Introgressing *cry1Ac* for Pod Borer Resistance in Chickpea through Marker Assisted Backcross Breeding

Ajinder Kaur¹, Urvashi Sharma¹, Sarvjeet Singh², Ravinder Singh², Yogesh Vikal¹, Satnam Singh³, Palvi Malik¹, Khushpreet Kaur³, Inderjit Singh², Shayla Bindra², Bidyut Kumar Sarmah⁴ and Jagdeep Singh Sandhu^{1*}

¹School of Agricultural Biotechnology, Punjab Agricultural University, Ludhiana, Punjab 141004, India,

² Pulses Section, Department of Plant Breeding and Genetics, Punjab Agricultural University, Ludhiana, Punjab 141004, India, ³Punjab Agricultural University, Regional Research Station, Faridkot, Punjab 151203, India, ⁴ Department of Biotechnology-Assam Agricultural University Centre, Assam Agricultural University, Jorhat, Assam 785013, India

* Correspondence: Jagdeep Singh Sandhu

js_sandhu@pau.edu

SUPPLEMENTARY FIGURE 1 Schematic map of transgene construct showing *cry1Ac* under the control of *Arabidopsis* SSU promoter and tobacco SSU terminator. RB is right border, LB is left border.



SUPPLEMENTARY FIGURE 2 *crylAc* expressing cultivated chickpea backcross populations grown under contained conditions during 2017-18. (A) BC_1F_3 derived from Cross B (L552 × BS 100E). (B) Closer view of BC_1F_3 plant. (C) Non-transgenic recipient parent L552. (D) Transgenic donor parent BS 100E. (E) BC_2F_2 derived from Cross C (PBG7 × BS 100E). (F) Closer view of BC_2F_2 plant. (G) Non-transgenic recipient parent PBG7.



SUPPLEMENTARY FIGURE 3 Foreground selection of BC_1F_1 population derived from Cross A (PBG7 × BS 100B) through PCR using *cry1Ac* specific primers. P₁ indicates non-transgenic recipient parent PBG7; P₂ represents transgenic donor parent BS 100B; C refers to control PCR reaction without template DNA; the numbers 1 to 130 denote BC_1F_1 plants; M represents 50 bp DNA ladder (Cat. No. DM1100, Smobio Technology, Inc., Taiwan); forty six plants, namely 4, 5, 6, 7, 8, 14, 16, 17, 18, 21, 22, 24, 27, 28, 30, 35, 36, 42, 44, 52, 59, 60, 68, 72, 73, 77, 78, 81, 83, 84, 86, 87, 88, 89, 90, 93, 97, 100, 101, 102, 105, 106, 108, 119, 121 and 122 carried *cry1Ac* gene.



SUPPLEMENTARY FIGURE 4 Foreground selection of BC_1F_1 population derived from Cross B (L552 × BS 100E) through PCR using *cry1Ac* specific primers. P₁ indicates non-transgenic recipient parent L552; P₂ represents transgenic donor parent BS 100E; C refers to control PCR reaction without template DNA; the numbers 1 to 50 denote BC_1F_1 plants; M represents 50 bp DNA ladder (Cat. No. DM1100); twenty five plants, namely 1, 2, 4, 5, 7, 8, 10, 11, 12, 15, 24, 25, 28, 31, 33, 34, 36, 40, 41, 42, 44, 45, 46, 49 and 50 carried *cry1Ac* gene.

 $M \hspace{0.1in} P_1 \hspace{0.1in} P_2 \hspace{0.1in} C \hspace{0.1in} 1 \hspace{0.1in} 2 \hspace{0.1in} 3 \hspace{0.1in} 4 \hspace{0.1in} 5 \hspace{0.1in} 6 \hspace{0.1in} 7 \hspace{0.1in} 8 \hspace{0.1in} 9 \hspace{0.1in} 10 \hspace{0.1in} 11 \hspace{0.1in} 12 \hspace{0.1in} 13 \hspace{0.1in} 14 \hspace{0.1in} 15 \hspace{0.1in} 16 \hspace{0.1in} 17 \hspace{0.1in} 18 \hspace{0.1in} 19 \hspace{0.1in} 20 \hspace{0.1in} 21 \hspace{0.1in} 22 \hspace{0.1in} 2$



SUPPLEMENTARY FIGURE 5 Bioassay of BC_1F_1 plant derived from Cross A (PBG7 × BS 100B) expressing Cry1Ac for toxicity to *H. armigera* through detached leaf method. (A) Twig from BC_1F_1 plant showing healthy leaflets and mortality of neonate larvae. Arrows point dead larvae. (B) Twig from non-transgenic recipient parent PBG7 showing damage on the leaflets and survival of larva.



SUPPLEMENTARY FIGURE 6 Foreground selection of BC_1F_2 population derived from Cross A (PBG7 × BS 100B) through PCR using *cry1Ac* specific primers. P₁ indicates non-transgenic recipient parent PBG7; P₂ represents transgenic donor parent BS 100B; C refers to control PCR reaction without template DNA; the numbers 1 to 190 denote BC_1F_2 plants; M represents 50 bp DNA ladder (Cat. No. DM1100); sixteen plants, namely 6, 8, 12, 17, 18, 38, 40, 45, 47, 51, 53, 58, 88, 89, 90 and 94 carried *cry1Ac* gene.



SUPPLEMENTARY FIGURE 7 Foreground selection of BC_1F_2 population derived from Cross B (L552 × BS 100E) through PCR using *crylAc* specific primers. P₁ refers non-transgenic recipient parent L552; P₂ indicates transgenic donor parent BS 100E; C refers to control; the numbers 1 to 17 represent BC₁F₂ plants; M denotes 50 bp DNA ladder (Cat. No. DM1100); thirteen plants, namely 1, 2, 3, 6, 8, 9, 10, 12, 13, 14, 15, 16 and 17 carried *crylAc* gene.

- M P₁ P₂ C 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 M

SUPPLEMENTARY FIGURE 8 Assessment of parental polymorphism through PCR using SSR markers. Lanes 1, 2 showing polymorphism using GA 6 marker; lanes 3, 4 displaying polymorphism with TA 59; lanes 5, 6 revealing polymorphism by GA 20 marker; lanes 7, 8 exhibiting polymorphism with GAA 40 marker; lanes 9, 10 displaying polymorphism with GAA 41 marker; lanes 11, 12 revealing polymorphism by TA 146 marker; lanes 13, 14 exhibiting polymorphism with CGMM 008 marker; lanes 15, 16 showing polymorphism using CGMM 016 marker; lanes 17, 18 displaying polymorphism by CGMM 022 marker; lanes 19, 20 revealing polymorphism with TA 34 marker; lanes 21, 22 exhibiting polymorphism using TA 64 marker; lanes 23, 24 displaying polymorphism with TAASH marker. P_1 indicates non-transgenic recipient parent PBG7; P_2 represents transgenic donor parent BS 100E.



