

# Effect of long-term fertilization in soil and soilless culture of tomatoes on the microbial and fungal community structure

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18 Supplementary table 1: Overview of the chemical composition of soilplant and soilanimal before the start of the experiment (8/1/2015) and

19 throughout the whole experimental period. n=1. As bulk density 1.25 t ha<sup>-1</sup> (Vlaamse zandstreek; [Arthur et al. [58]]) was chosen for the 0.3

20 *m top soil layer.* 

Treatment Date analysis		NO3-N kg ha <sup>-1</sup>	NH4-N kg ha <sup>-1</sup>	(N-mineral)
SOILANIMAL	8/01/2015	243	< 4	247
SOILPLANT	8/01/2015	196	< 4	200
SOILANIMAL	3/2015	68	<4	72
SOILPLANT	3/2015	97	9	106
SOILANIMAL	5/2015	25	<4	29
SOILPLANT	5/2015	6	7	13
SOILANIMAL	6/2015	18	<4	22
SOILPLANT	6/2015	42	8	50
SOILANIMAL	27/7/2015	16	7	23
SOILPLANT	27/7/2015	17	18	35
SOILANIMAL	4/11/2015	10	<4	14
SOILPLANT	4/11/2015	9	<4	13

Fertilizer	Composition (%)								
	Ntotal	NO <sub>3</sub> <sup>-</sup> -N	NH4 <sup>+</sup> -N	P2O5	K <sub>2</sub> O	MgO	CaO	Cl	SO3
Blood meal*	14	-	-	0	0	-	-	-	-
Patentkali*	-	-	-	-	30	10	-	-	42
Magnesium sulphate*	-	-	-	-	-	16	-	-	32
Malt sprouts*	3	-	-	0	0	-	-	-	-
Antys MgS**	0	-	0	0	0	9	0	0	18
Biosyr**	9.35	-	0.53	3.96	9.08	0.11	3.98	0.68	10.9
Nutrikali**	2.4	-	0.1	0.3	4.9	0.1	0.1	1.1	1.1
SP**	0	-	0	0.001	58.6	0	0	0	0
Calcium chloride 33 %*	0	-	0	0	0	0	16.5	21	0
Calsal vlb 51%*	8.7		-	-	-	-	8.7	-	-
Amnitra vlb 51%*	18	9	9	-	-	-	-	-	-
Potassium nitrate*	13	13	-	-	45	-	-	-	-
Magnesium sulphate*	-	-	-	-	-	16	-	-	32
Monopotassium phosphate*	-	-	-	52	34	-	-	-	-
Potassium sulphate *	-	-	-	-	50	-	-	-	45

Supplementary table 2: Overview of the chemical composition of the different fertilizers used. "-" means that the elements was not analyzed
 or specified. "\*" means according to the specifications of the supplier. "\*\*" means that the chemical composition was actually analyzed.



25 Supplementary table 3: Overview of the fertilizers used and the total amount used for the different

26 treatments. GBOF = soilless culture system with organic growing medium and organic fertilizer.

27 GBFISH = soilless culture system with organic growing medium and fish. SOILANIMAL= organic

- 28 soil with animal-derived material as fertilizer and SOILPLANT= organic soil with plant-derived
- 29 material as fertilizer

Treatment	Type of fertilizer used	Total amount of fertilizer used (g m <sup>-2</sup> )	
	Calsal vlb 51% (Ca(NO <sub>3</sub> ) <sub>2</sub>	1855	
GBFISH	Amnitra vlb 51% (NH4NO3)	557	
	Potassium nitrate (KNO <sub>3</sub> )	959	
	Magnesium sulphate (MgSO <sub>4</sub> )	279	
	Monopotassium phosphate (KH <sub>2</sub> PO <sub>4</sub> )	368	
	Potassium sulphate (K <sub>2</sub> SO <sub>4</sub> )	288	
	Antys MgS	827	
	Biosyr	1292	
CDOE	Nutrikali	1870	
GBOF	SP	404	
	CaCl <sub>2</sub>	921	
	Libremix	49	
SOILANIMAL	Blood meal	180	
	Patentkali	163	
	Malt sprouts	1001	
SOILPLANT	Patentkali	163	

