

Supplemental Materials

ATPase assay

The rate NADH oxidation was monitored over several minutes by measuring the absorbance changes at 340 nm, based on NADH an extinction coefficient of $\epsilon_{340} = 6,220 \text{ M}^{-1}\cdot\text{cm}^{-1}$ using a spectrophotometer. The rate of NADH oxidation was plotted and fit to a line with the slope taken as the steady-state value and subsequently converted to $\text{Pi/sec/myosin head}$ based on the HMM concentration in the assay. These measurements were obtained over a range of actin concentrations (1-50 μM) and used to obtain values of V_{max} and K_m . To compare among conditions the ATPase rate at saturating actin concentrations were used in the ANOVA.

Analysis of mini-ensemble laser trap assay data

These data were analyzed using a custom Matlab program previously described (Longyear et al. 2017). Briefly, the program employs a threshold algorithm that scores displacements that had greater than 8 nm from a defined baseline that have a duration ≥ 10 ms. These values were chosen based on the lower limit of the average displacement for two-headed skeletal muscle myosin (Tyska and Warshaw 1999) and the expected lifetime of actomyosin strong-binding at 100 μM ATP 10 ms (White and Taylor 1976). The highest force sustained for ≥ 10 ms during a binding event was taken as the peak force. Event duration was determined based on the time spent $> 8\text{nm}$ from baseline and event frequency was determined by dividing the number of events by the total time of the displacement recording (typically 10-30sec each).

Statistical Analysis

Differences among the different conditions in the laser trap data were determined using a non-parametric Kruskal-Wallis ANOVA and associated post hoc test was used to locate differences. Non-parametric tests were used because the data force and lifetime data were not Normally-distributed (Shapiro-Wilk test) and had unequal variances (Unequal Variance test).

Differences among the difference conditions of the ATPase assay were determined using a one-way ANOVA followed by a Tukey's HSD to locate the differences. The ATPase data were fit with the Michealis-Menten relation to derive values for V_{\max} and K_m . All statistical analyses were performed using SigmaPlot® 11.0.