

Table S1. Bacterial strains used in this study.

Strains	Genotype and relevant features ^a	References
<i>E. coli</i>		
TG1	<i>supE hsdΔ5 thi Δ(lac-proAB) F'[traD36 proAB+ lacIq lacZΔM15]</i>	(Gibson, 1984)
TG1Nal	Nal ^R derivative of TG1	(Kiss et al., 2012)
TG2	<i>supE hsdΔ5 thi Δ(lac-proAB)Δ(srl-recA)306::Tn10(Tc^R) F'[traD36 proAB+ lacIq lacZΔM15]</i>	(Sambrook et al., 1989)
TG90	<i>pcn B80 zad::Tn10 (Tc^R)</i> derivative of TG1	(Gonzy-Treboul et al., 1992)
TG90Nal	Nal ^R derivative of the TG90, Tc ^R , Nal ^R	(Kiss et al., 2012)
TG1/R55	TG1 strain containing R55, Ap ^R , Cm ^R , Flo ^R , Sul ^R , Km ^R , Gm ^R	(Kiss et al., 2015)
TG1Nal/R55	TG1Nal strain containing R55, Nal ^R , Ap ^R , Cm ^R , Flo ^R , Su ^R , Km ^R , Gm ^R	(Kiss et al., 2015)
TG1Nal/R16a	TG1Nal strain containing R16a, Nal ^R , Ap ^R , Km ^R , Sul ^R	(Szabó et al., 2016)
TG1Nal::SGII-C	TG1Nal strain containing SGII-C variant integrated into <i>E. coli thdF</i> , Nal ^R , Sm ^R , Sp ^R , Sul ^R	(Kiss et al., 2015)
TG1Nal::SGII-C ^{ΔoriT}	TG1Nal strain containing the <i>ΔoriT</i> mutant SGII-C, in which the 18016-18140 bp region was replaced, Nal ^R , Sm ^R , Sp ^R , Sul ^R	this work
TG1Nal::SGII-C ^{ΔS019}	TG1Nal strain containing the <i>mpsB</i> (S019) KO mutant SGII-C, in which the 16656-16739 bp region was replaced, Nal ^R , Sm ^R , Sp ^R , Sul ^R	this work
TG1Nal::SGII-C ^{ΔS020}	TG1Nal strain containing the <i>mpsA</i> (S020) KO mutant SGII-C, in which the 17571-17709 bp region was replaced, Nal ^R , Sm ^R , Sp ^R , Sul ^R	this work
TG1Nal::mob _{SGII} /R55	TG1Nal::[miniTn10::mob _{SGII} -Km ^R] mobilization helper strain containing R55 and the 16447-18680 bp SGII region integrated into the chromosome by miniTn10 transposition, Nal ^R , Km ^R , Ap ^R , Cm ^R , Flo ^R , Sul ^R , Gm ^R	this work
S17-1 λpir	S17-1 λpir, a λ lysogen derivative of S17-1 (<i>pro thi recA hsdR</i> (r ⁻ m ⁺) Tp ^R Sm ^R Km ^S [Ω RP4-2-Tc::Mu-Km::Tn7]) expressing Π protein from <i>pir</i> gene of R6K	(Simon et al., 1983)
BM14	J5-3 derivative, <i>pro met azi</i> , Az ^R	Inst.Pasteur, France
<i>Salmonella enterica</i>		
<i>S. Agona</i> 47SA97	SGII-C ^{WT} , Sm ^R Sp ^R Sul ^R	(Boyd et al., 2002)
<i>S. Agona</i> 47SA97 SGII ^{Δint}	Derivative of <i>S. Agona</i> 47SA97 harbouring Δint mutant SGII-C, Sm ^R Sp ^R Sul ^R	(Doublet et al., 2005)

Table S2. Relevant features of plasmids used in this study.