| Year | Activity | Parent/Cross (Female × Male) | Seeds sown | Plants obtained | | Popula | | Designation | | |
|---------|---|--|------------|-----------------|---------------|--------|---------|------------------------|-----------|---|
| | | | (Number) | (Number) | FS | ELISA | SA BS P | | IB | of harvested seeds |
| 2013-14 | Seed multiplication | Male parent BS 100B | 15 | 10 | × | × | × | × | × | Transgenic |
| | | Male parent BS 100E | 15 | 11 | × | × | × | × | × | donor parents |
| | | Female parent PBG7 | 15 | 14 | × | × | × | × | × | Non-transgenic |
| | | Female parent L552 | 15 | 13 | × | × | × | × | × | recipient parents |
| | Generation of crosses | PBG7 × BS 100B (55) | - | - | × | × | × | × | × | Cross A-F ₁ |
| | | $L552 \times BS \ 100E \ (35)$ | - | - | × | × | × | × | × | Cross B-F ₁ |
| | | PBG7 × BS 100E (35) | - | - | × | × | × | × | × | Cross $C-F_1$ |
| 2014.15 | | | 10 | - | | | | | 1.573 | |
| 2014-15 | Raising of F_1 plants | $Cross A-F_1$ | 13 | 7 | × | × | × | х | N[/] | Cross $A-F_2$ |
| | | Cross $B-F_1$ | 10 | 7 | × | × | × | × | N[/] | Cross $B-F_2$ |
| | | Cross C-F ₁ | 0 | 3 | × | Х | × | X | √[3] | Cross C-F ₂ |
| | Generation of 1 st backcross | Cross A-F ₁ × PBG7 (250) | - | - | × | × | × | × | × | Cross A-BC ₁ F ₁ |
| | | Cross $B-F_1 \times L552$ (125) | - | - | × | × | × | × | × | Cross $B-BC_1F_1$ |
| | | Cross C- $F_1 \times PBG7$ (75) | - | - | × | × | × | × | × | Cross C-BC ₁ F ₁ |
| 2015-16 | Raising of BC.F. | Cross A-BC, F. | 150 | 130 | √[46] | √[13] | ~ | √[13] | √[13] | Cross A-BC ₁ E ₂ |
| 2015 10 | | Cross B-BC ₁ F ₁ | 55 | 50 | $\sqrt{[25]}$ | √[9] | ~ | √[9] | √[7] | Cross B-BC ₁ F ₂ |
| | | Cross C-BC ₁ F_1 | 30 | 18 | × | × | × | √[5] | √[4] | $\frac{Cross}{C} \frac{D}{B} \frac{D}{C_1} \frac{D}{F_2}$ |
| | Generation of 2 nd backcross | Cross C-BC ₁ $F_1 \times PBG7$ (70) | - | - | × | × | × | × | × | Cross C-BC ₂ F ₁ |
| 2016 17 | Daising of PC F | Cross A DC E | 280 | 100 | 1[16] | | | | | Cross A DC E |
| 2010-17 | Kaising of $BC_1\Gamma_2$ | Cross B BC E | 200 | 190 | $\sqrt{10}$ | × | × | × √ [0] | × √[0] | Closs A- BC_1F_3 |
| | | $CIOSS B-BC_1\Gamma_2$ | 30 | 17 | v[13] | X | X | ۲[۶] | v[9] | Closs D-DC ₁ P ₃ |
| _ | Raising of BC ₂ F ₁ | Cross C-BC ₂ F ₁ | 31 | 31 | × | × | × | × | × | Cross C-BC ₂ F ₂ |
| 2017-18 | Raising of BC ₁ F ₃ | Cross A-BC ₁ F ₃ | 210 | 201 | × | × | × | × | × | Cross A-BC ₁ F ₄ |
| | | Cross $B-BC_1F_3$ | 27 | 26 | × | Х | × | √[6] | √[2] | Cross B-BC ₁ F ₄ |
| | Raising of BC ₂ F ₂ | Cross C-BC ₂ F ₂ | 120 | 83 | √[10] | × | √[10] | $\sqrt{[7]^{\Delta}}$ | √[5] | Cross C-BC ₂ F ₃ |
| 2018-19 | Raising of BC ₂ F ₃ | Cross C-BC ₂ F ₃ | 128 | 128 | √[106] | × | × | $\sqrt{[12]^{\Delta}}$ | √[9] | Cross C-BC ₂ F ₄ |

SUPPLEMENTARY TABLE 1 Introgression of *crylAc* from pod borer resistant transgenic chickpea lines into two elite commercial cultivars through marker assisted backcross breeding.

FS, Foreground Selection; ELISA, Enzyme Linked Immunosorbent Assay; BS, Background Selection; P, Phenotype; IB, Insect Bioassay; figures in round brackets are number of pollinations attempted. Figures in square brackets are number of plants identified after analysis; $\sqrt{}$, Yes; \times , No; ^{Δ} Represents plants identified on basis of agronomic traits.

 $\label{eq:supplementary table 2} \begin{array}{l} \textbf{SUPPLEMENTARY TABLE 2} \\ \textbf{Estimation of Cry1Ac concentration in leaf tissues of BC_1F_1 populations} \\ \textbf{derived from Cross A (PBG7 \times BS 100B) and Cross B (L552 \times BS 100E) through ELISA.} \end{array}$

| Crv1Ac calibrator/Leaf tissue sample | | Optical density | | | | |
|--|----------------|-----------------|----------------|--|--|--|
| orgine canorator/neur assue sample | R ₁ | R ₂ | R ₃ | | | |
| Negative control (NC) | 0.05 | 0.05 | 0.05 | | | |
| 1.5 ppb Cry1Ac calibrator (C1) | 0.45 | 0.45 | 0.45 | | | |
| 10 ppb Cry1Ac calibrator (C2) | 1.95 | 1.98 | 1.96 | | | |
| 25 ppb Cry1Ac calibrator (C3) | 3.50 | 3.56 | 3.53 | | | |
| BC ₁ F ₁ plant number (derived from Cross A) | | | | | | |
| 4 | 3.98 | 3.99 | 3.99 | | | |
| 6 | 3.99 | 3.99 | 3.99 | | | |
| 7 | 3.86 | 3.86 | 3.86 | | | |
| 16 | 3.98 | 3.98 | 3.98 | | | |
| 17 | 3.99 | 3.99 | 3.99 | | | |
| 18 | 3.99 | 3.99 | 3.99 | | | |
| 21 | 3.95 | 3.96 | 3.96 | | | |
| 22 | 3.86 | 3.89 | 3.87 | | | |
| 24 | 3.97 | 3.96 | 3.97 | | | |
| 77 | 3.97 | 3.97 | 3.97 | | | |
| 81 | 3.88 | 3.88 | 3.88 | | | |
| 89 | 3.97 | 3.97 | 3.97 | | | |
| 90 | 3.79 | 3.78 | 3.79 | | | |
| BS 100B | 3.89 | 3.88 | 3.88 | | | |
| PBG7 | 0.05 | 0.05 | 0.05 | | | |
| BC ₁ F ₁ plant number (derived from Cross B) | | | | | | |
| 1 | 3.96 | 3.96 | 3.96 | | | |
| 2 | 3.95 | 3.95 | 3.95 | | | |
| 4 | 3.93 | 3.93 | 3.93 | | | |
| 25 | 3.93 | 3.93 | 3.93 | | | |
| 34 | 3.98 | 3.98 | 3.98 | | | |
| 36 | 3.96 | 3.96 | 3.96 | | | |
| 41 | 3.98 | 3.98 | 3.98 | | | |
| 42 | 3.93 | 3.93 | 3.93 | | | |
| 45 | 3.94 | 3.93 | 3.94 | | | |
| BS 100E | 3.98 | 3.96 | 3.97 | | | |
| L552 | 0.05 | 0.05 | 0.05 | | | |