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## systems

Supplementary table 4: Overview of the chemical composition of the four different organic fertilizers (Nutrikali. ANTYS MgS. Biosyr and SP). "-": means that this element was not determined in the 

fertilizer

	Nutrikali	ANTYS MgS	Biosyr	SP
Total Nitrogen (%)	$2.35\pm0.1\ 2$	-	$9.35\pm0.23$	-
Organic nitrogen (%)	$2.25\pm0.11$	-	$8.81 \pm 0.22$	-
NH4 <sup>+</sup> -N (%)	$0.056\pm0.003$	-	$0.532\pm0.027$	-
NO3 <sup>-</sup> -N (%)	$0.042\pm0.002$	-	$0.012\pm0.001$	-
P <sub>2</sub> O <sub>5</sub> (%)	$0.257\pm0.013$	-	$3.96\pm0.20$	$0.0007 \pm 0.0001$
K <sub>2</sub> O (%)	$4.93\pm0.25$	-	$9.08\pm0.23$	$58.6 \pm 1.5$
CaO (%)	$60.113\pm0.006$	-	$3.98\pm0.20$	-
MgO (%)	$0.121 \pm 0.006$	$0.725\pm0.036$	$0.110\pm0.005$	-
$SO_3^{2-}$ (%)	$1.11\pm0.06$	$1.28\pm0.06$	$10.9\pm0.3$	-
Na <sub>2</sub> O (%)	$1.97\pm0.10$	-	$0.836\pm0.042$	-
Cl (%)	$0.839\pm0.042$	-	$0.676\pm0.034$	-
Organic matter (%)	$0.839\pm0.042$	-	$57.4 \pm 1.4$	-



46 Supplementary table 5: Correlations between microbial community composition and chemical characteristics in four different tomato cultivating systems across time points. indicated by the Multiple Factor Analysis. Dimensions of the MFA can be described by the categorical 47 48 variables included in the analysis. For each categorical variable (growing medium and time point). a one-way analysis of variance was 49 performed with the coordinates of the samples on the axis. explained by the time point or growing medium type. Then, for each level of the category (i.e. time point 1. time point 2 or time point 3 or growing medium GB). a Hotelling  $T^2$ -test was used to compare the average of the 50 51 category with the general average (using the constraint P i  $\alpha i = 0$ .  $\alpha i = 0$ ). For instance, the coordinates of the relative abundance of family "x" at GB at time point 1 were compared with the average coordinates of the relative abundance of family "x" in GB. The P value associated 52 53 to this test is transformed to a normal quantile to assess whether the mean of the category is significantly less or greater than 0. Negative 54 values indicate negative correlations.

Dimension	Variance	Descriptor	Estimate (R <sup>2</sup> )	P value	Taxon and or chemical characteristics	Correlation	P value
					Κ	0.9505387	< 0.0001
					Protozoa	0.882805	< 0.0001
					EC	0.8713766	< 0.0001
		GBOF	1.5492	0.00261	Fungi.18.3	0.7156968	1E-07
		GBFISH	1.451668	0.015256	Na	0.7015117	3E-07
					Total	0.6796157	0.000001
					Ca	0.6779943	1.1E-06
					Р	0.6499159	4.3E-06
DIM 1					NO <sub>3</sub> —N	0.6348602	8.3E-06
					Fungi.18.2	0.5960548	3.92E-05
					Cl	0.548466	0.000205
					Mg	0.4452108	0.003536
					SO4 <sup>2-</sup>	0.4374311	0.004237
					Actinomycetes.	-0.3618954	0.020071
					pH(H2O)	-0.6273887	1.13E-05
					Bacteria Fungi18.2 ratio.	-0.6355404	8.00E-06
					Gram-positive bacteria	-0.7987867	< 0.0001
DIM 2				AMF	0.729531	1E-07	
				Gram-negative bacteria	0.7288208	1E-07	

