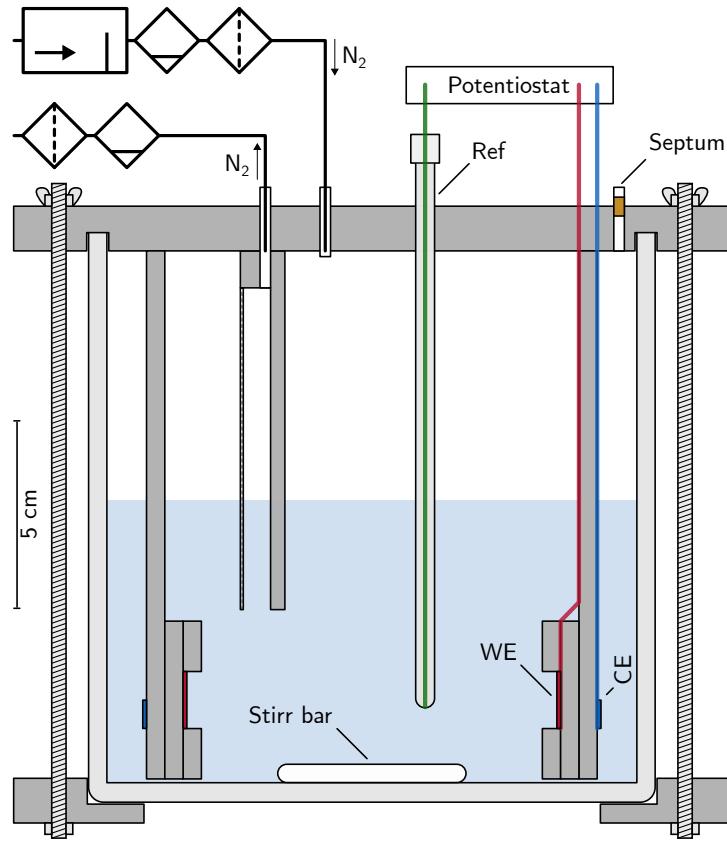


## Supplementary Material

### 1 BIOELECTROCHEMICAL REACTOR



**Figure S1.** Schematic of the bioelectrochemical reactor with electrical peripherals and gas supply system. The working electrode is connected to the potentiostat in a two-wire configuration in order to reduce the uncompensated resistance. Abbreviations: working electrode (WE), counter electrode (CE), reference electrode (Ref).

## 2 (GROWTH) MEDIA

**Table S1.** Components of the different media. The symbol ‘–’ indicates no change compared to the reference medium RM.

Component	Medium type					
	RM	25xPBS	100xRF	IM	AM	WB
Na <sub>2</sub> HPO <sub>4</sub>	10 mM	250 mM	–	40 mM	–	–
KH <sub>2</sub> PO <sub>4</sub>	1.76 mM	44 mM	–	7.04 mM	–	–
Sodium D/L lactate	50 mM	–	–	–	–	none
NaCl	137 mM	–	–	77 mM	–	–
KCl	2.7 mM	–	–	–	–	–
Casamino acids	1 g L <sup>-1</sup>	–	–	–	–	–
Riboflavin	none	–	100 µM	1 µM	–	–
Fumarate	none	–	–	–	100 mM	–
Trace elements	See Tab. S2	–	–	–	–	–

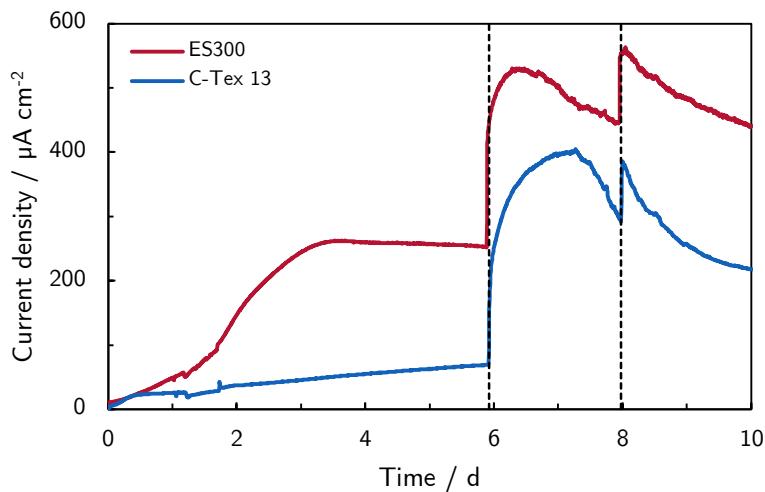
**Table S2.** Concentration of the trace elements in the media.

