

Table S1. DNMTs from *F. graminearum* share identity with RID2 and DIM-2 from *Neurospora tetrasperma*. Protein sequences were obtained from Uniprot (<https://www.uniprot.org/>) under accession numbers Q8NJV8 (*NtRID*), G4UYB9 (*NtDIM-2*), I1RWH8 (FGSG_08648) and I1S1Y7 (FGSG_10766). The sequences were subjected to reciprocal protein-protein BLASTp searches on NCBI (<https://blast.ncbi.nlm.nih.gov/Blast.cgi>).

<i>F. graminearum</i> Strain	<i>Nt</i> RID (Q8NJV8) Identity (e-value)	<i>Nt</i> DIM-2 (G4UYB9) Identity (e-value)
FGSG_08648 (I1RWH8)	35.7% (2e-95)	23.6% (2e-7)
FGSG_10766 (I1S1Y7)	26% (8e-13)	48.3% (0.0)

Table S2. Whole genome bisulfite sequencing statistics for WT and $\Delta FgDim-2/\Delta FgRid$ strains sequenced using Illumina HiSeq X PE150. Sequencing reads were deposited under accession # PRJNA587083 and mapped to the reference genome (Accession # SPRZ000000000) using CLC-Genomics Workbench.

Sample	Sequencing Statistics	PN		WT NPN		PN		$\Delta FgDim-2/\Delta FgRid$ NPN				
		BR 1	BR 2	BR 1	BR 2	BR 3	BR 1	BR 2	BR 3	BR 1	BR 2	BR 3
Genome size	36,766,638											
%C+G	47.91											
Gene numbers	14190											
Method	PE ^b											
Conversion rate(%) ^a	99.5											
Total reads (Mbp)		68,974,006	65,251,418	66,751,428	62,125,272	59,077,414	60,359,444	66,775,820	47,251,660	70,047,640	70,366,392	70,048,580
Number of mapped reads (Mbp)		61,192,341	58,245,915	58,451,778	55,195,708	52,689,907	54,702,044	59,086,782	41,952,959	62,363,043	62,072,370	62,541,206
Read Length (bp) ^c		128.76	129.2	128.85	127.77	127.37	128.84	128.83	128.77	128.75	128.74	128.68
Mapping efficiency (%)		88.72	89.26	87.57	88.85	89.19	90.8	88.63	88.93	89.22	88.37	89.51
Coverage (%)		100	100	100	100	100	100	100	100	100	100	100
Mean sequencing depth per strand		212.05	202.87	201.42	187.21	177.66	189.85	203.7	144.65	213.41	212.74	214.51

BR: Biological replicates; PN: Preferred Nutrient conditions, and NPN: Non-Preferred nutrient conditions

^a Conversion Rate: the conversion rate of BS-seq for *F. graminearum* is calculated from an un-methylated lambda DNA added to the BS-seq library

^b PE: Paired End Reads

^c Post Trimming

Table S3: DNA methylation is present under all three cytosine contexts, CpG, CHG and CHH in WT and $\Delta FgDim-2/\Delta FgRid$ strains, under both 24 hrs PN and 6 hrs NPN environmental conditions. Methylation level remained consistent between strains and environmental conditions. Methylation level is defined as the proportion of methylation at any given site in a population.

	WT					$\Delta FgDim-2/\Delta FgRid$					
	PN		NPN			PN			NPN		
Cytosine Context	BR 1	BR 2	BR 1	BR 2	BR 3	BR 1	BR 2	BR 3	BR 1	BR 2	BR 3
CpG(%)	4.45	4.72	4.97	5.11	5.29	4.88	4.64	6.00	4.62	4.77	4.50
CHG(%)	4.45	4.71	4.94	5.14	5.38	4.89	4.65	6.00	4.63	7.72	4.53
CHH(%)	4.68	4.90	5.10	5.52	5.80	5.20	4.73	6.03	4.95	4.97	4.86

BR: Biological replicates; PN: Preferred nutrient conditions; NPN: non-preferred nutrient conditions.

Table S4. DNA methylation exists in WT and $\Delta FgDim-2/\Delta FgRid$ strains with minor differences between strains at the genome wide level. DNA methylation density was defined as the number of methylated cytosine as a percentage of the total cytosine. DNA methylation was predominantly identified in the asymmetrical CHH context in both strains.

