

## *Supplementary Material*

### **Maternal and fetal mechanisms of B cell regulation during pregnancy: human Chorionic Gonadotropin stimulates B cells to produce IL-10 while alpha-fetoprotein drives them into apoptosis**

Franziska Fettke<sup>1,5</sup>, Anne Schumacher<sup>1</sup>, Andrea Canellada<sup>2</sup>, Natalia Toledo<sup>2</sup>, Isabelle Bekerédjian-Ding<sup>3</sup>, Albert Bondt<sup>4</sup>, Manfred Wuhrer<sup>4</sup>, Serban-Dan Costa<sup>5</sup>, Ana Claudia Zenclussen<sup>1\*</sup>

#### **\*Correspondence:**

Corresponding Author: Prof. Dr. Ana Claudia Zenclussen, Experimental Obstetrics and Gynecology, Medical Faculty, Otto-von-Guericke University Magdeburg, Gerhart-Hauptmann-Straße 35, 39108 Magdeburg, Germany. Tel.: +49 -391-67-17460, Fax +49-391-67-17440. E-mail: [ana.zenclussen@med.ovgu.de](mailto:ana.zenclussen@med.ovgu.de)

***Supplementary Figure 1: CD19<sup>+</sup> B cell purity following magnetic isolation was above 95 %***

B cells were isolated from PBMCs using the B cell isolation Kit, using MS/LS columns and MidiMACS Separator. Our flow cytometry analysis showed that CD19<sup>+</sup> (FITC) B cells had a purity above 95 %.

***Supplementary Figure 2: Schematic diagram of 3 culture conditions***

Schematic diagram of the three culture conditions for B cells and their main characteristics; adding recombinant pregnancy hormones and AFP concentrations for flow cytometry assessment and IL-10 quantification (A), Ig quantification (B) and co-culture of B cells and JEG-3 cells.

***Supplementary Figure 3: IL-10 cytokine secretion was unaffected by pregnancy hormones***

(A) CD19<sup>+</sup> B cells were magnetically isolated from human PBMCs. Stimulated B cells (CD40L/CpG) were co-cultured with JEG-3 trophoblast cells for 24 h in charcoaled medium in the presence or absence of an anti-hCG antibody. IL-10 secretion in supernatants was analyzed by ELISA. JEG-3 co-culture increased IL-10 secretion.

***Supplementary Figure 4: Supernatant from JEG-3 cells stimulated IL-10 production by CD19<sup>+</sup> cells. hCG and CD40L/CPG had also a stimulatory effect on IL-10 production, while their combined addition the culture boosted IL-10 production***

CD19<sup>+</sup> B cells were magnetically isolated from PBMCs from female donors (n=5) and cultured either with JEG-3 supernatant, or in medium (DMEM containing 10% Fetal Bovine Serum, 1% Penicillin/Streptomycin) with hCG (100 mIU/ml), or stimulated with (CD40L/CpG, 1 µg/mL and

10 µg/mL respectively) or CD40L/CPG with hCG (same dose) for 48 hs. The addition of JEG-3 supernatant boosted the capacity of B cells to produce IL-10 as analyzed by flow cytometry. The addition of hCG or CD40L/CPG provoked the same effect, while the simultaneous Statistical analysis was carried out by repeated measures one-way ANOVA followed by Bonferroni correction for multiple analysis.

***Supplementary Figure 5: IL-10 secretion after hormonal stimulation***

Addition of recombinant hCG (100mIU/ml), P4 (30 ng/ml), E2 (1000 pg/ml) or the combination of P4 and E2 did not significantly alter IL-10 secretion by stimulated B cells. Unstimulated B cells served as controls. Statistical analysis was carried out by repeated measures one-way ANOVA followed by Bonferroni correction for multiple analysis.

