



FIG S1. pTIC10a-*phoU1* leaky expression. RNA was extracted from the indicated strains grown to mid-logarithmic phase in complete 7H9 medium supplemented with the indicated anhydrotetracycline (ATc) concentration. Quantitative RT-PCR was performed to determine abundance of the *phoU1* transcripts relative to the *sigA* housekeeping control. Results are the mean of three biological replicates $\pm$ standard deviations. Asterisks indicate statistically significant differences compared to the mc²155 control: * $P < 0.05$.

Table S1. Plasmids used in this study.

Plasmid	Genotype	Reference
pCR2.1 TOPO	PCR cloning vector Amp ^R Kan ^R	Invitrogen
pJG1100	Allelic exchange suicide vector Kan ^R Hyg ^R <i>sacB</i>	(1)
pBE101	pJG1100::Δ <i>phoU1</i> (<i>Msmeg_5776</i>), Kan ^R , Hyg ^R , <i>sacB</i>	This work
pBE102	pJG1100::Δ <i>phoU2</i> (<i>Msmeg_1605</i>), Kan ^R , Hyg ^R , <i>sacB</i>	This work
pMV261	Episomal vector with <i>hsp60</i> promoter Kan ^R	(2)
pMV <i>phoU1</i>	pMV261:: <i>phoU1</i> , Kan ^R	This work
pMV <i>phoU2</i>	pMV261:: <i>phoU2</i> , Kan ^R	This work
pTIC10a	Integrating vector with codon-optimized TetR, P _{<i>smyc</i>} -TetO, Kan ^R	(3)
pTIC <i>phoU1</i>	pTIC10a:: <i>phoU1</i> , Kan ^R	This work
pJT6a	Integrating vector with codon-optimized TetR, P _{<i>smyc</i>} -TetO, Hyg ^R	(4)
pJT <i>phoU1</i>	pJT6a:: <i>phoU1</i> , Hyg ^R	This work

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2. **Stover CK, de la Cruz VF, Fuerst TR, Burlein JE, Benson LA, Bennett LT, Bansal GP, Young JF, Lee MH, Hatfull GF, Snapper SB, Barletta RG, Jacobs WR, Jr., Bloom BR.** 1991. New use of BCG for recombinant vaccines. *Nature* **351**:456-460.
3. **Glover RT, Kriakov J, Garforth SJ, Baughn AD, Jacobs WRJ.** 2007. The two-component regulatory system *senX3-regX3* regulates phosphate-dependent gene expression in *Mycobacterium smegmatis*. *J Bacteriol* **189**:5495-5503.
4. **Rosen BC, Dillon NA, Peterson ND, Minato Y, Baughn AD.** 2017. Long-chain fatty acyl coenzyme A ligase FadD2 mediates intrinsic pyrazinamide resistance in *Mycobacterium tuberculosis*. *Antimicrob Agents Chemother* **61**:e02130-16.

Table S2. Oligonucleotide primers used for cloning or strain construction in this study.