Plasmid name	Relevant features ^a	References
R55	IncC Type2, tra+, Cm ^R , Flo ^R , Sul ^R , Ap ^R , Km ^R , Gm ^R	(Chabbert et al., 1972)
R55 ^{ΔTn6187}	R55 derivative, Tn6187 was deleted, tra+, Cm ^R , Flo ^R , Sul ^R , Ap ^S , Km ^S , Gm ^S	this work
R16a	IncC Type1, tra+, Ap ^R , Km ^R , Sul ^R	(Chabbert et al., 1972)
R16a ^{Δtral}	IncC Type1, tra-, Δtral::Cm ^R , Ap ^R , Km ^R , Sul ^R	(Hegyí et al., 2017)
pACYC184	p15A-based Tc ^R Cm ^R cloning vector	(Rose, 1988)
pBluescript II-SK	pMB1-based Ap ^R cloning vector	(Short et al., 1988)
pEMBL19	pMB1-based Ap ^R cloning vector	(Dente et al., 1988)
pKD3	R6Kγ-based PCR template plasmid with FRT-flanked <i>cat</i> gene for one-step recombination gene-KO Cm ^R , Ap ^R	(Datsenko and Wanner, 2000)
pKD46	Ap ^R ara-inducible expression vector of λ Red recombinase, temperature-sensitive pSC101 replication system	(Datsenko and Wanner, 2000)
pCP20	Thermo-inducible FLP recombinase expression (λ p _R :: <i>FLP</i>), temperature-sensitive pSC101 replication system, λ <i>cl857</i> , Ap ^R , Cm ^R	(Cherepanov and Wackernagel, 1995)
pACYC-1	145-1351 bp region of SGII cloned in pACYC184	this work
pACYC-2	1331-2324 bp region of SGII cloned in pACYC184	this work
pACYC-3	2354-3874 bp region of SGII cloned in pACYC184	this work
pACYC-4	3921-6536 bp region of SGII cloned in pACYC184	this work
pACYC-5	6516-8851 bp region of SGII cloned in pACYC184	this work
pACYC-6	8860-11754 bp region of SGII cloned in pACYC184	this work
pACYC-7	11734-13434 bp region of SGII cloned in pACYC184	this work
pACYC-8	13414-15164 bp region of SGII cloned in pACYC184	this work
pACYC-9	15144-16913 bp region of SGII cloned in pACYC184	this work
pACYC-10	17005-19833 bp region of SGII cloned in pACYC184	this work
pACYC-11	19852-21930 bp region of SGII cloned in pACYC184	this work
pACYC-12	21981-23590 bp region of SGII cloned in pACYC184	this work
pACYC-13	23570-25250 bp region of SGII cloned in pACYC184	this work
pACYC-14	25230-28141 bp region of SGII cloned in pACYC184	this work
pACYC-10A	17005-18348 bp region of SGII cloned in pACYC184	this work

pACYC-10D	17534-18348 bp region of SGII cloned in pACYC184	this work
pACYC-10D2	17534-17939 bp region of SGII cloned in pACYC184	this work
pACYC-10D4	17799-18348 bp region of SGII cloned in pACYC184	this work
pACYC-10D4B	18017-18348 bp region of SGII cloned in pACYC184	this work
pACYC-10D4C	18132-18348 bp region of SGII cloned in pACYC184	this work
pACYC-10D4E	18017-18261 bp region of SGII cloned in pACYC184	this work
pACYC-10D7	18017-18151 bp region of SGII cloned in pACYC184	this work
pFOL1343	Sm ^R derivative of pJKI671.	this work
pFOL1362	15444-22496 bp (S015-S025) region of SGII cloned in pJK708.	this work
pFOL1365	BglII deletion derivative of pFOL1362 containing 15444-16807 bp and 21056-22496 bp regions of SGII.	this work
pFOL1372	EcoRI deletion derivative of pFOL1362 containing 15444-19843bp (S015-S023) region of SGII.	this work
pJKI88	p15A-based Km ^R cloning vector deriving from pACYC177 (Rose, 1988).	(Kiss and Olasz, 1999)
pJKI391	p15A-based Km ^R expression vector deriving from pJKI88.	(Kiss et al., 2015)
pJKI669	d1 deletion derivative of SGII-C (SGII-C-d1, (Kiss et al., 2012)) containing DRL-S004 and S013-DRR regions of SGII-C cloned in the pJKI88-derived vector pJKI633.	this work
