

SUPPLEMENTARY MATERIAL

Expression of *Castanea crenata* Allene Oxide Synthase in Arabidopsis Improves the Defense to *Phytophthora cinnamomi*

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	Wild type	CcAOS1	CcAOS2	CcAOS3
Rosette area (cm ²)	0.61 ± 0.14	0.71 ± 0.19 *	0.79 ± 0.30 *	0.83 ± 0.22 *
Root (dry weight; g)	0.0006±0.0002	0.0009±0.0005	0.0007±0.0005	0.0005±0.0002
Aerial (dry weight; g)	0.0049±0.0009	0.0069±0.0006 *	0.0098±0.0034 *	0.0078±0.0006 *
Time of flowering	73±6.67	81±14.67	94±6.0	100±0.00 *
Primary root length (cm)	2.28±1.22	3.30±1.02 *	2.77±1.14	3.55±1.38 *
Lateral root number (between 5-10mm)	6.4±3.5	11±4.08 *	7.7±6.2	10.8±5.5 *
Root hair number (between 5-10mm)	67.3±33.1	43.6±31.2	37.5±25.2 *	49.3±40.3
Root hair length (μm) (between 5-10mm)	243.24±47.79	209.13±47.7	226.27±38.22	228.46±33.14

Supplementary Table 1. Phenotypical parameters of transgenic CcAOS Arabidopsis plants.

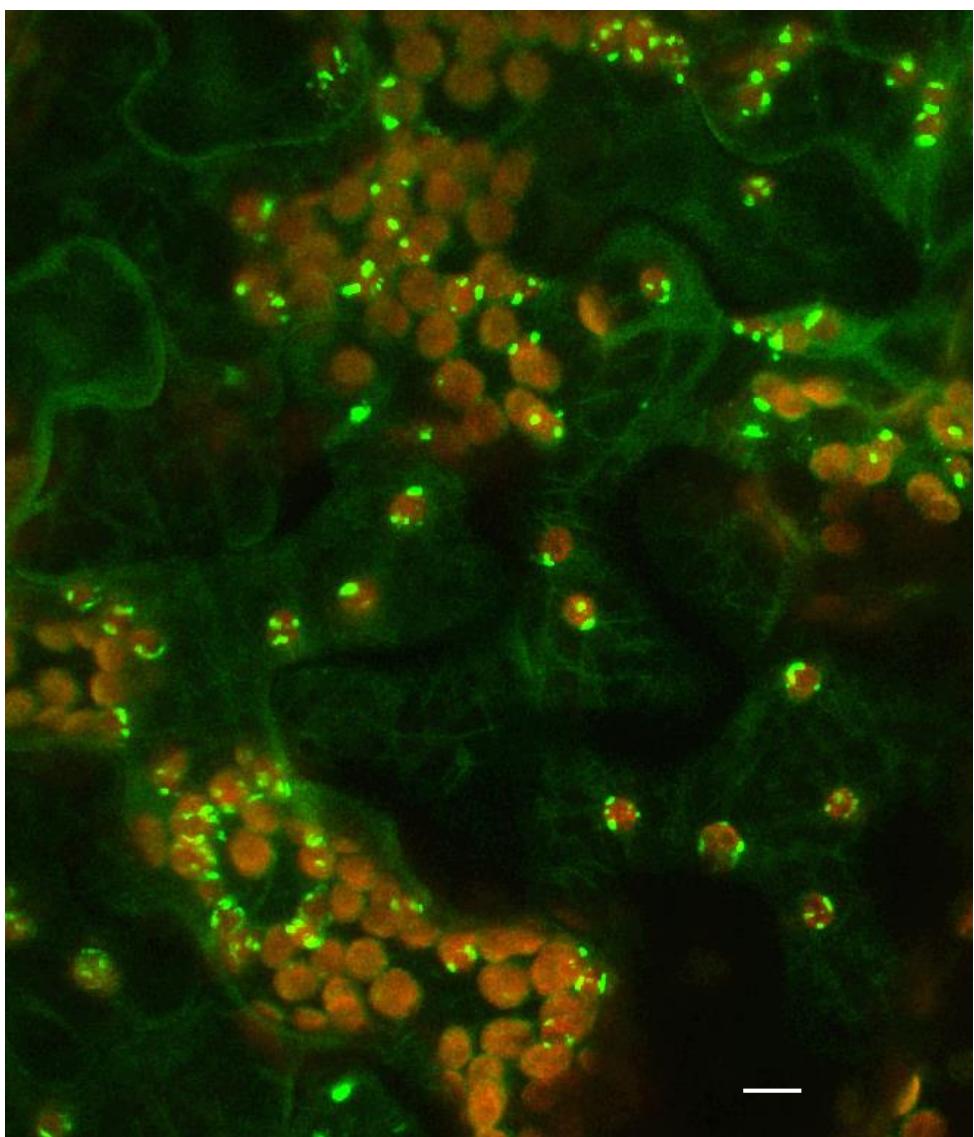
Rosette area and dry weight were measured 1 month after germination. Time of flowering was measured 2 weeks after germination. The remaining root parameters were measured 13 days after germination. Measurements (means ± standard deviation) are from three independent assays, each using 5 plants per genotype. Asterisks refers to significant differences from the Ler-0 wild type ($P<0.05$, *t*-test).