Name	Purpose	Sequence (5'-3')^a
1605F1	Upstream $\Delta phoU2$	ATGCTTAATTAACACTGCGCCTGCTCAAC
1605R1	Upstream $\Delta phoU2$	ATGCCCTAGGAGCTCG CAT CGCCCATGAC
1605F2	Downstream $\Delta phoU2$	GCATCCTAGGCTCGCCT TGATT ACCGGCTAGC
1605R2	Downstream $\Delta phoU2$	ATGCGGCGCGCCCGTTGATGCGCTCTGCGAACT
5776F1	Upstream $\Delta phoU1$	GCATTTAATTAACCGCATCACGTTCTGCACCAT
5776R1	Upstream $\Delta phoU1$	ATGCCCTAGGATGGTACTGGATCCG CAT GCA
5776F2	Downstream $\Delta phoU1$	GCATCCTAGGAAGGTCACCACGCAGCAG
5776R2	Downstream $\Delta phoU1$	ATGCGGCGCGCCCCCTTGGTGAGGTTGGTGAG
1605F3	Check $\Delta phoU2$	GGAAACAGCTGCTGCGCAAC
1605R3	Check $\Delta phoU2$	CGAGCTTTTCGGCAAACCTCGA
1605F4	Check $\Delta phoU2$	GGATGGGTACCTACGTCCTCA
1605R4	Check $\Delta phoU2$	CCTGGTGATCTGCTCCGTAAC
5776F3	Check $\Delta phoU1$	ACCAAGGATCTCGTGGACCTC
5776R3	Check $\Delta phoU1$	CGACGGGAAGCTCGATCTCCT
5776F4	Check $\Delta phoU1$	GGTGTTACCGCCTAATCTGG
5776R4	Check $\Delta phoU1$	CGCGACTTCCTGATGCGCTA
1605CF	pMV $phoU2$ cloning	ATGAATTCCGGTACCTACGTCCTCATCCGG
1605CR	pMV $phoU2$ cloning	ATAAGCTTCTGGTGATCTGCTCCGTACC
5776CF	pMV $phoU1$ cloning	ATGAATTCCACCAGCAGCTTCGTGACTG
5776CR	pMV $phoU1$ cloning	ATAAGCTTCCGATCAGCCGTAGGTCA
TIC5776F	pTIC10a- $phoU1$ cloning	ATAAGCTTCCACCAGCAGCTTCGTGACTG
TIC5776R	pTIC10a- $phoU1$ cloning	ATGAATTCCCGATCAGCCGTAGGTCA
pTfor	Check plasmid switch	CATCCCGGCGTTGATCTGTG
pTIC6a_R	Check plasmid switch	TTTTCTTAAGGAGCAAGACGTTTCCCGTT
1387F2	Verify point mutation	TGCGACATCTTCCTGGTGC
1387R2	Verify point mutation	AGCACGTTGCGCATCAGC
DM504_pstSF	Verify point mutation	ATGAGCGGCGAATACGTTGC
DM504_pstSR	Verify point mutation	GGGAACCTGTCGGTCATGTG
DM518_pstBF	Verify point mutation	CGCTCACGCTCATCCTGC

DM518_pstBR	Verify point mutation	GTCCATGGTGGATGCCGG
DM521_pstBF	Verify point mutation	GATCGCCCTGCTGAATGTCTG
DM521_pstBR	Verify point mutation	GAAGTCGCCGTCGGGAAC
DM625_pstSF	Verify point mutation	TACGTTGCCGGGGAGTCTG
DM625_pstSR	Verify point mutation	GTCACTGTTATCCCGTCGGG
DM664_pstCF	Verify point mutation	GCCCAACGCCTTCAAAGAGC
DM664_pstCR	Verify point mutation	GTGTGGTCGGTCAGCCTG

^aRestriction enzyme sites used for cloning are underlined. Start and stop codons in primers used for construction of in-frame deletions are indicated in bold.

Table S3. Oligonucleotides used for qRT-PCR.

Gene	F primer sequence 5'-3'	R primer sequence 5'-3'
<i>Msm_sigA</i>	CCAAGGGCTACAAGTTCTCG	CCATGTGCACCGGGATAC
<i>Msm_pstS</i>	GGCGTCGACAAGCTGGTACT	GGTGATCTGGCCTTGGAAGA
<i>Msm_regX3</i>	TCCCGTGCGCATGGA	GGCAACGTGATCGGTTCAC
<i>Msm_phoA</i>	CGCAGAAGGCCATCGATCT	ATCGACGCGCCTTCCA
<i>Msm_1605</i>	GCAGACGGCGTTCAAAC	ACGACCATACGCAGCTCACT
<i>Msm_5776</i>	AGAGGTCAACGGCTACTTCG	CGTCGTCTTCCTCCTGGAT