

***Supplementary Material***

**Role of Sterylglucosidase 1 (Sgl1) on the pathogenicity of  
*Cryptococcus neoformans*: potential applications for vaccine  
development**

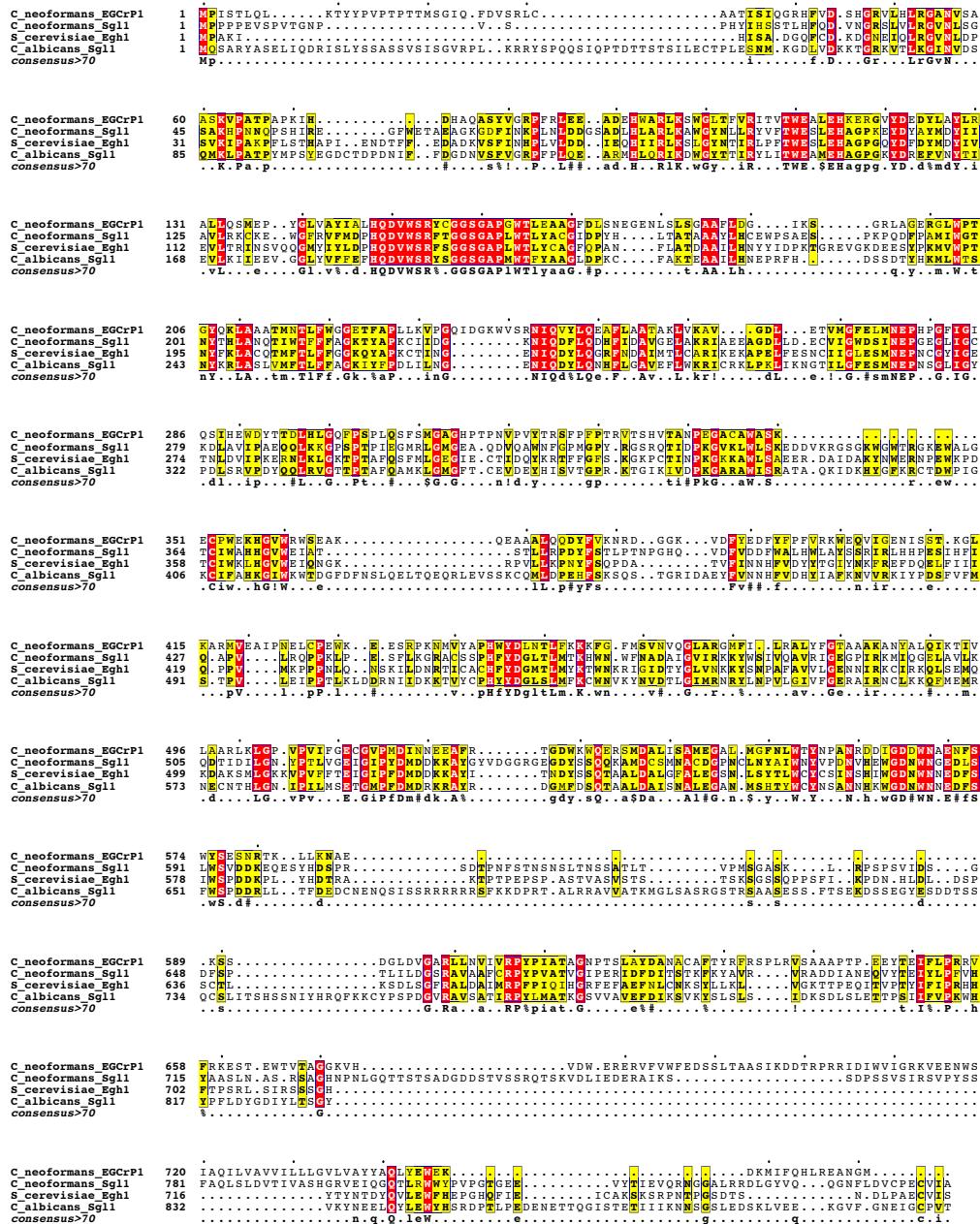
Antonella Rella<sup>1</sup>, Visesato Mor<sup>1</sup>, Amir M. Farnoud<sup>1</sup>, Ashutosh Singh<sup>1</sup>, Achraf A. Shamseddine<sup>2</sup>, Elitza Ivanova<sup>1</sup>, Nicholas Carpino<sup>1</sup>, Maria Teresa Montagna<sup>3</sup>, Chiara Luberto<sup>4</sup>, and Maurizio Del Poeta<sup>1,\*</sup>

