Supplementary Table S2. The examples of application the separation techniques in analysis of alkaloids from *C. majus*

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| **Method** | **Sample/analytes** | **Stationary phase** | **Mobile phase** | **Detection** | **References** |
| 2D-HPTLC | herb, chelerythrine, protopine, berberine | CN F254 | 1) methanol- water (60:40) - 2% ammonia 2) diisopropyl ether - methanol (80:20) - 2% ammonia | DAD-densitometry | Petruczynik et al., 2007 |
| NP-TLC | leaf, stem, flower, root/coptisine, chelidonine, chelerythrine, sanguinarine, berberine | Si 60 F254 | methylene chloride-methanol (97:3); chloroform-methanol (60:30) | UV light at 254 and365 nm, densitometry | Sárközi et al., 2006 |
| NP-TLC | aerial parts/coptisine, chelidonine, | Si 60 GF 254 | 1-propanol–formic acid–water (90:1:9) | densitometry | Wagner et al., 1984 Then et al., 2000 |
| NP-TLC | aerial parts, roots/ sanguinarine, chelerythrine, berberine, coptisine, chelidonine, protopine | Si 60 F254 | two-step elution:1) chloroform- methanol - water (70:30:4)  2) toluene-ethyl acetate - methanol (83:15:2)  | UV light at 254 and365 nm.densitometry | Gadzikowska and Gołkiewicz, 1998;Bogucka-Kocka and Zalewski, 2016 |
| NP-TLC in magnetic field  | *C. majus* extracts/ allocryptopine, protopine, homochelidonine, chelidonine | Si 60 F254 | toluene-ethyl acetate-methanol (70:15:15) | densitometry | Malinowska et al., 2017 |
| NP-OPLC | *C. majus* extracts/ allocryptopine, protopine, chelidonine chelerythrine, chelilutine, sanguinarine, chelirubine | Si 60 F254 | Tertiary alkaloids, toluene–ethyl acetate-methanol, 70 + 15 + 15 (v/v) as mobile phase, quaternary alkaloids with toluene–ethyl acetate–methanol,83 + 15 + 2 (v/v) | densitometry | Malinowska et al., 2005 |
| HPLC | roots/ sanguinarine, chelerythrine, berberine, | Silica (250 x 4.6mm, 10 µm) | 0.005 M sodium acetate in methanol-1,4-dioxane-acetic acid (88:10:2), flow rate of 1.5 ml/min; | UV-Vis at 280 nm | Bugatti et al., 1987 |
| HPLC | *C. majus* plants/ corysamine, methoxychelidonine, allocryptopine, protopine, chelerythrine, berberine,chelidonine, homochelidonine, oxysanguinarine, sanguinarine, dihydrochelerythrine, dihydrosanguinarine  | Hypersil ODS column (100 x 4.6 mm, 5 µm) | water (adjusted to pH 7.5 with propylamine) –acetonitrile - methanol with 0.15 mM potassium iodide, gradient elution from 50:20:30 to 15:55:30 in 15 min. flow rate: from 0.8 to 1.5 ml/min in 15 min. | DAD | Han et al., 1991 |
| HPLC | *C.majus* tincture/ chelidonine, sanguinarine, chelerythrine, protopine | Discovery HS C18 (150 x 4.6 mm, 3 µm) | A: acetonitrile B: 0.030 mol/L formic acid in water, gradient elution: from 0 to 5 min 15% A; from 5 to 20 min increase from 15% to 90% A; from 20 to 22 min 90% A, flow rate: 0.7 ml/min | DADMS/MS | Prosen Pendry, 2016 |
| HPLC | aerial parts, tincture/ chelidonine, sanguinarine, protopine, berberine, coptisine | Luna C18 (250 × 4.6 mm, 5 μm) | acetonitrile–methanol–30 mM ammonium formate, pH 2.80(14.7:18:67.3); flow rate: 1 mL/min, temperature: 30ºC | DAD | Kursinszki et al., 2006 |
