

Supplementary Material

Immobilization of the highly active UDP-glucose pyrophosphorylase from *Thermocrispum agreste* provides a highly efficient biocatalyst for the production of UDP-glucose

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The supplemental material comprises the following information; codon optimized gene encoding the employed biocatalyst as well as activity data regarding divalent cations, buffers, potential inhibitors, solvents and kinetic data describing soluble and immobilized enzyme preparations.

The *Ta*GalU gene (accession number: WP_028847555) was codon optimized (accession number: MT321102) and synthesized prior cloning.

>codon optimized gene plus restrictions sites for cloning (NdeI and NotI; underlined)

CAT ATG ACG GCG AGT GCC GAG CAT CAG ACC TTC ACC ACT GCA ATT GTG CCT GCA GCT GGC CTC GGT ACG CGC TTT CTG CCC ACC ACC AAA TCG GTG CCG AAG GAA TTG CTG CCT GTC GTG GAT ACA CCG GCG ATT GAG CTT GTG GCG GAC GAA GCG CGT CAA GCG GGC GCG GAA CGT CTG GTG ATC GTG ACG TCA CCA GCT AAA CAG AGC ATT GCT GCC TAC TTTCGC CCA GCT CCG GAA CTG GAA CGC TCG TTG GAA GAG AAG GGC AAA ACC GGA CAA CTC GCG AAA ATT CGC CGT GCA CCA GAA CTG CTT GAG GTA GAG GTT GCG ATC CAG GAA CAG GCT CTG GGG TTA GGT CAC GCA GTT GCC TGT GCC GAA CCT AAC CTG GGT CCG GAA GAT GAC GTA GTT GCC GTT CTG CTG CCG GAT GAT CTG GTC CTT CCA CAC GGC ATC CTG GAA CGC ATG GCG AAA GTT CGT GCC GAA CAT GGC GGC TCT GTG CTC TGC GCG TTT GAC ATC CCG AAA GAG GAA ATT AGC GCG TAT GGG GTT TTC GAT GTG AGC GAC ACG GAT GAT GCG GAC GTG AAA CGG GTG CAC GGT ATG GTC GAG AAA CCG CCT GCA GAA CAA GCA CCC TCC ACT GCC GCC GCA GGC CGT TAC TTG CTG GAT CGT GCG TTTATC TTC GAT GCG TTA CGT CGC ATT GAA CCC GGT GCG GGT GGA GAG CTG CAG CTG ACA GAT GCC GTC GCT TTG CTG ATT CAG GAA GGA CAT CCG GTA CAC GTC GTC GTA CAT CGC GGT GAT CGG CAT GAC CTC GGC AAT CCG GGT GGG TTC CTG CGC GCT GCA GTT GAC TTTGCA CTG CAG GAT CCG GAC TAT GGC CCG GAA TTA CGC GCC TGG TTA ACC GAT CGC ATT GCC CGT CCG TGA GCG GCC GC



Supplementary Figure 1. Relative specific *Ta*GalU activity depending on the applied magnesium chloride concentration. Reaction solution contained 2 mM UTP, 2 mM G1P, 0-100 mM MgCl₂, 50 mM Hepes, pH 7.0, 0.013 μ g mL⁻¹, 50 °C, 1 mL reaction volume. 100 % relative activity corresponds to 2111 U mg⁻¹. Means with standard deviations of triplicate measurements are shown.



Supplementary Figure 2. Different metals as cofactors tested in the *Ta*GalU reaction. 1 mL reaction solution contained 2 mM UTP, 30 mM G1P, 3 mM metal salt, 50 mM Hepes, pH 7.0 and 0.013 μ g mL⁻¹ *Ta*GalU. Reaction was carried out at 50 °C. 100 % corresponds to 1256 U mg⁻¹. Means with standard deviations of triplicate measurements are shown.



Supplementary Figure 3. Different buffers tested for the *Ta*GalU reaction. 1 mL of reaction mixture contains 2 mM UTP, 30 mM G1P, 3 mM MgCl₂, 50 mM buffer, pH 7.0 and 0.013 μ g mL⁻¹ *Ta*GalU. The reaction temperature was 50 °C. 100 % corresponds to 1258 U mg⁻¹. Means with standard deviations of triplicate measurements are shown.



Supplementary Figure 4. Effect of inhibitors or solvents to *Ta*GalU. The reaction mixture contained: 2 mM UTP, 30 mM G1P, 3 mM MgCl₂, 50 mM Hepes, pH 7.0, 1 mM and 50 mM EDTA, respectively or 1 M solvent/inhibitor, 0.013 μ g mL⁻¹ *Ta*GalU. Reaction was carried out at 50 °C in 200 μ L reaction volume. Means with standard deviations are shown. Mercapto: 2-mercaptoethanol, DMSO: dimethyl sulfoxide, ACN: acetonitrile, MeOH: methanol, EtOH: ethanol, PrOH: isopropanol, EDTA: ethylenediaminetetraacetic acid.

kinetic constant	Free		Immobilized		Immobilized and multiplied by factor	
	UTP	G1P	UTP	G1P	UTP	G1P
$K_{\rm m}[{ m mM}]$	0.15	0.12	0.40	0.79	0.40	0.79
<i>V</i> _{max} [U mg ⁻¹]	1698	1109	226	157	3480 ^a	2418 ^a
$k_{\mathrm{cat}}[\mathrm{s}^{\text{-1}}]$	914	597	121	84	1874 ^a	1302 ^a
<i>K</i> _I [mM]	3.9	-	3.9	-	3.9	-
$k_{ m cat}/K_{ m m}$ [$\mu { m M}^{-1} { m s}^{-1}$]	6.09	4.98	0.3	0.11	4.69 ^a	1.65 ^a

Supplementary Table 1. Kinetic constants of the free and immobilized TaGalU calculated by fits

^a: For data collection of the kinetics for the immobilized *Ta*GalU an old batch of enzyme with reduced specific activity was used. Therefore, we recalculated the values for V_{max} with the time factor 15.4. This factor emerged from the fact that the initial specific activity of the free enzyme was 1016 U mg⁻¹ directly after purification. After storage of the enzyme for 1.5 years the activity decreased to only 66.03 U mg⁻¹, which is a reduction by the factor 15.4. Therefore, these values are of theoretical nature.