pJKI672	BssHII deletion derivative of pJKI669 containing DRL-S004, S019 and <i>intI1</i> -DRR regions of SGII-C.	this work
pJKI678	MfeI deletion derivative of pJKI669 containing DRL and S025-DRR regions of SGII-C.	this work
pJKI708	Sm ^R derivative of the p15A-based cloning vector, pJKI88	(Hegyi et al., 2017)
pJKI710	16807-19427 bp (S020-S023) region of SGII cloned in pJK708.	this work
pJKI725	PstI-SacI deletion derivative of pFOL1372 containing the 16594-19843 bp (S020-S023) region of SGII.	this work
pJKI726	15439-16595 bp (S015-S019) region of SGII cloned in pJK708.	this work
pJKI731	15439-18050 bp (S015-S021) region of SGII cloned in pJK708.	this work
pJKI737	pFOL1372 derivative carrying KO mutation in S020 in the 15444-19843bp (S015-S023) region of SGII.	this work
pJKI772	pFOL1372 derivative carrying KO mutation in S019 in the 15444-19843bp (S015-S023) region of SGII.	this work
pJKI774	pFOL1372 derivative carrying KO mutation in S022 in the 15444-19843bp (S015-S023) region of SGII.	this work
pJKI775	15439-18680 bp (S015-S022) region of SGII cloned in pJK708.	this work
pJKI776	15849-18680 bp (S018-S022) region of SGII cloned in pJK708.	this work
pJKI777	16087-18680 bp (S019-S022) region of SGII cloned in pJK708.	this work
pJKI780	16447-18680 bp mob _{SGII} region (S019-S022) of SGII cloned in pJK708.	this work
pJKI781	16447-18140 bp (S019-S021) region of SGII cloned in pJK708.	this work
pJKI791	18304-18680 bp of SGII (upstream region of S022) cloned in pJK708.	this work
pJKI796	pLOFKm (Herrero et al., 1990) derivative R6K-based delivery plasmid containing the 16447-18680 bp (S019-S022) region of SGII with a Km ^R gene in the transposable mini-Tn10 unit. R6K _{ori} , <i>oriT_{RK2}</i> , Ap ^R , <i>lac^R</i> , <i>P_{lac}::Tn10</i> transposase, miniTn10::SGII ₁₆₄₄₇₋₁₈₆₈₀ -Km ^R .	this work
pJKI810	17713-18140 bp region of SGII cloned in pJK708.	this work
pJKI811	17713-18050 bp region of SGII cloned in pJK708.	this work
pJKI818	17961-18140 bp region of SGII cloned in pJK708.	this work
pJKI833	Cm ^R , Sm/Sp ^S derivative of pJKI737 carrying KO mutation in <i>mpsA</i> in the 15444-19843bp (S015-S023) region of SGII.	this work
pJKI835	Cm ^R , Sm/Sp ^S derivative of pJKI772 carrying KO mutation in <i>mpsB</i> in the 15444-19843bp (S015-S023) region of SGII.	this work
pJKI836	Cm ^R , Sm/Sp ^S derivative of pFOL1372.	this work
pJKI842	Tc ^R , Ap ^S derivative of pKD46, the ara-inducible expression vector of λ Red recombinase with temperature-sensitive pSC101 replication system	this work
pJKI871	18016-18140 bp region of SGII (<i>oriT_{SGII}</i>) cloned in pJK708.	this work
pJKI872	18016-18119 bp region of SGII (<i>oriTΔIR3R</i>) cloned in pJK708. The truncated <i>oriT</i> lacks the right copy of IR3.	this work
pJKI873	18016-18140 bp region of SGII (<i>oriTΔIR2R</i>) cloned in pJK708. The right copy of IR2 in <i>oriT</i> is eliminated by base substitutions.	this work
pJKI874	18035-18140 bp region of SGII (<i>oriTΔIR1L</i>) cloned in pJK708. The truncated <i>oriT</i> lacks the left copy of IR1.	this work
pJKI935	Cm ^R pJKI391 derivative p15A-based vector expressing <i>mpsA</i> under the control of <i>P_{lac}</i> promoter	this work
pJKI937	Cm ^R pJKI391 derivative p15A-based vector expressing <i>mpsB</i> under the control of <i>P_{lac}</i> promoter	this work