Step 2: Calculate subtracted values (calculate mean OD of NC and subtract it from OD of each calibrator and leaf tissue sample)

| Cry1Ac calibrator/Leaf tissue sample | Su | ibtracted | values |
|--|-----------------------|----------------|----------------|
| | R ₁ | \mathbf{R}_2 | R ₃ |
| NC | 0 | 0 | 0 |
| C1 | 0.40 | 0.40 | 0.40 |
| C2 | 1.90 | 1.93 | 1.91 |
| C3 | 3.45 | 3.51 | 3.48 |
| BC ₁ F ₁ plant number (derived from Cross A) | | | |
| 4 | 3.93 | 3.94 | 3.94 |
| 6 | 3.94 | 3.94 | 3.94 |
| 7 | 3.81 | 3.81 | 3.81 |
| 16 | 3.93 | 3.93 | 3.93 |
| 17 | 3.94 | 3.94 | 3.94 |
| 18 | 3.94 | 3.94 | 3.94 |
| 21 | 3.90 | 3.91 | 3.91 |
| 22 | 3.81 | 3.84 | 3.82 |
| 24 | 3.92 | 3.91 | 3.92 |
| 77 | 3.92 | 3.92 | 3.92 |
| 81 | 3.83 | 3.83 | 3.83 |
| 89 | 3.92 | 3.92 | 3.92 |
| 90 | 3.74 | 3.73 | 3.74 |
| BS 100B | 3.84 | 3.83 | 3.83 |
| PBG7 | 0 | 0 | 0 |
| BC ₁ F ₁ plant number (derived from Cross B) | | | |
| 1 | 3.91 | 3.91 | 3.91 |
| 2 | 3.90 | 3.90 | 3.90 |
| 4 | 3.88 | 3.88 | 3.88 |
| 25 | 3.88 | 3.88 | 3.88 |
| 34 | 3.93 | 3.93 | 3.93 |
| 36 | 3.91 | 3.91 | 3.91 |
| 41 | 3.93 | 3.93 | 3.93 |
| 42 | 3.88 | 3.88 | 3.88 |
| 45 | 3.89 | 3.88 | 3.89 |
| BS 100E | 3.93 | 3.91 | 3.92 |
| L552 | 0 | 0 | 0 |
| | | | 11 |

Step 3: Generation of linear scale graph of mean OD of each calibrator against its Cry1Ac concentration (first calculate mean OD of each calibrator C1, C2 and C3; then generate 'y' equation and R^2 based on mean ODs of calibrators; thereafter insert scatter chart in excel sheet)

| Cry1Ac concentration in ppb | Mean OD |
|-----------------------------|-----------|
| (x value) | (y value) |
| 1.5 (C1) | 0.45 |
| 10 (C2) | 1.96 |
| 25 (C3) | 3.53 |

To generate 'y' equation (y = mx + b), where m = slope and b = y intercept), select 'x' and 'y' values given in Step 3. To generate linear scale graph, insert scatter chart in Microsoft Excel sheet:



Step 4: Estimation of Cry1Ac concentration of each calibrator and leaf tissue sample was done as follows: OD value of calibrator or leaf tissue sample (value as per Step 2) - 0.425/ 0.127

| Cry1Ac calibrator/ Leaf tissue sample | Cry1Ac concentration (ppb) | | | | |
|---|----------------------------|----------------|-----------------------|--|--|
| | R ₁ | \mathbf{R}_2 | R ₃ | | |
| NC | 0 | 0 | 0 | | |
| C1 | 0 | 0 | 0 | | |
| C2 | 11.61 | 11.85 | 11.69 | | |
| C3 | 23.82 | 24.29 | 24.05 | | |
| BC ₁ F ₁ plant number (derived from Cro | ss A) | | | | |
| 4 | 27.60 | 27.68 | 27.68 | | |
| 6 | 27.68 | 27.68 | 27.68 | | |
| 7 | 26.65 | 26.658 | 26.65 | | |
| 16 | 27.60 | 27.60 | 27.60 | | |
| 17 | 27.68 | 27.68 | 27.68 | | |
| 18 | 27.68 | 27.68 | 27.68 | | |
| 21 | 27.36 | 27.44 | 27.44 | | |
| 22 | 26.65 | 26.89 | 26.74 | | |
| 24 | 27.52 | 27.44 | 27.52 | | |
| 77 | 27.52 | 27.52 | 27.52 | | |
| 81 | 26.81 | 26.81 | 26.81 | | |
| 89 | 27.52 | 27.52 | 27.52 | | |
| 90 | 26.10 | 26.02 | 26.10 | | |
| BS 100B | 26.89 | 26.81 | 26.81 | | |
| PBG7 | 0 | 0 | 0 | | |
| BC1F1 plant number (derived from Cros | ss B) | | | | |
| 1 | 27.44 | 27.44 | 27.44 | | |
| 2 | 27.36 | 27.36 | 27.36 | | |
| 4 | 27.20 | 27.20 | 27.20 | | |
| 25 | 27.20 | 27.20 | 27.20 | | |
| 34 | 27.60 | 27.60 | 27.60 | | |
| 36 | 27.44 | 27.44 | 27.44 | | |
| 41 | 27.60 | 27.60 | 27.60 | | |
| 42 | 27.20 | 27.20 | 27.20 | | |
| 45 | 27.28 | 27.20 | 27.28 | | |
| BS 100E | 27.60 | 27.44 | 27.52 | | |
| L552 | 0 | 0 | 0 | | |

Step 5: Estimation of Cry1Ac concentration in ppm or $\mu g g^{-1}$ by applying the formula: OD value (as per Step 4) × dilution factor 1 (38.46) × dilution factor 2 (11, as samples were diluted at 1:11)/ 1000

| Cry1Ac | Cry1 | g g ⁻¹ leaf tissue) | |
|--|------------------|--------------------------------|----------------|
| calibrator/Leaf | \mathbf{R}_1 | \mathbf{R}_2 | \mathbf{R}_3 |
| tissue sample | | | |
| NC | 0 | 0 | 0 |
| C1 | 0 | 0 | 0 |
| C2 | 4.91 | 5.01 | 4.94 |
| C3 | 10.08 | 10.28 | 10.17 |
| BC ₁ F ₁ plant nun | nber (derived fr | om Cross A) | |
| 4 | 11.68 | 11.71 | 11.71 |
| 6 | 11.71 | 11.71 | 11.71 |
| 7 | 11.27 | 11.28 | 11.27 |
| 16 | 11.68 | 11.68 | 11.68 |
| 17 | 11.71 | 11.71 | 11.71 |
| 18 | 11.71 | 11.71 | 11.71 |
| 21 | 11.57 | 11.61 | 11.61 |
| 22 | 11.27 | 11.38 | 11.31 |
| 24 | 11.64 | 11.61 | 11.64 |
| 77 | 11.64 | 11.64 | 11.64 |
| 81 | 11.34 | 11.34 | 11.34 |
| 89 | 11.64 | 11.64 | 11.64 |
| 90 | 11.04 | 11.01 | 11.04 |
| BS 100B | 11.38 | 11.34 | 11.34 |
| PBG7 | 0 | 0 | 0 |
| BC ₁ F ₁ plant nun | nber (derived fr | om Cross B) | |
| 1 | 11.61 | 11.61 | 11.61 |
| 2 | 11.57 | 11.57 | 11.57 |
| 4 | 11.51 | 11.51 | 11.51 |
| 25 | 11.51 | 11.51 | 11.51 |
| 34 | 11.68 | 11.68 | 11.68 |
| 36 | 11.61 | 11.61 | 11.61 |
| 41 | 11.68 | 11.68 | 11.68 |
| 42 | 11.51 | 11.51 | 11.51 |
| 45 | 11.54 | 11.51 | 11.54 |
| BS 100E | 11.68 | 11.61 | 11.64 |
| L552 | 0 | 0 | 0 |