	WT					$\Delta FgDim-2/\Delta FgRid$					
	PN		NPN			PN			NPN		
Cytosine Context	BR 1	BR 2	BR 1	BR 2	BR 3	BR 1	BR 2	BR 3	BR 1	BR 2	BR 3
CpG	0.41	0.44	0.6	0.35	0.36	0.72	0.37	0.42	0.29	0.43	0.4
CHG	0.37	0.39	0.52	0.32	0.32	0.6	0.33	0.36	0.27	0.38	0.37
CHH	1.3	1.39	1.71	1.07	1.09	1.94	1.16	1.21	0.95	1.33	1.26
Average	2.15		2.11			2.37			1.89		

BR: Biological replicates; PN: Preferred Nutrient conditions, and NPN: Non-Preferred nutrient conditions. Values represent methylation density (#5mC/#GenomicC*100).

Table S5. Primers Used in This Study

Primer Name	Sequence (5'-3')	
P1 - DIM2	gggtttaaugcagcctatcctcatgaagtga	KO
P2 - DIM2	ggacttaaugtcctatgttaatgaatatgcacc	
P3 - DIM2	ggcattaaugtcggaacagtgctcgcg	
P4 - DIM2	ggctttaauacccaacatcgtttc	
P1 - RID	gggtttaaugtggattagtgtgttgaggaaac	Compl
P2 - RID	ggacttaauccaaggtaggtagcaacgaatg	
P3 - RID	ggcattaautgattgaggcgaggagtatc	
P4 - RID	ggctttaauaggaaatgaaggagcccgtg	
P5 - DIM2	ggacttaaucgagaacattcttggttgg	
P6 - DIM2	gggtttaauctactcatttgaaacgctg	
P5 - RID	ggacttaaugtgaacatggatttctgatc	
P6 - RID	gggtttaauttagtcaattcgatc	
DIM2 Int. F	gtggtcgatctgagccttgc	Confirm
DIM2 Int. R	gttccgccattggatcac	
RID Int. F	caagacgaagcaaagctca	
RID Int. R	atgtacggatgcatgagtgt	
HYG F	agctgcgccgatggtttctacaa	
HYG R	gcgcgtctgctgtccatacaa	
Gen F	tcatcaatcccagccttttc	
Gen R	cagtcgatgaatccagaaaagc	
TRI6 ORF F	atgattacatggaggccg	
TRI6 ORF R	acacttatgtatccgcctatagtg	
gpdA F	gaagtggaaaggctggtgtg	qPCR
gpdA R	ataagggatgggaaggatgg	
FGSG_09530 F (<i>β tubulin</i>)	gttgatctccaagatccgtg	
FGSG_09530 R (<i>β tubulin</i>)	catgcaaatgctgtagagg	
FGSG_16627 F (<i>GAPDH</i>)	tgacttgactgttcgcctcgagaa	
FGSG_16627 R (<i>GAPDH</i>)	atggaggagttggtgttgccgtta	
DIM2 qPCR F	ggagggtggtatcgcgatcg	
DIM2 qPCR R	tcgatcgagcccaagaatgg	
RID qPCR F	actcgtcttctctcaccgga	
RID qPCR R	aaacactgtggctcaaacgc	
FGSG_02322 qPCR F	aagtgatgtgcctacgggtg	BSseqAnalysis
FGSG_02322 qPCR R	tcgcaacatcaatcccgtca	
FGSG_09595 qPCR F	ttacaccattcccctcgtgc	
FGSG_09595 qPCR R	gtgtcgggttgagggtgat	
Primers used in BSseq Analysis		
Loci_Contig1_9033981-9034488 R	gtattttataaatagatttaaagttataaagt	
Loci_Contig1_9033981-9034488 F	aacatactacatttcctaa	
Loci_Contig6_127178-127396 F	gttaatggttttggaatggtat	
Loci_Contig6_127178-127396 R	ataaaaacctattaaaaataataaaaatt	
M13F	tgtaaaacgacggccagt	
M13R	caggaaacagctatgacc	