

Supplemental table 2: commands for SNP caller workflows

NGM

- 1. Call SNPs
samtools pileup -cvf TAIR9_ref_file sorted.bam_file > output.pileup
- 2. Generate.emap file
SAM2NGM.pl output.pileup

SAMTOOLS

- 1. Prepare input file for SNP calling
samtools mpileup -C50 -Q30 -Bf TAIR9_ref_file sorted_bam_file | bcftools view -bvcg - > raw.bcf
- 2. Call SNPs
bcftools view raw.bcf | vcfutils.pl varFilter -Q0 > filtered.vcf

GATK

- 1. Add read groups (Picard tools)
AddOrReplaceReadGroups.jar I=sorted.bam_file O=s1.rg.bam RGLB=genome RGPL=ILLUMINA
RGPU=GATKv4 RGSM=sample_name VALIDATION_STRINGENCY=LENIENT
- 2. Mark duplicates (Picard tools)
MarkDuplicates.jar INPUT=s1.rg.bam OUTPUT=s2.dedup.bam ASSUME_SORTED=TRUE
VALIDATION_STRINGENCY=LENIENT METRICS_FILE=s2.dedup.metrics
- 3. Index (samtools)
samtools index s2.dedup.bam
- 4. Realign reads (create intervals first, then do IndelRealigner) (GATK)
GenomeAnalysisTK.jar -I s2.dedup.bam -R ref_file -T RealignerTargetCreator -o s3.intervals
GenomeAnalysisTK.jar -T IndelRealigner -I s2.dedup.bam -R ref_file -targetIntervals s3.intervals -o
s4.realn.bam
- 5. Unified genotyper (GATK)
GenomeAnalysisTK.jar -T UnifiedGenotyper -R ref_file -I s4.realn.bam -glm BOTH -o s5.UG1.vcf -mbq 30 -nt
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- 6. Base score recalibrator (GATK)
GenomeAnalysisTK.jar -T BaseRecalibrator -I s4.realn.bam -R ref_file -knownSites s5.UG1.vcf -o s6.recal
- 7. Print Reads (GATK)
GenomeAnalysisTK.jar -T PrintReads -R ref_file -I s4.realn.bam -BQSR s6.recal -o s7.recal.bam
- 8. Unified Genotype (GATK)
GenomeAnalysisTK.jar -T UnifiedGenotyper -R ref_file -I s7.recal.bam -glm BOTH -o s8.UG2.vcf -mbq 30
- 9. Base score recalibrator (GATK)
GenomeAnalysisTK.jar -T BaseRecalibrator -I s4.realn.bam -R ref_file -knownSites s8.UG1.vcf -o s9.recal
- 10. Print Reads (GATK)
GenomeAnalysisTK.jar -T PrintReads -R ref_file -I s4.realn.bam -BQSR s9.recal -o s10.recal.bam
- 11. Unified Genotyper (GATK)
GenomeAnalysisTK.jar -T UnifiedGenotyper -R ref_file -I s10.recal.bam -glm BOTH

SHOREMAP backcross

- 1. Preprocess the reference file
shore preprocess -f TAIR9_ref_file -i TAIR9 -W
- 2. Import reads
shore import -v Fastq -a genomic -x read1.fastq -y read2.fastq --rplot --disable-illumina-filter -k 50 -o
sample_name
- 3. Map reads using shore
shore mapflowcell -f sample_name -i TAIR9_ref_file.shore -v bwa -n 10% -g 7% -p -c 2 -b 250000 -M 2 -P
replace
- 4. Merge
shore merge -p -m sample_name -o sample_name/merge
- 5. Call SNPs
shore consensus -n sample_name -f TAIR9_ref_file.shore -o sample_name/consensus -i
sample_name/merge/map.list.gz -g 4 -a scoring_matrix_het.txt -v -r
- 6. Use SHOREmap.pl backcross (require TAIR9 chr sizes; require steps 1 to 5 on parent line for “--bg”
option)
SHOREmap.pl backcross --marker sample_name/consensus/ConsensusAnalysis/quality_variant.txt --out
shoremapout --chrsize TAIR9_chr_sizes.txt --bg
parent_name/consensus/ConsensusAnalysis/quality_variant.txt --marker-score 25 --marker-freq 95 --
marker-cov 8 --bg-freq 20