SUPPLEMENTARY TABLE 3 SSR markers used to analyze parental polymorphism and carry out background selection of BC_2F_2 plants derived from Cross C (PBG7 × BS 100E).

| S. No | Marker | Status of | Reference | Chromosomal | Reference for | PIC | Reference for PIC value |
|----------|----------------------|-----------|--|-------------|-----------------------------------|-------------|-------------------------|
| 1 | | D | Cruiceria at al. (2011) | ocation | chromosomal location | value | |
| 2 | CGMM 001* | r M | Gujaria et al. (2011) Gujaria et al. (2011) | 0 | https://plantgarden.jp>list>CAPS | NA | |
| 3 | CGMM 003 | M | Gujaria et al. (2011) | | | | |
| 4 | CGMM 004 | М | Gujaria et al. (2011) | | | | |
| 5 | CGMM 005 | М | Gujaria et al. (2011) | | | | |
| 6 | CGMM 006 | М | Gujaria et al. (2011) | | | | |
| 7 | CGMM 007 | М | Gujaria et al. (2011) | | | | |
| 8 | CGMM 008 | Р | Gujaria et al. (2011) | NA | https://plantgarden.jp>list>CAPS | NA | - |
| 9 | CGMM 009 | M | Gujaria et al. (2011) | | | | |
| 10 | CGMM 010 | M | Gujaria et al. (2011) Guiaria et al. (2011) | | | | |
| 12 | CGMM 012* | P | Gujaria et al. (2011) | 2 | Guiaria et al. (2011) | NA | |
| 12 | CGMM 012 | M | Gujaria et al. (2011) | 2 | Gujaria et al. (2011) | 1471 | |
| 14 | CGMM 014 | М | Gujaria et al. (2011) | | | | |
| 15 | CGMM 015 | М | Gujaria et al. (2011) | | | | |
| 16 | CGMM 016 | Р | Gujaria et al. (2011) | 3 | https://plantgarden.jp>list>CAPS | NA | - |
| 17 | CGMM 017 | М | Gujaria et al. (2011) | | | | |
| 18 | CGMM 018 | М | Gujaria et al. (2011) | | | | |
| 19 | CGMM 020 | М | Gujaria et al. (2011) | | | | |
| 20 | CGMM 021 | M | Gujaria et al. (2011) | ~ | https://slagt_l_t_t_t_t_t_t_t_t_t | NT 4 | |
| 21 | CGMM 022 | ۲ ۲ | Gujaria et al. (2011) | 5 | nttps://plantgarden.jp>list>CAPS | NA | - |
| 22 | CGMM 023 | M | Gujaria et al. (2011) | | | | |
| 23 24 | CGMM 024 CGMM 025 | M | Gujaria et al. (2011) Gujaria et al. (2011) | | | | |
| 24 25 | CGMM 025 | M | Gujaria et al. (2011) Gujaria et al. (2011) | | | | |
| 26 | CGMM 027 | M | Gujaria et al. (2011) | | | | |
| 27 | CGMM 028 | M | Gujaria et al. (2011) | | | | |
| 28 | CGMM 029 | М | Gujaria et al. (2011) | | | | |
| 29 | CGMM 030 | М | Gujaria et al. (2011) | | | | |
| 30 | CGMM 031 | М | Gujaria et al. (2011) | | | | |
| 31 | CGMM 032 | М | Gujaria et al. (2011) | | | | |
| 32 | CGMM 033 | М | Gujaria et al. (2011) | | | | |
| 33 | CGMM 062 | M | Gujaria et al. (2011) | | | | |
| 34 | CGMM 063 | M | Gujaria et al. (2011) | | | | |
| 35 36 | CGMM 064 | M | Gujaria et al. (2011) Gujaria et al. (2011) | | | | |
| 30 | CGMM 066 | M | Gujaria et al. (2011) | | | | |
| 38 | CGMM 067 | M | Gujaria et al. (2011) | | | | |
| 39 | CGMM 068 | M | Gujaria et al. (2011) | | | | |
| 40 | CGMM 069 | М | Gujaria et al. (2011) | | | | |
| 41 | CGMM 070 | М | Gujaria et al. (2011) | | | | |
| 42 | CGMM 071 | М | Gujaria et al. (2011) | | | | |
| 43 | CGMM 072 | М | Gujaria et al. (2011) | | | | |
| 44 | CGMM 073 | М | Gujaria et al. (2011) | | | | |
| 45 | CGMM 074 | M | Gujaria et al. (2011) | | | | |
| 46 | CGMM 075 | M | Gujaria et al. (2011) | | | | |
| 4/ 18 | CGMM 077 | M | Gujaria et al. (2011) Gujaria et al. (2011) | | | | |
| 40 49 | CGMM 077 | M | Gujaria et al. (2011) | | | | |
| 50 | CGMM 138 | M | Gujaria et al. (2011) | | | | |
| 51 | CaM 0038 | М | Thudi et al. (2011) | | | | |
| 52 | CaM 0046 | М | Thudi et al. (2011) | | | | |
| 53 | CaM 0244 | М | Thudi et al. (2011) | | | | |
| 54 | CaM 0594 | М | Thudi et al. (2011) | | | | |
| 55 | CaM 0805 | М | Thudi et al. (2011) | | | | |
| 56 | CaM 1101 | M | Thudi et al. (2011) | - | | | |
| 57 | CaM 1125* | Р | Thudi et al. (2011) | 6 | Thudi et al. (2011) | NA | |
| 58 50 | CaM 1502 | IVI M | Thuch et al. (2011) | | | | |
| 59 60 | CaM 1002* | M D | Thuch et al. (2011) | А | Nevel (2010) | 0.52 | Sachdeva et al. (2019) |
| 61 | CaM 2049 | r M | Thudi et al. (2011) Thudi et al. (2011) | 4 | INAYAK (2010) | 0.52 | Sachueva et al. (2018) |
| 62 | GA 2 | M | Winter et al. (1992) | | | | |
| 63 | GA 4 | М | Winter et al. (1992) | | | | |
| 64 | GA 6 | Р | Winter et al. (1992) | 8 | Millan et al. (2010) | 0.47 | Sachdeva et al. (2018) |
| 65 | GA 8* | Р | Winter et al. (1992) | NA | | 0.24 | Vashist et al. (2019) |
| 66 | GA 9 | М | Winter et al. (1992) | | | | |
| 67 | GA 11 | М | Winter et al. (1992) | | | | |
| 68 | GA 13 | М | Winter et al. (1992) | | | | |
| 69 | GA 14 | М | Winter et al. (1992) | | | | |
| 70 | GA 17 | М | Winter et al. (1992) | | | | |
| 71 | GA 20 | Р | Winter et al. (1992) | 2 | Millan et al. (2010) | 0.49 | Ghaffari et al. (2014) |
| 72 | GA 21 | М | Winter et al. (1992) | | | | |