***Supplementary Figure 6: Total IgM, IgG and IgA were unaffected by pregnancy hormones and AFP***

Quantification of human IgM, IgG and IgA assessed in supernatants after treatment with pregnancy hormones (A) or AFP (B) and incubation for 12 d was performed by ELISA. No significant differences for all Igs could be detected following treatment with hCG, P4, E2 and P4/E2 or treatment with AFP (A and B). Results represent means ± SEM. Statistical analysis was carried out by repeated measures one-way ANOVA followed by Bonferroni correction for multiple analysis. \*p ≤ 0,05.

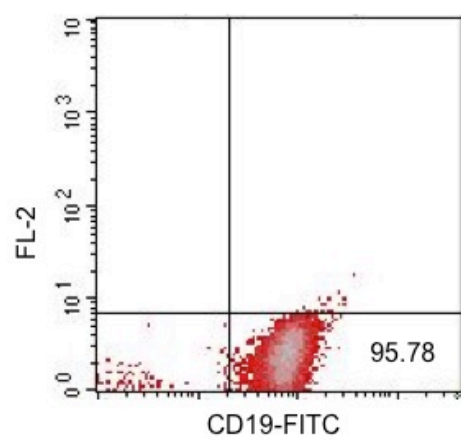
***Supplementary Figure 7: IgG glycosylation at their Fc region was not affected by pregnancy hormones or AFP treatment***

B cells were cultured for 12 d in charcoaled medium (control) or charcoaled medium with stimulation (CD40L/CpG) in the presence of recombinant hCG (100 mIU/ml), P4 (30 ng /ml), E2 (1000 pg/ml)

or the combination of P4 and E2 (**A**) or with AFP at a concentration found in maternal serum (1<sup>st</sup> trimester: 0.015 µg/ml, 2<sup>nd</sup> trimester: 0.06 µg/ml, 3<sup>rd</sup> trimester: 0.2 µg/ml), or AFP at a concentration found in fetal serum (50 µg/ml) (**B**). Glycopeptide analysis did not reveal statistical differences with regards to IgG1 galactosylation, bisection, fucosylation, or sialylation, and neither for combined IgG2 and 3 glycosylation traits (data not shown).

***Supplementary Figure 8: Dot plots depicting Annexin V/PI positive CD19<sup>+</sup> cells after stimulation with CD40L/CPG and CD40L/CPG + hCG.***

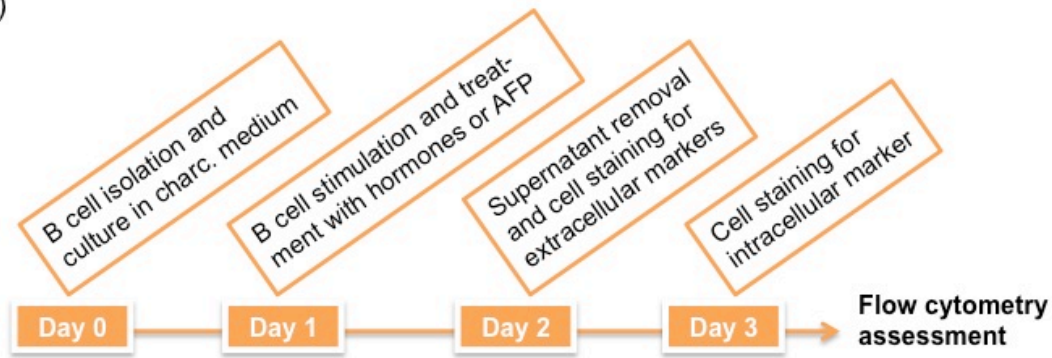
Isolated CD19<sup>+</sup> B cells were stimulated by CD40L/CpG, stimulated and exposed to 50 µg/ml AFP or incubated in standard medium as a control for 24 h. The percentage of viable, apoptotic and necrotic cells in the total B cell population was assessed using the FITC Annexin V Apoptosis Detection kit with flow-cytometric analysis. Dot plots indicate representative dot plots for early apoptotic Annexin<sup>+</sup> cells (right bottom panel) or late apoptotic/necrotic Annexin<sup>+</sup>/PI<sup>+</sup> cells (right upper panel).