pJKI948	16447-18680 bp mob _{SGII} region (S019-S022) of SGII, a Cm ^R derivative of pJKI780	this work
pJKI990	ColE1-based cloning vector for β-gal assays, containing promoterless <i>lacZ</i> gene preceded by pHP45Ω (Prentki and Krisch, 1984) and <i>rrnB</i> terminators.	(Kiss et al., 2015)
pJKI1023	Sm ^R /Sp ^R derivative of the R6K _{ori} -based PCR template plasmid pSG76-CS (Kolisnychenko et al., 2002), where I-SceI cleavage sites flank the resistance cassette.	this work
pMNI41	Km ^R derivative of pJKI871, carrying the <i>oriT_{SGII}</i> .	this work
pMSZ934	Ap ^R , Km ^R mobilizable derivative of the I-SceI producer plasmid pSTKST (Kolisnychenko et al., 2002). Temperature-sensitive pSC101 replication system, Tc ^R , <i>P_{tet}::SCEI</i> , <i>oriT_{RK2}</i>	this work
pMSZ947	pJKI990-derivative β-galactosidase tester plasmid containing the non-coding upstream region of <i>mpsA</i> (17710-18681 bp) fused to the promoterless <i>lacZ</i> gene.	this work
pMSZ948	pJKI990-derivative β-galactosidase tester plasmid containing the non-coding upstream region of <i>mpsA</i> (17710-18050 bp) fused to the promoterless <i>lacZ</i> gene.	this work
pMSZ949	16447-18680 bp mob _{SGII} region (S019-S022) of SGII cloned in pJKI88, a Km ^R equivalent of pJKI780.	this work
pMSZ957	16447-18680 bp mob _{SGII} region (S019-S022) of SGII cloned in pJKI88, a Km ^R equivalent of pJKI780. pMSZ957 contains a single T insertion at 17816 th position, which generates a new StuI site and cause frameshift in S021.	this work
pMSZ976	16447-17805 bp region of SGII (<i>mpsAB</i> +94 bp upstream of <i>mpsA</i>) cloned in pJKI88.	this work
pMSZ980	16447-17881 bp region of SGII (<i>mpsAB</i> +170 bp upstream of <i>mpsA</i>) cloned in the pJKI88-analogue pasmid pMSZ973.	this work
pMSZ981	16447-17712 bp region of SGII (<i>mpsAB</i>) cloned in pJKI88.	this work
pMSZ984	16447-17781 bp region of SGII (<i>mpsAB</i> +70 bp upstream of <i>mpsA</i>) cloned in pJKI88.	this work
pMSZ988	16447-16743 bp region of SGII (<i>mpsB</i>) cloned in pJKI88.	this work
pMSZ989	18042-18140 bp region (<i>oriT ΔIR1</i>) of SGII cloned in pJK708. The truncated <i>oriT_{SGII}</i> fragment lacks the IR1 repeat.	this work
pMSZ990	18048-18140 bp region (<i>oriT ΔIR1</i> +spacer to IR2L) of SGII cloned in pJK708. The truncated <i>oriT_{SGII}</i> fragment lacks the IR1 repeat and the 6-bp spacer sequence to IR2L.	this work
pMSZ991	18024-18140 bp region <i>oriT_{SGII}</i> cloned in pJK708. This fragment lacks the 7 bp preceding IR1L and contains a single base (C) deletion at 18038 bp position in IR1R.	this work
pMSZ993	16447-17732 bp region of SGII (<i>mpsAB</i> +20 bp upstream of <i>mpsA</i>) cloned in pJKI88.	this work
pMSZ995	18024-18140 bp region <i>oriT_{SGII}</i> cloned in pJK708. This fragment lacks the 7 bp preceding IR1L and contains a single base	this work

	(G) deletion at 18037 bp position in IR1R.	
pMSZ996	16447-17619 bp region of SGI1 (<i>mpsB-mpsA</i> beginnig with the 2 nd inframe ATG codon) cloned in pJK188.	this work
pMSZ997	18024-18140 bp region <i>oriT</i> _{SGI1} cloned in pJK708. This fragment lacks the 7 bp preceding IR1L.	this work
pMSZ1017	pJK1990-derivative β -galactosidase tester plasmid containing the upstream region of <i>mpsB</i> (16741-16975 bp) fused to the promoterless <i>lacZ</i> gene.	this work

Table S3. List of oligonucleotides used.