| 73 | GA 22 | М | Winter et al. (1992) | | | | |
|------------|------------------|---|--|----|--|------|---|
| 74 | GA 26* | Р | Winter et al. (1992) | 6 | Winter et al. (2000) | 0.61 | Jha et al. (2018) |
| 75 | GA 31 | М | Winter et al. (1992) | | | | |
| 76 | GA 33 | М | Winter et al. (1992) | | | | |
| 77 | GA 105 | М | Winter et al. (1992) | | | | |
| 78 | GA 108 | M | Winter et al. (1992) | | | | |
| 79 | GA 117 | M | Winter et al. (1992) | | | | |
| 80 | GA 13/ | M | Winter et al. (1992) | 6 | N1- (2010) | 0.20 | Washist at al. (2010) |
| 81 | GAA 39* | P | Winter et al. (1992) | 0 | Nayak (2010) Winter at al. (2000) | 0.30 | Vasnist et al. (2019) Sofore et al. (2011) |
| 02 | GAA 40 | P | winter et al. (1992) | I | winter et al. (2000) | 0.37 | Selera et al. (2011) |
| 83 | GAA 41 | Р | Winter et al. (1992) | 6 | Nayak et al. (2010) | 0.82 | Ganguly et al. (2008 |
| 84 | GAA 42 | M | Winter et al. (1992) | | | | |
| 85 | GAA 43 | M | Winter et al. (1992) | | | | |
| 86 | GAA 44 | М | Winter et al. (1992) | | | | |
| 87 | GAA 45* | Р | Winter et al. (1992) | 3 | Jadhav et al. (2015) | 0.03 | Jadhav et al. (2015) |
| 88 | GAA 46 | M | Winter et al. (1992) | | | | |
| 09 | GAA 50 | M | Winter et al. (1992) | | | | |
| 90 01 | GAA 51 | M | Winter et al. (1992) | | | | |
| 91 | GAA 58 | M | Winter et al. (1992) | | | | |
| 03 | GAA 60 | M | Winter et al. (1992) | | | | |
| 93 | GAA 129h | M | Winter et al. (1992) | | | | |
| 94 | TA 2 | M | Winter et al. (1992) | | | | |
| 96 | TA 18 | M | Winter et al (1992) | | | | |
| 97 | TA 20 | M | Winter et al. (1992) | | | | |
| 98 | TA 25 | M | Winter et al. (1992) | | | | |
| 99 | TA 28 | M | Winter et al. (1992) | | | | |
| 100 | TA 34 | Р | Winter et al. (1992) | 3 | Winter et al. (2000) | 0.39 | Choudhary et al. (2012 |
| 101 | TA 36 | М | Winter et al. (1992) | - | | | |
| 102 | TA 37 | M | Winter et al. (1992) | | | | |
| 103 | TA 43 | M | Winter et al. (1992) | | | | |
| 104 | TA 44 | М | Winter et al. (1992) | | | | |
| 105 | TA 45 | М | Winter et al. (1992) | | | | |
| 106 | TA 46 | М | Winter et al. (1992) | | | | |
| 107 | TA 47 | М | Winter et al. (1992) | | | | |
| 108 | TA 53 | М | Winter et al. (1992) | | | | |
| 109 | TA 59 | Р | Winter et al. (1992) | 2 | Winter et al. (2000) | 0.36 | Choudhary et al. (2012 |
| 110 | TA 64 | Р | Winter et al. (1992) | 3 | Winter et al. (2000) | 0.34 | Choudhary et al. (2012 |
| 111 | TA 66 | М | Winter et al. (1992) | | | | |
| 112 | TA 71 | M | Winter et al. (1992) | | | | |
| 113 | TA 72 | М | Winter et al. (1992) | | | | |
| 114 | TA 76s* | Р | Winter et al. (1992) | NA | Sefera et al. (2011) | 0.57 | Rizvi et al. (2013) |
| 115 | TA 78 | Μ | Winter et al. (1992) | | | | |
| 116 | TA 80 | М | Winter et al. (1992) | | | | |
| 117 | TA 87 | М | Winter et al. (1992) | | | | |
| 118 | TA 89 | Μ | Winter et al. (1992) | | | | |
| 119 | TA 93 | Μ | Winter et al. (1992) | | | | |
| 120 | TA 103 | М | Winter et al. (1992) | | | | |
| 121 | TA 104 | М | Winter et al. (1992) | | | | |
| 122 | TA 106 | М | Winter et al. (1992) | | | | |
| 123 | TA 108 | М | Winter et al. (1992) | | | | |
| 124 | TA 113 | M | Winter et al. (1992) | | | | |
| 125 | TA 114 | M | Winter et al. (1992) | | | | |
| 126 | TA 116 | M | Winter et al. (1992) | | | | |
| 127 | TA 117 | M | Winter et al. (1992) | | | | |
| 128 | 1A 118 TA 127 | M | Millan et al. (2010) | | | | |
| 129 | TA 125 | M | Winter et al. (1992) | | | | |
| 130 | TA 127 | M | Winter et al. (1992) | | | | |
| 131 | 1A 155 | M | Winter et al. (1992) | | | | |
| 132 | TA 130 | M | Winter et al. (1992) | | | | |
| 133 | TA 140 | M | Winter et al. (1992) | | | | |
| 134 | TA 141 | M | Winter et al. (1992) | | | | |
| 135 | TA 142 | M | Winter et al. (1992) Winter et al. (1002) | | | | |
| 137 | TA 144 | P | Winter et al. (1992) | 4 | Winter et al. (2000) | 0.72 | Choudhary et al. (2017 |
| 130 | TA 140 | м | | - | (1) mor et al. (2000) | 0.72 | Choudhary et al. (201) |
| 130 | TA 139 | M | Winter at al. (2010) | | | | |
| 139 140 | TA 10/ | M | Winter et al. (1992) Winter et al. (1002) | | | | |
| 140 | TA 170 | M | Winter et al. (1992) Winter et al. (1002) | | | | |
| 142 | TA 180 | M | Winter et al. (1992) Winter et al. (1002) | | | | |
| 142 | TA 186 | M | Winter et al. (1992) | | | | |
| 144 | TA 199 | M | Winter et al. (1992) | | | | |
| 145 | TA 200 | M | Winter et al (1992) | | | | |
| 146 | TA 203 | M | Winter et al. (1992) | | | | |
| 147 | TA 206 | M | Winter et al. (1992) | | | | |
| 1/9 | TAA 55 | M | Winter et al. (1992) | | | | |
| 140 | | | | | | | |
| 148 | TAA 57 | M | Winter et al. (1992) | | | | |

