Supplementary Figure

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Supplementary Table

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Supplementary method

Genomic and clinical data sources and preprocessing.

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Cell line.



Figure S1. Heatmap of 22 immune cell types based on immune microenvironment clustering. **(A-D)** Heatmap of 22 immune cell types based on consensus clustering identifying the high- and low-immune subgroups in BLCA, SKCM, OV and UCEC. Abbreviation list of tumor cohorts from TCGA is given in **Table S4**.



Figure S2. Evaluation of the predictive efficiency of the prognostic model. ROC curves representing the predictive capacity for OS and PFS of prognostic model in the indicated immunotherapy cohorts. OS, overall survival; PFS, progression-free survival; ROC, receiver operating characteristic curve.



Figure S3. Difference of TME components between high- and low-risk groups. **(A-C)** The infiltrating levels of 22 immune cell types between the high- and low-risk groups in IMvigor210 **(A)**, IMmotion150 **(B)** and POPLAR **(C)**. **(D-M)** The correlation between risk score and the level of infiltrating immune cell in IMvigor210 **(D-J)** and IMmotion150 **(K-M)**. **(N-P)** Comparison of immune, stromal, and ESTIMATE score distribution between high- and low-risk groups in IMvigor210 **(N)**, IMmotion150 **(O)** and POPLAR **(P)**. TME, tumor microenvironment. * P < 0.05, ** P < 0.01, ***P < 0.001, ns no significant.



Figure S4. GSEA analysis based the genelist ordered by log2(FoldChange). **(A-D)** GSEA plot for significantly enriched hallmark gene sets associated with the risk score in IMvigor210 **(A)**, IMmotion150 **(B)**, POPLAR **(C)** and GSE91061 **(D)**. GSEA: Gene Set Enrichment Analysis



Figure S5. Heatmap of ssGSEA scores of TME and hallmark gene sets. **(A-F)** Heatmap showing the significantly different gene sets of TME (left panel) and hallmark (right panel) gene sets between risk groups in IMvigor210 **(A-B)**, IMmotion150 **(C-D)** and POPLAR **(E-F)**. ssGSEA, single-sample gene set enrichment analysis.



Figure S6. Difference of pathways activity between the high- and low-risk groups. **(A-C)** The pathway activities levels of 14 common malignant signaling pathways between high- and low-risk groups in IMvigor210 **(A)**, IMmotion150 **(B)** and POPLAR **(C)**. * P < 0.05, ** P < 0.01, ***P < 0.001, ns no significant.



Figure S7. Univariate Cox regression analysis of risk score. (A) Forest plots showed the relation between risk score and OS in TCGA pan-cancer. (B) Forest plots showed the relation between risk score and DSS in TCGA pan-cancer. Red color represents significant results (P < 0.05). DSS, Disease-specific survival; OS, overall survival; TCGA, The Cancer Genome Atlas. Abbreviation list of tumor cohorts from TCGA is given in **Table S4**.



Figure S8. Correlation analysis between risk score and ssGSEA score in in different cancer types from the TCGA dataset. **(A-B)** Correlation analysis between risk score and ssGSEA score of hallmark **(A)** and TME **(B)** in TCGA pan-cancer. ssGSEA, single-sample gene set enrichment analysis; TCGA, The Cancer Genome Atlas. Abbreviation list of tumor cohorts from TCGA is given in **Table S4**.



Lung normal

Figure S9.

RPLP0 protein expression profile in normal tissues and tumors.(A) Expression level of RPLP0 protein in TCGA cohorts based on the data from CPTAC database. (B) Images of immunohistochemistry staining of RPLP0 in indicated tumors and normal specimens from the HPA database. CPTAC, Clinical Proteomic Tumor Analysis Consortium ;HPA, the Human Protein Atlas; TCGA, The Cancer Genome Atlas. Abbreviation list of tumor cohorts from TCGA is given in **Table S4**. * P < 0.05, ** P < 0.01, ***P < 0.001.



Figure S10.

Bar charts representing the mRNA expression level of RPLP0 in tumor tissues from mice in indicated groups (n = 5 per group). One-way ANOVA followed by Dunnett's multiple comparisons tests were used to compare data in four groups. * P < 0.05, ** P < 0.01, ***P < 0.001.

Indication	IMvigor210	POPLAR	IMmotion150	IMmotion151	CheckMate	GSE91061
Sample count	348	81	162	407	170	51
Arm	Atezolizumab	Atezolizuma b	Atezolizumab (n=77)	Atezolizumab+ Bevacizumab	Nivolumab	Nivolumab
			Atezolizumab+ Bevacizumab (<u>n=85</u>)			
Gender Male (%)	272 (78.2)	56 (69.1)	118 (72.8)	281 (69.0)	128 (75.3)	NA
Age >=60 years (%)	NA	47 (58.0)	92 (56.8)	243 (59.7)	101 (59.8)	NA
Samples with WES	234	50	107	341	112	46
TMB (muts/Mb), median (IQR)	8.0 (9.0)	15.5 (13.2)	9.0 (6.4)	3.5 (2.6)	58 (33.5)	182.5 (361.5)
ORR, CR/PR, n(%)	68 (19.5)	47 (58.0)	102 (63.0)	150 (39.5)	37 (23.0)	10 (20.4)
OS (months), median (IQR)	8.0 (14.9)	12.6 (13.1)	NA	NA	22.8 (40.5)	16.1 (20.5)
PFS (months), median (IQR)	NA	2.8 (7.0)	8.1 (15.9)	9.7 (11.3)	3.8 (7.6)	NA

