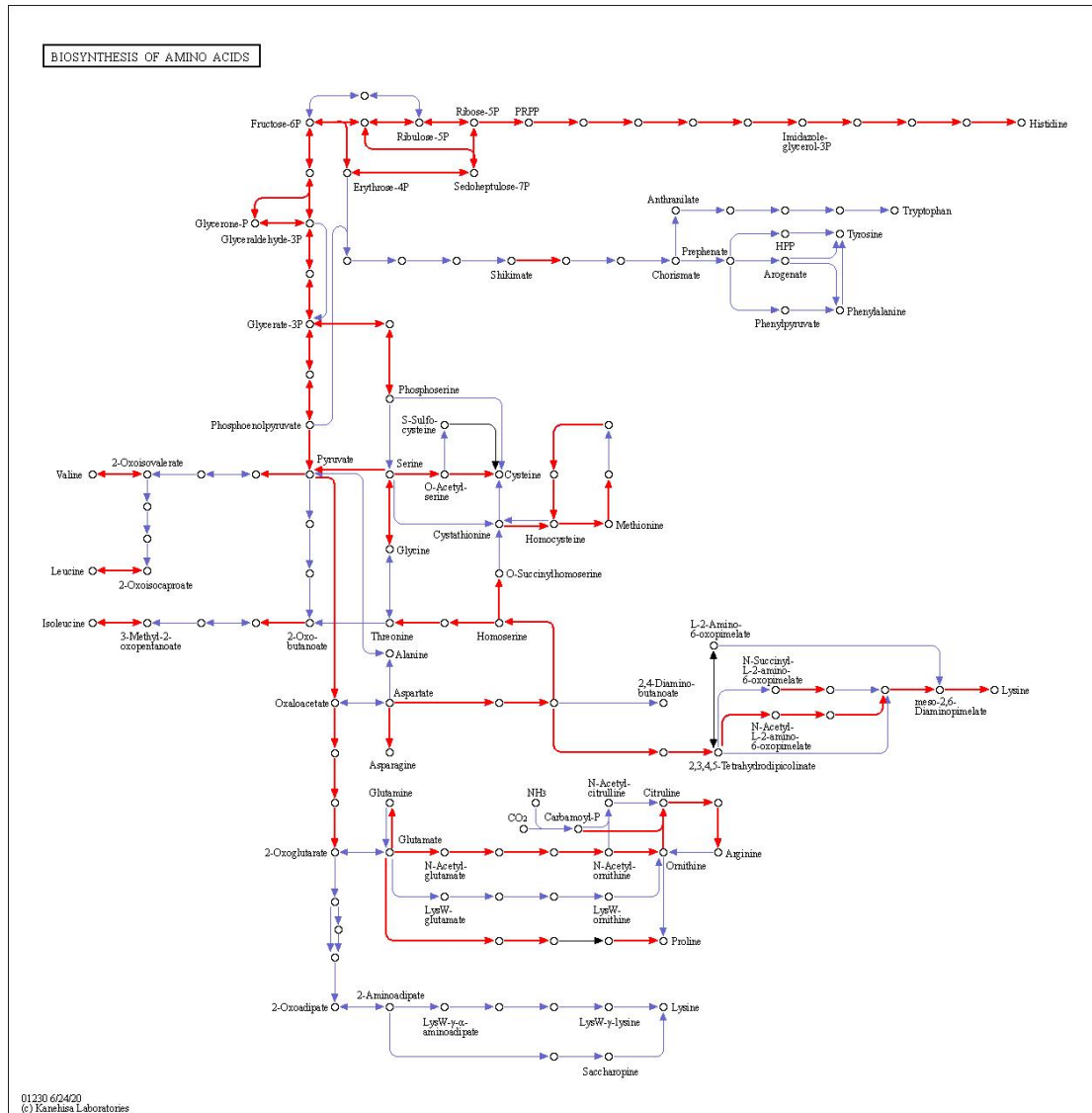


Figure S8. Biosynthesis pathway of amino acids in *Ligilactobacillus agilis* W70 using the KEGG database.