Primer	Sequence (5'-3')a	Reference
ampforXSP	gctctagagtcgacctgcagtagcattcaaatatgtatccgctc	this work
amprevXP	aatctagactgcagggtctgacagttaccaatgc	this work
attsgil for	gctctagagcggccgcatggaaggcggcttctctggc	(Kiss et al., 2012)
attsgil rev	gctctagagcggccgcaaatgtgaatcgaatcacaaatcg	(Kiss et al., 2012)
deloriTfor	gtgtcattctttgaaaggaaagcgcgaagcgcgctaaccgccgaaggcgGTGTAGGCTGGAGCTGCTTC	this work
deloriTrev	aagaggccctccctcatccgtcagaacgagtcgtgatttccggcttactCATATGAATATCCTCCTTAGTTC	this work
delS019for	gtgatgtgctggtgactccttttattggcggcaaatatcgttagagccGTGTAGGCTGGAGCTGCTTC	this work
delS019rev	gggtcatggtcgcacgatgtgacaaatagttatttgggtaattgatggCATATGAATATCCTCCTTAGTTC	this work
delS020for	tatgtcccgcatgtcctttatgccctgttcggacttaatecaagcgctaaGTGTAGGCTGGAGCTGCTTC	this work
delS020rev	acggggccaaagaagtaacaattttgattaacagagttagggggatcaatgCATATGAATATCCTCCTTAGTTC	this work
delS022for	gtgtttggcgcctcggaattgagcgcagggaacgcgacatagctgtcagcGTGTAGGCTGGAGCTGCTTC	this work
delS022rev	aggggcactcctcgtctaaaacctatctccccggaggaaaatcagatgtCATATGAATATCCTCCTTAGTTC	this work
FwEcoRI1	attgtgaattcttctgtattgggaagtaaat	this work
FwEcoRI10	attgtgaattcgtaggcttcttgcggaact	this work
FwEcoRI10D	attgtgaattcctttatgccctgttcggact	this work
FwEcoRI10D3A	attgtgaattcctctgcatgctaaggccaac	this work
FwEcoRI10D4B	tattgtgaattctataattcgcgcacattcgt	this work
FwEcoRI10D4C	tattgtgaattcggagcatagagtaagccgga	this work
FwEcoRI11	attgtgaattcaagaaaacatcgctgaagt	this work
FwEcoRI12	attgtgaattcacgcttgagttatctcttc	this work
FwEcoRI13	attgtgaattcatactaaagttgttaccggc	this work
FwEcoRI14	attgtgaattcctctgatgcatcttgctcta	this work
FwEcoRI2	attgtgaattcaaaacatgttacttccacg	this work
FwEcoRI3	attgtgaattcgcagtcactttcttaactt	this work
FwEcoRI4	attgtgaattcacaaaagtttaaggccaagt	this work
FwEcoRI5	attgtgaattcaacacatgttcgctgattaa	this work
FwEcoRI6	attgtgaattcgtcaccatattgtgaacaa	this work
FwEcoRI7	attgtgaattcgtgtacgcgtcctcaatag	this work
FwEcoRI8	attgtgaattcctcactactgttgcctcaat	this work
FwEcoRI9	attgtgaattctagaccagatcttgagcat	this work
oriTd1for	aaactgcagtgccggtgcgaaagcc	this work
oriTd2for	aaactgcagtcgcgcacattcgtgcggtgcgaaag	this work
oriTd2rev	ttgaattcgccttaggcgtaaacagag	this work
oriTd3for	aaactgcagtcgcaaaccttagagcccttg	this work
oriTd4for	aaactgcagaagccttagagcccttgaggc	this work
oriTfor	aaactgcagtataattcgcgcacattcgtg	this work
oriTIR2mutrev	cgacgggaagcttcactcctcaagggc	this work
pBRTcPstfor	aaactgcaggcgtatcacgagccctttc	this work
pBRTcPstrev	aaactgcagtggtgaatccgttagcagg	this work
R55_dTn6187seqfor	aaactgcagtcgcttttgcagcgttc	this work
R55-dTn6187ABfor	gtcttcgagttgccagctttccaacgcgtgaaagtaccctctctgatccatcgctgctacccaatcagcgtggatggaccgaaattgtccTCAACAGGTT GAACTGCGGATC	this work
R55-dTn6187Crev	ctcagaaaacggaaatctatgtgactccctttttgcaacaccgattttgGATTTAGGTGACACTATAGAATAC	this work
R55-dTn6187seqrev	tagtcgacagatttagaccatcatgcaacg	this work
RvNcoI1	attgtccatggcgtggaaagtaacatgtttt	this work
RvNcoI10	attgtccatggacgatcagcaatatgaact	this work
RvNcoI10A	attgtccatggatctccccggaggaaaatca	this work
RvNcoI10D2	attgtccatgggttgttccgtcgggaacagac	this work
RvNcoI10D3	attgtccatgggtcggcttactctatgctcc	this work