Table S1: Patients clinical characteristics

Taget		Primer(5' - 3')
ACTB	Forward	CATGTACGTTGCTATCCAGGC
(numan)	Reverse	CTCCTTAATGTCACGCACGAT
RPLP0	Forward	TGGCAGCATCTACAACCCTG
(numan)	Reverse	GACAAGGCCAGGACTCGTTT
Actb	Forward	GGCTGTATTCCCCTCCATCG
(mouse)	Reverse	CCAGTTGGTAACAATGCCATGT
Rplp0	Forward	TGAGATTCGGGATATGCTGTTGG
(mouse)	Reverse	CGGGTCCTAGACCAGTGTTCT

Table S2: Olige sequences used in quantitative real time BCB

Table S3: siRNA target sequence used for gene knockdown

Name	Sequence (5' - 3')
Negative control siRNA	UUCUCCGAACGUGUCACGUTT
si-Rplp0	CGGAGGAAUCAGAUGAGGAUA

Abbreviation	Detail	
ACC	Adrenocortical carcinoma	
BLCA	Bladder Urothelial Carcinoma	
BRCA	Breast invasive carcinoma	
0500	Cervical squamous cell carcinoma and endocervical	
CESC	adenocarcinoma	
CHOL	Cholangio carcinoma	
COAD	Colon adenocarcinoma	
DLBC	Lymphoid Neoplasm Diffuse Large B-cell Lymphoma	
ESCA	Esophageal carcinoma	
GBM	Glioblastoma multiforme	
HNSC	Head and Neck squamous cell carcinoma	
KICH	Kidney Chromophobe	
KIRC	Kidney renal clear cell carcinoma	
KIRP	Kidney renal papillary cell carcinoma	
LGG	Brain Lower Grade Glioma	
LIHC	Liver hepatocellular carcinoma	
LUAD	Lung adenocarcinoma	
LUSC	Lung squamous cell carcinoma	
MESO	Mesothelioma	
OV	Ovarian serous cystadenocarcinoma	
PAAD	Pancreatic adenocarcinoma	
PCPG	Pheochromocytoma and Paraganglioma	
PRAD	Prostate adenocarcinoma	
READ	Rectum adenocarcinoma	
SARC	Sarcoma	
SKCM	Skin Cutaneous Melanoma	
STAD	Stomach adenocarcinoma	
TGCT	Testicular Germ Cell Tumors	
THCA	Thyroid carcinoma	
THYM	Thymoma	
UCEC	Uterine Corpus Endometrial Carcinoma	
UCS	Uterine Carcinosarcoma	
UVM	Uveal Melanoma	

Table S4: Abbreviation of 32 TCGA cancer types

Symbol	Ensemble ID	Entrez ID	SeqNames	Length	N_Exons	
ABCC1	ENSG00000103222.17	4363	chr16	8677	34	
ABLIM2	ENSG00000163995.17	84448	chr4	6303	33	
ADAM7	ENSG0000069206.14	8756	chr8	5226	27	
ADAT2	ENSG00000189007.14	134637	chr6	7493	7	
ADCY7	ENSG00000121281.11	113	chr16	9749	36	
ADH1B	ENSG00000196616.11	125	chr4	6534	14	
ADORA1	ENSG00000163485.14	134	chr1	6625	11	
AHCY	ENSG00000101444.11	191	chr20	4065	14	
AHCYL2	ENSG00000158467.15	23382	chr7	5887	20	
ANKRD2	ENSG00000165887.10	26287	chr10	4370	26	
ANKRD7	ENSG00000106013.11	56311	chr7	1358	7	
ANXA13	ENSG00000104537.15	312	chr8	1593	12	
APLP2	ENSG0000084234 15	334	chr11	7415	35	
ARAP1	ENSG0000186635 13	116985	chr11	5646	36	
ARHGEF4	ENSG0000136002 15	50649	chr2	18645	34	
ARI 10	ENSG0000175414.6	285598	chr5	6194	8	
ARI 4D	ENSG00001759064	379	chr17	1585	2	
ASGR2	ENSG0000161944 15	433	chr17	3292	19	
	ENSG0000069849.9	483	chr3	1853	7	
	ENSG00000147614 3	245972	chr8	2370	8	
	ENSC0000169857.6	57000	chr15	1555	7	
B4GALNT3	ENSG0000139044.9	283358	chr12	3438	20	
ΒΔΔΤ	ENSG0000136881 10	570	chrQ	3528	5	
	ENSC0000130001.10	55330	chr4	15/6	1	
BLOC 134	ENSC0000160504 11	646	chr15	0875	12	
	ENSC00000222000 7	140479 chr1	chr1		12	
C10orf62	ENSC0000222009.7	149470	chr10	10/1	<u> </u>	
C100/102	ENSC0000154403 16	119611	chr10	6222	17	
C100190	ENSG00000134493.10	10011	ohr12	0222	52	
C1201134	ENSC0000182226 12	716	chr12	2094	14	
Clorf18	ENSC00000182520.13	51161	chr3	290 4 5047	14	
C8B	ENSG000000000000000000000000000000000000	732	chr1	3025	11	
	ENSC0000021052.11	03580	chr12	10600	75	
	ENSG00000151002.15	93369		2727	73	
	ENSC00000102909.10	10/02	chr10	1677	10	
	ENSC0000064012 20	9/1	chr?	1600/	79	
CASF0	ENSC0000004012.20	873	chr21	2684	10	
	ENSC00000139220.11	147272	chr19	2004	4	
	ENSC00000135736.5	02022	chi 10	2420	17	
	ENSC00000133730.3	151887	chr3	73/2	11	
	ENSC0000091980.14	6360	chr7	360	3	
	ENSC0000158477.6	0009	chr1	2006	5	
CD1A CD276	ENSC0000102255 16	909	ohr15	2090	0 12	
CD270	ENSG00000103835.10	51202	chi 15 obr10	1251	13	
	ENSC0000158825.5	070	chi 19	1201	5	
	ENSC0000162814.6	970	chi i	900	4	
	ENSC0000120610.1	1000	ohr12	10414	13	
	ENSC0000106477 47	05601	chillZ	902	0	
	ENSC0000000071 14	2075	GHL/	100/0	23 26	
	ENSC000002444445	2070		4027	∠U Q	
	ENSC0000116795 12	3070	ohr1	1492	0	
	ENSC0000124265 11	100/0	ohr1	4003	10	
	ENSC0000020540244	2426		3770	13	
	ENSC0000117071 10	3420	CIII4	2100	- 17	
	ENSCO000117971.10	0460	chi 15	0440 12666	/ E	
	ENSG0000122863.5	9469	cnr10	13000	5	