Figure S9. Propionate metabolism in *Ligilactobacillus agilis* W70 using the KEGG database. The red boxes represent the enzymes involved in propionate metabolism in strain W70. EC:1.1.1.27, L-lactate dehydrogenase; EC:2.3.1.54, formate C-acetyltransferase; EC:2.3.1.8, phosphate acetyltransferase; EC:2.7.2.1, acetate kinase.

Table S1. Primers of *gdhA* and *glnA* in RT-qPCR analysis

Target gene	Forward primer (5' to 3')	Reverse primer (5' to 3')	Product size (bp)
16S rDNA	ACTCCTACGGGAGGCA GCAG	ATTACCGCGGCTGCTG GC	180
<i>gdhA</i>	CTTACAAGGGCGGCTT ACGA	CGCCGATTGGAAGGGA AGTA	108
<i>glnA</i>	AGCAACGTAAACAGG GGCTT	ACGGCCCTAACTAACC CAAC	75

Table S2. The genes involved in ammonia assimilation of *Ligilactobacillus agilis* W70

Gene ID	gene1600	gene1769
Gene Name	<i>glnA</i>	<i>gdhA</i>
ChromosomeLocation	1577318-1578661	1753698-1755032
Description	type I glutamate--ammonia ligase	NADP-specific glutamate dehydrogenase
NR ¹	type I glutamate--ammonia ligase	NADP-specific glutamate dehydrogenase
Swiss-Prot	Glutamine synthetase OS= <i>Staphylococcus aureus</i> (strain MW2)	NADP-specific glutamate dehydrogenase OS= <i>Corynebacterium glutamicum</i> (strain ATCC 13032 / DSM 20300 / BCRC 11384 / JCM 1318 / LMG 3730 / NCIMB 10025)
KEGG ²	K01915 (glutamine synthetase [EC:6.3.1.2])	K00262 (glutamate dehydrogenase NADP ⁺) [EC:1.4.1.4])
COG ³	COG0174 (Glutamine synthetase)	COG0334 (Glutamate dehydrogenase/leucine dehydrogenase)
GO ⁴	GO:0006542;GO:0005524;GO:0004356 (glutamine biosynthetic process; ATP binding; glutamate-ammonia ligase activity)	GO:0006520;GO:0000166;GO:0004352 (cellular amino acid metabolic process; nucleotide binding; glutamate dehydrogenase NAD ⁺ activity)
Pfam	PF00120-Gln-synt_C;PF03951-Gln-synt_N (Glutamine synthetase, catalytic domain; Glutamine synthetase, beta-Grasp domain)	PF00208-ELFV_dehydrog; PF02812-ELFV_dehydrog_N (Glutamate/Leucine/Phenylalanine/Valine dehydrogenase; Glu/Leu/Phe/Val dehydrogenase, dimerisation domain)

¹NR: Non-Redundant Protein Database;

²KEGG: Kyoto Encyclopedia of Genes and Genomes;

³COG: Clusters of Orthologous Groups of proteins; ⁴GO: Gene Ontology

Table S3. The genes involved in amino acid biosynthesis of *Ligilactobacillus agilis* W70