| 151 | TAA 61* | Р | Winter et al. (1992) | NA | | NA | |
|-----|-------------------|--------|---|-----|-----------------------|-------|----------------------|
| 152 | TAA 104 | М | Winter et al. (1992) | | | | |
| 153 | TAA 107 | М | Winter et al. (1992) | | | | |
| 154 | TAA 137 | М | Winter et al. (1992) | | | | |
| 155 | TAA 169 | М | Winter et al. (1992) | | | | |
| 156 | TAA 194 | M | Winter et al. (1992) | | | | |
| 157 | TAASH | P | Winter et al. (1992) | 5 | Winter et al. (2000) | 0.86 | Joshi et al. (2013) |
| 158 | TS 5 | M | Winter et al. (1992) | 5 | (1 intel et un (2000) | 0.000 | 20011 et al. (2015) |
| 150 | TS 10 | M | Winter et al. (1992) | | | | |
| 159 | TS 10 | M | Winter et al. (1992) | | | | |
| 161 | TS 12 | M | Winter et al. (1992) | | | | |
| 162 | TS 16* | D | Winter et al. (1992) | NA | | NA | |
| 162 | TS 10 | r M | Winter et al. (1992) | INA | | INA | |
| 164 | TS 22 | M | Winter et al. (1992) | | | | |
| 165 | TS 23 | M | Winter et al. (1992) | | | | |
| 165 | TS 20 | M | Winter et al. (1992) | | | | |
| 167 | TS 25 | M | Winter et al. (1992) | | | | |
| 167 | 15 55 TE 20 | M | Winter et al. (1992) | | | | |
| 108 | 15 39 TE 42 | M | Winter et al. (1992) | | | | |
| 109 | 1545 | M D | Winter et al. (1992) | 0 | Sefere et al. (2011) | 0.00 | Sefere et el (2011) |
| 170 | 15 45* TC 47 | P | Winter et al. (1992) | 8 | Selera et al. (2011) | 0.66 | Selera et al. (2011) |
| 1/1 | 154/ | M | Winter et al. (1992) | | | | |
| 172 | 15 52 TE 52 | M | Winter et al. (1992) | | | | |
| 175 | 15 55 | N | Winter et al. (1992) | | N 1 (2010) | 0.01 | W 1 (2000) |
| 174 | 15 54* | P | Winter et al. (1992) | 4 | Nayak (2010) | 0.81 | winter et al. (2000) |
| 175 | 15 58 | M | Winter et al. (1992) | | | | |
| 176 | TS 72 | M | Winter et al. (1992) | | | | |
| 177 | TS 74 | M | Winter et al. (1992) | | | | |
| 1/8 | 15 /9 | M | Winter et al. (1992) | | | | |
| 1/9 | 15 83 | M | Winter et al. (1992) | | | | |
| 180 | TS 84 | M | Winter et al. (1992) | | | | |
| 181 | TS 104 | M | Winter et al. (1992) | | | | |
| 182 | IR I | M | Winter et al. (1992) | | | | |
| 183 | TR 2 | M | Winter et al. (1992) | | | | |
| 184 | TR 3 | M | Winter et al. (1992) | | | | |
| 185 | IR / | M | Winter et al. (1992) | | | | |
| 186 | IR 8 | M | Winter et al. (1992) | | | | |
| 18/ | TR 20 | M | Winter et al. (1992) | | | | |
| 188 | TR 26 | M | Winter et al. (1992) | | | | |
| 189 | TR 29 | M | Winter et al. (1992) | | | | |
| 190 | TR 31 | M | Winter et al. (1992) | | | | |
| 191 | TR 32 | M | Winter et al. (1992) | | | | |
| 192 | TR 35 | M | Winter et al. (1992) | | | | |
| 195 | TR 55 | M | Winter et al. (1992) | | | | |
| 194 | TR 40 TR 42 | M | Winter et al. (1992) | | | | |
| 195 | TR 45 | M | Winter et al. (1992) | | | | |
| 190 | 1 K 44 TD 45 | M | Winter et al. (1992) | | | | |
| 197 | TR 45 | M | Winter et al. (1992) | | | | |
| 198 | TR 55 | M | Winter et al. (1992) | | | | |
| 200 | TR 50 | M | Winter et al. (1992) | | | | |
| 200 | TR 39 | M | Winter et al. (1992) | | | | |
| 201 | IK 00 NCDCD 21 | M | winner et al. (1992) | | | | |
| 202 | NCPGP 127 | M | Varshnav et al. (2000) | | | | |
| 205 | NCPGP 141 | M | v arshney et al. (2014) Varshney et al. (2014) | | | | |
| 204 | NCPGP 171 | M | v arshney et al. (2014) Varshney et al. (2014) | | | | |
| 205 | NCPGP 247 | M | Varshney et al. (2014) | | | | |
| 200 | CaSTMS 11 | M | * arsinicy of al. (2014) Hüttel et al. (1000) | | | | |
| 207 | HIII6 | M | Lichtenzveig et al. (2005) | | | | |
| 200 | H1A10 | M | Lichtenzveig et al. (2005) | | | | |
| 209 | H4G11 | M | Lichtenzveig et al. (2005) | | | | |
| 210 | 110011 | 111 | Element verg et al. (2003) | | | | |

M, Monomorphic marker; P, Polymorphic marker; markers in bold depict polymorphic markers that had high reproducibility and were used to carry out background selection; *represents polymorphic markers that had low reproducibility; NA, Not Available.

- Jadhav, A. A., Rayate, S. J., Mhase, L. B., Thudi, M., Chitikineni, A., Harer, P. N., Jadhav, A. S., Varshney, R., and Kulwal, P. (2015). Marker-trait association study for protein content in chickpea (*Cicer arietinum* L.). J. Genet. 94, https://doi.org/10.1007/s12041-015-0529-6.
- Choudhary, S., Kaur, J., Chhuneja, P., Sandhu, J. S., Singh, I., Singh, S., and Sirari, A. (2012). Assessment of genetic diversity in *kabuli* chickpea (*Cicer arietinum* L.) genotypes in relation to seed size using SSR markers. *Journal of Food Legumes* 26, 1–4.
- Ganguly, A. K., Chawla, G., Yadav, R., and Kumar, R. (2008). STMS profiling of chickpea (*Cicer arietinum*) with regards to nematode resistance. *Indian J. Nematol.* 38, 209–217.

