

FIGURE S1

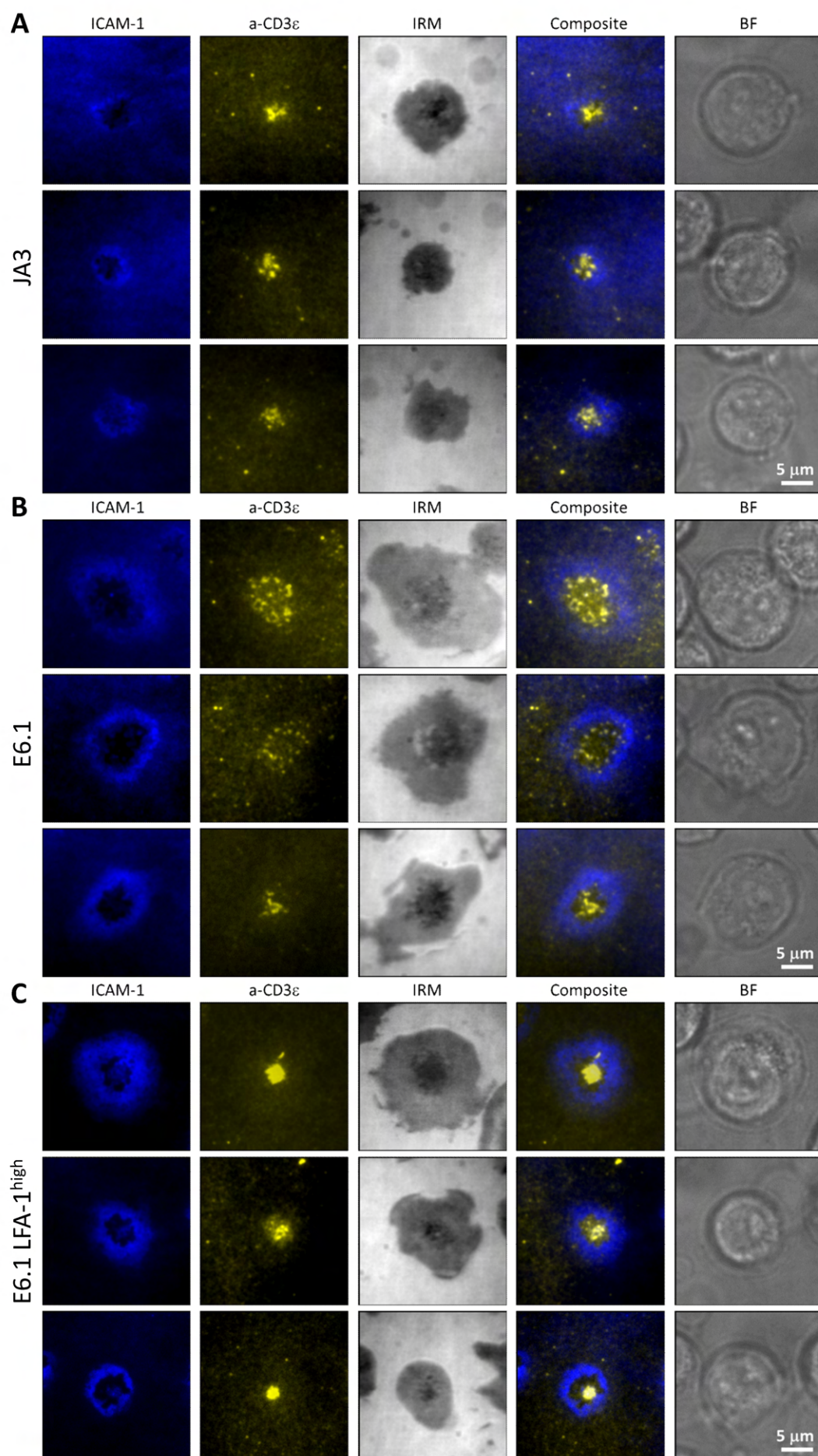


FIGURE S2

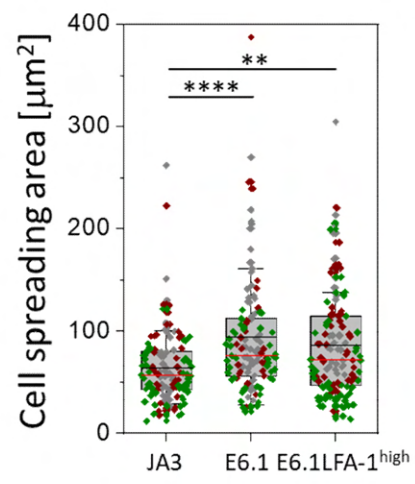