<sup>1</sup>Department of Molecular Genetics and Microbiology, Stony Brook University, Stony Brook, NY 11794, <sup>2</sup>Department of Medicine, Stony Brook University, Stony Brook, NY 11794, <sup>3</sup>Department of Biomedical Science and Human Oncology, Hygiene Section, University of Bari, Italy, <sup>4</sup>Department of Physiology and Biophysics, Stony Brook University, Stony Brook, NY. 11794.

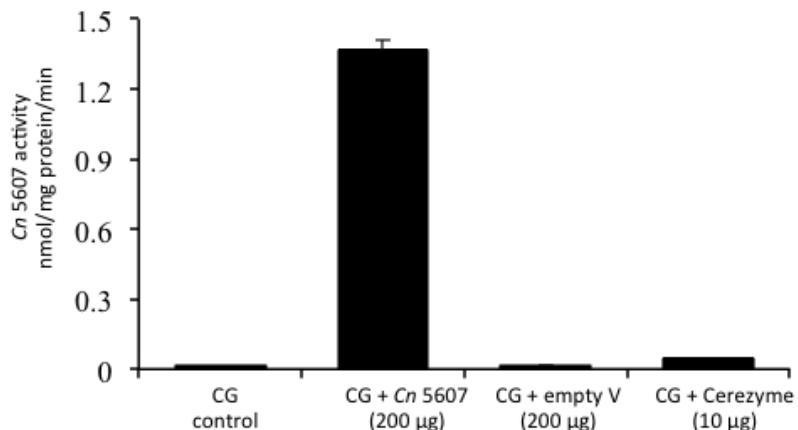
#Running Title: Sterylglucosides-accumulating *Cryptococcus* as fungal vaccine

**\*Correspondence:**

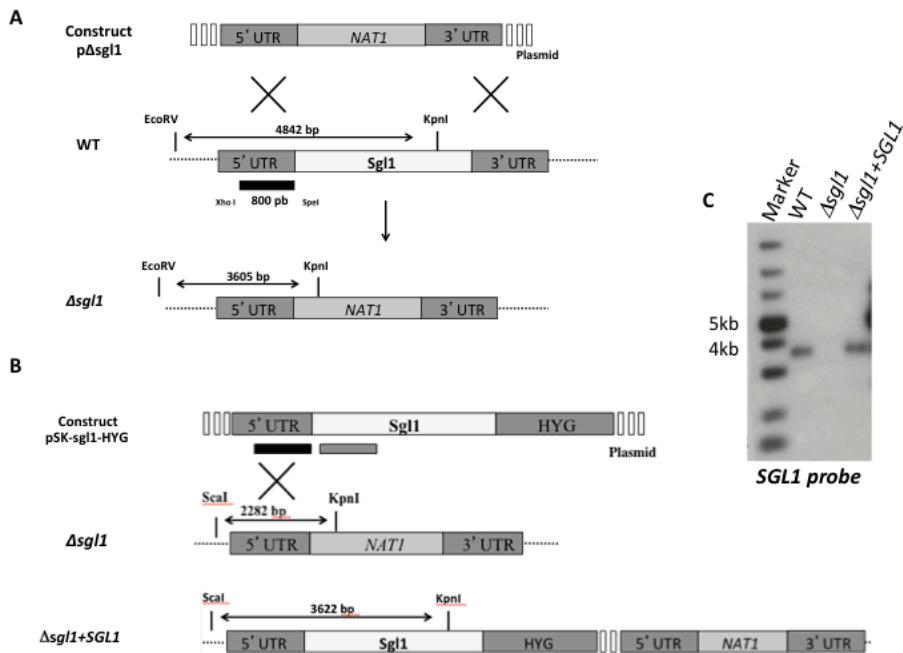
Dr. Maurizio Del Poeta,  
Stony Brook University,  
Department of Molecular Genetics and Microbiology,  
145 Life Sciences Building, Stony Brook, NY, USA, 11794  
Tel.: (631) 632-4024; Fax: (631) 632-9797;  
E-mail: [maurizio.delpoeta@stonybrook.edu](mailto:maurizio.delpoeta@stonybrook.edu)