## Microbial diversity of soil and soilless culture systems

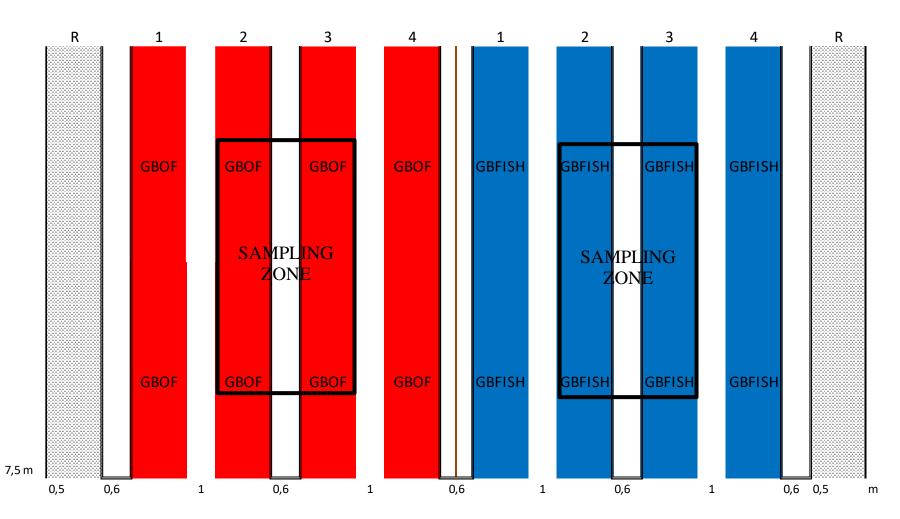
	-					
				Bacteria Fungi ratio.18.2.	0.670518	1.6E-06
	tpt4	1.178248	0.014011	Ca	0.5255753	0.000418
	SOILPLANT	0.8708145	0.016697	SO4 <sup>2-</sup>	0.4707461	0.001895
	SOILANIMAL	0.7376762	0.103984	Р	0.4189329	0.006407
				Gram-positive bacteria	0.4164783	0.006757
				pH(H2O)	0.4021037	0.009157
				Na	0.3751969	0.015642
				Cl	0.3520146	0.024004
				Fungi.18.3	-0.3528578	0.023645
				NH4 <sup>+</sup> -N	-0.517281	5.34E-04
				Actinomycetes	-0.5900243	4.91E-05
				Fungi.18.2	-0.6236232	1.33E-05
				Fungi.18.1	-0.8010289	< 0.0001
DIM 3				NH4 <sup>+</sup> -N	0.675832	1.2E-06
				Total	0.6403078	6.5E-06
	GBOF	1.27019	5.96E-05	Cl	0.5425307	0.000248
				pH(H2O)	0.4863632	0.001263
				Mg	0.3706436	0.017054
				Р	0.3569491	0.021966
				Fungi.18.2	-0.446345	0.003443
				Fungi.18.3	-0.49035	1.14E-03
				NO <sub>3</sub> -N	-0.722229	1.00E-07
				Actinomycetes	0.5783302	7.49E-05
DIM 4	SOILANIMAL	-0.6524775	0.005355	Gram-negative bacteria	0.5103223	0.000653
				SO4 <sup>2-</sup>	0.4863936	0.001262
				NH4 <sup>+</sup> -N	0.4423815	0.003778
	tpt3	0.4160831	0.051796	Mg	0.3565815	0.022113
DIM 5				Gram-negative bacteria	0.3443766	0.027467
	_			Cl	-0.313677	0.045812

## Microbial diversity of soil and soilless culture systems

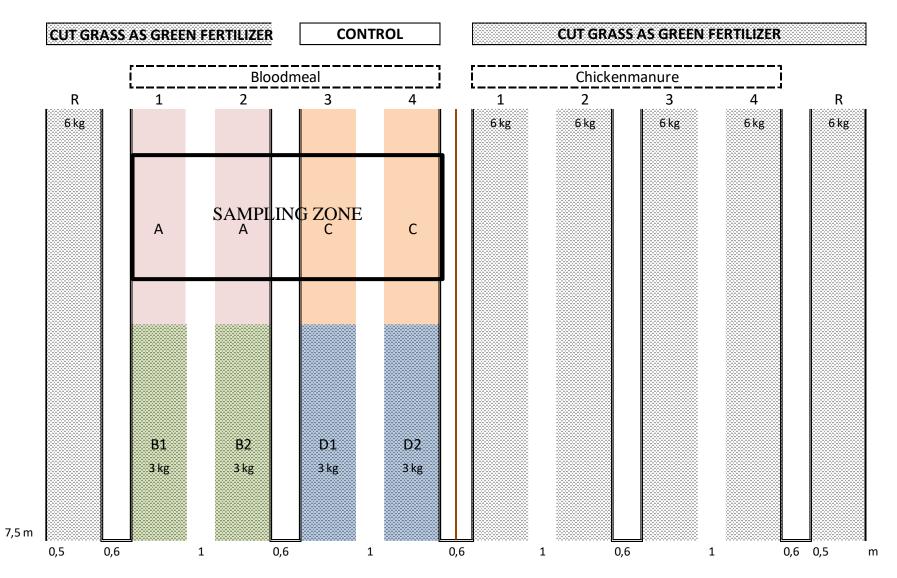
Na	-0.320351	0.041154
Fungi.18.1	-0.347512	0.025998
 Р	-0.39069	0.011552