FIGURE S3

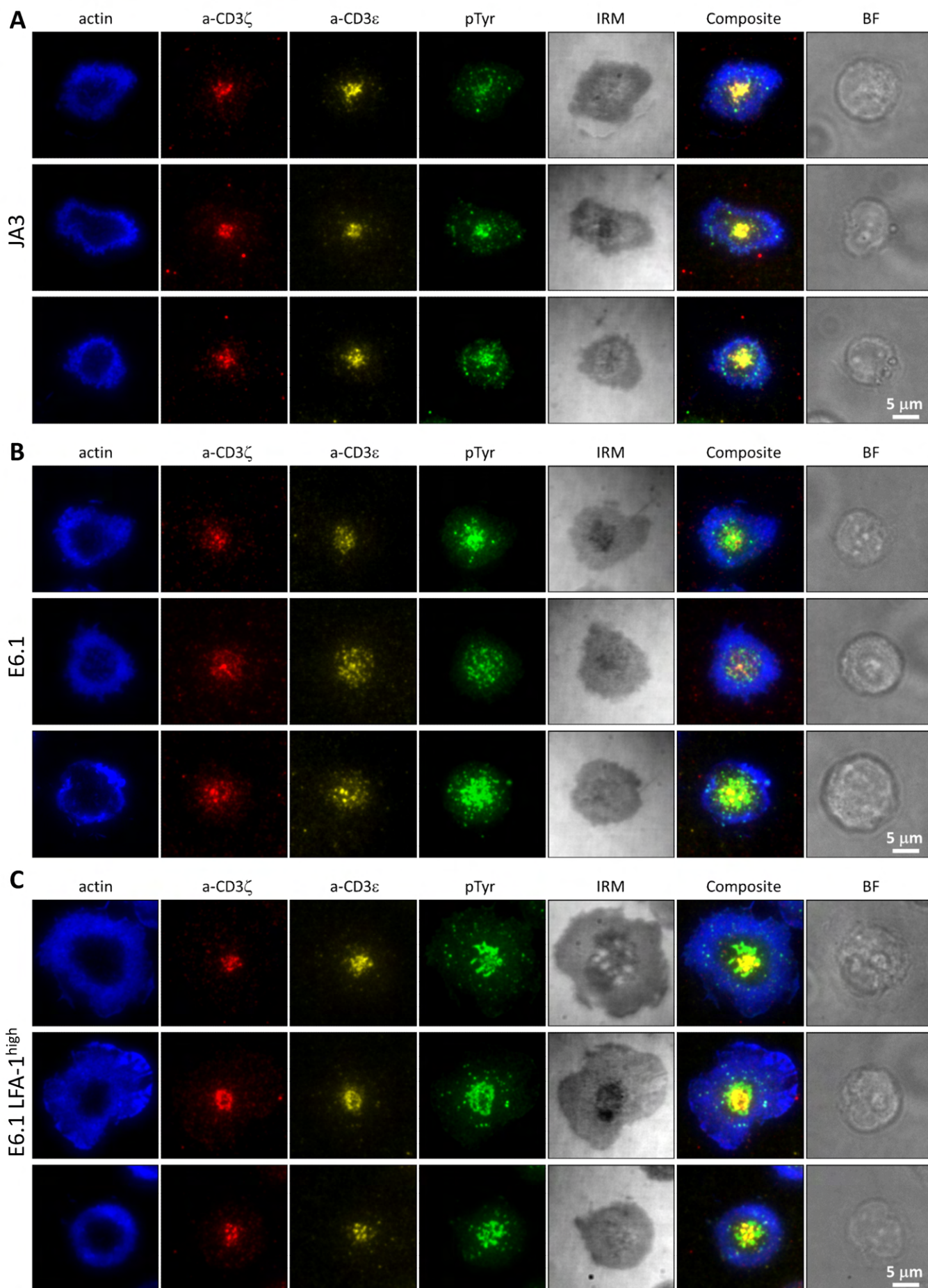


FIGURE S4

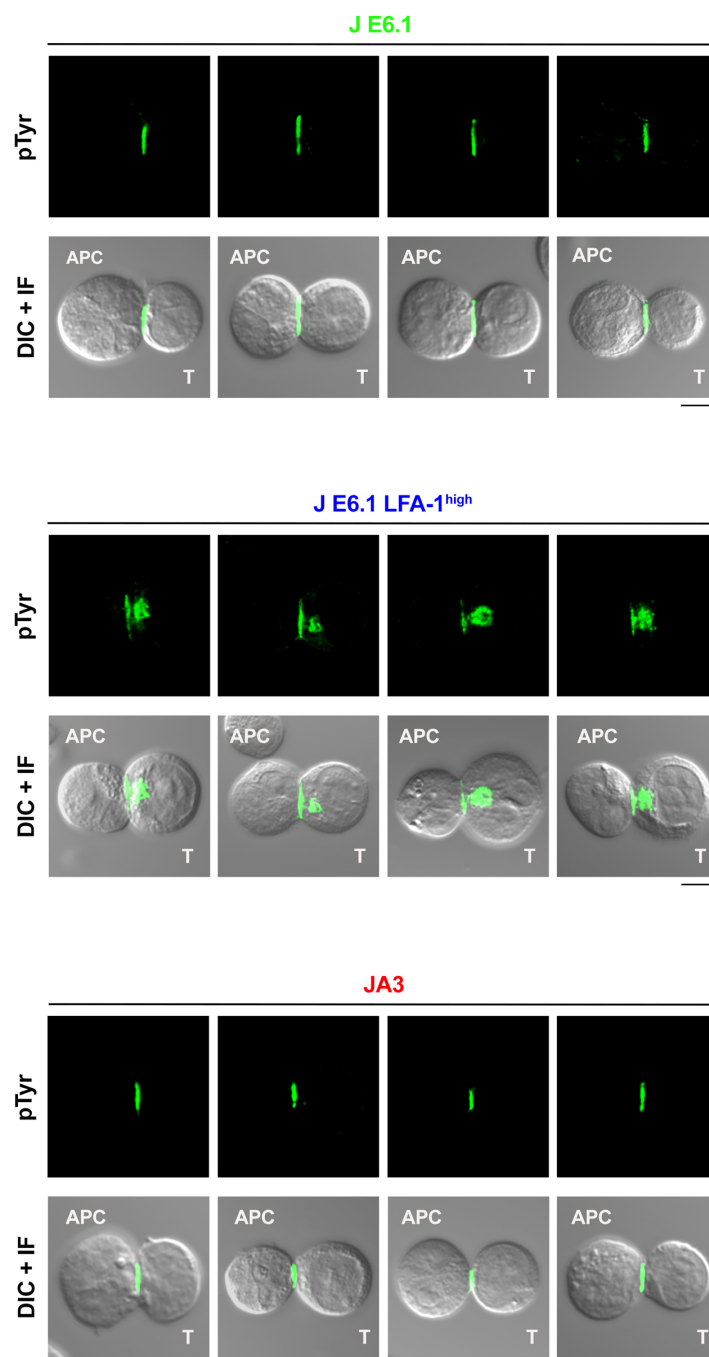