| HPLC | *C. majus* plant/ sanguinarine, chelerythrine | RP 18 (250 × 4.6 mm, 5 μm) | A : 0.1% phosphoric acidand 0.02% SDS (pH 3.5, adjusted by triethylamine); B: acetonitrile, gradient elution: 0–15 min,30%–35% B; and 15–25 min, 35%-45% B; flow rate 1 mL/min,temperature: 30ºC  | Fluorescence: excitation 330 nm, emission 555 nm. | Wu and Du, 2012 |
| HPLC | leaves/ chelidonine,berberine, sanguinarine, chelerythrine, coptisine,  | Nucleosil RP-18 (5 µm) | A: acetonitrile; B: 10 mM (NH4)2SO3 with 0.2% triethylamine adjusted with acetic acid to pH 4.0; C: methanol, gradient elution: 0 min: 5% A, 90% B, 5% C; 24 min: 70% A, 10% B,20% C; flow rate: 1.0 mL/min. | DADMS | Paulsen et al., 2015 |
| HPLC | herb/ chelidonine coptisine sanguinarine berberine chelerythrine | ZORBAX Poroshell 120 SB-C18 (3×100 mm, 2.7 μm) | A: 30 mM ammonium formate (pH 2.8) , B: acetonitrile:methanol 14.7:18.0; gradient elution: from 20% to 60% B in 16 min; flow rate 0.5 mL/min,temperature: 40ºC  | DAD | Seidler-Łożykowska et al., 2016 |
| HPLC | aerial parts, flowers, fruits, leaves, roots, stems/ protopine, chelidonine, coptisine, stylopine, sanguinarine, berberine, chelerythrine | Luna C18 (250 × 4.6 mm, 5 μm) | acetonitrile-methanol-30 mM ammonium formate (pH 2.8)150:180:670; flow rate 0.8 mL/min. | UV/vis detector | Borghini et al., 2015 |
| HPLC | aerial parts, terrestrial parts/ chelidonine, cheleritrine, sanguinarine, berberine | RP 18 (250 × 4.6 mm, 5 μm) |  A: heptanesulfonic acid (0.01 M) and triethylamine (0.1 M) in water acidified with formic acid (pH 2.5) B: acetonitrile;gradient elution: 0 min: 75% A; 1 min: 68% A; 2 min: 57.5% A; 4.5 min: 40% A; 12 - 40 min: 20% A; flow rate 1 mL/min  | DAD | Gañán et al., 2016 |
| HPLC | aerial parts/ dihydroberberine, protopine, allocryptopine, chelidonine, coptisine, tetrahydrocoptisine, tetrahydroberberine, berberine, norchelidonine, chelerythrine | Luna C18 (150 x 1.0 mm, 3 µm) | A: 1% acetic acid in water, B: methanol gradient elution: 20% to 50% B during 30 min, 80% B at 40 min. flow rate 20 µL/min  | DAD–ESI/MSn | Grosso et al., 2014 |
| CE | aerial parts/ sanguinarine, coptisine,chelerythrine,stylopine ,chelidonine, protopine, allocryptopine | polyimide-coated fused silica capillary, 50 cm x 75 µm  | 20 mM sodium phosphate pH 3.1 | ultraviolet light-emitting diode-induced native fluorescence: excitation 280 nm, emission 200 to 600 nm. ESI-MS | Kulp et al., 2011 |
| CE | *C. majus* plant/ sanguinarine, coptisine, chelerythrine, berberine, chelidonine, protopine,allocryptopine,stylopine | fused-silica capillary, 35 cm x 50 µm  | 500 mM Tris–H3PO4 buffer (pH 2.5) with 50% methanol and 2mM HP-β-cyclodextrin.  | DAD | Zhou et al., 2012 |
| CE | aerial parts/ coptisine,berberine, protopine, chelidonina, stylopine | fused-silica capillary, 70 cm × 50 µm  | citric acid-Na2HPO4 buffer (pH 5.5) and β-cyclodextrin (12.5 mM) | UV/vis detector | Stuppner and Ganzera, 1995 |

\*OPLC optimum performance laminar chromatography