*Supplementary Figure 1*



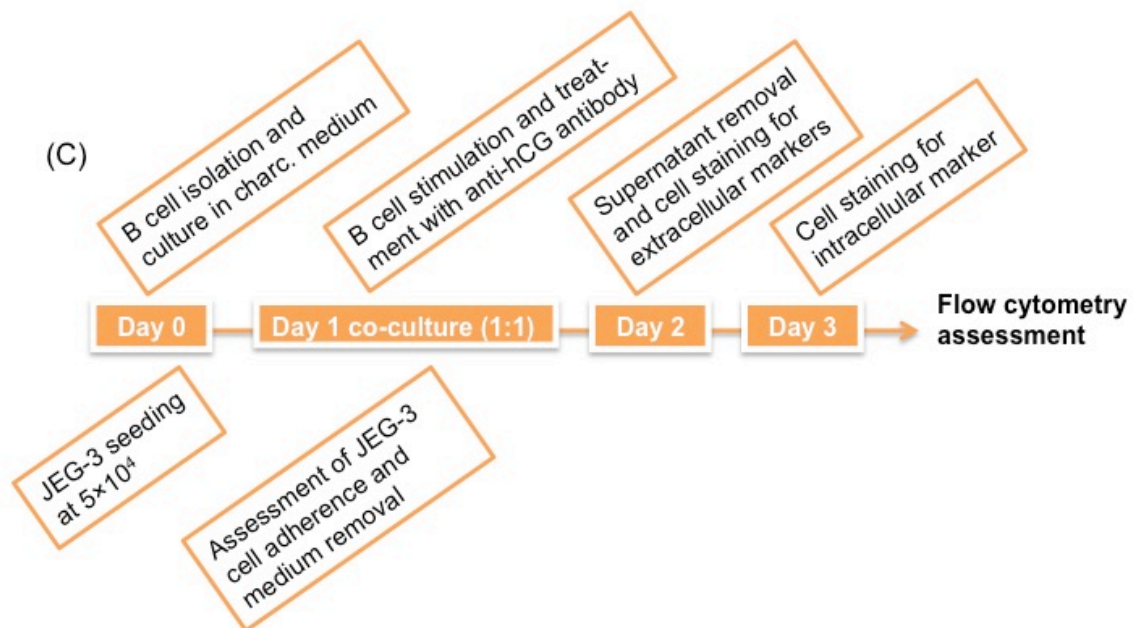
(A)



(B)

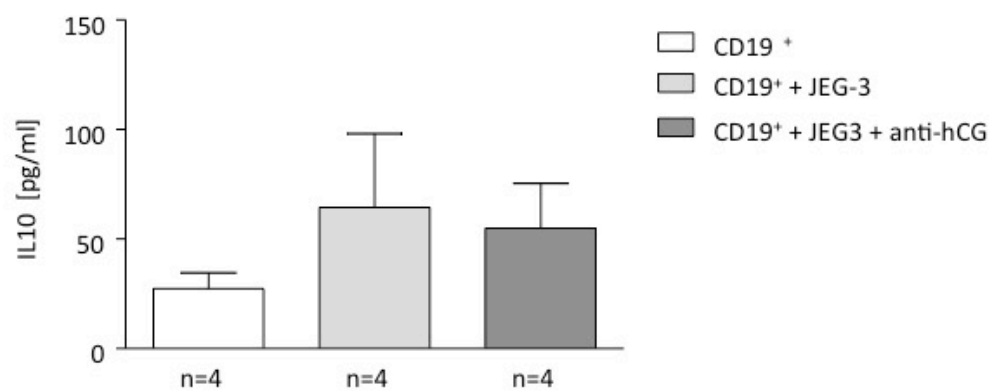


(C)

