Table S2. Primers used for species confirmation, multilocus sequence typing (MLST), and PCR typing of virulence genes.

Gene/Target	Primer	Sequence (5‘ to 3‘)	Amplicon size (bp)	T_a (°C)	References
Species confirmation, characterization and subtyping					
toxR	UtoxF	GASTTTGTTTGGCGYGARCAAGGTT	640	59	Bauer et al. (2007)
	VctoxR	GGTTAGCAACGATGCGTAAG			
ctxA	ctxA1	CTCAGACGGGATTTGTTAGGCACG	301		Shirai et al. (1991);
	ctxA2	TCTATCTCTGTAGCCCCTATTACG			Chatterjee et al. (2009)
rfb O1 cluster	O1F	GTTTCACTGAACAGATGGG	192		Hoshino et al. (1998);
	O1R	GGTCATCTGTAAGTACAAC			Mantri et al. (2006);
rfb O139 cluster	O139F	AGCCTCTTTATTACGGGTGG	449		Shuan et al. (2009)
	O139R	GTCAAACCCGATCGTAAAGG			
Multilocus Sequence Typing (MLST)					
adk	V-chol-adk F	CATCATTCTTCTCGGTGCTC	592	59	Octavia et al. (2013)
	V-chol-adk R	AGTGCCGTCAAACTTCAGGTA			
gyrB	V-chol-gyrB F	GTACGTTTCTGGCCTAGTGC	749	58	Octavia et al. (2013)
	V-chol-gyrB R	GGGTCTTTTTCCTGACAATC			
mdh	V-chol-mdh F	ATGAAAGTCGCTGTTATTGG			Octavia et al. (2013)
	V-chol-mdh R1[a]	GCCGCTTGGCCCATAGAAAG	692	62	
	V-chol-mdh R2	TAGCTTGATAGGTTGGG	810	51	
metE	V-chol-metE F	CGGGTGACTTTGCTTGGT	827	56	Octavia et al. (2013)
	V-chol-metE R	CAGATCGACTGGGCTGTG			
pntA	V-chol-pntA F1[a]	CTTTGATGGAAAAACTCTCA	740	52	Octavia et al. (2013)
	V-chol-pntA F2	GGCCAGCCCAAAATCCT	758	52	
	V-chol-pntA R	GATATTGCCGTCTTTTTCTT			
purM	V-chol-purM F	GGTGTCGATATTGATGCAGG	734	58	Octavia et al. (2013)
	V-chol-purM R	GGAATGTTTTCCCAGAAGCC			
pyrC	V-chol-pyrC F	ATCATGCCTAACACGGTTCC	726	57	Octavia et al. (2013)
	V-chol-pyrC R	TTCAAACACTTCGGCATA			
PCR typing of virulence genes					
chxA	VC-chxA-F	TGTGTGATGATGCTTCTGG	2000	52	Awasthi et al. (2013)
	VC-chxA-R	TTATTTCAGTTCATCTTTTCGC			
	VC-Cholix-fo	GCAACAACACTGAAAACGAG	397	56	This study
	VC-Cholix-re	TCCTCATCAATCGCCAATTC			
$hlyA^{CL/ET}$	hlyA-489F	GGCAAACAGCGAAACAAATACC	727/738	60	Rivera et al. (2001);
$hlyA^{ET}$	hlyA-744F	GAGCCGGCATTCATCTGAAT	481		Singh et al. (2001)
$hlyA^{CL/ET}$	hlyA-1184R	CTCAGCGGGCTAATACGGTTTA			
mshA	VC0409-F	ATTCTCGGTATCTTGGCCGTC	459	62	Rahman et al. (2008)
	VC0409-R	ACAAGCAGTTCCAGCAACCC			

Table continued

Gene/Target	Primer	Sequence (5‘ to 3‘)	Amplicon size (bp)	T_a (°C)	References
PCR typing of virulence genes					
ompU	ompU-F	ACGCTGACGGAATCAACCAAAG	869	60	Rivera et al. (2001);
	ompU-R	GCGGAAGTTTGGCTTGAAGTAG			Singh et al. (2001)
rstR^{Calc}	VC-rstR-calc-F	CCAGCATTTCTGTTTCTTTG	104	56	Rahman et al. (2008)
	VC-rstR-calc-R	GGCAACAAAGCACATTAAAG			
rstR^{CL}	VC-rstR-class-F	CTCATCAGCAAAGCCTCCATC	241	62	Rahman et al. (2008)
	VC-rstR-class-R	TAGCAAATGGTATCGGCGTTG			
rstR^{ET}	VC1455-F	AGCCAACCAAAGAAAGGCAAT	186	62	Rahman et al. (2008)
	VC1455-R	TCATCTGTGGCCCATCTTCC			
rtxA[b]	VC1451-F	GATTCTTCCGTTCAAGCTCCG	2571	63	Schirmeister et al. (2014)
	VC1451-R	TGGTTCAGGCTGTTGCACAC			
rtxC	VC1450-F	TGCAAATCTCACATTAGCGCA	430	63	Schirmeister et al. (2014)
	VC1450-R	CCACTGCACCTTTCGGATACA			
tcpA$^{CL/ET}$	tcpA-F_Class-ET	CACGATAAGAAAACCGGTCAAGAG		60	Mantri et al. (2006)
tcpA^{CL}	tcpA-R_class	TTACCAAATGCAACGCCGAATG	620		
tcpA^{ET}	tcpA-R_ET	AATCATGAGTTCAGCTTCCCGC	823		
TTSS *vcsC2*	TTSS_vcsC2-A	CGTTACCGATGCTATGGGT	535	60	Chatterjee et al. (2009)
	TTSS_vcsC2-B	AGAAGTCGGTTGTTTCGGTAA			
TTSS *vcsN2*	TTSS_vcsN2-A	CAGTTGAGCCAATTCCATT	484	55	Chatterjee et al. (2009)
	TTSS_vcsN2-B	GACCAAACGAGATAATG			
TTSS *vcsV2*	TTSS_vcsV2-A	TTTGGCTCACTTGATGGG	742	55	Chatterjee et al. (2009);
	TTSS_vcsV2-B	GCCACATCATTGCTTGCT			Dziejman et al. (2005)
TTSS *vspD*	TTSS_vspD-A	AACTCGAAGAGCAGAAAAAAGC	422	55	Chatterjee et al. (2009);
	TTSS_vspD-B	CTTCCCGCTTTTGATGAAATG			Dziejman et al. (2005)
VSP-1	Vch-VspI-fo	GCTCTCGCCAGCAAGGAGCTG	1700	60	Rahman et al. (2008)
	Vch-VspI-re	CCGTCGAAGTGAACGGCGAAC			
VSP-2	Vch-VspII-fo	TGCCCATTCCGCTAAGTGTTC	800	60	Rahman et al. (2008)
	Vch-VspII-re	GCAAAAGCACTGCGTAAACTG			

Calc, Calcutta; CL, Classical; ET, El Tor; T_a, annealing temperature.

[a] alternative primer

[b] VC1451 of *Vibrio cholerae* O1 biovar El Tor str. N16961.

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