

# Supplementary Material

“Precise excision of the CAG tract from the Huntingtin gene by Cas9 nickases”  
 Dabrowska M. et al., Frontiers in Neuroscience

**Supplementary Table S1. Oligonucleotides used in the study**

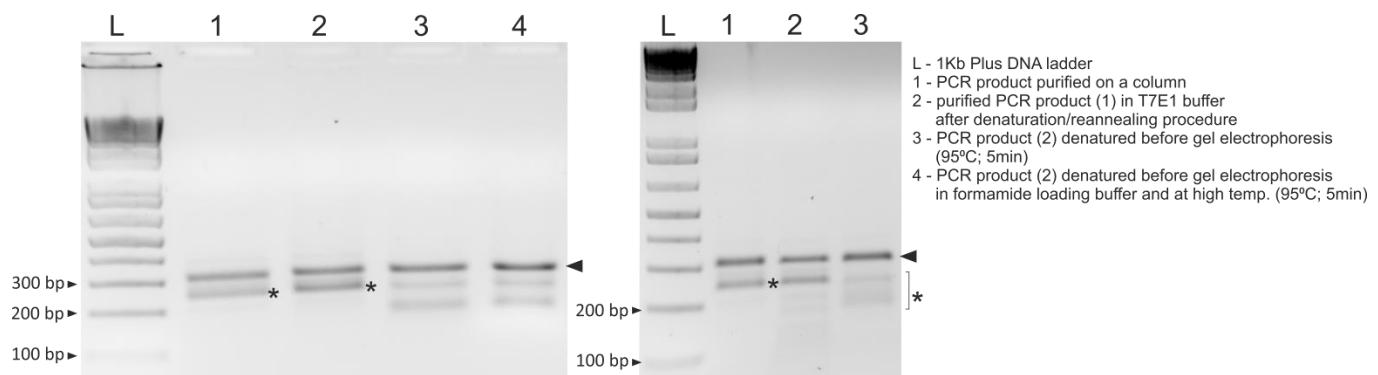
Oligonucleotide ID	Sequence (5'-3')	Description
sgRNA1s	CACCGCTGCTGCTGCTGCTGCTGGAA	oligo for HTT_sgRNA1 plasmid construction
sgRNA1a	AAACTCCAGCAGCAGCAGCAGCAGCAGC	oligo for HTT_sgRNA1 plasmid construction
sgRNA2s	CACCGAGCAGCAGCAGCAGCAGCAGCAG	oligo for HTT_sgRNA2 plasmid construction
sgRNA2a	AAACCTGCTGCTGCTGCTGCTGCTC	oligo for HTT_sgRNA2 plasmid construction
sgRNA3s	CACCGGAAGGACTTGAGGGACTCGA	oligo for HTT_sgRNA3 plasmid construction
sgRNA3a	AAACTCGAGTCCTCAAGTCCTCC	oligo for HTT_sgRNA3 plasmid construction
sgRNA4s	CACCGGCTTCCTCAGCCGCCGCCGC	oligo for HTT_sgRNA4 plasmid construction
sgRNA4a	AAACCGCGCGGCCGCTGAGGAAGCC	oligo for HTT_sgRNA4 plasmid construction
U6-Fwd	GAGGGCCTATTCCATGATTCC	Sequencing primer
HD1F	CCGCTCAGGTTCTGCTTTA	PCR primer, sequencing primer
HD1R	GGCTGAGGCAGCAGCGGCTG	PCR primer
GAPDH_F	GAAGGTGAAGGTCGGAGTC	qRT-PCR primer
GAPDH_R	GAAGATGGTATGGGATTTC	qRT-PCR primer
HD_F	CGACAGCGAGTCAGTGATTG	qRT-PCR primer
HD_R	ACCACTCTGGCTTCACAAGG	qRT-PCR primer
cDNAF	CCCTGGAAAAGCTGATGAAG	primer for <i>HTT</i> cDNA, sequencing primer
cDNAR	TCTTCGGGTCTCTGCTTGT	primer for <i>HTT</i> cDNA
ZFHX3-F	CCAAATAAACCGTCCTCAGC	primer for ZFHX gene- sgRNA1 off-target
ZFHX3-R	TTCCCTTGTCGTGCCTTTC	primer for ZFHX gene- sgRNA1 off-target
TEX13A-F	CGTCCTACCCTGCTTAGTGC	primer for TEX13A gene-sgRNA1 off-target
TEX13A-R	GGTTCGTGGTCCAGAGAAA	primer for TEX13A gene- sgRNA1 off-target
TJP2-F	GTAGCGGCCAATTGACAGT	primer for TJP2 gene- sgRNA4 off-target
TJP2-R	CACAAGGAGGCCTTACGC	primer for TJP2 gene- sgRNA4 off-target
FBXW7-F	CACAGAGCGAGGGAGACAG	primer for FBXW7 gene- sgRNA4 off-target
FBXW7-R	CCTCCTCAGCGTTCTCTCAC	primer for FBXW7 gene- sgRNA4 off-target