Table S5: Details of 304 immunosuppression-related genes (ISRGs)

Symbol	Ensemble ID	Entrez ID SeqNames Length		Length	N_Exons	
CHST6	ENSG00000183196.7	4166	chr16	9422	5	
CLEC11A	ENSG00000105472.11	6320	chr19	1407	4	
CLEC5A	ENSG00000258227.5	23601	chr7	8514	14	
CLIP4	ENSG00000115295.18	79745	chr2	10850	39	
CNN2	ENSG0000064666.13	1265	chr19	2458	7	
COL16A1	ENSG0000084636.16	1307	chr1	6264	73	
COL18A1	ENSG00000182871.13	80781	chr21	6058	43	
COMMD10	ENSG00000145781.7	51397	chr5	1424	7	
CORO2B	ENSG00000103647.11	10391	chr15	4410	16	
CPEB1	ENSG00000214575.8	64506	chr15	6874	22	
CRACR2A	ENSG00000130038.8	84766	chr12	7005	25	
CRB2	ENSG00000148204 10	286204	chr9	9771	23	
CRTAC1	ENSG0000095713 12	55118	chr10	3532	18	
CRYAB	ENSG00000109846.6	1410	chr11	2610	10	
CSTA	ENSG0000121552.3	1475	chr3	828	3	
CYP19A1	ENSG0000137869 12	1588	chr15	6618	16	
CYP2C18	ENSG0000108242 11	1562	chr10	2545	9	
CYP2C19	ENSG0000165841.8	1557	chr10	1473	9	
CYP2C9	ENSG0000138109.9	1559	chr10	2153	11	
CVP8B1	ENSC0000180432.5	1582	chr3	3050	1	
	ENSC000002057954	102668	chr2	2710		
	ENSC0000182308.6	285/20		<u> </u>	1	
	ENSC0000133083 13	203429	chr13	25010	24	
	ENSC0000135005.15	8604	chr8	2721	17	
	ENSC0000073737.15	10170	chr2	5822	17	
	ENSC000001073131.13	10170	ohr6	2206	19	
	ENSC0000196721.11	10400	chi0	3200	20	
	ENSG00000104329.10	1930	chio obr7	29190	2 29	
	ENSC00000100203.10	0002	ohr22	1040	15	
	ENSG00000100353.10	0004	CIIIZZ	1949	15	
	ENSG00000147077.9	0007	chio chr10	1270	0	
	ENSG00000105131.0	79002	chi 19	2909	10	
	ENSG00000144466.13	04141		4907	12 E	
	ENSG00000115303.12	04141	CHIZ	2102	5 65	
	ENSG00000112319.10	2070	CIIIO	20074	65	
	ENSG00000166659.0	149297		1401	<u>ک</u>	
	ENSG00000110001.9	20232	Chr I	3103	13	
	ENSG00000179639.9	2205	Chr I	2101	10	
FCGR3B	ENSG00000102747.8	2215		3101	10	
FCGRI	ENSG00000104870.11	2217	Chi 19	3000	13	
FCN2	ENSG00000160339.14	2220	cnr9	1003	10	
FGA	ENSG00000171560.13	2243	CNr4	5297	1	
FGF11	ENSG0000161958.9	2256	Chr17	2/15	C	
FGL1	ENSG00000104760.15	2267		2122	14	
FLNC	ENSG00000128591.14	2318	chr/	10065	50	
FMOD	ENSG00000122176.10	2331	chr1	5528	5	
FOLR2	ENSG00000165457.12	2350	chr11	1519	8	
FOXA2	ENSG00000125798.13	3170	chr20	4689	6	
GBG11	ENSG00000148288.10	26301	chr9	5045	12	
GEMIN5	ENSG0000082516.8	25929	cnr5	5521	29	
GFP12	ENSG0000131459.11	9945	chr5	4048	21	
GLI1	ENSG0000111087.8	2/35	chr12	40/5	16	
GLI2	ENSG00000074047.19	2736	chr2	11341	17	
GLRX3	ENSG00000108010.10	10539	chr10	3254	27	
GNAL	ENSG00000141404.14	2774	chr18	18016	36	
GOLGA8O	ENSG00000206127.9	728047	chr15	8967	21	
GSTP1	ENSG00000084207.14	2950	chr11	958	7	
HDAC10	ENSG00000100429.16	83933	chr22	2707	20	