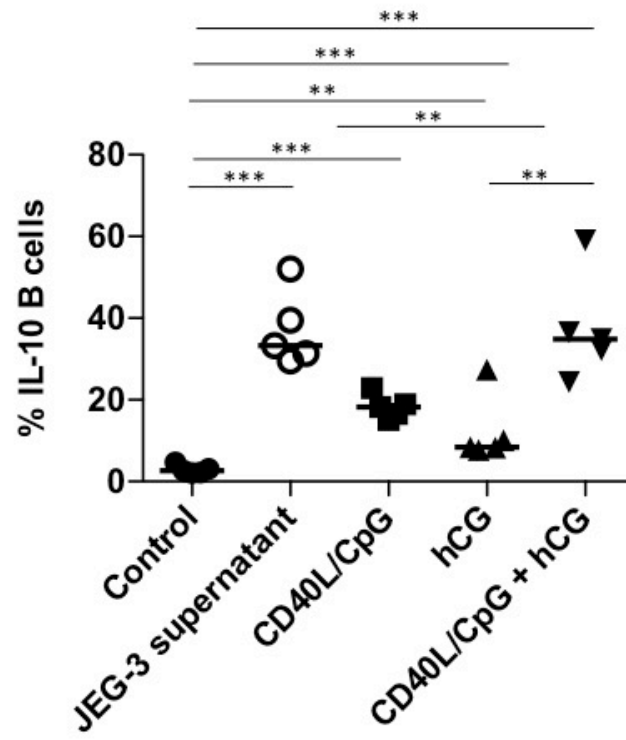


*Supplementary Figure 2*

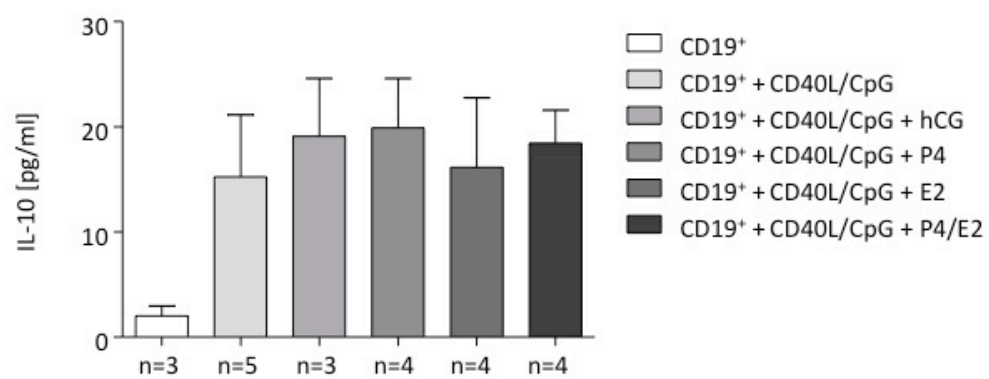
(A)