FIGURE S5

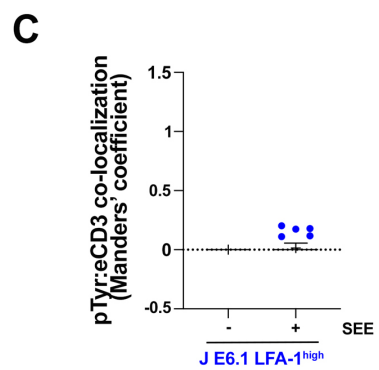
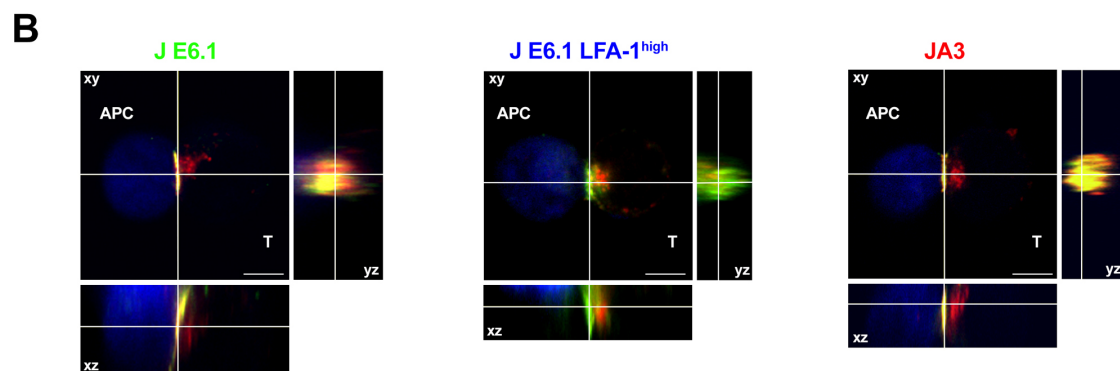