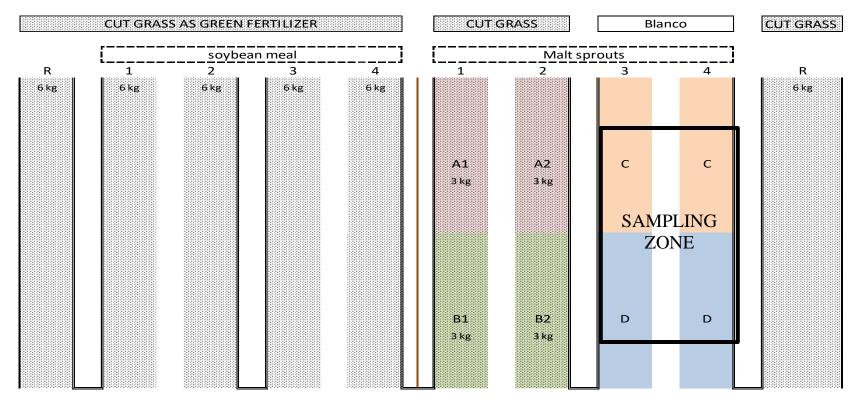


#### 9a SOILLESS CULTURE



### 9b SOILANIMAL





#### **9c SOILPLANT**

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60 Supplementary figure 1: Overview of the experimental set-up. The glass house was divided into three compartments (S91 = 9a GBOF and

61 *GBFISH.* S92=9b soilanimal and S93=9c soilplant) with a surface of 80 m<sup>2</sup>. S91 was subdivided into two part. i.e. red color = GBOF and

62 blue color = GBFISH; S92 was organic soil with animal (blood meal) derived material and S92 was the organic soil with plant-derived

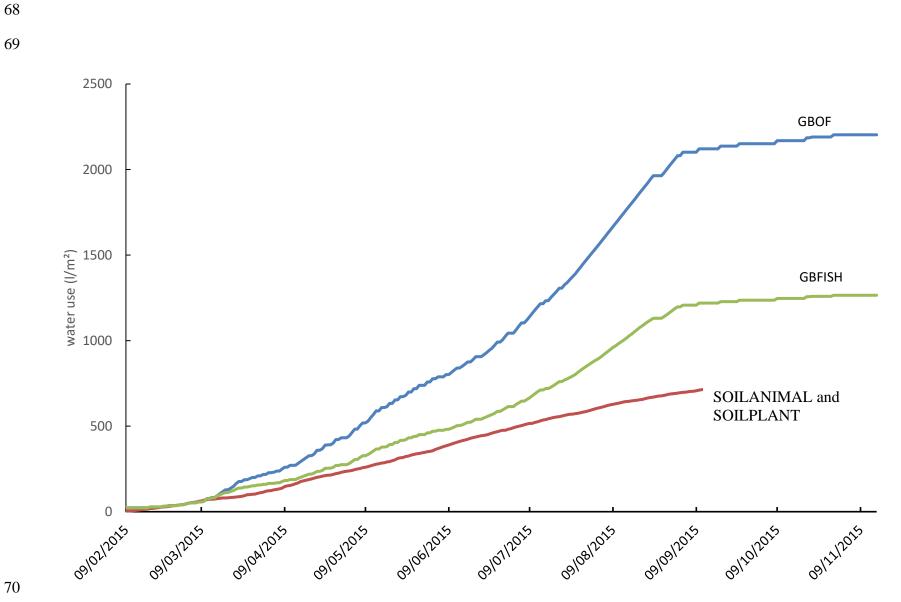
63 material (malt sprouts). In S92 and S93 the previous cultures were tomato in 2014. pepper in 2013 and cucumber in 2012. R= outer rows.

64 Plant density was the same for all the treatment 2.65 plants m<sup>-2</sup>. Samples were taken in the sampling zone with 40 plants per sampling zone.

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71 Supplementary figure 2: Overview of the cumulative water dosage (L per m<sup>2</sup>) for the different tomato cultivating systems.