**Supplementary Table S2. Predicted exonic off-targets regions for HTT\_sgRNA1 and HTT\_sgRNA4 (<http://crispor.tefor.net>)**

Off-target	Guide sequence <b>sgRNA_1</b> CTGCTGCTGCTGCTGCTGGA	PAM	Chromosome	Gene	Strand	Mismatches
1	CTGCTGCTGCTGCTGCTGGG	GGG	chr16	ZFHX3	+	1
2	CTGCTGCTGCTGCTGCTGGG	GGG	chr19/3'UTR	DMPK	-	1
3	CTGCTGCTGCTGCTGCTGCA	AGG	chr15	SEMA6D	+	1
4	CTGCTGCTGCTGCTGCTGGC	GGG	chr2	APOB/ex1	-	1
5	CTGCTGCTGCTGCTGCTGGC	GGG	chr1	SDC3/ex1	-	1
6	CTGCTGCTGCTGCTGCTGGC	CGG	chr4	SDAD1	-	1
7	CTGCTGCTGCTGCTGCTGGC	AGG	chr11	AP2A2	+	1
8	TTGCTGCTGCTGCTGCTGGC	TGG	chr22	TCF20	+	2
9	CTGCTGCTGCTGCTGCTGGA	GGA	chr1	NOS1AP	-	0
10	CTGCTGATGCTGCTGCTGGA	TGA	chr2	SOX11	-	1
11	CTGCTGATGCTGCTGCTGGA	TGA	chr2	HDAC4	+	1
12	CTGCTGCTGGTGTGCTGCTGGA	GGA	chrX	TEX13A	-	1
13	CTGCTGCTGGTGTGCTGCTGGA	GGA	chrX	TEX13B	-	1

Off-target	Guide sequence <b>sgRNA_4</b> GCTTCCTCAGCCGCCGCCGC	PAM	Chromosome	Gene	Strand	Mismatches
1	TCTTCCTCATCCACGCCAC	TGG	chr12	PXN-AS1	-	4
2	GCTCCTCAGCCGCCGCCGC	TGG	chr8	PLEC	+	3
3	GCTTCCGGAGCCGCCGCCGC	AGG	chr16	CDH5	-	2
4	GCTGCCAGCCGCCGCCGC	AGG	chr12	MBD6	-	2
5	ATTCCTGGCCGCCGCCGC	CGG	chr14	DIO3	+	4
6	GCTCTTTAGCCACCGCCGC	CGG	chr12	YBX3	-	4
7	GCTTCCCAGCAGCCACTGC	TGG	chr17	CDC42EP4	+	4
8	GCTGCCACGCCGCCGCCGC	AGG	chr20	DUSP15/TT LL9	+	3
9	GGTCCTGAGCCGCCGCCGC	GGG	chr17	MMP28	-	3
10	TCCCTCTCAGCCGCCGCCCTC	AGG	chr12	CLEC1A	-	3
11	GCTGCCGCCGCCGCCGCCGC	TGA	chr4	FBXW7	-	3
12	GCTGACGCCGCCGCCGCCGC	GGG	chr9	TJP2	+	4

