

## Supporting information

# **Structural and functional characterization of a unique AP endonuclease from *Deinococcus radiodurans***

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**Supplementary Table1. Strains and plasmids used in this study**

Strain and plasmid	Description	Reference and source
<b>strains</b>		
<i>Deinococcus radiodurans</i>		
R1	ATCC 13939	Laboratory Stock
$\Delta xth$	R1 but <i>xth::kan</i>	This study
$\Delta xth/pk-xth$	$\Delta xth$ but pRADK:: <i>xth</i>	This study
<i>Escherichia coli</i>		
DH5 $\alpha$	<i>E. coli</i> cloning strain	TransGen
BL21(DE3)	<i>E. coli</i> expression strain	TransGen
<b>plasmids</b>		
pET28a	T7 promoter, T7 transcription start, His•Tag coding sequence, T7•Tag coding sequence, Multiple cloning sites(BamH I -Xho I), T7 terminator, <i>lacI</i> coding sequence, pBR322 origin, <i>Kanr</i> , fl origin, 6His-tag coding sequence	Novagen
pET28S	pET28 plasmid modified with a Strep-tag sequence (WSHPQFEK)	Laboratory Stock
pET28aXth	pET28a containing wild type <i>xth</i> gene	This study
pET28aXth $\Delta$ 22	pET28a containing wild type N-terminus domain of <i>xth</i> gene	This study
pET28S-Xth	pET28S containing wild type <i>xth</i> gene	
pET28aD177N	pET28a containing <i>xth</i> directed site D155N mutation gene	This study
pET28a S143A/N234A/R235A	pET28a containing <i>xth</i> directed site S121A/N212A/R213A triple mutation gene	This study
pET28aG198H	pET28a containing <i>xth</i> directed site G176H mutation gene	This study
pET28aG198A	pET28a containing <i>xth</i> directed site G176A mutation gene	This study
pET28aAPE	pET28a containing <i>ape</i> gene	This study
pET28aPolA-C	pET28a containing C-terminus domain of <i>drpolA</i> gene	This study
pET28aPolX	pET28a containing <i>polx</i> gene	Laboratory Stock
PRADK	<i>D. radiodurans</i> shuttle vector	Laboratory Stock
pk-xth	pRADK:: <i>xth</i>	This study

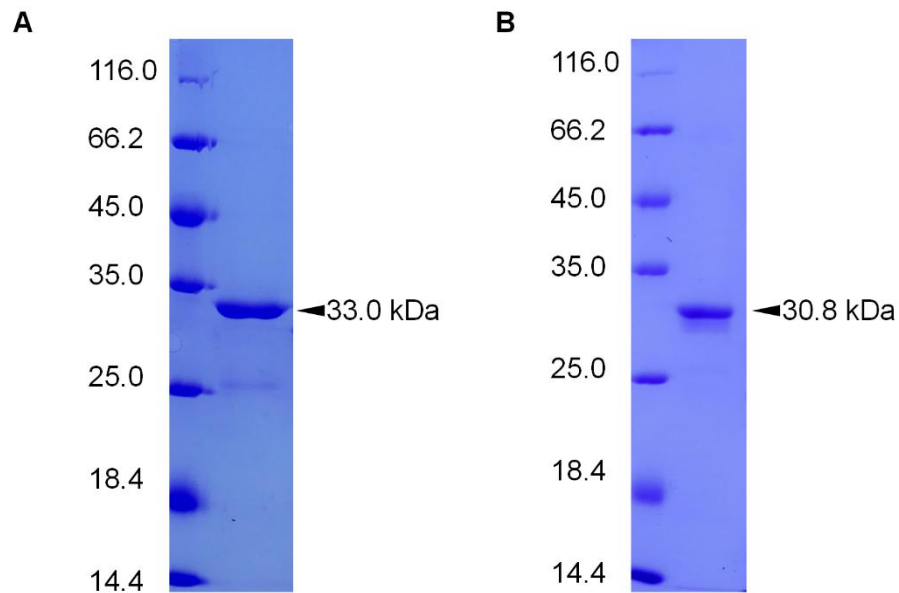
**Supplementary Table2.** Primes and oligonucleotides used in this study

