

# **Invariant natural killer T cells shape the gut microbiota and regulate neutrophil recruitment and function during intestinal inflammation**

Sj Shen<sup>1</sup>, Kathryn Prame Kumar<sup>1</sup>, Dragana Stanley<sup>2</sup>, Robert J. Moore<sup>3,4</sup>, Thi Thu Hao Van<sup>4</sup>,  
Shu Wen Wen<sup>1</sup>, Michael J. Hickey<sup>1</sup> and Connie H. Y. Wong<sup>1,\*</sup>

<sup>1</sup> Centre for Inflammatory Diseases, Monash University Department of Medicine, Monash Medical Centre, Clayton, Victoria, Australia;

<sup>2</sup> School of Health Medical and Applied Sciences, Central Queensland University, Australia;

<sup>3</sup> Infection and Immunity Program, Monash Biomedicine Discovery Institute and Department of Microbiology, Monash University, Australia;

<sup>4</sup> School of Science, RMIT University, Australia;

\*Address for correspondence: Connie H. Y. Wong, PhD., Centre for Inflammatory Diseases, Department of Medicine, School of Clinical Sciences at Monash Health, Monash Medical Centre, Monash University, Clayton, VIC 3168 Australia; Email: [connie.wong@monash.edu](mailto:connie.wong@monash.edu)

Running title: iNKT cells regulate neutrophil recruitment and function

Key words: neutrophils, colitis, inflammation, mucosal immunology, innate immunity

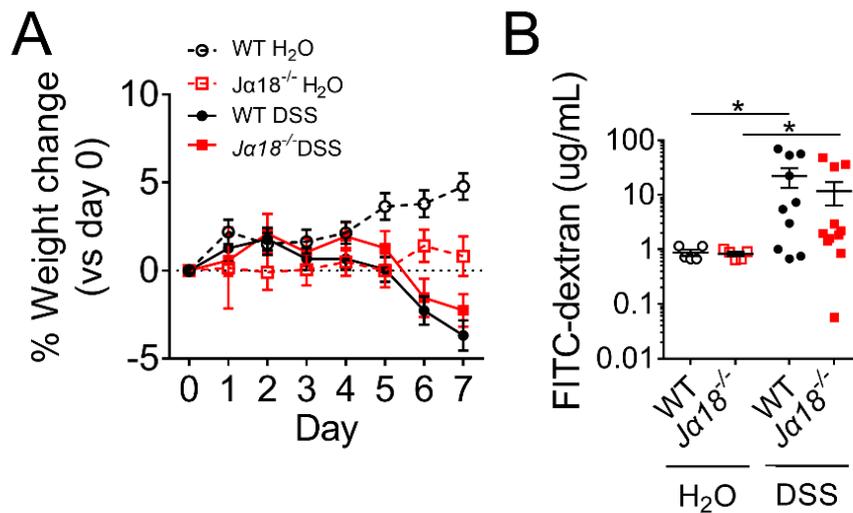
**Supplemental materials**

## Supplemental tables

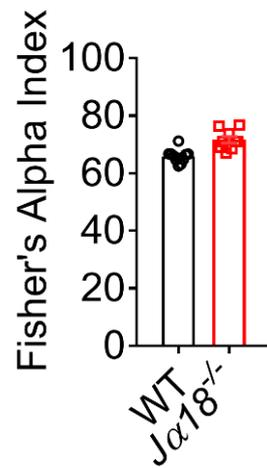
**Supplemental Table 1. Primer sets used for gene expression analysis by qPCR**