RvNcoI10D4D	attgtccatggaggcttccatcgcgatct	this work
RvNcoI11	attgtccatgggttacgggtatcgccctaagt	this work
RvNcoI12	attgtccatgggcccggtaacaacttagatat	this work
RvNcoI13	attgtccatgggtagagcaagatgcatcagag	this work
RvNcoI14	attgtccatgggaatatcgctgtatggcttca	this work
RvNcoI2	attgtccatggattccataccgtaattgact	this work
RvNcoI3	attgtccatggggatattgattgagtaagg	this work
RvNcoI4	attgtccatgggttaatcagcgaacatgtgtt	this work
RvNcoI5	attgtccatggcagatagcaatggatactgc	this work
RvNcoI6	attgtccatggctattgaggacgcgtacaac	this work
RvNcoI7	attgtccatggattggagcaacagtagaga	this work
RvNcoI8	attgtccatggatgctccaagatctggtcta	this work
RvNcoI9	attgtccatggcggtaaaagcaggtgttaaa	this work
S019Ndefor	gatgatcatatggaaaatgggtaagagaaagaagc	this work
S019promfor	atccatggattaccccaataactattgtcacatcg	this work
S020for1	tactcatcctgcaggcttttaaac	this work
S020for2	aaactgcagtcgatacataaaatctccctc	this work

S020for3	<u>a</u> actgcagaatcgaagccttatttagtag	this work
S020for4	<u>a</u> actgcagttaatcagcagagccggtgtttttg	this work
S020Nde_for2	<u>g</u> atgatcatatgaagagtttcagtcctatagcc	this work
S020Ndefor	<u>g</u> atgatcatatgcgttcagagcggactaatccggat	this work
S020promfor_Nc	<u>a</u> accatggatccccctaactctgttaate	this work
S020promrev	<u>a</u> agaattctttcgcaccgcgcacgaatg	this work
S021for	<u>a</u> actgcagtgcacccctaactctgttaate	this work
S021for_Stu,Sph	<u>c</u> tgcactgaagcctaacgcctgggac	this work
S021for2	<u>c</u> ggctgcagctgtcgtgtcattctttgg	this work
S021promrev	<u>a</u> agaattcctatgctccaccgcgtctctg	this work
S022promfor	<u>c</u> actgcagtgcacgacacatgatgataaacatc	this work
S022promrev	<u>a</u> agaattcgttaagttagtgagcatccaac	this work
S022promrev_P	<u>a</u> actgcagtaagttagttgagcatccaac	this work
sgi_17781rev	<u>g</u> tgaattcggatccgtcgactttcattaccacggatacggac	this work
sgi_orf019rev	<u>a</u> actgcagggatccttaatacagcagagccggtgtttttg	this work
sgi_orf020for	<u>a</u> acatagtcgttcagagcggactaatc	this work
sgi_S020rev	<u>a</u> aggatccttaccaccaataactattgtcac	this work
SGI1orf020_17119for	<u>g</u> aacagtgcgcgccggccac	this work
SmRforSmP	<u>c</u> gctgcagcccggtgtccgggtgacgcac	this work
SmRevSmP	<u>a</u> actgcagcccggtcggctgaacgaattgttagac	this work

^a Uppercase shows the template plasmid sequence in primers used for producing the KO amplicons. Restriction cleavage sites are underlined.

Supplementary References

- Boyd, D., Cloeckert, a., Chaslus-Dancla, E., and Mulvey, M. R. (2002). Characterization of Variant Salmonella Genomic Island 1 Multidrug Resistance Regions from Serovars Typhimurium DT104 and Agona. *Antimicrob. Agents Chemother.* 46, 1714–1722. doi:10.1128/AAC.46.6.1714-1722.2002.
- Chabbert, Y. A., Scavizzi, M. R., Witchitz, J. L., Gerbaud, G. R., and Bouanchaud, D. H. (1972). Incompatibility Groups and the Classification of f- Resistance Factors. *J. Bacteriol.* 112, 666–675.
- Cherepanov, P. P., and Wackernagel, W. (1995). Gene disruption in Escherichia coli: TcR and KmR cassettes with the option of FLP-catalyzed excision of the antibiotic-resistance determinant. *Gene* 158, 9–14. doi:10.1016/0378-1119(95)00193-A.