Component	Concentration
(NH <sub>4</sub> ) <sub>2</sub> SO <sub>4</sub>	1 mM
CaCl <sub>2</sub>	0.1 mM
Mg <sub>2</sub> SO <sub>4</sub>	1 mM
CoCl <sub>2</sub>	5 µM
CuSO <sub>4</sub>	0.2 µM
K <sub>3</sub> BO <sub>3</sub>	57 µM
FeCl <sub>2</sub>	5.4 µM
MnSO <sub>4</sub>	1.3 µM
Na <sub>2</sub> EDTA	67.2 µM
Na <sub>2</sub> MoO <sub>4</sub>	3.9 µM
Na <sub>2</sub> SeO <sub>4</sub>	1.5 µM
NiCl <sub>2</sub>	5 µM
ZnSO <sub>4</sub>	1 µM

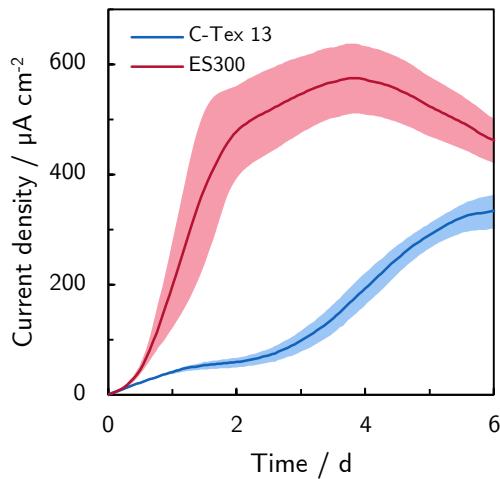
**Table S3.** Lysis buffer (LyB) composition. <sup>a</sup>Protease inhibitor cocktail for bacteria (Carl Roth, Germany)

Component	Concentration
Tris/HCl	20 mM
Triton X-100	16.5 mM
NaCl	137 mM
Protease inhibitors <sup>a</sup>	25 µL mL <sup>-1</sup>

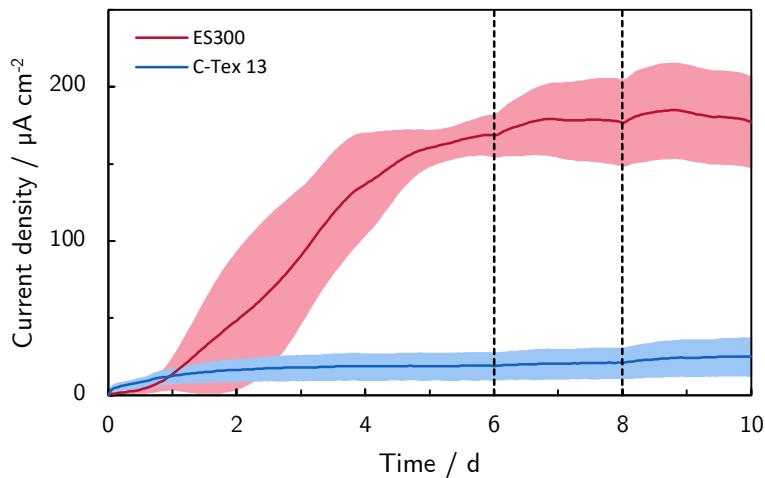
### 3 CHRONOAMPEROMETRY DATA



**Figure S2.** Current production with riboflavin spikes on day 6 (+500 nM) and 8 (+500 nM). The current does not stabilize within 2 days. Therefore, the effect of riboflavin on the current production is studied in individual experiments.

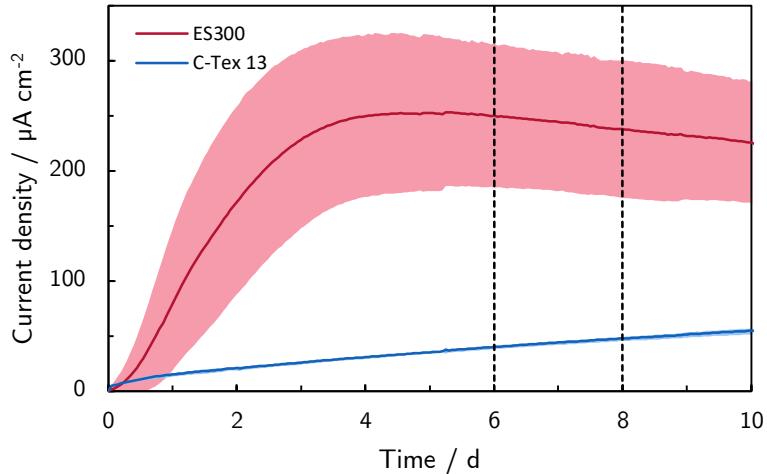


**Figure S3.** Current production with the improved medium (1  $\mu\text{M}$  riboflavin and 40 mM PBS buffer). The shaded area corresponds to the sample standard deviation of three anodes.



**Figure S4.** Current production with lactate spikes on day 6 (+25 mM) and 8 (+50 mM). The current density is evaluated before each lactate spike on day 6 and 8, and at the end of the experiment on day 10. The shaded area corresponds to the sample standard deviation of three anodes.