Primer		Sequence
<i>18S</i>	F	CTT AGA GGG ACA AGT GGC G
	R	ACG CTG AGC CAG TCA GTG TA
<i>Chil3</i> (Ym-1)	F	AAG CTC TCC AGA AGC AAT CCT G
	R	GAA GAA TTG CCA GAC CTG TGA C
<i>Cxcl1</i>	F	CCG AAG TCA TAG CCA CAC TCA A
	R	GCA GTC TGT CTT CTT TCT CCG TTA C
<i>Cxcl2</i>	F	CCA ACC ACC AGG CTA CAG G
	R	GCG TCA CAC TCA AGC TCT G
<i>Il1b</i>	F	ATG GCA ACT GTT CCT GAA CTC AAC T
	R	CAG GAC AGG TAT AGA TTC TTT CCT TT
<i>Il18</i>	F	CAG GCC TGA CAT CTT CTG CAA
	R	CTG ACA TGG CAG CCA TTG T
<i>Mrc1</i> (CD206)	F	AGG CTG ATT ACG AGC AGT GG
	R	CGC CCA GTA TCC ATC CTT GC
<i>Nos2</i> (iNOS)	F	GCT TGG GTC TTG TTC ACT CC
	R	GCA AGT GAA ATC CGA TGT GGC
<i>Tgfb</i>	F	GAG CCA GAA CGA GAA GTA CCG
	R	CCT CAA GAC GAG CAA TTT CAT CA
<i>Tnfa</i>	F	ATG AGC ACA GAA AGC ATG ATC CGC
	R	CCA AAG TAG ACC TGC CCG GAC TC

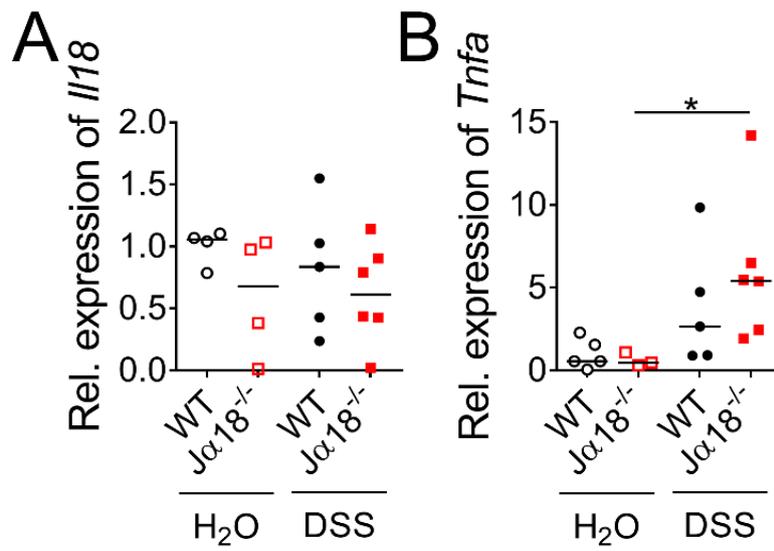
## Supplemental figures



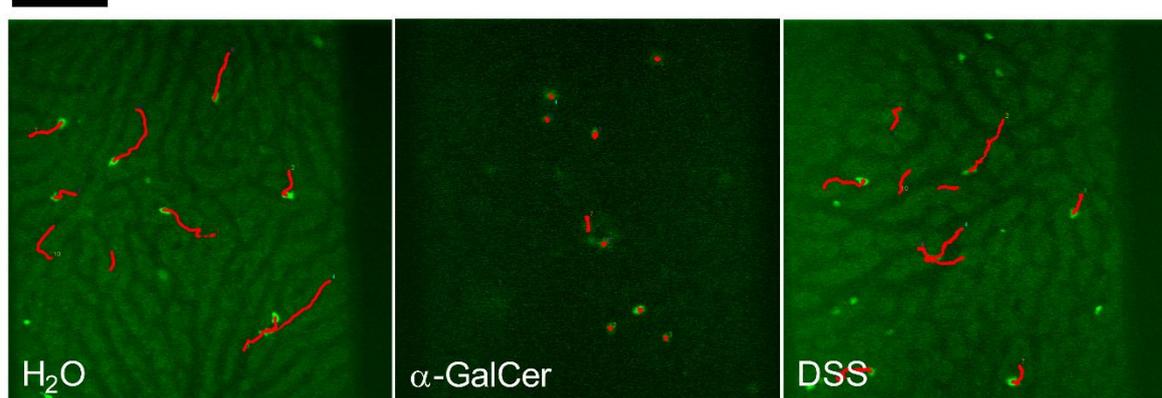
**Supplemental Figure 1.** (A) Changes in weight of WT and *Ja18*<sup>-/-</sup> mice provided with normal drinking water (H<sub>2</sub>O groups) or treated with 2% DSS (DSS groups) were normalised to day 0. (B) Gut permeability of WT and *Ja18*<sup>-/-</sup> mice. FITC-dextran was administered by oral gavage, and its levels in the serum was analysed in WT and *Ja18*<sup>-/-</sup> mice administered with H<sub>2</sub>O or 2% DSS. n≥5. Data represent mean ± SEM. Student's *t* test was performed. These data were obtained from at least 2 independent experiments.