- Datsenko, K. A., and Wanner, B. L. (2000). One-step inactivation of chromosomal genes in Escherichia coli K-12 using PCR products. *Proc. Natl. Acad. Sci. U. S. A.* 97, 6640–5. doi:10.1073/pnas.120163297.
- Dente, L., Cesareni, G., and Cortese, R. (1983). pEMBL: A new family of single stranded plasmids. *Nucleic Acids Res.* 11, 1645–1655. doi:10.1093/nar/11.6.1645.
- Doublet, B., Boyd, D., Mulvey, M. R., and Cloeckert, A. (2005). The Salmonella genomic island 1 is an integrative mobilizable element. *Mol. Microbiol.* 55, 1911–1924. doi:10.1111/j.1365-2958.2005.04520.x.
- Gibson, T. J. (1984). Studies on the Epstein-Barr virus genome. Thesis.
- Gonzy-Treboul, G., Karmazyn-Campelli, C., and Stragier, P. (1992). Developmental regulation of transcription of the Bacillus subtilis ftsAZ operon. *J. Mol. Biol.* 224, 967–979. doi:10.1016/0022-2836(92)90463-T.
- Hegyi, A., Szabó, M., Olasz, F., and Kiss, J. (2017). Identification of oriT and a recombination hot spot in the IncA/C plasmid backbone. *Sci. Rep.* 7, 10595. doi:10.1038/s41598-017-11097-0.
- Herrero, M., De Lorenzo, V., and Timmis, K. N. (1990). Transposon vectors containing non-antibiotic resistance selection markers for cloning and stable chromosomal insertion of foreign genes in gram-negative bacteria. *J. Bacteriol.* 172, 6557–6567. doi:10.1128/jb.172.11.6557-6567.1990.
- Kiss, J., Nagy, B., and Olasz, F. (2012). Stability, entrapment and variant formation of Salmonella genomic island 1. *PLoS One* 7, e32497. doi:10.1371/journal.pone.0032497.
- Kiss, J., and Olasz, F. (1999). Formation and transposition of the covalently closed IS 30 circle : the relation between tandem dimers and monomeric circles. *Mol. Microbiol.* 34, 37–52.
- Kiss, J., Papp, P. P. P., Szabó, M., Farkas, T., Murányi, G., Szakállas, E., et al. (2015). The master regulator of IncA/C plasmids is recognized by the Salmonella Genomic island SGI1 as a signal for excision and conjugal transfer. *Nucleic Acids Res.* 43, 8735–8745. doi:10.1093/nar/gkv758.
- Kolisnychenko, V., Plunkett, G., Herring, C. D., Fehér, T., Pósfai, J., Blattner, F. R., et al. (2002). Engineering a reduced Escherichia coli genome. *Genome Res.* 12, 640–7. doi:10.1101/gr.217202.
- Prentki, P., and Krisch, H. M. (1984). In vitro insertional mutagenesis with a selectable DNA fragment. *Gene* 29, 303–313. doi:10.1016/0378-1119(84)90059-3.
- Rose, R. E. (1988). The nucleotide sequence of pACYC184. *Nucleic Acids Res.* 16, 355. doi:10.1093/nar/16.1.356.
- Sambrook, J., Fritsch, E. F., and Maniatis, T. (1989). *Molecular Cloning: A Laboratory Manual*. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY.
- Short, J. M., Fernandez, J. M., Sorge, J. A., and Huse, W. D. (1988). Lambda ZAP: A bacteriophage lambda expression vector with in vivo excision properties. *Nucleic Acids Res.* 16, 7583–7600. doi:10.1093/nar/16.15.7583.
- Simon, R., Priefer, U., and Pühler, A. (1983). A Broad Host Range Mobilization System for In Vivo Genetic Engineering: Transposon Mutagenesis in Gram Negative Bacteria. *Bio/Technology* 1, 784–791. doi:10.1038/nbt1183-784.
- Szabó, M., Nagy, T., Wilk, T., Farkas, T., Hegyi, A., Olasz, F., et al. (2016). Characterization of Two Multidrug-Resistant IncA/C Plasmids from the 1960s by Using the MinION Sequencer Device. *Antimicrob. Agents Chemother.* 60, 6780–6786. doi:10.1128/AAC.01121-16.