Prime	Sequence(5'-3')	
construction and complement of drxth mutant		
xth p1	CCCCCGAACTCGACGTG	
xth p2(HindIII)	CCCAAGCTTCGGCGCCGACCATAGC	
xth p3(BamHI)	CGGGATCCACCCTACCTTCTCCCCAGTACC	
xth p4	TCTGCGCTGTCCTCGGTG	
xth p5	TGCTTCTGCAAGAAGTCCGC	
xth p6	GATTCCAGCTCCACCCACCC	
Expression of proteins		
Xth F (NdeI)	GGAATTCCATATGTTGAGCCTCCTTGCCCCA	
Xth R (BamHI)	CGGGATCCTCATTAGATTCCAGCTCCACC	
XthΔ22 (NdeI)	GGAATTCCATATGATGTCTGCCCCCGCCG	
XthΔ22 (BamHI)	CGGGATCCTCATTAGATTCCAGCTCCACC	
APE1F(NdeI)	GGAATTCCATATGATGCCGAAGCGTGGGAAA	
APE1R(BamHI)	CGGGATCCTCACAGTGCTAGGTATAGGGTGATAGG	
PolA-C(NdeI)	GGAATTCCATATGATGGGGCTGAACGGGGCCA	
PolA-C (BamHI)	CGGGATCCTCACTTCGTGTCAAACCAGTTTCG	
Site-directed mutagenesis		
xth G198H(F)	GCTCGTGGGGCAGGAAATGGCTGTTTTTCTGGTTGC	
xth G198H(R)	GCAACCAGAAAAACAGCCATTTCTGCCCCACGAGC	
Xth D177N(F)	GGCGATGTTGTAGTTGCCGCCGATGACGA	
Xth D177N(R)	TCGTCATCGGCGGCAACTACAACATCGCC	
XTH G198A(F)	CGTGGGGCAGGAAAGCGCTGTTTTTCTGG	
XTH G198A(R)	CCAGAAAAACAGCGCTTTCTGCCCCACG	
Xth S143A(F)	CGCCTCGCCGGCGCTGCCGCTCG	
Xth S143A(R)	CGAGCGGCAGCGCCGGCGAGGCG	
Xth N234R235A(F)	GGCGTAGGCATTGGCGGCGGCGCTCCACCAGGTGTAC	
Xth N234R235A(R)	GTACACCTGGTGGAGCGCCGCCCAATGCCTACGCC	
Oligonucleotides for enzyme activity assay		
Type of substrate	Sequence	label
THF.T	5'*GCTATGGACTAAFAATGACTGCGTG 3'	F(THF)
	3'CGATCCTGATTTTTACTGACGCAC5'	*(FAM)
Exo40.T	5'*ATGACAACATAAGCAACACC3' 5'GATAGAACGACCGCCAGTG3'	*(FAM)
	3'TACTGTTGATTTCTGTTGTGGTCTATCTTGCTGGCGGTCAC5'	
Exo40 <sup>P</sup> .T	5'*ATGACAACATAAGCAACACC <sup>P</sup> 3' 5'GATAGAACGACCGCCAGTG3'	*(FAM)
	3'TACTGTTGATTTCTGTTGTGGTCTATCTTGCTGGCGGTCAC5'	
Exo40 <sup>THF</sup> .T	5'*ATGACAACATAAGCAACACCF 5'GATAGAACGACCGCCAGTG3'	F(THF)
	3'TACTGTTGATTTCTGTTGTGGTCTATCTTGCTGGCGGTCAC5'	*(FAM)
αdA	5'*TGACTGCATAXGCATGTAGACGATGTGCAT3'	X(αdA)
	3'ACTGACGTATACGTACATCTGCTACACGT5'	*(FAM)