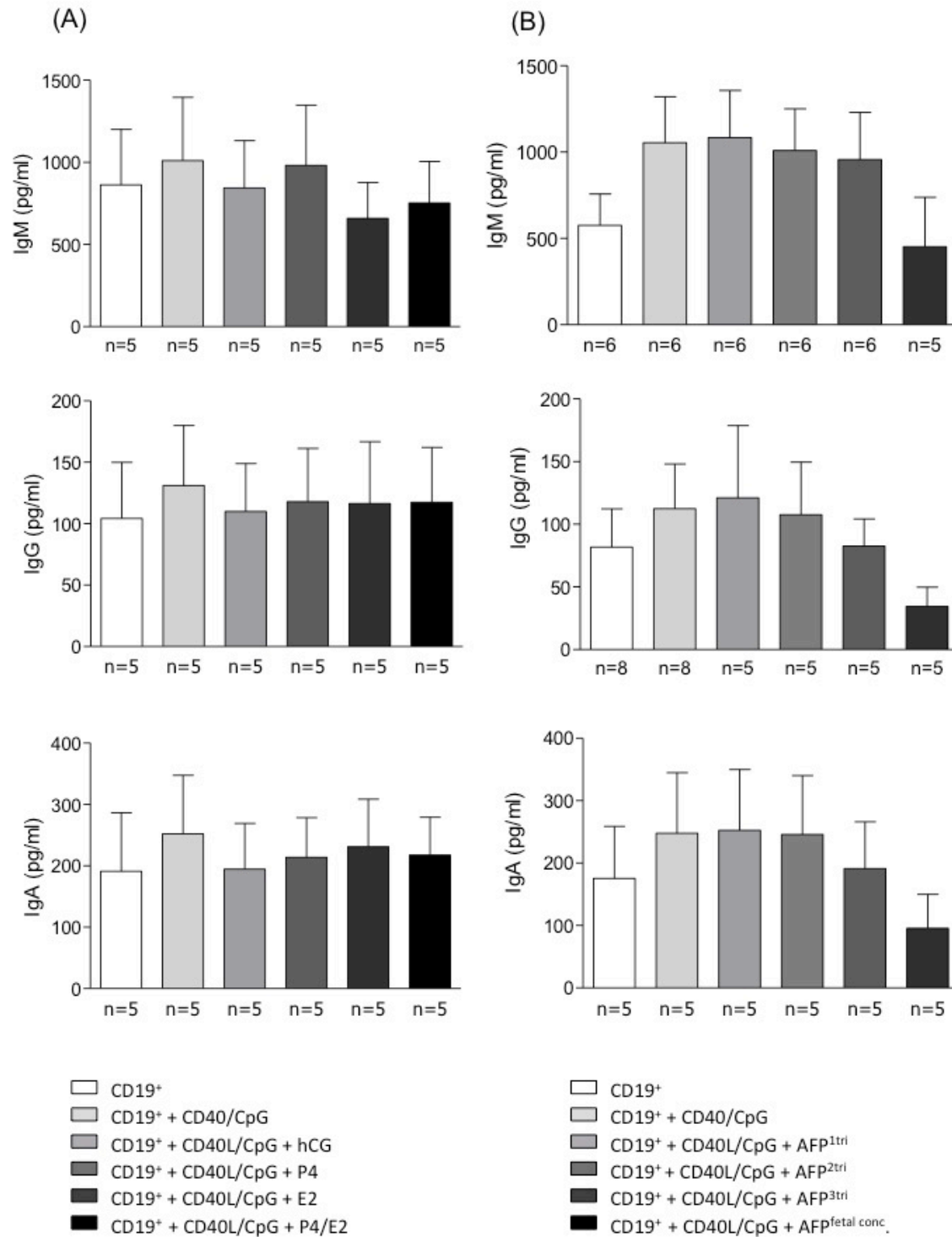




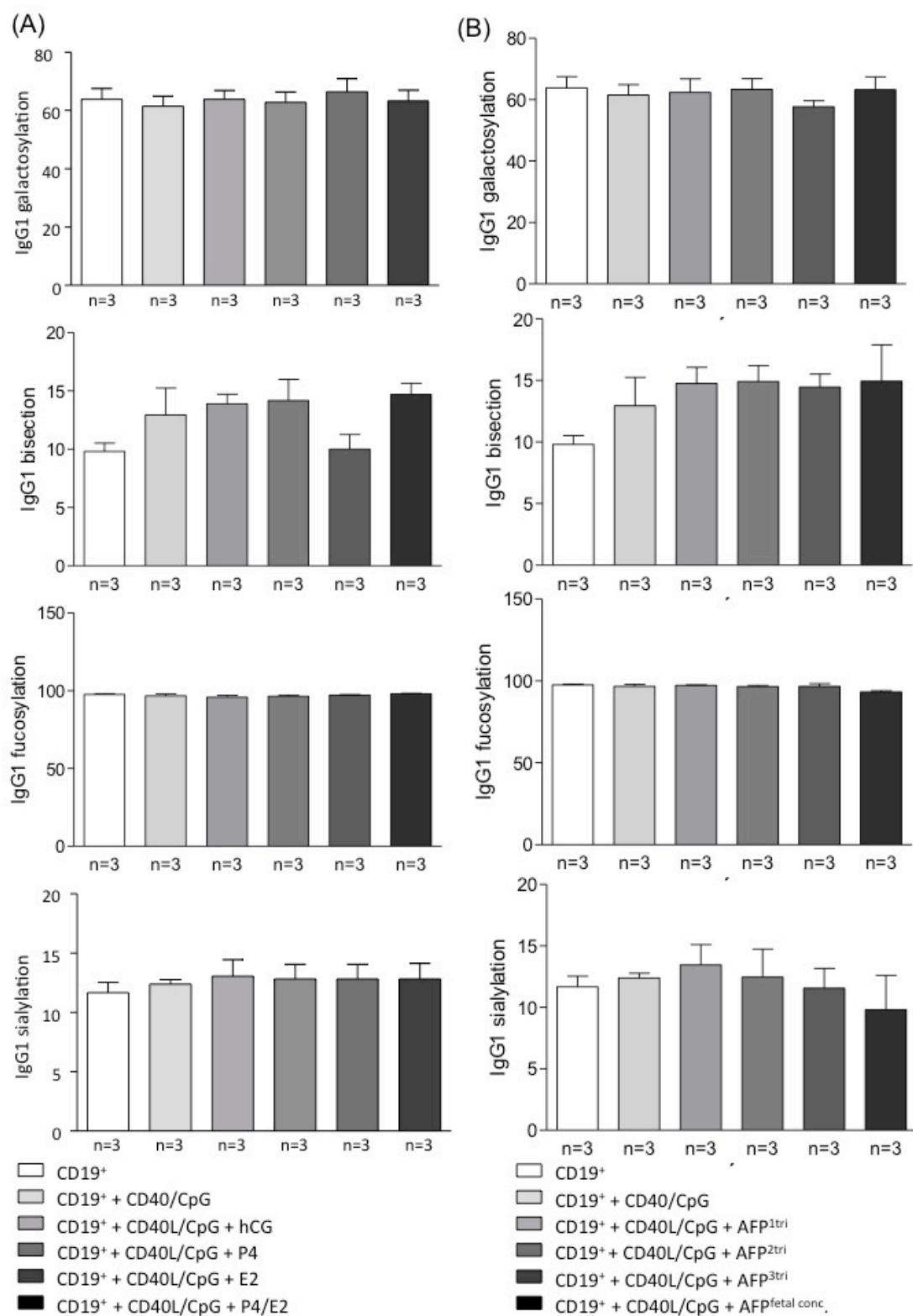
Supplementary Figure 4



