

SPECTRUM MATCH OF PEPTIDE FALGAGCGVK





SPECTRUM MATCH OF PEPTIDE KTAAAAAAAAAAAAAAAA

D.





SUPPLEMENTARY FIGURE 1. Na-PTP peptides identified from Hek293 cells. Na-PTP peptides identified from Hek293 cells. (A). Spectrum match of peptide AFLTTAILVGMK from sample 49-4B. Scan n. 33098; PEAKS score: 33.61; mass 1263.7261 Da; Precursor error -1.2 ppm; Retention time 46.04 min. (B). Spectrum match of peptide FALGAGCGVK from sample 49-7A. Scan n. 17075; PEAKS score: 34.72; mass 978.4957 Da; Precursor error -0.2 ppm; Retention time 27.96 min. (C). Spectrum match of peptide ALGPGGGAGAR from sample 49-7A. Scan n. 15840; PEAKS score: 32.97; mass 924.4777 Da; Precursor error 0.7 ppm; Retention time 26.83 min. (D). Spectrum match of peptide KTAAAAAAAAAAAAAAAAAAAAAA from sample 10-2B. Scan n. 24968; PEAKS score: 50.09; mass 1440.78 Da; Precursor error -6 ppm; Retention time 36.63 min. (E). Spectrum match of peptide LGEDFLER from sample 10-1B. Scan n. 20145; PEAKS score: 34.28; mass 977.4818 Da; Precursor error 2.5 ppm; Retention time 33.27 min. (F). Spectrum match of peptide TLLTETGAGR from sample 73-21 (Hek293). Scan n. 7058; PEAKS score: 36.25; mass 1017.5455 Da; Precursor error 1 ppm; Retention time 17.32 min.The Orbitrap Fusion mass spectrometer was used for the measurement. Relates to Figure 2.



A. SPECTRUM MATCH OF PEPTIDE TLLTETGAGR (H1299 cells)

SUPPLEMENTARY FIGURE 2. Na-PTP peptides identified from H1299 cells) (A). Spectrum match of peptide TLLTETGAGR from sample 02-3 (H1299). Precursor Id. 26826; PEAKS score: 26.85; mass 1017.5455 Da; Precursor error -4.4 ppm; Retention time 22.26 min. The TimsTOF SCP mass spectrometer was used for the measurement. Relates to Figure 2.

SUPP. FIGURE 3.

Β.

CGC TTT GCA CTG GGG GCT GGA TGT GGT GTG AAG GCT

R F A L G A G C G V K A

RFALGAGCGVKAAPTRPGPGLQRCGARLRPGESSHPSLALHPKFSLLLGLGRWP (53aa)

QKTAAAAAAAAAAAAAANRMTLQHVLFMPIVCFAKPSKSMSLITSY (45aa)

ALGPGGGAGARGREGGGQDEEGRARRRGRAGVCGRGRGGPAGAARGRVRSRGRARGLGFFVSGAAARSGQRRRRGAEAALWGSRGARPRPCSPRPSS ATLPPSPGISGFFKILGAPGRAPGIGVQVTSGARLAGRETEAGPQRSAPGSVGSGCAAWACLAPLVPVPHPASVGPIRATSPAAVPSPAVAPWAEPPPPWSL ALLLRGVGETPAPPGRVEREGARSAPGTAPATPPALSTREPPGRCHP **(246aa)**

Ε.

D.

AGA CTA GGA GAA GAT TTT CTG GAG CGG AGA

R L G E D F L E R R

RLGEDFLERRDLSTLWKVVLKLAGPWELGRFPGPPVCHTTSLSSSSSAPAQATVDTVPVSHAFPEPVGSTGCQVGGRERMRMRCFLALGCCTFRFASPFHYKPCP HLCKHFLSQLTSVPQLASQAEAVACAKARSQDLINMAQDAAA **(147aa)**

F.

