

Supplement to “B cell immune repertoire sequencing in smoking, vaping, and chronic obstructive pulmonary disease in the COPDGene cohort”

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Supplementary Methods

B cell receptor repertoire sequencing (BCR-seq) library preparation

Library preparation was performed according to Vollmers et al.¹. Briefly, a pooled set of 5 isotype-specific IGH constant region primers (RT_pool), containing 12 random nucleotides (nt) and partial Illumina adapters were added to the RNA samples, incubated at 72°C for 3 min then immediately placed on ice for 2 min. First-strand cDNA synthesis was then performed using Smartscribe reverse transcriptase (Takara) according to manufacturer's instructions. Second-strand cDNA synthesis was done using Phusion HiFi DNA polymerase (ThermoFisher) using a pool of six IGH variable region primers (V_FR1_pool) and a pool of five IGH constant region primers (C_pool), both containing 12 random nts and partial Illumina adapters. The reaction was incubated using the following heat cycle program: [98°C for 4 min, 52°C for 1 min, 72°C for 5 min]x2. The resulting double-stranded cDNA was purified and size-selected for molecules > 350bp using Zymo Select-a-Size columns. Finally, the double-stranded cDNA was amplified using 2x kapa hifi hotstart readymix (Roche) using primers containing complete Illumina adapters indexes (NexteraA_Index and NexteraB_Index primers) and the following heat cycle program: 95C for 3 mins, [98C for 20s, 67C for 15s, 72C for 1min]x26, 72C for 5 mins. PCR products were purified once using Ampure XP beads at a 0.7:1 ratio then pooled for multiplexed sequencing using the Illumina MiSeq sequencer and 2x300 run kits and flowcells.

AIRR-seq data processing

We downloaded from IMGT the reference germline sequences for IGH V (IMGT-gapped), D and J genes and created blast database files following instructions from NCBI IgBLAST. Blast database for IGH C genes were directly downloaded from NCBI IgBLAST release ftp site. Fasta files of preprocessed AIRR-seq reads were then aligned to IGH V, D, J and C genes using igblastn command line program from IgBLAST (version 1.19.0). We further filtered aligned reads keeping only those with non-missing v_support and j_support fields, v_support $\geq 1e-50$ and predicted to be productive. We used the spectralClones function from the scoper R package to infer clonal relationships between the sequence reads.

Supplementary Discussion

Our finding of higher levels of IgG1 and IgG3 in AA compared to NHW participants agrees with a previous report², which we extend by demonstrating this association in the context of smoking/vaping behaviors. However, there was no statistical interaction such that the combination of race and smoking/vaping variables demonstrated greater effects on isotype usage than the sum of their individual effects - though we note that our study is not well-powered to detect interactions of modest effect. Despite the lack of a large literature on racial differences in B-cell function, this topic is of substantial interest due to the increased risk of multiple myeloma in AA individuals³. IgG1 and IgG3 both have excellent complement activation and opsonization capabilities, and IgG3 is a potent immune effector, suggesting differences in response to pathogens and toxins. The degree to which our findings represent interactions between self-identified race and gene-by-environment interactions, specifically the effects of racism, are unclear.

References

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