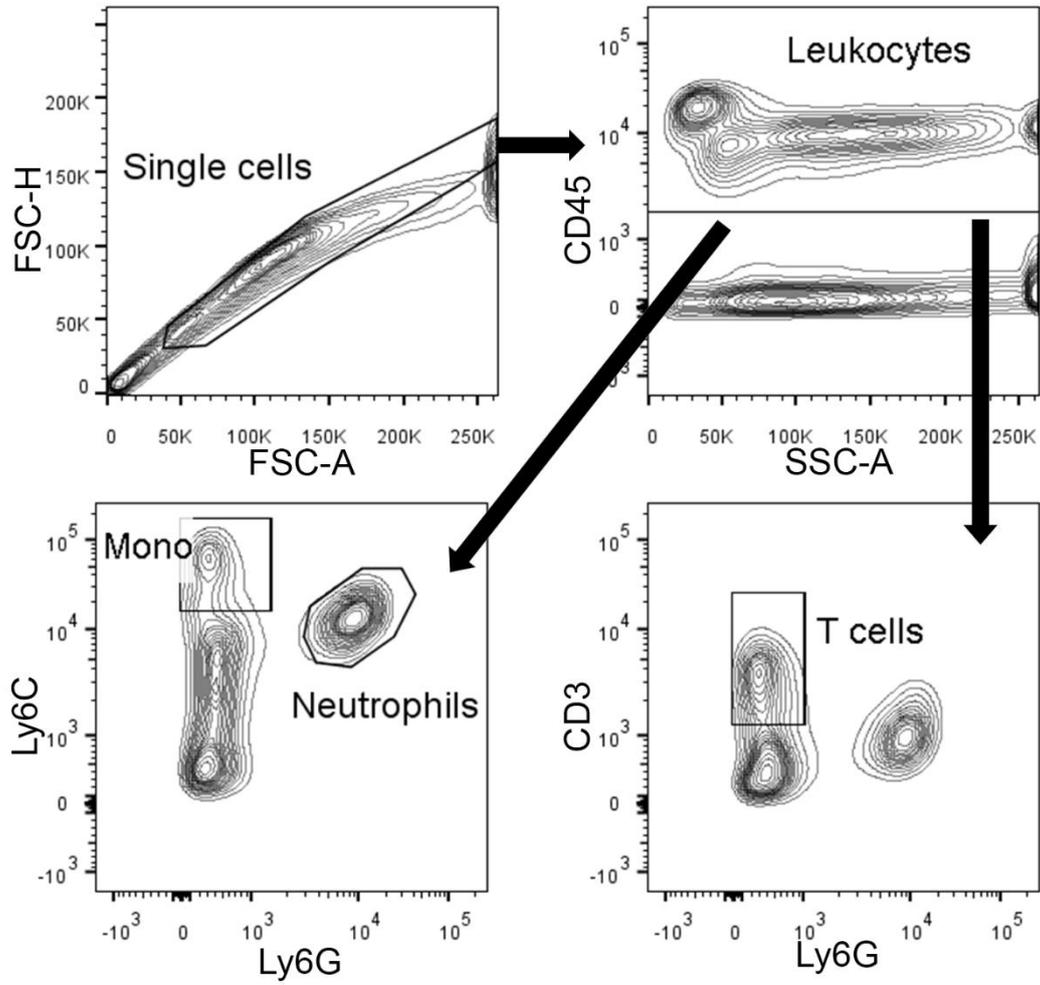
**Supplemental Figure 2.**  $\alpha$ -diversity of the gut microbiota in WT and  $J\alpha 18^{-/-}$  mice.  $n \geq 9$ . Data represent mean  $\pm$  SEM. These data were obtained from one experiment (2 independent cages/group).