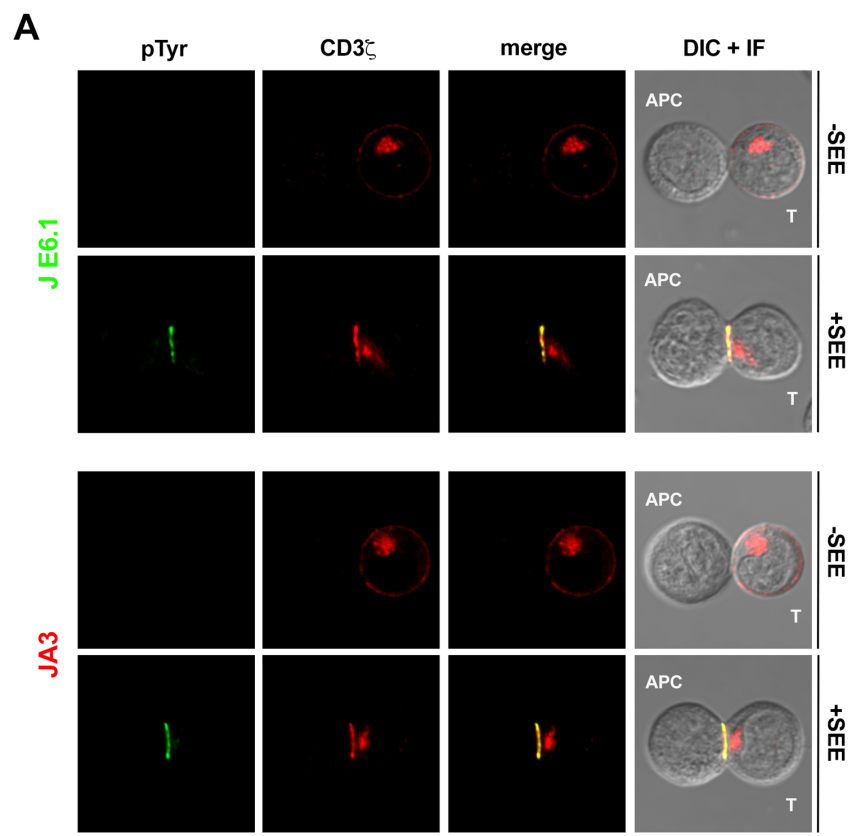
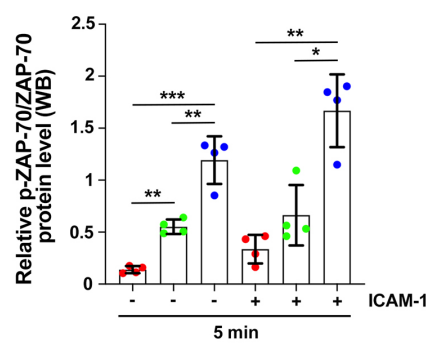
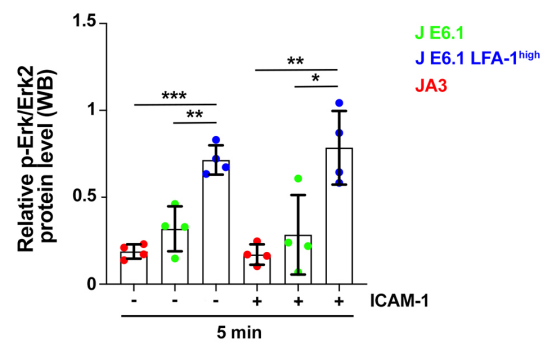