Gene ID	KO ID	KO Name	KO Description
gene 0016	K01953	<i>asnB</i>	asparagine synthase (glutamine-hydrolysing) [EC:6.3.5.4]
gene 1327	K01953	<i>asnB</i>	asparagine synthase (glutamine-hydrolysing) [EC:6.3.5.4]
gene 0036	K04072	<i>adhE</i>	acetaldehyde dehydrogenase / alcohol dehydrogenase [EC:1.2.1.10 1.1.1.1]
gene 0084	K00055	-	aryl-alcohol dehydrogenase [EC:1.1.1.90]
gene 0086	K01426	<i>amiE</i>	amidase [EC:3.5.1.4]
gene 0093	K00020	<i>mmsB</i>	3-hydroxyisobutyrate dehydrogenase [EC:1.1.1.31]
gene 0128	K01939	<i>purA</i>	adenylosuccinate synthase [EC:6.3.4.4]
gene 0153	K00826	<i>ilvE</i>	branched-chain amino acid aminotransferase [EC:2.6.1.42]
gene 0162	K00382	<i>pdhD</i>	dihydrolipoamide dehydrogenase [EC:1.8.1.4]
gene 0177	K01733	<i>thrC</i>	threonine synthase [EC:4.2.3.1]
gene 0178	K00003	<i>hom</i>	homoserine dehydrogenase [EC:1.1.1.3]
gene 0179	K00872	<i>thrB</i>	homoserine kinase [EC:2.7.1.39]
gene 0207	K23304	<i>cysE</i>	serine O-acetyltransferase [EC:2.3.1.30]
gene 0208	K01738	<i>cysK</i>	cysteine synthase [EC:2.5.1.47]
gene 0251	K01652	<i>ilvB</i>	acetolactate synthase I/II/III large subunit [EC:2.2.1.6]
gene 0258	K01752	<i>sdaA</i>	L-serine dehydratase [EC:4.3.1.17]
gene 0259	K01752	<i>sdaA</i>	L-serine dehydratase [EC:4.3.1.17]
gene 0330	K00145	<i>argC</i>	N-acetyl-gamma-glutamyl-phosphate reductase [EC:1.2.1.38]
gene 0331	K00620	<i>argJ</i>	glutamate N-acetyltransferase / amino-acid N-acetyltransferase [EC:2.3.1.35 2.3.1.1]
gene	K00	<i>argB</i>	acetylglutamate kinase [EC:2.7.2.8]

0332	930		
gene 0333	K00 821	<i>argD</i>	acetylornithine/N-succinyldiaminopimelate aminotransferase [EC:2.6.1.11 2.6.1.17]
gene 0334	K00 611	<i>argF</i>	ornithine carbamoyltransferase [EC:2.1.3.3]
gene 0353	K01 929	<i>murF</i>	UDP-N-acetylmuramoyl-tripeptide--D-alanyl-D-alanine ligase [EC:6.3.2.10]
gene 0362	K00 016	<i>ldh</i>	L-lactate dehydrogenase [EC:1.1.1.27]
gene 1774	K00 016	<i>ldh</i>	L-lactate dehydrogenase [EC:1.1.1.27]
gene 0388	K00 286	<i>proC</i>	pyrroline-5-carboxylate reductase [EC:1.5.1.2]
gene 0428	K00 549	<i>metE</i>	5-methyltetrahydropteroyltriglutamate--homocysteine methyltransferase [EC:2.1.1.14]
gene 0429	K00 549	<i>metE</i>	5-methyltetrahydropteroyltriglutamate--homocysteine methyltransferase [EC:2.1.1.14]
gene 0430	K00 651	<i>metA</i>	homoserine O-succinyltransferase/O-acetyltransferase [EC:2.3.1.46 2.3.1.31]
gene 0431	K01 740	<i>metY</i>	O-acetylhomoserine (thiol)-lyase [EC:2.5.1.49]
gene 0505	K07 173	<i>luxS</i>	S-ribosylhomocysteine lyase [EC:4.4.1.21]
gene 0758	K07 173	<i>luxS</i>	S-ribosylhomocysteine lyase [EC:4.4.1.21]
gene 0572	K01 424	<i>ansA</i>	L-asparaginase [EC:3.5.1.1]
gene 0630	K05 822	<i>dapH</i>	tetrahydrodipicolinate N-acetyltransferase [EC:2.3.1.89]
gene 0631	K05 823	<i>dapL</i>	N-acetyldiaminopimelate deacetylase [EC:3.5.1.47]
gene 0688	K00 789	<i>metK</i>	S-adenosylmethionine synthetase [EC:2.5.1.6]
gene 0714	K01 756	<i>purB</i>	adenylosuccinate lyase [EC:4.3.2.2]
gene 0723	K23 265	<i>purQ</i>	phosphoribosylformylglycinamidine synthase subunit PurQ / glutaminase [EC:6.3.5.3 3.5.1.2]
gene 0725	K00 764	<i>purF</i>	amidophosphoribosyltransferase [EC:2.4.2.14]
gene 0863	K01 919	<i>gshA</i>	glutamate--cysteine ligase [EC:6.3.2.2]
gene 0879	K01 919	<i>gshA</i>	glutamate--cysteine ligase [EC:6.3.2.2]