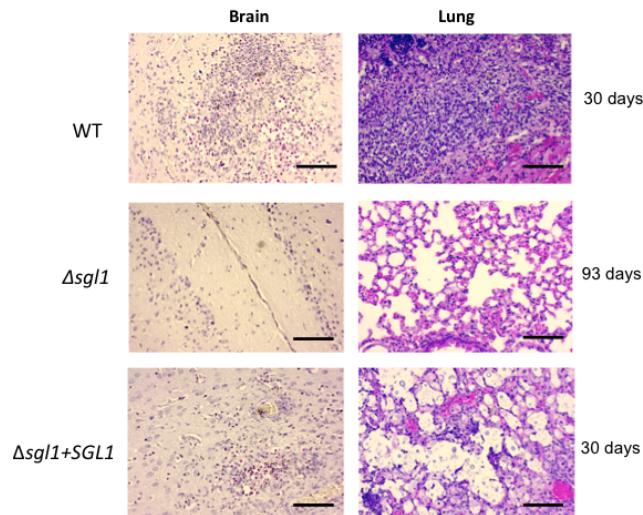
**Supplementary Figure 1.** Alignment of the entire protein sequences of *C. neoformans* protein EGCrP1, *C. neoformans* protein Sgl1, *S. cerevisiae* protein Egh1 and *C. albicans* protein Sgl1 (not characterized yet). Identical conserved residues are shown in red, homologous residues in at least 3 of the sequences are shown in yellow. The protein homology scores were 45% for *C. neoformans* EGCrP1 and *C. neoformans* Sgl1, 42% for *C. neoformans* EGCrP1 and *S. cerevisiae* Egh1, and 45% for *C. neoformans* EGCrP1 and *C. albicans* putative Sgl1.



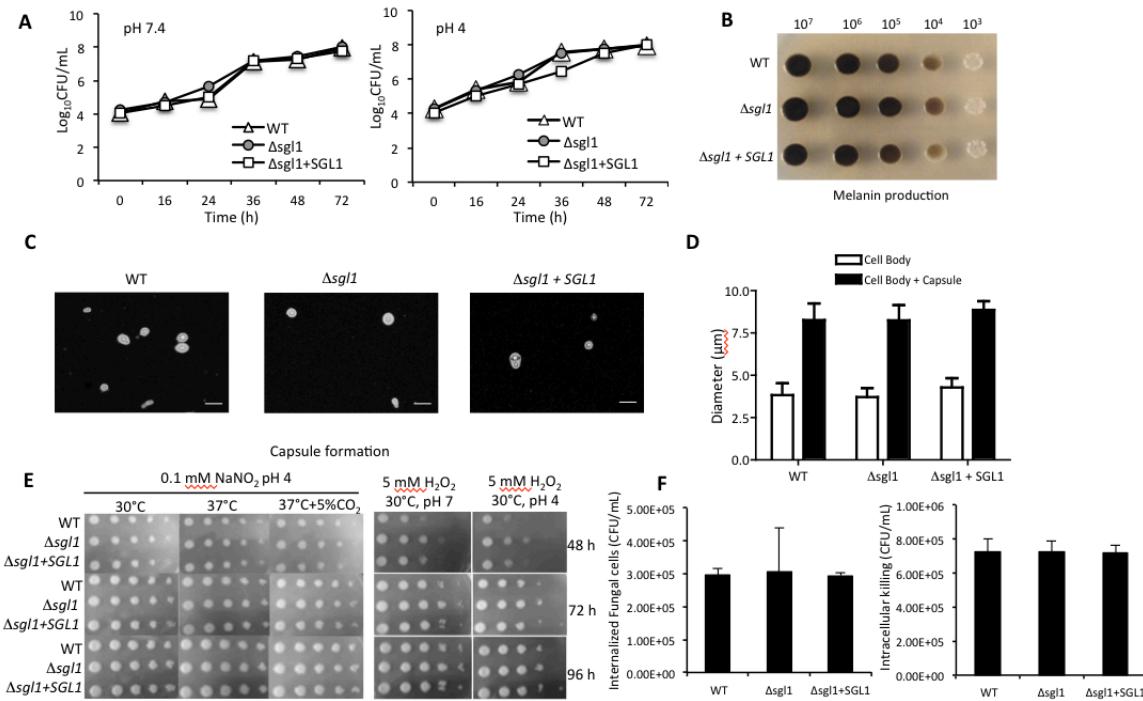
**Supplementary Figure 2.** CNAG\_05607 enzyme has cholesterol glucosidase activity. The release of cholesterol from cholesterol glycoside by CNAG\_05607 enzyme was monitored using GC-MS. Empty vector and Cerezyme were used as negative control. CG, Cholesterol glucoside; Empty V, empty vector.