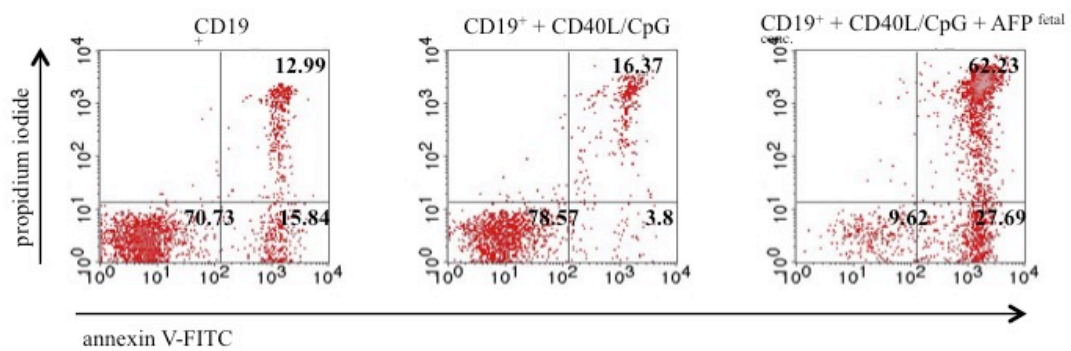
Supplementary Figure 5



Supplementary Figure 6



Supplementary Figure 7



*Supplementary Figure 8*