## FIGURE S1



**Analysis of a non-specific band generated during agarose gel electrophoresis of HTT PCR product.** Genomic DNA from HEK293 cells (controls from Cas9 experiments, see Fig. 1C) was amplified using Phusion High-Fidelity PCR Master Mix with primers HD1F and HD1R spanning CAG repeats in exon 1 of the HTT gene. The two-step PCR amplification program was used as follows: an initial denaturation at 98°C for 3 min; 12 cycles at 98°C for 15 s, 72°C for 15 s; 21 cycles at 98°C for 15 s, 62°C for 15 s, and 72°C for 15 s; and a final elongation at 72°C for 5 min. PCR products were purified using the GeneJET PCR Purification Kit. 400 ng of the purified PCR product (1), PCR product after T7E1 annealing reaction (2) or PCR product after denaturation at high temperature (3) and formamide (4) were separated in 1.3% agarose gels and detected using G-BOX. Two gels represent two independent experiments. The main product (~305 bp) is indicated with an arrowhead. Faster migrating bands are secondary structure forms of the main product (marked with a star) and their contribution is significantly reduced after denaturation of a sample directly before gel electrophoresis.

## FIGURE S2

The sequence targeted by Cas9n/HTT\_sgRNAs within exon 1 of the *HTT* gene

**a) Human huntingtin 3144 aa, 345kDa**

ATG GCG ACC CTG GAA AAG CTG ATG AAG GCC TTC GAG TCC CTC AAG  
TCC TTC **CAG** CAG  
**CAG** CAG CAG CAG CAG CAG CAA CAG CCG CCA CCG CCG CCG CCG  
CCG CCG CCT CCT CAG CTT CCT CAG CCG CCG **CCG** CAG GCA CAG CCG  
CTG CTG CCT CAG CCG CAG CCC CCG CCG CCG CCC CCG CCG CCA  
CCC GGC CCG GCT GTG GC**T GAG** GAG CCG CTG CAC CGA CC**AAAGAAAGA**  
**ACTTTCAG...**

**Cas9n/HTT\_sgRNA1+4; 43 aa 4,73kDa**

ATG GCG ACC CTG GAA AAG CTG ATG AAG GCC TTC GAG TCC CTC AAG  
TCC TTC **CACG** GCA GGC ACA GCC GCT GCT GCC TCA GCC GCA GCC  
GCC CCC GCC GCC CCC GCC ACC CGG CCC GGC TGT GGC **TGA** GGA  
GCC GCT GCA CCG ACC**AAAGAAAGA**ACTTTCAG...

**b) Human huntingtin 3144 aa, 345kDa**

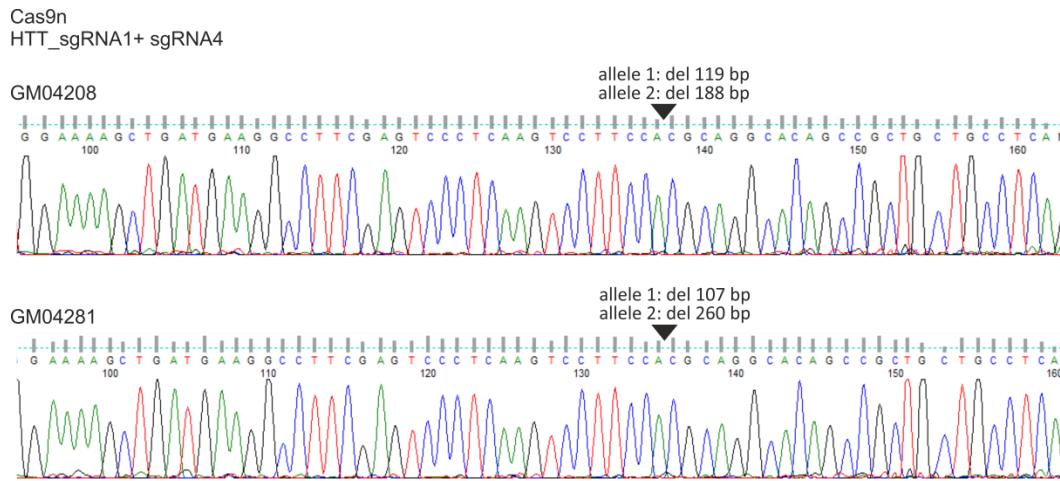
ATG GCG ACC CTG GAA AAG CTG ATG AAG GCC TTC **GAG** TCC CTC AAG  
TCC TTC CAG  
**CAG** CAG CAG CAG CAG CAG CAA CAG CCG CCA CCG CCG CCG CCG  
CCG CCG CCT CCT CAG CTT CCT CAG CCG CCG **CCG** CAG GCA CAG CCG  
CTG CTG CCT CAG CCG CAG CCC CCG CCG CCG CCC CCG CCG CCA  
CCC GGC CCG GCT GTG GC**T GAG** GAG CCG CTG CAC CGA CC**AAAGAAAGA**  
**ACTTTCAG...**