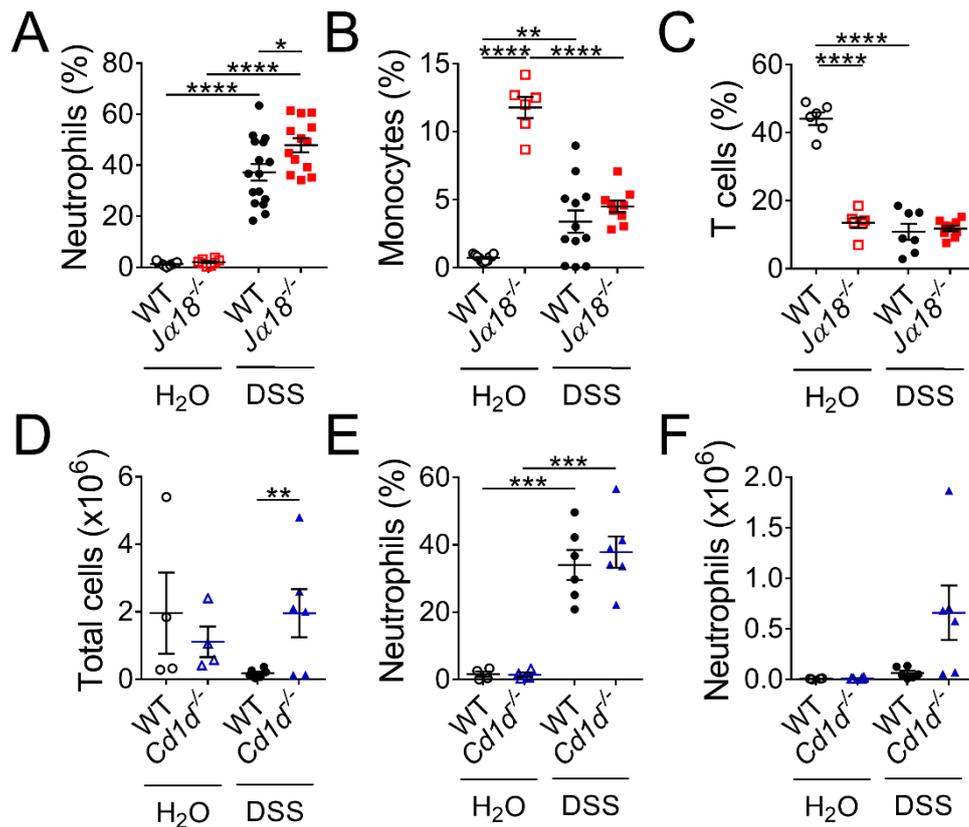
**Supplemental Figure 3.** Local effects of DSS administration in WT and *Ja18*<sup>-/-</sup> mice. (A, B) Expressions of genes in the colon were quantified by qPCR and expressed relative to WT H<sub>2</sub>O controls. n<sub>≥</sub>4. Lines represent medians, and Mann-Whitney U-test was performed. Significance is represented by \*p<0.05. These data were obtained from at least 2 independent experiments.



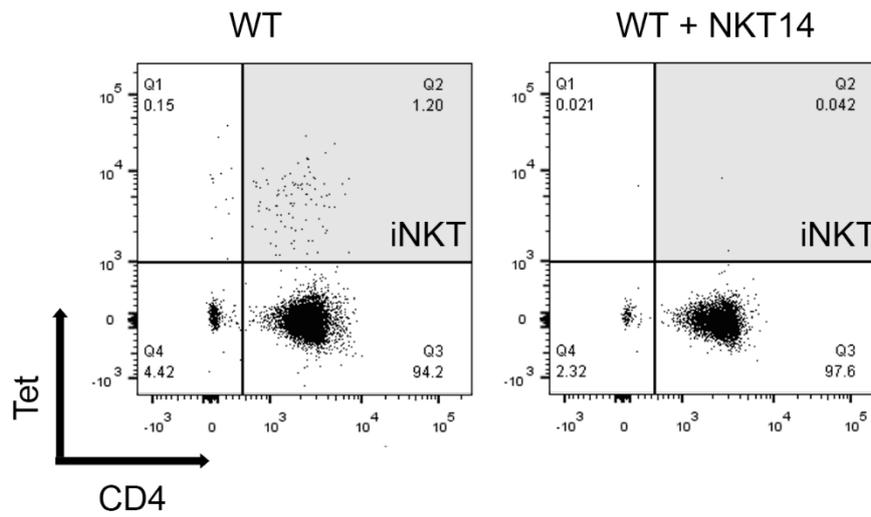
**Supplemental Figure 4.** Representative images showing tracking of iNKT cells in the liver of *Cxcr6<sup>gfp/+</sup>* mice. The use of MTrackJ plugin enabled the tracking of displacement of iNKT cells in the liver of *Cxcr6<sup>gfp/+</sup>* mice on H<sub>2</sub>O control, injected with α-GalCer i.p., or after 7 days of 2% DSS. Bright green cells are iNKT cells; red lines represent paths. Scale bar represents 100 μm.