- Ghaffari, P., Talebi, R., and Keshavarzi, F. (2014). Genetic diversity and geographical differentiation of Iranian landrace, cultivars, and exotic chickpea lines as revealed by morphological and microsatellite markers. *Physiol. Mol. Biol. Plants* 20, 225–233.
- Gujaria, N., Kumar, A., Dauthal, P., Dubey, A., Hiremath, P., Prakash, A. B., Farmer, A., Bhide, M., Shah, T., Gaur, P. M., Upadhyaya, H. D., Bhatia, S., Cook, D. R., May, G. D., and Varshney, R. K. (2011). Development and use of genic molecular markers (GMMs) for construction of a transcript map of chickpea (*Cicer arietinum L.*). *Theor. Appl. Genet.* 122, 1577–1589.
- Rizvi, H., Babu, B. K., and Agrawal, P. K. (2013). Molecular analysis of *kabuli* and *desi* type of Indian chickpea (*Cicer arietinum* L.) cultivars using STMS markers. J. Plant Biochem. Biotech., https://doi.org/10.1007/s13562-012-0187-1.
- Hüttel, B., Winter, P., Weising, K., Choumane, W., Weigand, F., and Kahl, G. (1999). Sequence-tagged microsatellite site markers for chickpea (*Cicer arietinum* L.). *Genome* 42, 210–217.
- Lichtenzveig, J., Scheuring, C., Dodge, J., Abbo, S., and Zhang, H. B. (2005). Construction of BAC and BIBAC libraries and their applications for generation of SSR markers for genome analysis of chickpea, *Cicer arietinumL. Theor. Appl. Genet.* 110, 492–510.
- Millan, T., Winter, P., Jüngling, R., Gil, J., Rubio, J., Cho, S., Cobos, M. J., Iruela, M., Rajesh, P. N., Tekeoglu, M., Kahl, G., and Muehlbauer, F. J. (2010). A consensus genetic map of chickpea (*Cicer arietinum* L.) based on 10 mapping populations. *Euphytica* 175, 175–189.
- Sethy, N. K., Shokeen, B., Edwards, K. J., and Bhatia, S. (2006). Development of microsatellite markers and analysis of intraspecific genetic variability in chickpea (*Cicer arietinum L.*). *Theor. Appl. Genet.* 112, 1416–1428.
- Nayak, S. N. (2010). Identification of QTLs and genes for drought tolerance using linkage mapping and association mapping approaches in chickpea (*Cicer arietinum* L.). Ph.D. thesis, Osmania University, Hyderabad, India.
- Nayak, S. N., Zhu, H., Varghese, N., Datta, S., Choi, H. K., Horres, R., Jüngling, R., Singh, J., Kishor, P. K., Sivaramakrishnan, S., Hoisington, D. A., Kahl, G., Winter, P., Cook, D. R., and Varshney, R. K. (2010). Integration of novel SSR and gene-based SNP marker loci in the chickpea genetic map and establishment of new anchor points with *Medicago truncatula* genome. *Theor. Appl. Genet*. 120, 1415–1441.
- Sachdeva, S., Bharadwaj, C., Sharma, V., Patil, B. S., Soren, K. R., Roorkiwal, M., Varshney, R. K., and Bhat, K. V. (2018). Molecular and phenotypic diversity among chickpea (*Cicer arietinum*) genotypes as a function of drought tolerance. *Crop & Pasture Sci.* 69, 142–153.
- Sefera, T., Abebie, B., Gaur, P. M., Assefa, K., and Varshney, R. K. (2011). Characterisation and genetic diversity analysis of selected chickpea cultivars of nine countries using simple sequence repeat (SSR) markers. *Crop & Pasture Sci.* 62, 177–187.
- Thudi, M., Bohra, A., Nayak, S. N., et al. (2011). Novel SSR markers from BAC-end sequences, DArT arrays and a comprehensive genetic map with 1,291 marker loci for chickpea (*Cicer arietinum*). *PLoS ONE* 6, e27275.
- Jha, U. C., Jha, R., Bohra, A., Parida, S. K., Kole, P. C., Thakro, V., Singh, D., and Singh, N. P. (2018). Population structure and association analysis of heat stress relevant traits in chickpea (*Cicer arietinum L.*). 3 Biotech 8, 43.
- Varshney, R. K., Thudi, M., Nayak, S. N., et al. (2014) Genetic dissection of drought tolerance in chickpea (*Cicer arietinum* L.). *Theor. Appl. Genet*. 127, 445–462.
- Vashist, U., Boora, K. S., and Kumar, M. (2019). Evaluation of genetic diversity among chickpea (*Cicer arietinum* L.) genotypes using PCR based simple sequence repeats markers. *The Pharma Innovation Journal* 8, 182–188.
- Winter, P., Benko-Iseppon, A. M., Hüttel, B., Ratnaparkhe, M., Tullu, A., Sonnante, G., Pfaff, T., Tekeoglu, M., Santra, D., Sant, V. J., Rajesh, P. N., Kahl, G., and Muehlbauer, F. J. (2000). A linkage map of the chickpea (*Cicer arietinum* L.) genome based on recombinant inbred lines from a *C. arietinum* × *C. reticulatum* cross: localization of resistance genes for *Fusarium* wilt races 4 and 5. *Theor. Appl. Genet.* 101, 1155–1163.
- Winter, P., Pfaff, T., Udupa, S. M., Hüttel, B., Sharma, P. C., Sahi, S., Arreguin-Espinoza, R., Weigand, F., Muehlbauer, F. J., and Kahl, G. (1992). Characterization and mapping of sequence-tagged microsatellite sites in the chickpea (*C. arietinum* L.) genome. *Mol. Gen. Genet.* 262, 90–101.
- Joshi, N., Sharma, S., Subramanian, R. B., and Rao, K. S. (2013). Genetic fingerprinting of chickpea (*Cicer arietinum* L.) germplasm using morphological and molecular markers. *Asian J. Exp. Biol. Sci.* 4, 398–405.

| Marker | Plant number | | | | | | | | | |
|--------------------------------------|--------------|------|------|------|------|------|------|------|------|------|
| name | 1 | 2 | 8 | 9 | 12 | 20 | 26 | 33 | 39 | 44 |
| CGMM 008 | - | А | А | А | А | А | А | А | - | А |
| CGMM 016 | А | А | А | А | А | А | А | А | А | А |
| CGMM 022 | А | А | А | А | А | А | А | А | А | А |
| GA 6 | - | А | А | А | А | А | А | А | А | А |
| GA 20 | А | А | А | А | А | А | А | А | А | А |
| GAA 40 | А | - | А | А | - | - | - | - | - | - |
| GAA 41 | А | А | А | А | А | А | А | А | А | А |
| TA 34 | А | А | А | А | А | А | А | А | А | А |
| TA 59 | А | - | А | А | А | А | - | А | А | А |
| TA 64 | А | А | А | А | А | А | А | А | А | А |
| TA 146 | А | - | А | А | А | А | - | А | А | А |
| TAASH | А | А | А | А | А | А | А | А | А | А |
| <i>cry1Ac</i> specific PCR primer | В | В | В | В | В | В | В | В | В | В |
| A (%) | 90.9 | 90.0 | 92.3 | 92.3 | 91.7 | 91.7 | 90.0 | 91.7 | 90.9 | 91.7 |

SUPPLEMENTARY TABLE 4 Recurrent parent genome recovery in *cry1Ac* positive BC_2F_2 plants derived from Cross C (PBG7 × BS 100E) using polymorphic SSR markers.

'A' represents presence of allele corresponding to recurrent parent PBG7,

'B' denotes presence of allele for donor parent BS 100E.