gene 0873	K00891	<i>aroK</i>	shikimate kinase [EC:2.7.1.71]
gene 0882	K08968	<i>msrC</i>	L-methionine (R)-S-oxide reductase [EC:1.8.4.14]
gene 1006	K00609	<i>pyrB</i>	aspartate carbamoyltransferase catalytic subunit [EC:2.1.3.2]
gene 1008	K01956	<i>carA</i>	carbamoyl-phosphate synthase small subunit [EC:6.3.5.5]
gene 1009	K01955	<i>carB</i>	carbamoyl-phosphate synthase large subunit [EC:6.3.5.5]
gene 1041	K00600	<i>glyA</i>	glycine hydroxymethyltransferase [EC:2.1.2.1]
gene 1072	K00215	<i>dapB</i>	4-hydroxy-tetrahydrodipicolinate reductase [EC:1.17.1.8]
gene 1186	K00841	<i>patA</i>	aminotransferase [EC:2.6.1.-]
gene 1188	K00817	<i>hisC</i>	histidinol-phosphate aminotransferase [EC:2.6.1.9]
gene 1189	K01523	<i>hisE</i>	phosphoribosyl-ATP pyrophosphohydrolase [EC:3.6.1.31]
gene 1190	K01496	<i>hisI</i>	phosphoribosyl-AMP cyclohydrolase [EC:3.5.4.19]
gene 1191	K02500	<i>hisF</i>	imidazole glycerol-phosphate synthase subunit HisF [EC:4.3.2.10]
gene 1192	K01814	<i>hisA</i>	phosphoribosylformimino-5-aminoimidazole carboxamide ribotide isomerase [EC:5.3.1.16]
gene 1193	K02501	<i>hisH</i>	imidazole glycerol-phosphate synthase subunit HisH [EC:4.3.2.10]
gene 1194	K01693	<i>hisB</i>	imidazoleglycerol-phosphate dehydratase [EC:4.2.1.19]
gene 1195	K00013	<i>hisD</i>	histidinol dehydrogenase [EC:1.1.1.23]
gene 1196	K00765	<i>hisG</i>	ATP phosphoribosyltransferase [EC:2.4.2.17]
gene 1197	K02502	<i>hisZ</i>	ATP phosphoribosyltransferase regulatory subunit
gene 1198	K04486	-	histidinol-phosphatase (PHP family) [EC:3.1.3.15]
gene 1307	K01586	<i>lysA</i>	diaminopimelate decarboxylase [EC:4.1.1.20]
gene 1357	K00133	<i>asd</i>	aspartate-semialdehyde dehydrogenase [EC:1.2.1.11]
gene 1362	K01641	-	hydroxymethylglutaryl-CoA synthase [EC:2.3.3.10]

gene 1373	K01 714	<i>dapA</i>	4-hydroxy-tetrahydrodipicolinate synthase [EC:4.3.3.7]
gene 1400	K01 243	<i>mtnN</i>	adenosylhomocysteine nucleosidase [EC:3.2.2.9]
gene 1403	K01 778	<i>dapF</i>	diaminopimelate epimerase [EC:5.1.1.7]
gene 1415	K01 928	<i>murE</i>	UDP-N-acetylmuramoyl-L-alanyl-D-glutamate--2,6-diaminopi melate ligase [EC:6.3.2.13]
gene 1420	K00 928	<i>lysC</i>	aspartate kinase [EC:2.7.2.4]
gene 1422	K00 928	<i>lysC</i>	aspartate kinase [EC:2.7.2.4]
gene 1600	K01 915	<i>glnA</i>	glutamine synthetase [EC:6.3.1.2]
gene 1735	K01 834	<i>gpm A</i>	2,3-bisphosphoglycerate-dependent phosphoglycerate mutase [EC:5.4.2.11]
gene 1874	K01 834	<i>gpm A</i>	2,3-bisphosphoglycerate-dependent phosphoglycerate mutase [EC:5.4.2.11]
gene 1769	K00 262	<i>gdhA</i>	glutamate dehydrogenase (NADP+) [EC:1.4.1.4]
gene 1868	K00 135	<i>gabD</i>	succinate-semialdehyde dehydrogenase / glutarate-semialdehyde dehydrogenase [EC:1.2.1.16 1.2.1.79 1.2.1.20]
gene 2005	K00 135	<i>gabD</i>	succinate-semialdehyde dehydrogenase / glutarate-semialdehyde dehydrogenase [EC:1.2.1.16 1.2.1.79 1.2.1.20]
gene 1875	K14 155	<i>patB</i>	cysteine-S-conjugate beta-lyase [EC:4.4.1.13]
gene 1887	K00 824	<i>dat</i>	D-alanine transaminase [EC:2.6.1.21]
gene 1896	K17 363	<i>urdA</i>	urocanate reductase [EC:1.3.99.33]
gene 1956	K01 755	<i>argH</i>	argininosuccinate lyase [EC:4.3.2.1]
gene 1957	K01 940	<i>argG</i>	argininosuccinate synthase [EC:6.3.4.5]
gene 1973	K00 147	<i>proA</i>	glutamate-5-semialdehyde dehydrogenase [EC:1.2.1.41]
gene 1974	K00 931	<i>proB</i>	glutamate 5-kinase [EC:2.7.2.11]
gene 1978	K01 581	<i>speC</i>	ornithine decarboxylase [EC:4.1.1.17]
gene 1998	K00 058	<i>serA</i>	D-3-phosphoglycerate dehydrogenase / 2-oxoglutarate reductase [EC:1.1.1.95 1.1.1.399]