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**Figure S5.** Control experiment for the spike experiments without perturbations. The current density is evaluated at on day 6 and 8, and at the end of the experiment on day 10. The shaded area corresponds to the sample standard deviation of four anodes.

**Table S4.** Experimental results as numerical values with sample standard deviation. Initial and final  $OD_{600}$  values are given in brackets.

<b>Riboflavin</b>		0 nM (0.05 / 0.05)	500 nM (0.04 / 0.04)	1000 nM (0.05 / 0.04)
ES300	$i_{\text{Max}} / \mu\text{A cm}^{-2}$	217.5 ± 9.8	254 ± 35	326 ± 41
	$i_{\text{Final}} / \mu\text{A cm}^{-2}$	199 ± 13	200 ± 22	248 ± 20
	Dry weight / mg	5.51 ± 0.99	4.76 ± 0.45	5.42 ± 0.67
	Fold change ( $i_{\text{Max}}$ )	–	1.17 ± 0.16	1.50 ± 0.20
	Fold change ( $i_{\text{Final}}$ )	–	1.00 ± 0.11	1.25 ± 0.13
C-Tex 13	$i_{\text{Max}} / \mu\text{A cm}^{-2}$	60.0 ± 3.2	207 ± 10	353 ± 19
	$i_{\text{Final}} / \mu\text{A cm}^{-2}$	59.2 ± 3.1	204.8 ± 9.8	334 ± 15
	Dry weight / mg	2.089 ± 0.073	7.23 ± 0.95	13.9 ± 2.8
	Fold change ( $i_{\text{Max}}$ )	–	3.45 ± 0.17	5.89 ± 0.32
	Fold change ( $i_{\text{Final}}$ )	–	3.46 ± 0.17	5.64 ± 0.26
<b>Buffer capacitance (0.07 / 0.05)</b>		10 mM	+10 mM	+20 mM
ES300	Current density / $\mu\text{A cm}^{-2}$	327 ± 37	412 ± 56	590 ± 25
	Dry weight / mg	–	–	12.1 ± 1.8
	Fold change	–	1.260 ± 0.041	1.8 ± 0.2
C-Tex 13	Current density / $\mu\text{A cm}^{-2}$	83 ± 31	88 ± 34	88 ± 37
	Dry weight / mg	–	–	5.1 ± 1.9
	Fold change	–	1.051 ± 0.023	1.213 ± 0.019
<b>Inoculation cell density (0.06 / 0.20)</b>		0.05	+0.05	+0.10
ES300	Current density / $\mu\text{A cm}^{-2}$	176 ± 22	188 ± 15	188 ± 25
	Dry weight / mg	–	–	5.7 ± 2.0
	Fold change	–	1.068 ± 0.038	1.214 ± 0.090
C-Tex 13	Current density / $\mu\text{A cm}^{-2}$	19.90 ± 0.86	24.2 ± 3.7	24.21 ± 0.84
	Dry weight / mg	–	–	1.823 ± 0.065
	Fold change	–	1.223 ± 0.038	2.088 ± 0.090
<b>Lactate (0.05 / 0.04)</b>		25 mM	+25 mM	+50 mM
ES300	Current density / $\mu\text{A cm}^{-2}$	167 ± 14	177 ± 28	178 ± 29
	Dry weight / mg	–	–	4.6 ± 1.2
	Fold change	–	1.046 ± 0.097	1.05 ± 0.11
C-Tex 13	Current density / $\mu\text{A cm}^{-2}$	19.0 ± 9.2	21 ± 10	25 ± 13
	Dry weight / mg	–	–	1.491 ± 0.098
	Fold change	–	1.1089 ± 0.0060	1.306 ± 0.049
<b>Control (0.05 / 0.05)</b>		Day 6	Day 8	Day 10
ES300	Current density / $\mu\text{A cm}^{-2}$	250 ± 66	237 ± 63	226 ± 55
	Dry weight / mg	–	–	7.2 ± 2.3
	Fold change	–	0.950 ± 0.025	0.906 ± 0.030
C-Tex 13	Current density / $\mu\text{A cm}^{-2}$	40.0 ± 1.6	47.3 ± 2.3	53.9 ± 9.4
	Dry weight / mg	–	–	3.2 ± 1.1
	Fold change	–	1.185 ± 0.048	1.350 ± 0.062
<b>Improved medium (0.06 / 0.06)</b>				
ES300	$i_{\text{Max}} / \mu\text{A cm}^{-2}$	575 ± 63		
	$i_{\text{Final}} / \mu\text{A cm}^{-2}$	461 ± 40		
	Dry weight / mg	9.50 ± 0.60		
C-Tex 13	$i_{\text{Max}} / \mu\text{A cm}^{-2}$	334 ± 32		
	$i_{\text{Final}} / \mu\text{A cm}^{-2}$	334 ± 32		
	Dry weight / mg	7.97 ± 0.68		