Gene name, locus and relation to pathway (JA – jasmonate; SA – salicylate)	Forward (5'-3')	Reverse (5'-3')	Melting temperature (°C)
ALLENE OXIDE SYNTHASE AT5G42650.1 (JA)	CGACGGTGGGAATAAACAA	TCGCCGGAAAATCTCAATCA	60
LIPOXYGENASE 1 AT1G55020.1 (JA)	GATGGGCTTGAGGTTGGTA	TTCACGGTTGCATTTAGG	62
JASMONATE-ZIM-DOMAIN PROTEIN 10 AT5G13220.1 (JA)	CGATTCCTCGGACTTGAGA	GGAACCGAACGAGATTAGC	60
PLANT DEFENSIN 1.2 AT5G44420.1 (JA) (Proietti et al., 2018)	CACCCATTATCTCGCTGCTCTT	GCCGGTGCCTCGAAAG	58
PATHOGENESIS-RELATED GENE 5 AT1G75040.1 (SA)	ACTCCAGGTGCTCCGACA	ACTCCGCCGCCGTTACATCTT	60
NONEXPRESSER OF PR GENES 1 AT1G64280.1 (SA)	TTATCTGGCCGCCGAACAAG	AATCATCCGGCGAGTCAAAG	60

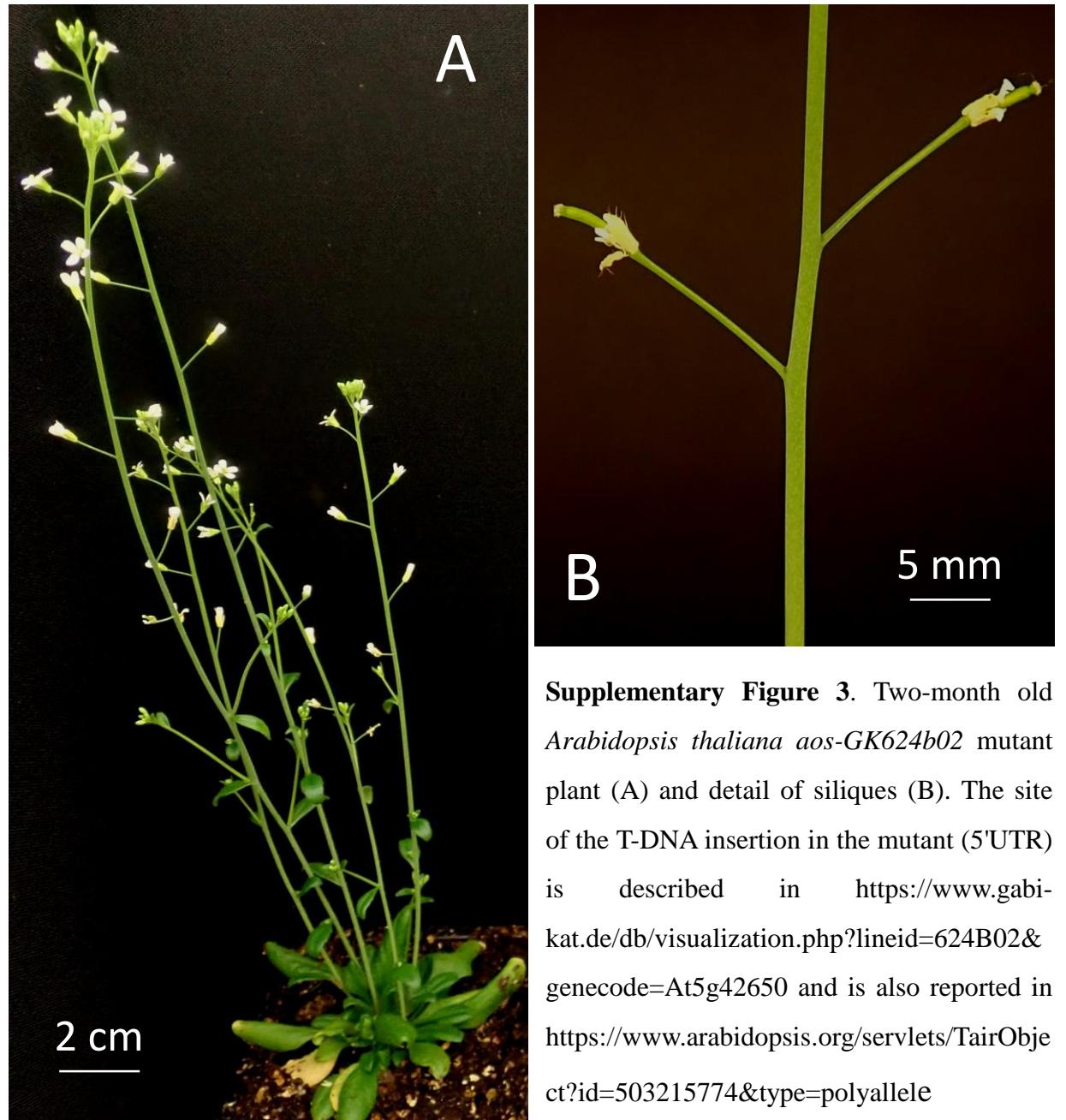
Supplementary Table 2. Primers used in RT-qPCR analysis of jasmonic acid and salycilic acid pathway genes.

