

Supplementary Material

Exogenous and endogenous phosphoethanolamine transferases differently affect colistin resistance and fitness in *Pseudomonas aeruginosa*

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1. Supplementary Figures and Tables

1.1. Supplementary Figures



Supplementary Figure 1. MALDI-TOF spectra of lipid A extracted from PAO1 pME*eptA* or PA14 pME*eptA* cultured in the absence or presence of IPTG at 0.125 or 0.5 mM. The m/z values of peaks corresponding to phosphoethanolaminated lipid A forms are highlighted in yellow. Spectra are representative of three biological replicates giving similar results.



Supplementary Figure 2. Growth curves of (A) PAO1 or (B) PA14 carrying pME6032, pMEmcr-1 or pME*eptA* in the presence of increasing IPTG concentrations (0-2 mM). Data represent the mean (\pm standard deviation) of three independent experiments.

Supplementary Figure 3. Full-length alignment of selected EptA orthologs from *Acinetobacter* baumannii (SSI74383.1), Enterobacter cloacae (WP_059385803.1), Escherichia coli (OWW57050.1), Helicobacter pylori (WP_189394748.1), Klebsiella pneumoniae (CCI78008.1), Neisseria meningitidis (AKM91408.1), Pseudomonas aeruginosa (WP_003113468.1), Salmonella enterica (QNL54535.1), Shigella flexneri (QLG55136.1), Vibrio cholerae (TYC39410.1). The MCR-1 protein has been included as control (WP_049589868.1). The highly conserved catalytic residue mutated in the EptA^{T278A} variant and the 22 residues deleted in the EptA^{ΔC-ter} variant are highlighted with a black box and a red box, respectively.

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Supplementary Figure 4. MALDI-TOF spectra of lipid A extracted from PAO1 or PA14 cells expressing EptA variants mutated in a catalytic residue (EptA^{T278A}) or deleted of 22 amino acids at the C-terminus (EptA^{ΔC -ter}) cultured in the presence of 0.5 mM IPTG. The m/z values of peaks corresponding to phosphoethanolaminated lipid A forms are highlighted in yellow. Spectra are representative of three biological replicates giving similar results.

1.2. Supplementary Table

Strain or plasmid	Genotype and/or relevant characteristics	Reference or source
P. aeruginosa		
PAO1 (ATCC15692)	Reference isolate	American type culture collection
PA14	Reference isolate	Rahme et al., 1995
PAO1 PrpsA::arn	PAO1 derivative in which the promoter of the <i>arn</i> operon is replaced by the promoter of the housekeeping gene <i>rpsA</i>	Lo Sciuto et al., 2020
PA14 PrpsA::arn	PA14 derivative in which the promoter of the <i>arn</i> operon is replaced by the promoter of the housekeeping gene <i>rpsA</i>	Lo Sciuto et al., 2020
E. coli		
DH5aF'	recA1 endA1 hsdR17 supE44 thi-1 gyrA96 relA1 ∆(lacZYA- argF)U169[φ80 dlacZ∆M15], Nal ^R	Liss, 1987
Plasmid		
pHNSHP45	<i>mcr-1</i> carrying plasmid	Liu et al., 2016
pBluescript II KS (pBS)	Sequencing vector; ColE1 replicon; Ap ^R	Stratagene
pBS <i>eptA</i>	pBS containing the coding sequence of the <i>eptA</i> gene from <i>P. aeruginosa</i> PAO1 (PA1972)	This study
pBSmcr-1	pBS containing the coding sequence of the <i>mcr-1</i> gene from plasmid pHNSHP45	This study
pBSeptA ^{T278A}	pBS containing the <i>eptA</i> coding sequence with a point mutation in the codon 278 leading to a T278A substitution	This study
$pBSeptA^{\Delta C-ter}$	pBS containing a truncated <i>eptA</i> allele encoding for an EptA variant lacking the last 22 amino acids	This study
pME6032	IPTG-inducible expression vector, <i>lacI^Q</i> , Tc ^R .	Heeb and Haas, 2001
pME <i>eptA</i>	pME6032 containing the <i>eptA</i> (PA1972) coding sequence from pBS <i>eptA</i> cloned downstream of the IPTG-inducible promoter	This study
pMEmcr-1	pME6032 containing the <i>mcr-1</i> coding sequence from pBS <i>mcr-1</i> cloned downstream of the IPTG-inducible promoter	This study
pMEeptA ^{T278A}	pME6032 containing the $eptA^{T278A}$ allele from pBS $eptA^{T278A}$ cloned downstream of the IPTG-inducible promoter	This study
$pMEeptA^{\Delta C-ter}$	pME6032 containing the $eptA^{\Delta C\text{-ter}}$ allele from pBS $eptA^{\Delta C\text{-ter}}$ cloned downstream of the IPTG-inducible promoter	This study

Supplementary Table 1. Bacterial strains and plasmids used in this study.

Primer name	Sequence (5'-3') ¹	Restriction sites ²	Application	
mcr-1_pME6032_FW	cggaattcATGATGCAGCATACTTCTGTG	EcoRI	Convertion of pDSmar 1	
mcr-1_pME6032_RV	ccc <u>ctcgag</u> TCAGCGGATGAATGCGGTG	XhoI	Generation of pbSmcr-1	
eptA_pME6032_FW	cggaattCATGTCGAAAGCCCGCGC	EcoRI	Generation of pBS <i>eptA</i>	
eptA_pME6032_RV	ccc <u>ctcgaG</u> TATCAGGAAGCCGGCGG	XhoI		
eptA_T278A_FW	CGGTACCGAGgCCGCGGTGTC		Convertion of a DS and T278A	
eptA_T278A_RV	CAGGAGTGCACGTTGGAGAAGTTGATC		Generation of pbSeptA	
$eptA_\Delta 22aa_RV^3$	ccc <u>ctcgag</u> tcaCGCCTGCAGGACCTCGGG	XhoI	Generation of $pBSeptA^{\Delta C-ter}$	
$eptA_RT_FW^4$	TGCCCTGCATGTTCTCCAAC			
$eptA_RT_RV^4$	GATCCTTGCTCTCGCTCAGG		qRT-PCR	
rpoD_RT_FW	GGGCGAAGAAGGAAATGGTC			
rpoD_RT_RV	CAGGTGGCGTAGGTGGAGAA			
M13_FW	GTTTTCCCAGTCACGAC		DNA sequencing from pBS	
M13_RV	AACAGCTATGACCATG			

Supplementary Table 2. Primers used in this study.

¹ Lowercase letters indicate the region of the primer that does not anneal to the template.

² The restriction site used for cloning is underlined in the primer sequence.

³ This primer was paired with *eptA_pME6032_FW* to amplify the truncated *eptA*^{Δ C-ter} variant.

⁴ Primers described and used in Nowicki et al., 2015.

Additional references (not included in the main text)

Liss, L. (1987). New M13 host: DH5 F' competent cells. Focus 9, 13.

Rahme, L.G., Stevens, E.J., Wolfort, S.F., Shao, J., Tompkins, R.G., Ausubel, F.M. (1995). Common virulence factors for bacterial pathogenicity in plants and animals. *Science* 268, 1899-1902.