FIGURE S6

A**B****FIGURE S7**

SUPPLEMENTAL FIGURE LEGENDS

Figure S1. Jurkat cell lines express comparable surface levels of CD3 ϵ . **A.** Flow cytometric analysis of the surface expression of LFA-1 on JA3, E6.1 and E6.1 LFA-1^{high} Jurkat cells. The histograms represent Mode of LFA-1. **B.** Flow cytometric analysis of the surface expression of CD3 ϵ on JA3, E6.1 and E6.1 LFA-1^{high} Jurkat cells. Representative FACS plots of CD3 ϵ are shown. The histograms represent GMFI, Mode and percentage of CD3 ϵ . Bars and error bars represent mean \pm SD. Data are from 4 independent experiments; each dot and colour represents an independent experiment. One-way analysis of variance (ANOVA) with Tukey's post-hoc test. Only significant differences are shown. *, p<0.05; **, p<0.01; ***, p<0.001.

Figure S2. Representative TIRFM images of JA3 (**A**), E6.1 (**B**) or E6.1 LFA-1^{high} (**C**) Jurkat cells interacting with activating [ICAM-1-AF405 (blue) + anti-CD3 ϵ UCHT1-CF568 Fab' (yellow)] SLB for 15 min. IRM – Interference reflection microscopy, BF – Bright Field. Scale bar, 5 μ m.

Figure S3. Quantification of the cell spreading area of JA3, E6.1 and E6.1 LFA-1^{high} Jurkat cells interacting with activating SLB for 15 min. Horizontal lines and error bars represent mean \pm SD (black line) and median (red line). Gray boxes represent 25-75 percentile. Data are from minimum of 120 cells from 3 independent experiments; each dot represents a cell; each colour represents an independent experiment. One-way analysis of variance (ANOVA) with Tukey's post-hoc test. Only significant differences are shown. **, p<0.01; ****, p<0.0001.

Figure S4. Representative TIRFM images of JA3 (**A**), E6.1 (**B**) or E6.1 LFA-1^{high} (**C**) Jurkat cells interacting with activating [ICAM-1 + anti-CD3 ϵ UCHT1-CF568 Fab' (yellow)] SLB for 15 min. Cells were permeabilized and stained with directly conjugated primary antibodies against anti-CD3 ζ -AF647 (red), anti-phosphotyrosine pTyr-AF488 (green) and the cell actin cytoskeleton was labelled with phalloidin-AF405 (blue). IRM – Interference reflection microscopy, BF – Bright Field. Scale bar, 5 μ m.