Symbol	Ensemble ID	Entrez ID SeqNames Lengt		Length	N_Exons	
HHIPL2	ENSG00000143512.11	79802	chr1	2574	9	
HOXA11	ENSG0000005073.5	3207	chr7	2653	2	
HOXA3	ENSG00000105997.21	3200	chr7	6645	11	
HR	ENSG00000168453.13	55806	chr8	6310	21	
HSPA1A	ENSG00000204389.9	3303	chr6	2429	1	
HSPA1B	ENSG00000204388.6	3304	chr6	2520	1	
HSPD1	ENSG00000144381.15	3329	chr2	2659	14	
HTR7	ENSG00000148680.14	3363	chr10	7530	7	
IFI27L2	ENSG00000119632.3	83982	chr14	467	4	
IFNE	ENSG00000184995.7	338376	chr9	1475	1	
IGFBP1	ENSG00000146678.8	3484	chr7	1650	4	
II 18	ENSG0000150782 10	3606	chr11	1147	6	
II 6R	ENSG0000160712 11	3570	chr1	16064	19	
IMPDH1	ENSG0000106348 15	3614	chr7	5523	29	
IRS2	ENSG00000185950.8	8660	chr13	6998	2	
IRX6	ENSG0000159387 7	79190	chr16	3432	8	
ISIR	ENSG0000129009 11	3671	chr15	2385	3	
	ENSG00000076513	380/3/	chr6	23478	23	
	ENSC0000140854 11	10300	chr16	12/10	20	
KIEG	ENSC0000164627.16	221/58	chr6	4603	26	
	ENSC0000187961 12	330/51	chr1	6/27	20	
	ENSG00000167901.12	0003	chr1	6024	50	
	ENSC0000102413.13	9903	chi i	6409	20	
	ENSC00000112612	2012	chito chr7	0490 5946	29	
	ENSG0000091130.12	3912	CIII /	2040 2055	34	
	ENSG00000125872.7	104312	Chr20	2900	5	
LY6E	ENSG0000160932.9	4061	chr8	<u>chr8 1280 5</u>		
LY6K	ENSG00000160886.12	54742	chr8	5289	× ×	
LZIS2	ENSG00000107816.16	84445	chr10	3454	8	
MAP4K4	ENSG0000071054.14	9448	chr2	9837	41	
MARCO	ENSG0000019169.10	8685	chr2	1835	1/	
MARS2	ENSG00000247626.4	92935	chr2	3087	1 8	
MATN3	ENSG00000132031.11	4148	chr2	2583		
MCTP1	ENSG00000175471.18	79772	chr5	22366	81	
MEIOB	ENSG00000162039.13	254528	chr16	2372	16	
METTL8	ENSG00000123600.17	79828	chr2	9622	18	
MGMT	ENSG00000170430.9	4255	chr10	1900 9		
MLLT11	ENSG00000213190.3	10962	chr1	2158	2	
MMADHC	ENSG00000168288.11	27249	chr2	1437	8	
MMP7	ENSG00000137673.7	4316	chr11	1119	6	
MRPS27	ENSG00000113048.15	23107	chr5	2900	13	
MYBPC2	ENSG0000086967.9	4606	chr19	3598	28	
NBL1	ENSG00000158747.12	4681	chr1	3620	12	
NCKAP5L	ENSG00000167566.15	57701	chr12	5544	19	
NEFM	ENSG00000104722.12	4741	chr8	3627	4	
NKAIN4	ENSG00000101198.13	128414	chr20	2517	10	
NLRP1	ENSG0000091592.14	22861	chr17	6180	19	
NOP56	ENSG00000101361.13	10528	chr20	4206	24	
NPAS2	ENSG00000170485.15	4862	chr2	8660	31	
NRBP2	ENSG00000185189.14	340371	chr8	8574	29	
OBSL1	ENSG00000124006.13	23363	chr2	15986	32	
OSM	ENSG0000099985.3	5008	chr22	3855	5	
ОХТ	ENSG00000101405.3	5020	chr20	513	3	
PARVB	ENSG00000188677.13	29780	chr22	2964	23	
PAX8	ENSG00000125618.15	7849	chr2	6778	14	
PCDHA10	ENSG0000250120 5	56139	chr5	12752	9	
PDCD4	ENSG00000150593 14	27250	chr10	5737	22	
PDE11A	ENSG00000128655 15	50940	chr2	10019	26	
		· · · · · · ·				