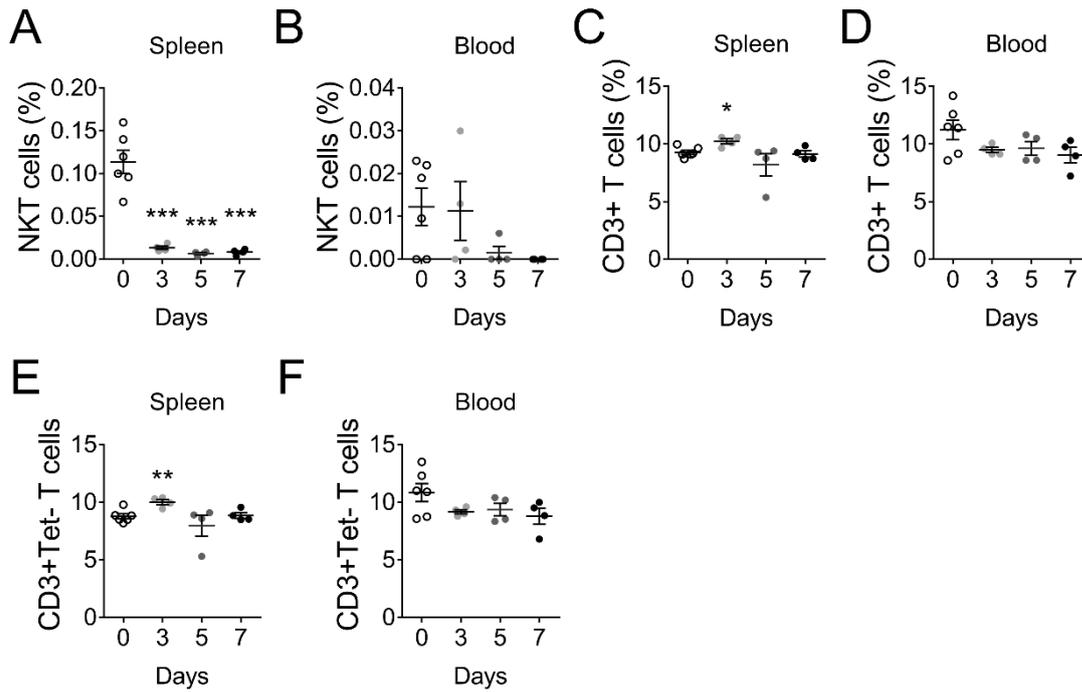
**Supplemental Figure 5.** Gating strategy used to examine leukocyte populations following DSS-induced colitis. Mono: Ly6C<sup>hi</sup> monocytes