Figure S5. Gallery of examples of SEE-specific conjugates of E6.1, E6.1 LFA-1^{high} and JA3 Jurkat cells. Representative images of SEE-specific conjugates stained with anti-pTyr antibody as described for the representative images in figure 3.

Figure S6. Neither E6.1 nor JA3 cells have an intracellular pTyr pool associated with endosomal CD3 ζ , as opposed to E6.1 LFA-1^{high} cells. A. Immunofluorescence analysis of 15-min conjugates of E6.1 or JA3 cells and Raji cells (APC) in the presence or absence of SEE. Cells were co-stained with anti-pTyr and anti-CD3 ζ antibodies. The histogram showing the quantification of the co-localization of pTyr and the endosomal CD3 ζ pool in E6.1 LFA-1^{high} conjugates is presented in figure 3E. **B.** Representative xy and orthogonal views of conjugates formed by E6.1, E6.1 LFA-1^{high} or JA3 with SEE-pulsed Raji B cells (3D reconstructions of representative z-stacks are shown in supplemental videos 1-3). Cells were co-stained with anti-pTyr and anti-CD3 ζ antibodies. **C.** Control of co-localization specificity of pTyr and the endosomal CD3 ζ pool in E6.1 LFA-1^{high} cells. Co-localization was measured after tilting one staining relative to the other of 90 degrees (mean \pm SD; Kruskal-Wallis test; n \geq 20 conjugates/sample from 3 independent experiments). Graph boxes represent 10-90 percentile and the mean is shown as "+". Only significant differences are shown. Scale bar, 5 μ m.

Figure S7. LFA-1 engagement does not enhance basal signaling in E6.1, E6.1 LFA-1^{high} or JA3 Jurkat cells. A,B. Immunoblot analysis with anti-p-ZAP-70 (A) or anti-p-Erk1/2 (B) antibodies of lysates from E6.1, E6.1 LFA-1^{high} or JA3 Jurkat cells either unstimulated or plated on glass-immobilized ICAM-1 for 5 min. Anti-ZAP-70 and anti-Erk2 antibodies were used as respective loading controls. The histograms show the quantification of the relative levels of p-ZAP-70 and p-Erk1/2, normalized to the respective loading controls (mean \pm SD; paired Student's *t*-test; n=4). Only significant differences are shown. * p<0.05; ** p<0.01; *** p<0.001.

Video 1_J E6.1. The movie shows a representative confocal z-stack of a E6.1-APC conjugate. CD3 ϵ , red; pTyr, green; SEE-pulsed Raji, blue.

Video 2_J E6.1 LFA-1^{high}. The movie shows a representative confocal z-stack of a E6.1 LFA-1^{high}-APC conjugate. CD3 ϵ , red; pTyr, green; SEE-pulsed Raji, blue.

67 **Video 3_JA3.** The movie shows a representative confocal z-stack of a JA3-APC conjugate.
68 CD3 ϵ , red; pTyr, green; SEE-pulsed Raji, blue.

69

70 **Video 4_J E6.1.** The movie shows a representative confocal z-stack and orthogonal view
71 of the distribution of CD3 ϵ and pTyr in E6.1 Jurkat cell line under activating conditions. CD3 ϵ ,
72 red; pTyr, green; actin, blue.

73

74 **Video 5_J E6.1 LFA-1^{high}.** The movie shows a representative confocal z-stack and
75 orthogonal view of the distribution of CD3 ϵ and pTyr in E6.1 LFA-1^{high} Jurkat cell line under
76 activating conditions. CD3 ϵ , red; pTyr, green; actin, blue.

77

78 **Video 5_JA3.** The movie shows a representative confocal z-stack and orthogonal view of
79 the distribution of CD3 ϵ and pTyr in JA3 Jurkat cell line under activating conditions. CD3 ϵ ,
80 red; pTyr, green; actin, blue.

81