Symbol	Ensemble ID	Entrez ID SeqNames Lengt		Length	N_Exons	
PHACTR3	ENSG0000087495.15	116154	chr20	3541	18	
PI15	ENSG00000137558.6	51050	chr8	8091	10	
PID1	ENSG00000153823.17	55022	chr2	2822	4	
PILRB	ENSG00000121716.17	29990	chr7	1438	4	
PIP5KL1	ENSG00000167103.10	138429	chr9	3532	15	
PKNOX2	ENSG00000165495.14	63876	chr11	4140	18	
PLA2G12B	ENSG00000138308.5	84647	chr10	3942	9	
PLA2G2A	ENSG00000188257.9	5320	chr1	1258	8	
PLA2G4E	ENSG00000188089.12	123745	chr15	4779	20	
PLP2	ENSG00000102007.9	5355	chrX	1049	5	
PLTP	ENSG00000100979.13	5360	chr20	3352	20	
POU5F1B	ENSG0000212993.4	5462	chr8	1599	1	
POU5F2	ENSG0000248483.5	134187	chr5	1295	1	
POU6F2	ENSG00000106536.18	11281	chr7	2826	12	
PPP1R14B	ENSG0000173457.9	26472	chr11	987	4	
PPP4R4	ENSG0000119698 10	57718	chr14	17373	107	
PRAM1	ENSG0000133246 10	84106	chr19	2602	12	
PRSS3	ENSG0000010438 15	5646	chr9	1558	10	
PSMD14	ENSG0000115233 10	10213	chr2	1717	10	
	ENSC0000106246 16	26024	chr7	5505	8	
PTGIR	ENSC00001600137	5730	chr10	5583	11	
PVGB	ENSC0000100013.7	583/	chr20	1731	23	
	ENSC0000128340 13	5880	chr20	2172	13	
	ENSC00000120340.13	720475	chr2	7450		
	ENSC0000108774 4	0192	ohr12	1706	0	
	ENSC000007294 14	64295	ohr16	6526	<u> </u>	
	ENSG0000007364.14	04200	ohr11	7000	43	
	ENSC0000160439.5	200000100459.5 204225 1117 7		7023	0	
	ENSC0000136260.11	00.11 300391 CIII1 7003		31 10		
	ENSG00000142541.15	23521	chi 19	2017	10	
	ENSG00000003177.11	6141	chr19	1/00	14	
	ENSG00000105040.11	0142	CIII 19	0/1	о О	
RPL19	ENSG00000108298.8	0143	Chr 17	1200	0 F	
	ENSG00000131409.11	0100	CIII 17	490	5 10	
RPL20	ENSG00000106107.11	0100	CHI 19	10100	10	
	ENSG00000174444.13	0124		1441	10	
	ENSG0000089157.14	0175	Chr12	1300	9	
	ENSG00000137818.10	6000		512	4	
RPS10	ENSG00000231500.5	6222	CIIIO	04Z	0	
RPS19	ENSG00000105372.5	0223	chr19	849	0	
RP52	ENSG00000140988.14	6187	CNF16	942	1	
RP523	ENSG00000186468.11	6228		3314	4	
RP524	ENSG00000138326.17	6229	chr10	9349	45	
RP527A	ENSG00000143947.11	6233		1469	8	
RPS28	ENSG0000233927.4	6234	cnr19	382	4	
RPS3	ENSG00000149273.13	6188	chr11	7592	27	
RPS5	ENSG0000083845.7	6193	chr19	/41	6	
RPS8	ENSG00000142937.10	6202	chr1	705	6	
RPS9	ENSG00000170889.12	6203	chr19	35697	220	
	ENSG0000052749.12	23223	chr10	4426	34	
	ENSG0000132275.9	23378	chr11	1/51	(
S100A1	ENSG00000160678.10	62/1	chr1	594	3	
S100A9	ENSG00000163220.10	6280	chr1	577	3	
SAA1	ENSG00000173432.9	6288	chr11	775	6	
SBSN	ENSG00000189001.9	374897	chr19	2665	6	
SDCBP2	ENSG00000125775.13	27111	chr20	1955	11	
SEMA7A	ENSG00000138623.8	8482	chr15	3780	17	
SERPINA3	ENSG00000273259.2	12	chr14	1590	5	