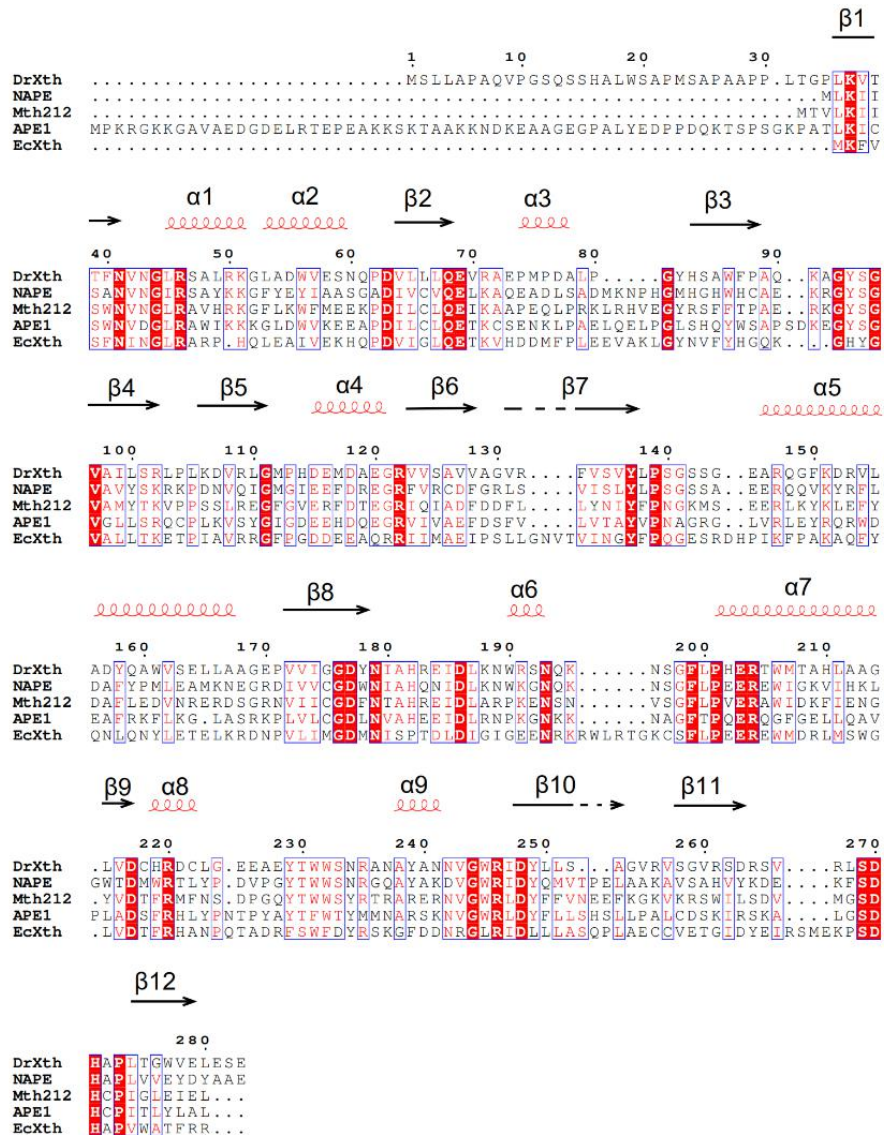
**Supplementary Table3.** Data collection, phasing and refinement statistics

	DrXth
<b>Data collection</b>	
Space group	$P2_12_12_1$
Cell dimensions	
<i>a</i> , <i>b</i> , <i>c</i> (Å)	57.42
	58.86
	74.28
Wavelength (Å)	0.9792
Resolution (Å)	30.0-1.50
<i>R</i> <sub>sym</sub> (%)	6.6 (47.3)
<i>I</i> / <i>σI</i>	14.6 (4.3)
Completeness (%)	98.2
	(91.0)
Redundancy	5.1 (3.4)
<b>Refinement</b>	
Resolution (Å)	30.0-1.50
No. reflections	40705
<i>R</i> <sub>work</sub> / <i>R</i> <sub>free</sub>	18.6/20.4
No. atoms	
Protein	1941
Water	243
B-factors	
Protein	19.3
Water	32.0
R.m.s deviations	
Bond lengths (Å)	0.005
Bond angles (°)	0.865