	5	15	25	35	45	55
Cc AOS	MASTSLAfp- SLQPKFQSSR KPSKPSTRRF IVRPITASVS EKPSVSVPFA TVVEQAEPK						
At AOS	MASISTPFPI SLHPKTVRS- ---KPLKFRV LTRPIKASGS E----- TPDL TVATRTGSKD						
Consensus	MAS S FP SL PK S KP R RPI AS S E P TV						
	65	75	85	95	105	115
Cc AOS	LPIKKIIPGNY GLPFVGPIRD RFDFFYNQGR DEYFKSRAHK YQSTVFRANM PPGPLIASNP						
At AOS	LPIRNIPGNY GLPIVGPIKD RWDYFYDQGA EEEFFKSRIRK YNSTVYRVNM PPGAFIAENP						
Consensus	LPI IPGNY GLP VGPI D RDYQG E FKS R K YSTVRNM PPG IA NP						
	125	135	145	155	165	175
Cc AOS	NVVVLLDGKS FPVLFDVTKV EKKDLFTGTF MPSTELETGGY RVLSYLDPSE PNHGKLKRLL						
At AOS	QVVALLDGKS FPVLFDVDKV EKKDLFTGTY MPSTELETGGY RILSYLDPSE PKHEKLKNLL						
Consensus	VV LLDGKS FPVLFDV KV EKKDLFTGT MPSTELETGGY R LSYLDPSE PHKLKLL						
	185	195	205	215	225	235
Cc AOS	FFHLKARRDH VIPEFHSSYT ELFEGLENEL ATKGKAAFGE PSDLAAFNFL ARSLYGTNPV						
At AOS	FFLLKSSRNR IFPEFQATYS ELFDSLEKEL SLKGKADFGG SSDGTAFNFL ARAFYGTNP						
Consensus	FF LK R PEF Y ELF LE EL KGKA FG SD AFNFL AR YGTNP						
	245	255	265	275	285	295
Cc AOS	DTKLGLDAPK MIDKWLFWQI SPLFSLGLPK HLDDLLLRTV RLPPALVKAD YQKLYDFFYA						
At AOS	DTKLKADAPG LITKWLFNL HPLL SIGLPR VIEEPLIHTF SLPPALVKSD YQRLYEFFLE						
Consensus	DTKL DAP I KW PL S GLP L T LPPALVK D YQ LY FF						
	305	315	325	335	345	355
Cc AOS	SSGFVLDEAE RLGISREEAC HNLLFATCFN SFGGMKFLFP NMVKLIGRAG VKLHTQLAEE						
At AOS	SAGEILVEAD KLGISREEAT HNLLFATCFN TWGGMKILFP NMVKRIGRAG HQVHNRLAEE						
Consensus	S G L EA LGISREEA HNLLFATCFN GGMK LFP NMVK IGRAG H LAEE						
	365	375	385	395	405	415
Cc AOS	IRSVIRSNNG NITMAAMEQM PLMKSVVYES LRIEPPVALQ YGKAKKDLVI ESHDAAFRKV						
At AOS	IRSVIKSNNG ELTMGAIKEM ELTKSVVYEC LRFEPPVTAQ YGRAKKDLVI ESHDAAFKV						
Consensus	IRSVI SNNG TMAEM L KSVVYE LR EPPV Q YG AKKDLVI ESHDAAF VK						
	425	435	445	455	465	475
Cc AOS	EGERMLFGYQP FATKDPKIFE RAEEFVADRF VGEEGEKLLK HVLWSNGPES ESPSVGNKQC						
At AOS	AGEMLYGYQP LATRDPKIFD RADEFVPERF VGEEGEKLLR HVLWSNGPET ETPTVGNKQC						
Consensus	GEML GYQP AT DPKIF RA EFV RF VGEEGEKLL HVLWSNGPE E P VGNKQC						
	485	495	505	515	525	
Cc AOS	AGKDFVVLVA RLLVVELFLR YDSIEIEVDS SPLGAALTWT SLKKASF						
At AOS	AGKDFVVLVA RLFVIEIFRR YDSFDIEVGT SPLGSSVNFS SLRKASF						
Consensus	AGKDFVVLVA RLVEFR YDS IEV SPLG SL KASF						