Symbol	Ensemble ID	Entrez ID	SeqNames	Length	N_Exons
SERPINA5	ENSG00000188488.12	5104	chr14	2337	6
SERPINB4	ENSG00000206073.9	6318	chr18	3426	16
SERPINE2	ENSG00000135919.11	5270	chr2	6558	25
SERPINF2	ENSG00000167711.12	5345	chr17	5573	29
SERPINH1	ENSG00000149257.12	871	chr11	5908	10
SHANK1	ENSG00000161681.14	50944	chr19	10491	28
SHH	ENSG00000164690.6	6469	chr7	1576	3
SIGLEC12	ENSG00000254521.5	89858	chr19	2732	9
SIX1	ENSG00000126778.8	6495	chr14	2687	2
SIX5	ENSG00000177045.7	147912	chr19	3331	3
SLAMF9	ENSG00000162723.8	89886	chr1	1740	6
SLC11A1	ENSG0000018280.15	6556	chr2	11886	38
SLC18A2	ENSG00000165646.10	6571	chr10	3872	16
SLC25A3	ENSG00000075415.11	5250	chr12	2314	10
SLC38A5	ENSG00000017483.13	92745	chrX	5360	42
SLC39A13	ENSG00000165915.12	91252	chr11	13614	58
SLC39A4	ENSG00000147804.8	55630	chr8	3407	16
SLC3A2	ENSG00000168003.15	6520	chr11	4687	23
SLC6A20	ENSG00000163817.14	54716	chr3	5952	13
SLC7A5	ENSG00000103257.7	8140	chr16	4679	13
SLPI	ENSG00000124107.5	6590	chr20	596	4
SNAI1	ENSG00000124216.3	6615	chr20	1709	3
SNRPE	ENSG00000182004.11	6635	chr1	1547	5
SPDYE4	ENSG00000183318.10	388333	chr17	3353	20
SPIN2A	ENSG00000147059.8	54466	chrX	5045	10
SQLE	ENSG00000104549.10	6713	chr8	2989	11
SRPX	ENSG00000101955.13	8406	chrX	2463	11
SRXN1	ENSG00000271303.1	140809	chr20	2698	2
SSTR5	ENSG00000162009.8	6755	chr16	7991	5
SULT1A1	ENSG00000196502.10	6817	chr16	4065	19
SYT12	ENSG00000173227.12	91683	chr11	3820	11
SYT13	ENSG00000019505.6	57586	chr11	9630	11
TAF1D	ENSG00000166012.13	79101	chr11	1312	6
TAS1R2	ENSG00000179002.5	80834	chr1	2521	6
TBC1D2	ENSG0000095383.18	55357	chr9	7091	27
TBX15	ENSG0000092607.12	6913	chr1	6938	12
TBX18	ENSG00000112837.15	9096	chr6	7046	10
TCIRG1	ENSG00000110719.8	10312	chr11	9804	51
TCN1	ENSG00000134827.6	6947	chr11	1567	9
TENM2	ENSG00000145934.14	57451	chr5	11410	38
TFB2M	ENSG00000162851.7	64216	chr1	1786	8
TFE3	ENSG0000068323.15	7030	chrX	4462	14
TGFB1I1	ENSG00000140682.17	7041	chr16	1982	14
THBS2	ENSG00000186340.13	7058	chr6	5881	23
TICAM1	ENSG00000127666.9	148022	chr19	2722	2
TIGD6	ENSG00000164296.6	81789	chr5	4682	3
TLR2	ENSG00000137462.6	7097	chr4	7460	12
TM4SF19	ENSG00000145107.14	116211	chr3	1677	7
TM4SF4	ENSG00000169903.6	7104	chr3	1607	5
TMEM179	ENSG00000258986.5	388021	chr14	3652	5
TMEM59L	ENSG00000105696.7	25789	chr19	1970	10
TNFSF14	ENSG00000125735.9	8740	chr19	7659	8
TNNI1	ENSG00000159173.17	7135	chr1	6143	9
TNNT2	ENSG00000118194.17	7139	chr1	2035	25
TP63	ENSG0000073282.11	8626	chr3	14539	22
TRIM29	ENSG00000137699.15	23650	chr11	4183	12
TSHZ3	ENSG00000121297.6	57616	chr19	4913	2

Symbol	Ensemble ID	Entrez ID	SeqNames	Length	N_Exons
TUBA1C	ENSG00000167553.13	84790	chr12	1553	4
TUBA3E	ENSG00000152086.8	112714	chr2	1540	5
UBTD1	ENSG00000165886.4	80019	chr10	1557	3
UCHL3	ENSG00000118939.16	7347	chr13	1850	15
UCN2	ENSG00000145040.3	90226	chr3	1485	2
UGT3A1	ENSG00000145626.10	133688	chr5	7667	17
UNC45A	ENSG00000140553.15	55898	chr15	4800	27
UST	ENSG00000111962.7	10090	chr6	5731	15
VCX3B	ENSG00000205642.8	425054	chrX	1175	3
VENTX	ENSG00000151650.7	27287	chr10	2412	3
VSTM1	ENSG00000189068.8	284415	chr19	9457	91
WDFY2	ENSG00000139668.8	115825	chr13	6371	24
WNT3	ENSG00000108379.8	7473	chr17	3355	5
WT1	ENSG00000184937.11	7490	chr11	3399	12
ZDHHC12	ENSG00000160446.17	84885	chr9	1891	7
ZFP92	ENSG00000189420.8	139735	chrX	7395	8
ZFYVE27	ENSG00000155256.16	118813	chr10	7821	23
ZNF20	ENSG00000132010.14	7568	chr19	3188	5
ZNF692	ENSG00000171163.14	55657	chr1	3215	18
ZSCAN12	ENSG00000158691.13	9753	chr6	18297	26
ZYX	ENSG00000159840.14	7791	chr7	2691	12