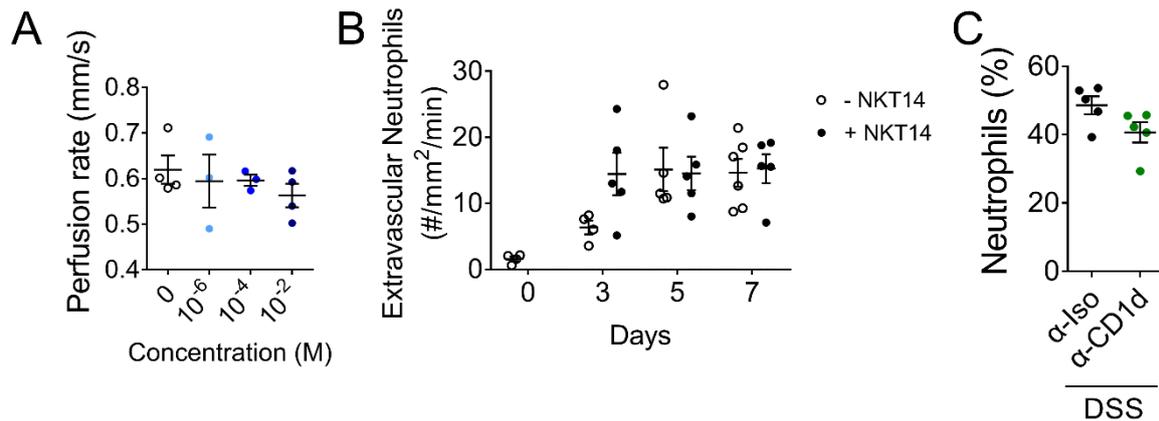
**Supplemental Figure 6.** Changes in leukocyte populations following DSS-induced colitis in WT, *Ja18<sup>-/-</sup>* and *Cd1d<sup>-/-</sup>* mice. (A-C) At day 7, leukocytes were isolated from colon of WT and *Ja18<sup>-/-</sup>* mice and stained with fluorochrome-conjugated antibodies against CD45, CD3, Ly6C, and Ly6G. The proportion (of CD45<sup>+</sup> cells) of (A) Ly6G<sup>+</sup> neutrophils, (B) Ly6C<sup>+</sup>Ly6G<sup>-</sup> monocytes, and (C) CD3<sup>+</sup> T cells were analysed by flow cytometry. (D) At day 7, total leukocyte numbers from colon of WT and *Cd1d<sup>-/-</sup>* mice were counted. (E) The percentage (of CD45<sup>+</sup> cells) and (F) number of Ly6G<sup>+</sup> neutrophils were analysed by flow cytometry.  $n \geq 2$ . Data represent mean  $\pm$  SEM. Student's *t* test was performed. Significance is represented by \* $p < 0.05$ , \*\* $p < 0.01$ , \*\*\* $p < 0.001$  and \*\*\*\* $p < 0.0001$ . These data were obtained from at least 2 independent experiments.



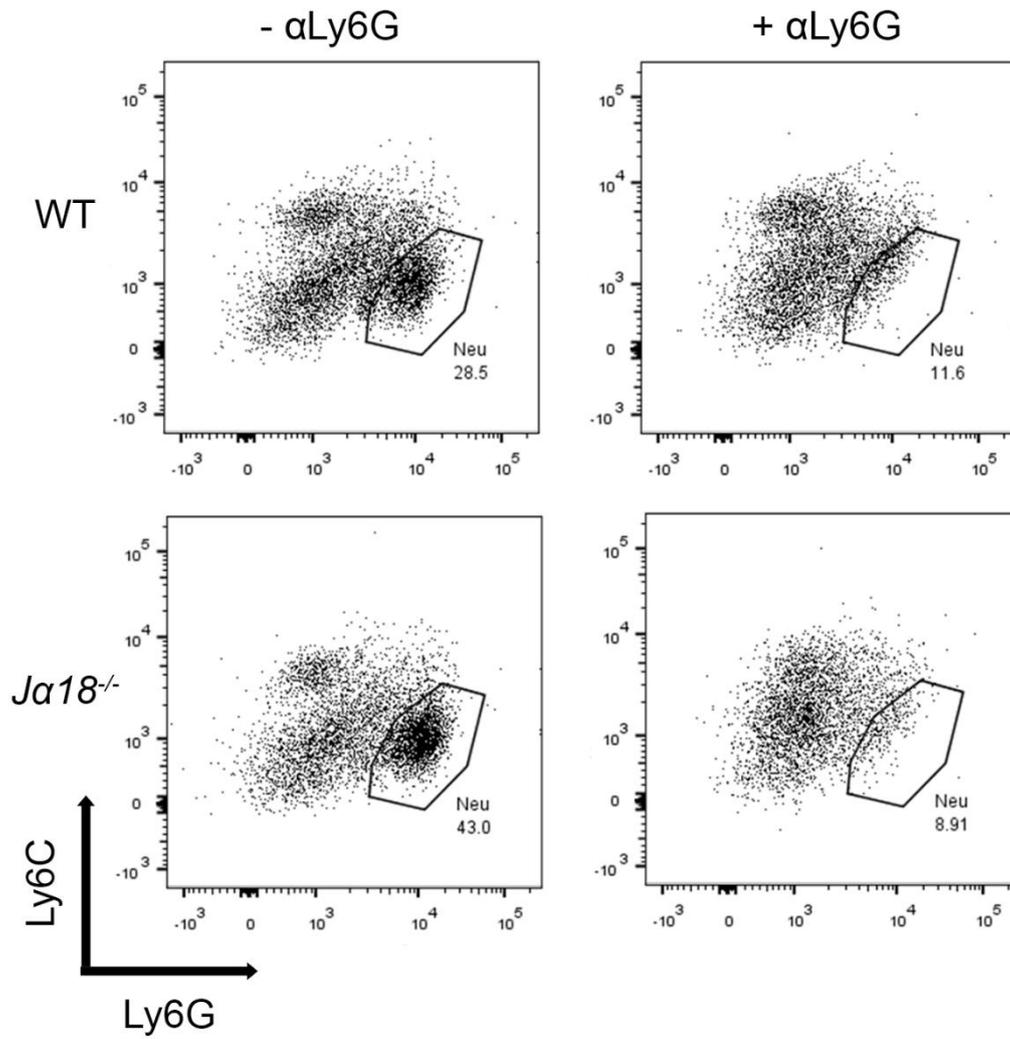
**Supplemental Figure 7.** Representative plot showing depletion of CD4<sup>+</sup> Tetramer (Tet)<sup>+</sup> iNKT cells in the spleen of WT mice 7 days after NKT14 injection. The population is gated from CD45<sup>+</sup> CD3<sup>+</sup> T cells