gene 1999	K00 831	<i>serC</i>	phosphoserine aminotransferase [EC:2.6.1.52]
gene 2008	K00 626	<i>atoB</i>	acetyl-CoA C-acetyltransferase [EC:2.3.1.9]
gene 2123	K00 547	<i>mmuM</i>	homocysteine S-methyltransferase [EC:2.1.1.10]
gene 2143	K00 820	<i>glmS</i>	glutamine---fructose-6-phosphate transaminase (isomerizing) [EC:2.6.1.16]
gene 2145	K00 865	<i>glxK</i>	glycerate 2-kinase [EC:2.7.1.165]

Table S4. The genes involved in propionate metabolism of *Ligilactobacillus agilis* W70

Gene ID	KO ID	KO Name	KO Description
gene0362	K00016	<i>ldh</i>	L-lactate dehydrogenase [EC:1.1.1.27]
gene1774	K00016	<i>ldh</i>	L-lactate dehydrogenase [EC:1.1.1.27]
gene0590	K00656	<i>pflD</i>	formate C-acetyltransferase [EC:2.3.1.54]
gene0525	K00625	<i>pta</i>	phosphate acetyltransferase [EC:2.3.1.8]
gene0708	K00925	<i>ackA</i>	acetate kinase [EC:2.7.2.1]
gene1856	K00925	<i>ackA</i>	acetate kinase [EC:2.7.2.1]
gene2133	K00925	<i>ackA</i>	acetate kinase [EC:2.7.2.1]

Appendix I.