**Cas9n/HTT\_sgRNA3+4; 37 aa, 4kDa**

ATG GCG ACC CTG GAA AAG CTG ATG AAG GCC TTC **GACG** GCA GGC ACA  
GCC GCT GCT GCC TCA GCC GCA GCC CCC GCC GCC GCC CCC GCC  
GCC ACC CGG CCC GGC TGT GGC **TGA** GGA GCC GCT GCA CCG ACC**AAAG**  
**AAAGAAACTTTCAG...**

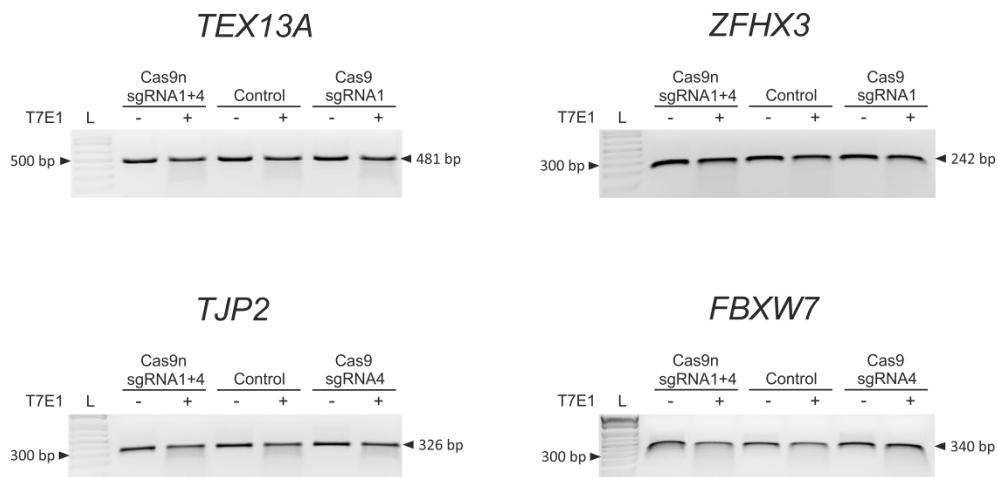
The sequence removed from exon 1 of the *HTT* gene by paired Cas9 nickases is highlighted in red and the nucleotides directly flanking the Cas9n-induced nicks are underlined and indicated in bold. As a result of the CAG repeat excision and frameshift mutation, a TGA STOP codon is generated (highlighted in yellow). The intronic sequence is indicated in blue.

## FIGURE S3



Sanger sequencing analysis of *HTT* gene editing in human fibroblast cell lines.

## FIGURE S4



**Analysis of potential off-target loci by T7E1 mismatch detection assay.** DNA from HEK293T cells treated with Cas9n/HTT\_sg1+4, Cas9/HTT\_sgRNA1, and Cas9/HTT\_sgRNA4 and from the control cells transfected with empty plasmid without sgRNAs was amplified using primers specific for HTT\_sgRNA1 (TEX13A, ZFHX3) and HTT\_sgRNA4 (TJP2, FBXW7) off-target genes. Next, T7E1 analysis was performed to examine cleavage activity in the potential off-target sites. Purified PCR products for appropriate genes were treated (+) and untreated (-) with the T7E1 enzyme and separated on agarose gels with a 1 kb Plus DNA ladder (L).