

## SUPPLEMENTARY MATERIALS

### Supplemental Figures

**Figure S1.** fMLP-induced calcium response in CTL and *plcg2<sup>kd</sup>* cells. Calcium responses in the individual CTL (**A**) and *plcg2<sup>kd</sup>* (**B**) cells in response to fMLP stimulation. See supplementary Video 8. Calcium response is visualized by the fluorescent calcium indicator Fluo-4. Cells were stained with Fluo-4 (green) 30 min prior to the experiment. CTL and *plcg2<sup>kd</sup>* cells were stimulated with fMLP at a final concentration of 10 nM, 1 nM, or 0.1 nM at time 0 s, respectively. The numbers in **A** and **B** indicate an individual cell.

**Figure S2.** Comparison of peak PIP<sub>3</sub> response in CTL and *plcg2<sup>kd</sup>* cells. Peak PH-GFP membrane translocation in the CTL and *plcg2<sup>kd</sup>* cells were measured. Mean Mean  $\pm$  SD is shown. N = 3 in each group of cells. A student's *t*-test was used to calculate the *p*-values. Statistical significance is indicated as follows: *ns* (not significant  $p > 0.05$ ), \* ( $p < 0.05$ ), \*\* ( $p < 0.01$ ).

### Supplementary videos:

**Video S1-S3.** Calcium response in the control (CTL, top) and *plcg2<sup>kd</sup>* (bottom) human neutrophil-like cell HL60 upon 10 nM (**S1**), 1 nM (**S2**), or 0.1 nM (**S3**) fMLP, respectively. Cells were stained with the calcium indicator Fluo-4 and stimulated with 10, 1, or 0.1 nM fMLP at the beginning of the movies, respectively. Scale bar = 50  $\mu$ m.

**Video S4-S5.** Membrane translocation of CAPRI-GFP in the control (CTL, top) and *plcg2<sup>kd</sup>* (bottom) HL60 cells upon 10 nM (**S4**) or 0.1 nM (**S5**) fMLP, respectively. Cells expressing CAPRI-GFP (green) were stimulated with 10, 1, or 0.1 nM fMLP at the beginning of the movies, respectively. To visualize the application of the stimuli, fMLP was mixed with a red fluorescent dye Alexa 594 (red). Scale bar = 5  $\mu$ m.

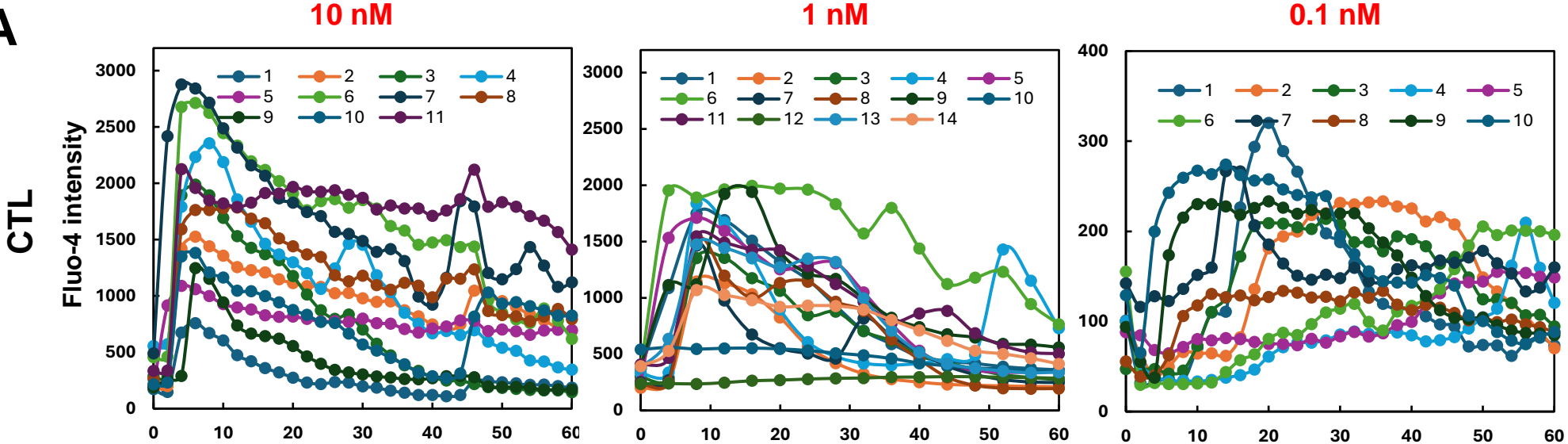
**Video S6-S7.** Membrane translocation of active Ras biosensor, RBD-RFP, in the control (CTL, top) and *plcg2<sup>kd</sup>* (bottom) HL60 cells upon fMLP stimulation. Cells expressing RBD-RFP (red) were stimulated with 10 nM (**S7**) or 0.1 nM (**S8**) fMLP at the beginning of the movies. To visualize the application of the stimuli, fMLP was mixed with a green, fluorescent dye Alexa 488 (green). Scale bar = 5  $\mu$ m.

**Video S8-S9.** Membrane translocation of PIP<sub>3</sub> biosensor, PH-GFP, in the control (CTL, top) and *plcg2<sup>kd</sup>* (bottom) HL60 cells upon fMLP stimulation. Cells expressing PH-GFP (green) and a PM marker (red) were stimulated with 10 nM (**S9**) or 0.1 nM (**S10**) fMLP, respectively, at the beginning of the movies. Scale bar = 5  $\mu$ m.

**Video S10-11.** Monitoring fMLP-induced actin polymerization in CTL (top) and *plcg2<sup>kd</sup>* (bottom) cells upon fMLP stimulation. Actin polymerization is monitored by the membrane translocation of GFP-F-tractin in cells. 10 nM (**S11**) or 0.1 nM (**S12**) fMLP was added to the cells at the beginning of the movies. Scale bar = 5  $\mu$ m.

**Video S12.** Montaged movie showing the migration behavior of CTL (left) and *plcg2<sup>kd</sup>* (right) cells under different conditions. Cells were exposed to either no gradient (top row) or to chemoattractant gradients generated from localized sources of fMLP (top two pairs), SDF-1 $\alpha$  (middle two pairs), or LTB4 (bottom two pairs). For each chemoattractant, the upper panel represents a gradient of 0.1 nM, and the lower panel represents a gradient of 100 nM.

**A**



**B**