> the partial sequence of the 16S rRNA gene of *Ligilactobacillus agilis* W70

CGAACGGGTGAGTAACACGTGGGTAACCTGCCAAAAGAGGGGGGATAACA
CTTGGAACAGGTGCTAATACCGCATAACCATGATGACCGCATGGTCATTAT
GTAAAAGATGGTTTCGGCTATCACTTTTGGATGGACCCGCGGCGTATTA
TGTGTTGGTGGGGTAACGGCCTACCAAGGTAATGATACGTAGCCGA
GGTTGATCGGCCACATTGGGACTGAGACACGGCCCAAACCTCTACGGGAG
GCAGCAGTAGGGAATCTTCCACAATGGGCGCAAGCCTGATGGAGCAACGC
CGCGTGAGTGAAGAAGGTCTTCGGATCGTAAAACTCTGTTGTTAGAGAAG
AACATGCAGGAGAGTAACTGTTCTTGTATTGACGGTATCTAACCAGAAAGC
CACGGCTAACTACGTGCCAGCAGCCGCGGTAATACGTAGGTGGCAAGCGTT
GTCCGGATTTATTGGGCGTAAAGGGAACGCAGGCGGTCCTTTAAGTCTGAT
GTGAAAGCCTTCGGCTTAACCGAAGAATTGCATTGGAACTGGAGGACTT
GAGTGCAGAAGAGGAGAGTGGAACTCCATGTGTAGCGGTGAAATGCGTAG
ATATATGGAAGAACACCAGTGGCGAAAGCGGCTCTCTGGTCTGTA
GCTGAGGTTTCGAAAGTGTGGGTAGCAAACAGGATTAGATACCCTGGTAGTC
CACACCGTAAACGATGAATGCTAAGTGTGAGGGTTTCCGCCCTTCAGTG
CTGCAGCTAACGCAATAAGCATTCCGCCTGGGGAGTACGACCGCAAGGTTG
AAACTCAAAGGAATTGACGGGGGGCCCGCACAAGCGGTGGAGCATGTGGTT
TAATTCGAAGCAACGCGAAGAACCTTACCAGGTCTTGACATCTTTTGACCA
TCTTAGAGATAAGATTTTCCCTTCGGGGACAAAATGACAGGTGGTGCATGG
CTGTCGTCAGCTCGTGTGTCGTGAGATGTTGGGTAAAGTCCCGCAACGAGCGC
AACCCTTGTTGTCAGTTGCCAGCATTAAAGTTGGGCACTCTGGCGAGACTGC
CGGTGACAAACCGGAGGAAGGTGGGGACGACGTCAAGTCATCATGCCCT
TATGACCTGGGCTACACACGTGCTACAATGGACGGTACAACGAGTCGCAAA
CTCGCGAGGGCAAGCTAATCTCTTAAAGCCGTTCTCAGTTTCGGATTGTAGG
CTGCAACTCGCCTACATGAAGTCGGAATCGCTAGTAATCGCGAATCAGCAT
GTCGCGGTGAATACGTTCCCGGGCCTTGTACACACCGCCCGTCACACCATG
AGAGTTTGTAACACCCAAAGCCGGTGGGGTAACCTTTAGGAGCTAGC

> *gdhA* encoded protein sequence of *Ligilactobacillus agilis* W70

MSYVDEVYNRVVAQNPSQPEFHQAVKEVLESLRPVIEADEAKYRKEALLERL
TTPDRQLLFRVSWVDDQGGVQVNNGYRVQFNNAIGPYKGGLRLHPSVYLGII
KFLGFEQVFKNALTSPIGGAKEGSDFDPKGKSDREIMAFQCSFMTELYKYIG
ADIDVPAGDIGTGGREIGYLFQYKRLKSTYEGVLTGKGLTFGGSLARTEATG
YGLLYLVDALLKDHGHDLQGKTVTVSGAGNVAIYAIEKAQQLGAKVVTCSDS
TGWVYDPEGVDVALLKEVKEVKRARLTEYAANRPSAVYHEGRGVWTVKTD
VALPCATQNELNLDDAKQLVANGVVAVCEGANMPTTLEATKYLQENKVLFP
GKASNAGGVAVSALEMSQNSERLAWSFEEVDGKLKDIMVNIYHNIAAAKK
YGLDGDYVAGANIAGFLKVAEAMEAQGVV

> *glnA* encoded protein sequence of *Ligilactobacillus agilis* W70

MAKATYTKDDIRRIAKEEDVKFLRLMFTDLYGTIKNVEVPISQLDKLLDNKL
MFDGSSIDGFVRIEESDMWLYPDLSTWMIFPWGNEHGKVARIICEVYNADRT
FYGDPRNNLIRILDEMKELGFTDFNIGPEPEFFLFKLPETGKPTTHLNDHGSY
FDLAPMDLGENCRDIVLELEKLGFDVEASHHEVAPGQHEVDFKYADALTA
CDNIQTFKLTVARKHGLHATFMPKPLERINGSGMHLNMSLFTKDGNAFF
DENGQEQLSKEAYYFLGGILKHARNFTALTNPTVNSYKRLVPGYEAPVYVAW
SGHNRSMPMVRVPVARGASTRLELRSDPSANPYMAVAVLAAGLDGLKNKIE

PAQPVDRNIYSMTEDERKAAGIVDLPSTLHNALKAMREDEVIKDALGNHLYS
NFMEGKRLEWDAYRLEVSQWERDQYLEMY