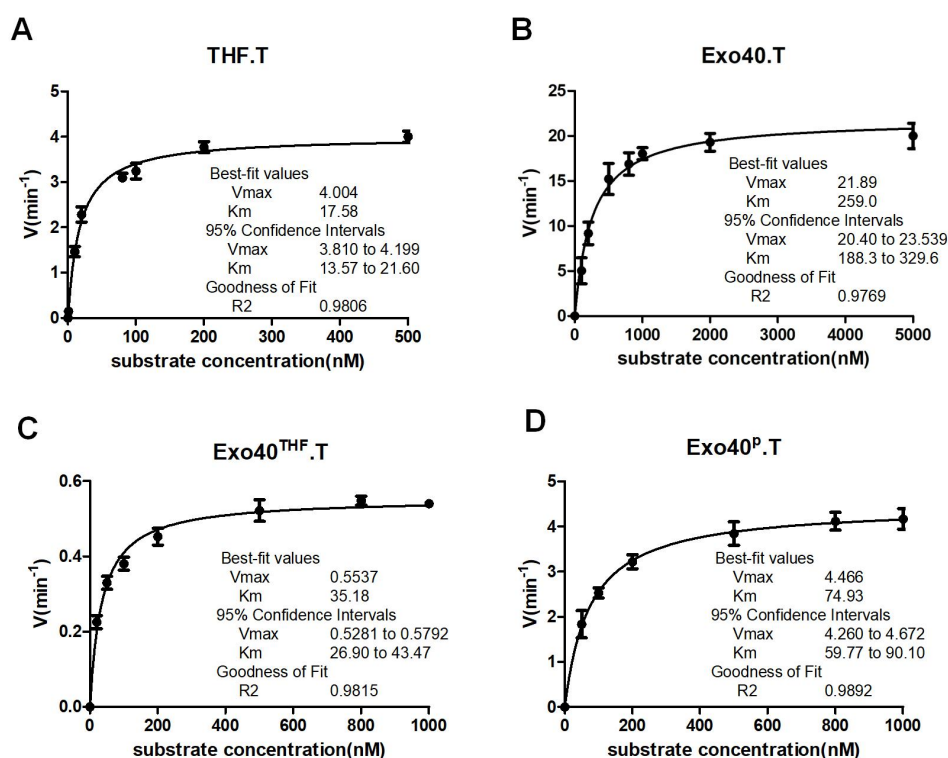
\*Highest resolution shell is shown in parenthesis.



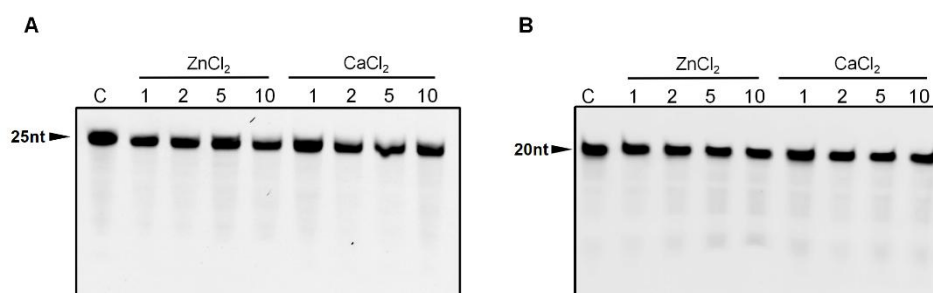
**Supplementary Figure1.** SDS-PAGE analysis of the purified full-length type and N-terminal truncated DrXth. (A) Molecular weight of full-length type DrXth protein with His-tag is 33.0 kDa. (B) Molecular weight of N-terminal truncated DrXth protein (N $\Delta$ 22DrXth) with His-tag is 30.8 kDa.



**Supplementary Figure2.** Sequence alignments among DrXth and other representative ExoIII family AP endonucleases. DrXth, *Deinococcus radiodurans*; NApe, *Neisseria meningitidis*; Mth212, *Methanobacterium thermoautotrophicum*; APE1, *Homo sapiens*; EcXth, *Escherichia coli*.