**Supplemental Figure 8.** Effect of NKT14 treatment in WT mice on T cell populations over 7 days. (A) CD45<sup>+</sup> CD3<sup>+</sup> CD4<sup>+</sup> Tetramer (Tet)<sup>+</sup> NKT cells in the spleen and (B) blood, (C) all T cells in the spleen and (D) blood, and (E) T cells that are Tet<sup>-</sup> in the spleen and (F) blood were assessed at days 0, 3, 5, and 7 by flow cytometry.  $n \geq 4$ . Data represent mean  $\pm$  SEM. Student's *t* test was performed. Significance (vs day 0) is represented by \* $p < 0.05$ , \*\* $p < 0.01$ , \*\*\* $p < 0.001$ . These data were obtained from at least 2 independent experiments.



**Supplemental Figure 9.** Use of atropine superfusion in intravital imaging of colonic neutrophils. (A) The colon of WT mice was superfused with increasing concentrations of atropine, and blood perfusion rate was measured by fluorescent beads using spinning disk confocal microscopy. (B) Extravascular neutrophils were quantified in *LysM<sup>eGFP</sup>* mice administered with or without NKT14. (C) The percentage of neutrophils in DSS-treated WT mice with or without CD1d-blocking antibody was assessed by flow cytometry.  $n \geq 3$ . Data represent mean  $\pm$  SEM. Student's *t* test was performed relative to (A) 0 M, (B) - NKT14 group, or (C) isotype control ( $\alpha$ -Iso). No statistical significances were observed. These data were obtained from at least 2 independent experiments.



**Supplemental Figure 10.** Representative plot showing depletion of Ly6C<sup>+</sup> Ly6G<sup>+</sup> neutrophils in the spleen of WT and *Ja18*<sup>-/-</sup> mice following injections of αLy6G and 7 days of DSS treatment.

**Supplemental Movie 1.** Multiphoton time-lapse video of *LysM<sup>eGFP</sup>* mouse colon microvasculature at baseline. There are minimal neutrophil-endothelial interactions, and few cells outside of the vasculature and in the tissue. The time-lapse covers a period of 3 mins, with approximately 15 s elapsed per second of movie.

**Supplemental Movie 2.** Multiphoton time-lapse video of *LysM<sup>eGFP</sup>* mouse colon microvasculature at day 7 post-DSS administration. Neutrophils are interacting with the endothelium, resulting in increased rolling and adhesion. Additionally, there are also increased extravascular neutrophils. The time-lapse covers a period of 3 mins, with approximately 15 s elapsed per second of movie.