IMvigor210 (OS)				
variable	HR	95% CI	<i>P</i> value	
Sex				
Female	1			
Male	0.785	0.583-1.059	0.113	
IC level ¹				
IC0	1			
IC1	0.872	0.642-1.185	0.383	
IC2	0.566	0.405-0.780	<0.001	
TC level ²				
IC0	1			
TC1	1.111	0.656-1.882	0.694	
TC2	0.992	0.691- 1.425	0.967	
PDCD1	0.789	0.645-0.966	0.021	
CD274	0.869	0.752-1.003	0.055	
TMB	0.956	0.583-0.980	<0.001	
risk score	1.331	1.194-1.484	<0.001	

Table S6: Univariate Cox regression of commonly used characteristics
and risk score in immune checkpoint inhibitor (ICI) treatment sets

POPLAR (OS)				
variable	HR	95% CI	<i>P</i> value	
Age				
<=60	1			
>60	0.875	0.504-1.518	0.635	
Sex				
Female	1			
Male	1.063	0.587-1.925	0.84	
IC level				
IC0	1			
IC1	0.606	0.332-1.108	0.103	
IC2	0.571	0.232-1.406	0.223	
IC3	0.431	0.101-1.833	0.254	
TC level				
TC0	1			
TC1	0.847	0.329-2.182	0.731	
TC2	1.061	0.417-2.7	0.901	
TC3	0.343	0.106-1.114	0.075	
PDCD1	0.611	0.409-0.911	0.016	
CD274	0.803	0.618-1.042	0.099	
TMB	1.005	1.001-1.008	0.006	
risk score	1.239	1.114-1.379	<0.001	

IMmotion150 (PFS)				
variable	HR	95% CI	<i>P</i> value	
Age				
<=60	1			
>60	0.777	0.54- 1.118	0.174	
Sex				
Female	1			
Male	1.029	0.685- 1.546	0.891	
IC level				
IC0	1			
IC1	0.778	0.517-1.171	0.229	
IC2	1.032	0.588-1.811	0.913	
IC3	1.013	0.480-2.137	0.973	
TC level				
TC0	1			
TC1	1.093	0.552-2.166	0.798	
TC2	2.108	0.917-4.845	0.079	
TC3	0.952	0.387-2.340	0.915	
PDCD1	0.777	0.629-0.960	0.02	
CD274	0.871	0.708-1.071	0.19	
TMB	1.005	0.999-1.012	0.113	
risk score	1.631	1.340-1.985	<0.001	

- ¹ PD-L1 expression on immune cells measure by immunohistochemistry
 ² PD-L1 expression on tumor cells measured by immunohistochemistry

Supplementary Method

Genomic and clinical data sources and preprocessing

RNA-seq data obtained before immunotherapy and matched clinical data of patients who received anti-PD1/PDL1 immunotherapy in several cohorts across multiple cancer types and other public data analyzed in this work were collected.

 RNA-seq data in Transcripts Per Million (TPM) and matched clinical data of patients with metastatic urothelial cancer (mUC) treated with atezolizumab (anti-PD-L1 antibody) were retrieved from the R package IMvigor210CoreBiologies (http://research-pub.gene.com/IMvigor210CoreBiologies)[1].

 $(\underline{\mathrm{Intp.//Tesearch-pub.gene.com/intvigor210CoreBiologies})[1].$

- (2) Transcriptomic and clinical data of 657 patients from three phase II clinical trials and one phase I clinical trial of atezolizumab were collected from the European Genome-Phenome Archive (EGA) (Study ID EGAS00001004343[2]; https://ega-archive.org/studies/EGAS00001004343). These included 208 patients from a phase II clinical trial in mUC (IMvigor210[3], NCT02108652), 81 patients from a phase II clinical trial in non-small cell lung cancer (NSCLC) (POPLAR[4], NCT01903993), and 162 patients from a phase II clinical trial with atezolizumab or atezolizumab + bevacizumab treatment in renal cell carcinoma (RCC) (IMmotion150[5], NCT01984242), and a total of 206 patients with mUC, NSCLC, or RCC from a phase I basket trial (PCD4989g[6], NCT01375842). Expression matrices of TPM-normalized counts were download and convert to log2(TPM +1).
- ③ Log2(TPM+1) transformed counts data and matched survival data from 407 patients treated with atezolizumab + bevacizumab from a phase III clinical trial in RCC (IMmotion151, NCT02420821) were collected from the EGA (Study ID EGAS00001004353[7], https://ega-archive.org/studies/EGAS00001004353].
- (4) Data from 170 patients with advanced clear cell RCC from three clinical trials of nivolumab (anti-PD-1 antibody) were obtained from a report by Braun et al[8]. (https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7499153/bin/NIHMS1611472-su pplement-Table_S4_RNA_expression_normalized_expression_matrix_.xlsx). These patients were from CheckMate 009[9](NCT01358721), CheckMate 010[10](NCT01354431), and CheckMate 025[11](NCT01668784).
- (5) Data from 51 patients with advanced melanoma from clinical trials of nivolumab (CA209-038, NCT01621490) were obtained from the Gene Expression Omnibus through the accession number GSE91061 [12](<u>https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE91061</u>). The transcription profiles (FPKM normalized) of GSE91061 were transformed into TPM.
- (6) RNA-seq data (TPM normalized) of The Cancer Genome Atlas (TCGA)

Pan-Cancer processed on 2016-09-01 were obtained from the University California Santa Cruz Xena browser TCGA hub (<u>https://tcga.xenahubs.net/</u>). Corresponding clinical data updated on 2018-09-13 were downloaded from supplementary data of a published TCGA Pan-Cancer research[13] (<u>https://www.cell.com/cms/10.1016/j.cell.2018.02.052/attachment/bbf46a06-1fb0-417a-a259-fd47591180e4/mmc1.xlsx</u>).