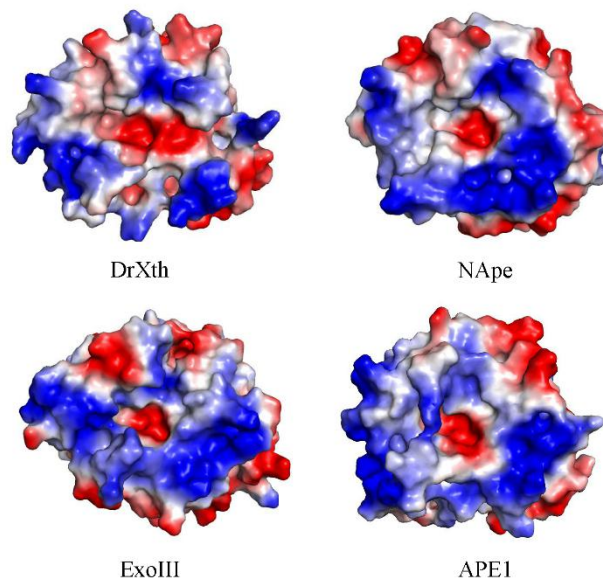


**Supplementary Figure3.** Kinetic of DrXth nuclease activity. (A) DrXth (1 nM) was incubated with increasing amounts of THF.T substrate (1, 10, 50, 80, 100, 200, 500 nM) at 37°C for 3 min. (B) DrXth (3 nM) was incubated with increasing amounts of Exo40.T substrate (200, 500, 800, 1000, 2000, 5000 nM) at 37°C for 3 min. (C) DrXth (3 nM) was incubated with increasing amounts of Exo40THF.T substrate (20, 50, 100, 200, 500, 800, 1000 nM) at 37°C for 3 min. (D) DrXth (3 nM) was incubated with increasing amounts of Exo40P.T substrate (20, 50, 100, 200, 500, 800, 1000 nM) at 37°C for 3 min. The data were fitted by the Michaelis–Menten equation in GraphPad Prism 5 in order to obtain the K<sub>cat</sub> and K<sub>m</sub>. Best-fit values, 95% confidence intervals and goodness of fit are shown.

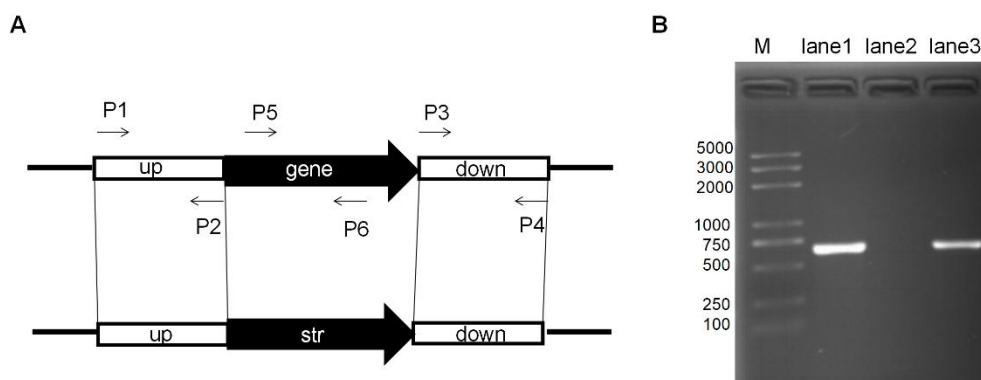


**Supplementary Figure4.** Analysis of ion effects on AP endonuclease activity. (A) 100 nM THF.T duplex was incubated with 2 nM DrXth in the presence of CaCl<sub>2</sub> or ZnCl<sub>2</sub> (1, 2, 5 or 10 mM) at 37°C for 5 min. (B) 100 nM Exo40.T 1nt gap duplex was incubated

with 10 nM DrXth in the presence of  $\text{CaCl}_2$  or  $\text{ZnCl}_2$  (1, 2, 5 or 10 mM) at 37°C for 5 min.

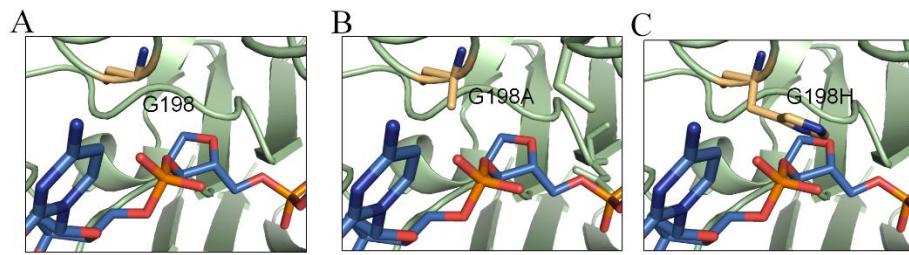


**Supplementary Figure5.** Structural comparison among DrXth, NApe, ExoIII and APE1. The structure shows the distribution of the electrostatic surface. Blue and red represent negative and positive charge potential at + and  $-70 \text{ kTe}^{-1}$  scale, respectively.