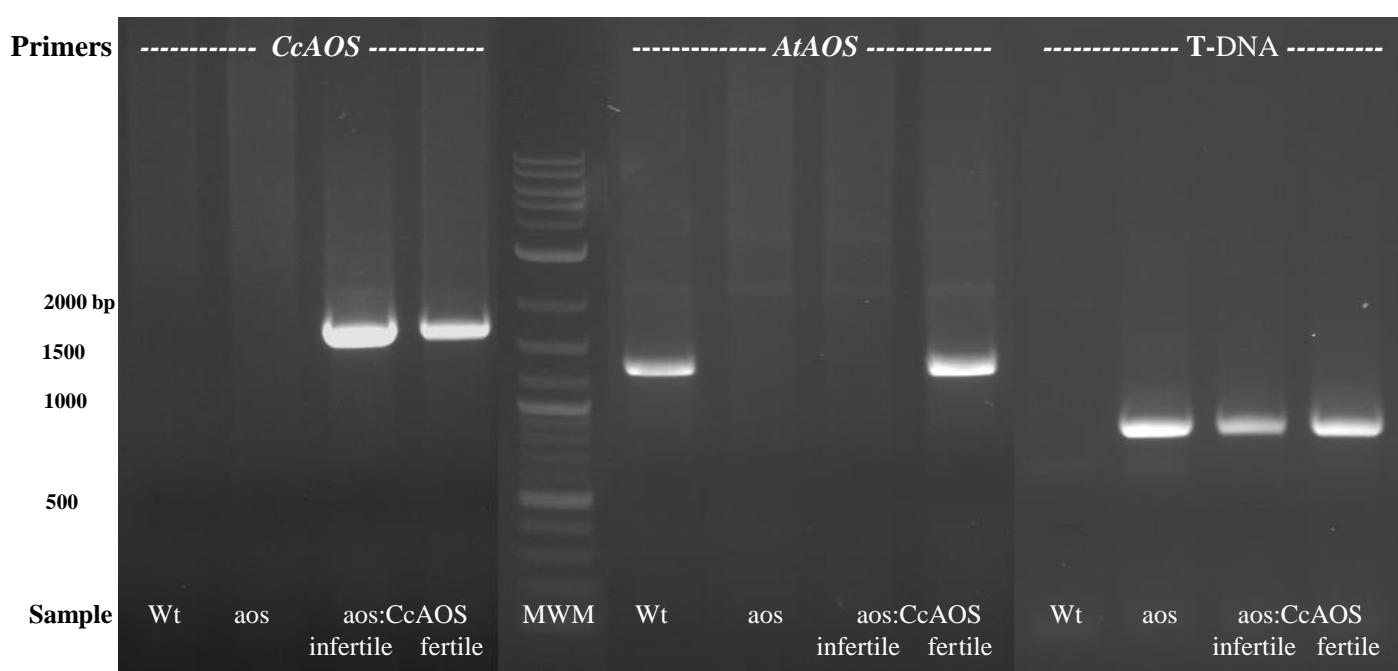
Supplementary Figure 1. Alignment of amino acid sequences of *Castanea crenata* (CcAOS) and *Arabidopsis thaliana* (AtAOS) allene oxide synthase. Clustal W alignment was conducted using the BioEdit Sequence Alignment Editor. Numbers refer to the amino acid position.



Supplementary Figure 2. 2D stack of subcellular imaging upon transient CcAOS-eGFP expression in leaves of *N. benthamiana*. Although CcAOS clearly accumulates at chloroplasts, a reticulate-like signal in the cytosol suggests that trafficking of the protein may involve the endomembrane compartment. Scale Bar = 10 μ m.



Supplementary Figure 3. Two-month old *Arabidopsis thaliana* *aos-GK624b02* mutant plant (A) and detail of siliques (B). The site of the T-DNA insertion in the mutant (5'UTR) is described in <https://www.gabi-kat.de/db/visualization.php?lineid=624B02&genecode=At5g42650> and is also reported in <https://www.arabidopsis.org/servlets/TairObject?id=503215774&type=polyallele>



Supplementary Figure 4. Genotyping of plants after expression of *CcAOS* gene in *aos-GK624b02* mutants. Genomic DNA of F2 *aos:CcAOS*-infertile and *aos:CcAOS*-fertile plants was screened for *CcAOS*, *AtAOS* and T-DNA insertion and further compared with wild-type Col-0 (Wt) and *aos* plants (used as controls). The expected band sizes are 1581 bp for the amplification of *CcAOS* ORF, 1345 bp for the amplification of a fragment of non-disrupted *AtAOS* and 850 bp for the amplification of a fragment of the T-DNA left border fused to *AtAOS* (T-DNA). MWM: molecular weight marker N3200L (New England Biolabs, Ipswich, MA, USA).