AAA ACT TTA TTA ACA GAA ACA GGT GCT GGG AGA TTT

K T L L T E T G A G R F

aaaactttattaacagaaacaggtgctgggagatttagtccatgggctctagttgcccaactgtga

KTLLTETGAGRFSPWALVAQL (20aa)

<u>SUPPLEMENTARY FIGURE 3</u>. Na-PTP peptides (green) from Hek293 cells identified by mass spectrometry analysis and verified by SFDB. The peptides and corresponding genomic sequences are shown. Below is shown the sequence until stop codon to calculate minimal Na-PTP length. Red indicates flanking amino acids used for predicting MHC-I binding peptides. (A) Peptide generated from anti-sense strand of gene CWF19L1. (B) Peptide generated from anti-sense strand of gene IQSEC2. (C) Peptide generated from intron 24 of gene LRRC7. Relates to figure 2 and Table 1. (D) Peptide generated from intron 1 of gene SNN. (E) Peptide derived from intron two of gene SPECC1. (F) Peptide derived from anti-sense strand of gene ZNF615. Note that peptide TLLTETGAGR was also detected from anti-sense strand of gene ZNF615 in H1299 cells. Relates to Figure 2 and Table 1.

SUPP. FIGURE 4.



SUPPLEMENTARY FIGURE 4. IFN- γ production by CD8+ T cells from PBMCs of individual healthy donors in response to Na-PTP peptides. Relative IFN- γ levels (fold change over vehicle) for each donor (D1 to D9) upon stimulation with peptides P1 (FAFLTTAIL), P2 (TTAILVGMKW), P3 (ALGAGCGVK), P4 (GEDFLERR), and P5 (LLTETGAGRF). Each panel represents the response to a specific peptide across donors, highlighting donor-specific IFN- γ production. Statistical significance was determined using a one-sample t-test comparing each donor's response to a theoretical mean of 1. Differences were considered statistically significant when P < 0.05. Relates to Figure 4.

SUPP. FIGURE 5.

IMMUNE RESPONSE ASSAYS



SUPPLEMENTARY FIGURE 5. (A). Immune response assay to peptide stimulation. PBMCs were isolated from buffy coats using a Ficoll gradient, cultured in complete medium with IL-2 and IL-7, and stimulated with specific peptides for 6 days (stimulation at days 1 and 5) to assess immune responses. IFN- γ levels in collected supernatants were quantified via ELISA. (B). Antigen presentation assay. Transiently co- transfected Hek293 cells with Kb and OVA constructs are co-cultured for 12h with OT-1 CD8+ T cell isolated from OT1-RAG mice. IL-2 concentration in the medium from co-cultured cells is analyzed by ELISA and is used as an indirect measure of antigenic peptide substrate synthesis. This figure was created in BioRender. Habault, J. (2024) https://BioRender.com/w63q319. Relates to Figures 4 and 5.

SUPP. FIGURE 6.





SUPPLEMENTARY FIGURE 6. Verification of the purity of synthetic peptides by high-performance liquid chromatography (HPLC). Representative chromatograms of the synthetic peptides show a single major peak, confirming their purity (A–G). Relates to Figure 4.

| Primer | Sequence (5' > 3') |
|-------------|---------------------------|
| CWF19L1 | |
| CWF19L1_Fw1 | GATGCTCTTATCATTAGCCATTATG |
| CWF19L1_Fw2 | GCACAATGATACACCACTTC |
| CWF19L1_Rev | GATTCCAATTCCTCCACATCC |
| IQSEC2 | |
| IQSEC2_Fw1 | GCTCCACAGCGCTGCAGC |
| IQSEC2_Fw2 | GCCTCGTGGGCGCAGCC |
| IQSEC2_Rev | CAGTGCAGGAGCTGGAACTG |
| OVA | |
| OVA-Fw | GATGAAGACACAAGCAAT |
| OVA-Rev | AAGCCATTGATGCCACTCTA |
| β-Actin | |
| Actin-Fw | TCACCCACACTGTGCCCATCTACGA |
| Actin-Rev | TGAGGTAGTCAGTCAGGTCCCG |

SUPPLEMENTARY TABLE 1. Specific primers used for RT-qPCR to evaluate the sense or anti-sense transcription of the CWF19L1 and IQSEC2 (peptides) or OVA (model) genes. Actin amplification was used to normalize data. Relates to Figures 3 and 5.