Table S1 : *C. albicans* strains and clinical isolates and *Candida* species used in this study .

|  |  |  |  |
| --- | --- | --- | --- |
| **Strain** | **Species** | **Source** | **Reference** |
| BWP17 + Clp 30 (Wild type) | *Candida albicans* | BWP17 | (Mayer et al, 2012; Wilson et al, 1999) |
| *pra1*Δ | *Candida albicans* | BWP17 | (Citiulo et al, 2012) |
| *pra1*Δ+*PRA1* | *Candida albicans* | BWP17 | (Citiulo et al, 2012) |
| RIH09 | *Candida albicans* | Blood | (MacCallum et al, 2009) |
| HUN96 | *Candida albicans* | Blood | (MacCallum et al, 2009) |
| IHEM16614 | *Candida albicans* | Oropharynx | (MacCallum et al, 2009) |
| AM2003/0182 | *Candida albicans* | Blood | (MacCallum et al, 2009) |
| J990102 | *Candida albicans* | Vagina | (MacCallum et al, 2009) |
| FC28 | *Candida albicans* | Vagina | (MacCallum et al, 2009) |
| LI086 | *Candida albicans* | Wound | (MacCallum et al, 2009) |
| S20176.079 | *Candida albicans* | Blood | (MacCallum et al, 2009) |
| Wü284 | *Candida dubliniensis* | Clinical (unknown) | (Morschhauser et al, 1999) |
| CD36 | *Candida dubliniensis* | Oral | (Sullivan et al, 1995) |
| MYA-3404 | *Candida tropicalis* | Reference strain | (Butler et al, 2009) |
| B31581/7/04 | *Candida tropicalis* | Urine | (Walker et al, 2013) |
| SCS40113 | *Candida parapsilosis* | Blood | (Walker et al, 2013) |
| ATCC 22019 | *Candida parapsilosis* | Reference strain | (Guerin et al, 1989) |
| ATCC 42720 | *Candida lusitaniae* | Reference strain | (Butler et al, 2009) |
| J960088 | *Candida famata* | Human skin lesions | Strain collection of Dr. Donna MacCallum, MRC Centre for Medical  Mycology at the University of Aberdeen |
| J960089 | *Candida famata* | Human skin lesions |

Table S2. Limited zinc media (LZM).

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| **Stock** | **Fold conc.** | **Component** | **Stock conc. (M)** | **Final conc. (M)** | **Vol.** |
| 1 | 500 | Na2EDTA.2H2O | 5.0x10-1 | 1.0x10-3 | 1ml |
| 2 | 100 | MgSO4.7H2O  NaCl | 5.0x10-1  1.0x10-1 | 5.0x10-3  1.0x10-3 | 5ml |
| 3 | 100 | CaCl2.2H2O | 1.0x10-1 | 1.0x10-3 | 5ml |
| 4 | 100 | uridine  L-histidine  L-leucine  L-lysine | 4.0x10-2  5.0x10-2  7.6x10-2  7.0x10-2 | 4.0x10-4  5.0x10-4  7.6x10-4  7.0x10-4 | 5ml |
| 5 | 100 | (NH4)2SO4 | 3.8 | 3.8x10-2 | 5ml |
| 6 | 100 | KH2PO4 | 1.0x10-1 | 1.0x10-3 | 5ml |
| 7 | 50 | Na3citrate.2H2O | 1.0 | 2.0x10-2 | 10ml |
| 8 | 20 | D-glucose | 2.2x10-1 | 1.1x10-2 | 25ml |
| 9 | 1000 | d-biotin  Ca pantothenate  myo-inositol  pyridoxin  thiamin.HCl | 1.6x10-5  1.7x10-3  1.0x10-2  2.0x10-3  1.0x10-3 | 1.6x10-8  1.7x10-6  1.0x10-5  2.0x10-6  1.0x10-6 | 0.5ml |
| 10 | 10000 | H3BO3  KI  Na2MoO4.2H2O | 1.0x10-1  5.0x10-3  1.0x10-2 | 1.0x10-5  5.0x10-7  1.0x10-6 | 50µl |

The pH values of stocks 1 and 7 were adjusted to 8.0 and 4.2, respectively, and Stock 10 was prepared in 0.1 M HCl. Solutions were filter-sterilized with 0.2 µm cellulose nitrate filters (Schleicher & Schuell) and stored in polycarbonate bottles. To prepare the different LXM media, the stock solutions were sequentially added to ultra-pure water, filter-sterilized and stored in polycarbonate bottles. To generate the respective metal limited media, the following transition metals were added: FeCl (6.17 µM), MnSO4 (13.24 µM), CuSO4 (0.3 µM) and ZnSO4 (25 µM). To generate LZM pH 7.3, media was alkalinised with NaOH and buffered with 50 mM HEPES pH 7.4

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