**Supplementary Figure 3.** Deletion and reconstitution of the *SGL1* gene. (A) Strategy for the deletion of *SGL1* in *C. neoformans* wild-type (WT) and creation of the mutant strain  $\Delta$ sgl1. (B) Strategy for the generation of the complemented strain  $\Delta$ sgl1+*SGL1*. (C) Southern Blot hybridization analysis of genomic DNA of WT,  $\Delta$ sgl1 and  $\Delta$ sgl1+*SGL1* digested with ScalI and KpnI and screened with the “SGL1” probe (gray bar in B). Results show that the *SGL1* gene has been deleted in the  $\Delta$ sgl1 mutant and reconstituted in the  $\Delta$ sgl1+*SGL1* strain. Gene deletion was confirmed using a second probe (5' UTR – black bar in B). 5' UTR, 5' untranslated region; 3' UTR, 3' untranslated region; NAT1, nourseothricin 1; Sgl1, sterylglucosidase 1; HYG, Hygromycin B.



**Supplementary Figure 4.** Histopathology of brains and lungs obtained from the CBA/J mice infected intranasally with *C. neoformans* wild-type (WT),  $\Delta sgl1$ , and  $\Delta sgl1+SGL1$  strains. The brains were stained with mucicarmine stain, the lungs were stained with Hematoxylin and Eosin (H&E) stain. Black bar, 50  $\mu$ m.



**Supplementary Figure 5.** Characterization of virulence phenotypes in the  $\Delta sgl1$  mutant strain. A) *C. neofomans* wild-type (WT),  $\Delta sgl1$ , and  $\Delta sgl1 + SGL1$  strains showed identical growth in DMEM at 37°C, 5%CO<sub>2</sub> and at pH 7.4 or pH 4. B) *C. neofomans* cells were plated on medium containing L-DOPA and the melanin produced by WT,  $\Delta sgl1$ , or  $\Delta sgl1 + SGL1$  strain was similar. C) Capsule size of WT,  $\Delta sgl1$ , or  $\Delta sgl1 + SGL1$  strains was similar when cells were incubated in DMEM at 37°C, 5%CO<sub>2</sub>. White bar, 10 μm D) Cell body and capsule size of WT,  $\Delta sgl1$ , or  $\Delta sgl1 + SGL1$  strains, reported by measuring the cell body and capsule size of 50 cells for each strain under the microscope. E) There were no major differences among WT,  $\Delta sgl1$ , and  $\Delta sgl1 + SGL1$  in their ability to grow under stressed conditions (hydrogen peroxide and nitrosative stress). F) Phagocytosis and intracellular killing within a macrophage-like cell line J774A.16 was identical between WT,  $\Delta sgl1$ , or  $\Delta sgl1 + SGL1$  strains at 2 and 24 hours, respectively.