Clinical samples collection

Bladder urothelial carcinoma and kidney renal clear cell carcinoma samples and corresponding adjacent normal tissues were resected from patients hospitalized in the First Affiliated Hospital of Sun Yat-sen University from 2020 to 2022. Patients are between the ages of 18 and 80. All patients were diagnosed through radiological examination and pathology methods. Before surgery, none of the patients received neoadjuvant chemotherapy or radiotherapy. The study was reviewed and approved by the Institutional Ethics Committee for Clinical Research and Animal Trials Ethical of the First Affiliated Hospital of Sun Yat-sen University [(2021)143], [(2021)144]. Written informed consent was obtained from participating patients. All experiments were performed following the ethical standards of the Helsinki Declaration.

Cell line

MB49 cell lines were purchased from Procell(Procell Life Science&Technology Co., Ltd). MB49 was raised in DMEM medium supplemented with 10% fetal bovine serum Cells were cultured in a humidified incubator with 5% CO2 at 37°C.

Protein expression profile of RPLP0

The difference of RPLP0 protein level between normal and tumor tissues in CPTAC database were explored through The University of Alabama at Birmingham Cancer data analysis Portal (UALCAN) (<u>http://ualcan.path.uab.edu</u>)[14]. Images of immunohistochemistry staining of RPLP0 in tumors and normal tissues were downloaded from the Human Protein Atlas (<u>https://www.proteinatlas.org/</u>).

Supplementary References

- 1. Mariathasan, S., et al., *TGFbeta attenuates tumour response to PD-L1 blockade by contributing to exclusion of T cells.* Nature, 2018. **554**(7693): p. 544-548.
- Banchereau, R., et al., *Molecular determinants of response to PD-L1 blockade across tumor types*. Nat Commun, 2021. **12**(1): p. 3969.
- 3. Balar, A.V., et al., *Atezolizumab as first-line treatment in cisplatin-ineligible patients with locally advanced and metastatic urothelial carcinoma: a single-arm, multicentre, phase 2 trial.* Lancet, 2017. **389**(10064): p. 67-76.
- 4. Fehrenbacher, L., et al., *Atezolizumab versus docetaxel for patients with previously treated non-small-cell lung cancer (POPLAR): a multicentre, open-label, phase 2 randomised controlled trial.* Lancet, 2016. **387**(10030): p. 1837-46.
- 5. McDermott, D.F., et al., *Clinical activity and molecular correlates of response to atezolizumab alone or in combination with bevacizumab versus sunitinib in renal cell*

carcinoma. Nat Med, 2018. **24**(6): p. 749-757.

- 6. Herbst, R.S., et al., *Predictive correlates of response to the anti-PD-L1 antibody MPDL3280A in cancer patients.* Nature, 2014. **515**(7528): p. 563-7.
- Rini, B.I., et al., Atezolizumab plus bevacizumab versus sunitinib in patients with previously untreated metastatic renal cell carcinoma (IMmotion151): a multicentre, open-label, phase 3, randomised controlled trial. Lancet, 2019. 393(10189): p. 2404-2415.
- Braun, D.A., et al., *Interplay of somatic alterations and immune infiltration modulates response to PD-1 blockade in advanced clear cell renal cell carcinoma*. Nat Med, 2020.
 26(6): p. 909-918.
- 9. Choueiri, T.K., et al., *Immunomodulatory Activity of Nivolumab in Metastatic Renal Cell Carcinoma*. Clin Cancer Res, 2016. **22**(22): p. 5461-5471.
- 10. Motzer, R.J., et al., *Nivolumab for Metastatic Renal Cell Carcinoma: Results of a Randomized Phase II Trial.* J Clin Oncol, 2015. **33**(13): p. 1430-7.
- Motzer, R.J., et al., *Nivolumab plus Ipilimumab versus Sunitinib in Advanced Renal-Cell Carcinoma*. N Engl J Med, 2018. **378**(14): p. 1277-1290.
- 12. Riaz, N., et al., *Tumor and Microenvironment Evolution during Immunotherapy with Nivolumab.* Cell, 2017. **171**(4): p. 934-949 e16.
- 13. Liu, J., et al., *An Integrated TCGA Pan-Cancer Clinical Data Resource to Drive High-Quality Survival Outcome Analytics.* Cell, 2018. **173**(2).
- 14. Chandrashekar, D.S., et al., *UALCAN: An update to the integrated cancer data analysis platform.* Neoplasia (New York, N.Y.), 2022. **25**: p. 18-27.