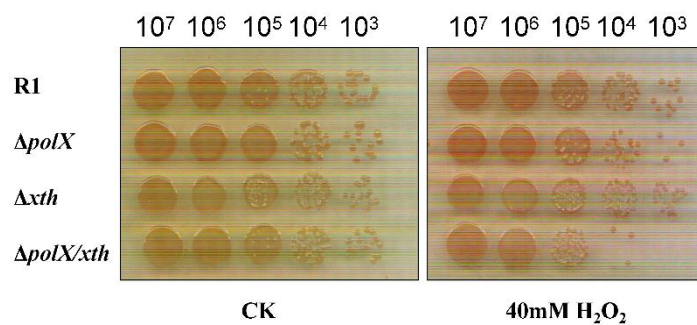


**Supplementary Figure6.** Deletion of *drxth* gene in *D.radiodurans* R1 strain (A) Scheme of gene mutation by homologous recombination that replaced the target ORFs with streptomycin-resistant fragment. P1, P2, P3, P4, P5 and P6 refer to the primer pairs (Supporting information Table2). (B) PCR analysis to confirm the mutation of *drxth*. An interior DNA fragment (652 bp) of the targeted gene was detected by amplification using primers P5 and P6. No products corresponding to the size of the fragment was observed from mutant (lane 2) but observed in the wild type (lane 1) and compensation type (lane 3), suggesting that the wild type alleles were completely replaced by streptomycin-resistance fragment in the mutant.

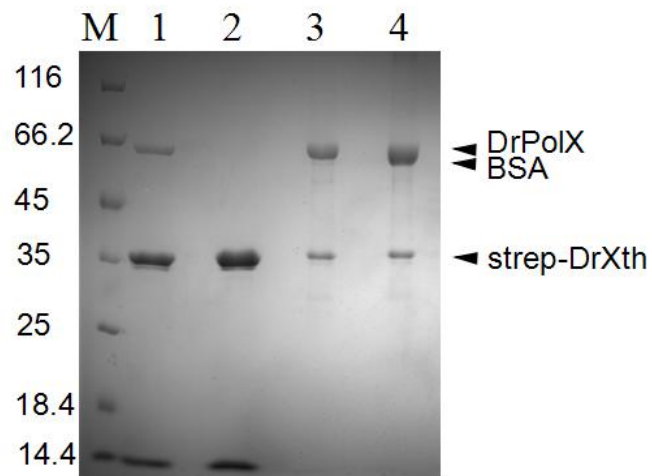




**Supplementary Figure7.** Predicted structure of Gly198 mutant DrXth. DNA from NApe-DNA complex were docked onto the DrXth by superposition between DrXth and NApe. (A) DrXth (B) Substitutue Ala for Gly198 (C) Substitute His for Gly198.



**Supplementary Figure8.** Phenotypes of R1 strain and mutant strain under H<sub>2</sub>O<sub>2</sub> stress. Growth and H<sub>2</sub>O<sub>2</sub> resistance features of wild type (R1), *drpolX* disruptant ( $\Delta polX$ ), *drxth* disruptant ( $\Delta xth$ ), *drpolX-drxth* double disruptant ( $\Delta polX/ xth$ ). Cells were incubated with H<sub>2</sub>O<sub>2</sub> (40 mM) for 30min and then the reaction were stopped by excess catalyase for 15 min. After treatment, the cells were serially diluted 1:10 and spotted on TGY agar plates, and then cultivated at 30°C for 3 days.



**Supplementary Figure9.** Interaction assay between His-Strep- DrXth and DrPolX. His-Strep-DrXth (N-terminal fused streptavidin tag) binding with strep-beads were rotating incubated with DrPolX and BSA at 4 °C for 3h. Lane1: His-Strep-DrXth and DrPolX were pulled-down by strep-beads. Lane2: His-Strep-DrXth and BSA (input control) were pulled by strep-beads. Lane3: His-Strep-DrXth and DrPolX. Lane4: His-Strep-DrXth and BSA (input control).