| S No | Plant | - | 0 | | - 2 JF - F | Agron | omic trait | (| | | | Recurrent |
|-----------|---------------|------------------|--------------------|-------------------|------------------|------------------|-------------------|------------------|---------------------------------------|----------------------|----------------------------|---------------------------------------|
| 5. 140. | number/ | Dave to 50 % | Number of branches | Dave to maturity | Plant height | Number of pode | Number of seeds | 100-cood woight | Biological viald | Seed vield per plant | Harvest Indev [€] | narent nhenome |
| | Parent | flowering | ner nlant | Days to maturity | (cm) | ner nlant | ner nlant | (g) | | (g) | (%) | recovery (%) |
| DOE | | nowering | per pluit | | (em) | per plant | per plant | (5) | (5) | (5) | (70) | recovery (70) |
| BC_2F_2 | | | | | | | | | | | | |
| 1 | 1 | 84 (93.33) | 13 (76.47) | 149 (96.13) | 51.3 (86.51) | 44 (73.33) | 80 (70.80) | 15.6 (93.97) | 40.22 (77.97) | 13.21 (76.36) | 32.84 (97.91) | 84.28 |
| 2 | 2 | 89 (98.89) | 15 (88.23) | 153 (98.71) | 55.5 (93.59) | 55 (91.67) | 102 (90.26) | 15.5 (93.37) | 47.16 (91.43) | 15.45 (89.31) | 32.76 (97.67) | 93.31 |
| 3 | 8 | 87 (96.67) | 16 (94.12) | 152 (98.06) | 57.5 (96.96) | 46 (76.67) | 85 (75.22) | 16.2 (97.59) | 44.36 (86.00) | 14.12 (81.62) | 31.83 (94.90) | 89.78 |
| 4 | 9 | 88 (97.78) | 12 (70.59) | 154 (99.35) | 58.1 (97.98) | 43 (71.67) | 78 (69.03) | 15.8 (95.18) | 38.34 (74.33) | 12.43 (71.85) | 32.42 (96.66) | 84.44 |
| 5 | 12 | 85 (94.44) | 12 (70.59) | 151 (97.42) | 50.2 (84.65) | 49 (81.67) | 94 (83.18) | 15.6 (93.97) | 43.78 (84.88) | 14.32 (82.77) | 32.71 (97.52) | 87.11 |
| 6 | 20 | 89 (98.89) | 15 (88.23) | 153 (98.71) | 58.3 (98.31) | 53 (88.33) | 98 (86.72) | 15.1 (90.96) | 49.56 (96.08) | 15.20 (87.86) | 30.67 (91.44) | 92.55 |
| 7 | 26 | 89 (98.89) | 14 (82.35) | 154 (99.35) | 57.6 (97.13) | 59 (98.33) | 112 (99.11) | 15.2 (91.57) | 51.00 (98.87) | 16.86 (97.46) | 33.06 (97.46) | 96.05 |
| 8 | 33 | 86 (95.55) | 16 (94.12) | 150 (96.77) | 51.4 (86.68) | 54 (90.00) | 99 (87.61) | 15.3 (92.17) | 47.62 (92.32) | 15.23 (88.03) | 31.98 (95.35) | 91.86 |
| 9 | 39 | 87 (96.67) | 15 (88.23) | 151 (97.42) | 55.6 (93.76) | 57 (95.00) | 108 (95.57) | 15.3 (92.17) | 50.50 (97.91) | 16.38 (94.68) | 32.43 (96.69) | 94.81 |
| 10 | 44 | 86 (95.55) | 15 (88.23) | 151 (97.42) | 56.2 (94.77) | 58 (96.67) | 108 (95.57) | 15.3 (92.17) | 51.02 (98.91) | 16.60 (95.95) | 32.54 (97.02) | 95.23 |
| 11 | BS 100E | 82 | 8 | 145 | 48.4 | 16 | 28 | 13.3 | 23.30 | 4.13 | 17.72 | - |
| 12 | PBG7 | 90 | 17 | 155 | 59.3 | 60 | 113 | 16.6 | 51.58 | 17.30 | 33.54 | - |
| | | | | | | | | | | Average recurrent p | arent phenome reco | very - 90 94 % |
| DC E | | | | | | | | | | riterage recarrent p | arent phenome reed | , , , , , , , , , , , , , , , , , , , |
| BC21-3 | | | | | | | | | | | | |
| 1 | 2-1 | 88 | 14 | 151 | 57.5 | 51 | 98 | 15.2 | 41.14 | 13.45 | 32.69 | |
| 2 | 2-2 | 88 | 13 | 150 | 54.6 | 48 | 94 | 15.6 | 46.20 | 13.23 | 28.64 | |
| 3 | 2-3 | 89 | 15 | 151 | 55.0 | 53 | 101 | 15.3 | 45.67 | 15.28 | 33.46 | |
| | Mean \pm SD | 88.33 ± 0.58 | 14.00 ± 1.00 | 150.67 ± 0.58 | 55.70 ± 1.58 | 50.67 ± 2.52 | 97.67 ± 3.51 | 15.36 ± 0.21 | 44.34 ± 2.78 | 13.99 ± 1.12 | 31.59 ± 2.59 | _ |
| 4 | 8-1 | 85 | 15 | 151 | 55.5 | 42 | 77 | 16.0 | 43.66 | 14.42 | 33.05 | |
| 5 | 8-2 | 87 | 16 | 153 | 58.6 | 45 | 86 | 15.7 | 41.20 | 14.10 | 34.22 | |
| 6 | 8-3 | 86 | 17 | 151 | 53.3 | 46 | 87 | 15.8 | 47.35 | 14.32 | 30.24 | |
| | Mean \pm SD | 86.00 ± 1.00 | 16.00 ± 1.00 | 151.67 ± 1.15 | 55.80 ± 2.66 | 44.33 ± 2.08 | 83.33 ± 5.51 | 15.83 ± 0.15 | 44.07 ± 3.09 | 14.28 ± 0.16 | 32.50 ± 2.04 | _ |
| 7 | 20-1 | 88 | 16 | 155 | 55.3 | 54 | 99 | 15.6 | 50.56 | 15.20 | 30.06 | |
| 8 | 20-2 | 86 | 15 | 154 | 57.4 | 55 | 100 | 14.8 | 51.23 | 15.51 | 30.27 | |
| 9 | 20-3 | 88 | 16 | 154 | 59.7 | 52 | 98 | 15.7 | 54.45 | 16.20 | 29.75 | |
| | Mean \pm SD | 87.33 ± 1.15 | 15.67 ± 0.58 | 154.30 ± 0.58 | 57.47 ± 2.20 | 53.67 ± 1.53 | 99.00 ± 1.00 | 15.37 ± 0.49 | 52.08 ± 2.08 | 15.64 ± 0.51 | 30.03 ± 0.26 | _ |
| 10 | 26-1 | 88 | 14 | 152 | 55.6 | 57 | 116 | 15.5 | 55.58 | 17.25 | 31.04 | |
| 11 | 26-2 | 89 | 13 | 151 | 58.5 | 51 | 107 | 15.1 | 52.34 | 16.45 | 31.43 | |
| 12 | 26-3 | 90 | 14 | 153 | 60.3 | 54 | 110 | 14.8 | 48.46 | 15.30 | 31.57 | |
| | Mean \pm SD | 89.00 ± 1.00 | 13.67 ± 0.58 | 152.00 ± 0.27 | 58.13 ± 2.37 | 54.00 ± 3.00 | 111.00 ± 4.58 | 15.13 ± 0.35 | 52.13 ± 3.56 | 15.33 ± 0.98 | 31.35 ± 0.27 | _ |
| 13 | 33-1 | 88 | 14 | 152 | 59.3 | 47 | 84 | 15.3 | 42.30 | 13.10 | 30.97 | |
| 14 | 33-2 | 86 | 13 | 154 | 51.2 | 41 | 76 | 15.7 | 38.75 | 12.65 | 32.65 | |
| 15 | 33-3 | 86 | 15 | 152 | 55.4 | 44 | 78 | 15.8 | 37.20 | 12.90 | 34.68 | |
| | Mean \pm SD | 86.67 ± 1.53 | 14.00 ± 1.00 | 152.67 ± 1.15 | 55.30 ± 4.05 | 44.00 ± 3.00 | 79.33 ± 4.16 | 15.60 ± 0.26 | 39.42 ± 2.61 | 12.88 ± 0.22 | 32.77 ± 1.86 | |
| 16 | 39-1 | 85 | 14 | 152 | 57.4 | 51 | 97 | 15.8 | 47.40 | 15.11 | 31.88 | - |
| 17 | 39-2 | 87 | 16 | 151 | 52.3 | 59 | 112 | 14.5 | 50.22 | 16.35 | 32.56 | |
| 18 | 39-3 | 88 | 16 | 154 | 51.6 | 55 | 102 | 15.4 | 47.45 | 15.76 | 33.21 | |
| | Mean \pm SD | 86.67 ± 1.53 | 15.33 ± 1.15 | 152.33 ± 1.53 | 53.76 ± 3.16 | 55.00 ± 4.00 | 103.67 ± 7.64 | 15.23 ± 0.66 | 48.36 ± 1.61 | 15.74 ± 0.62 | 32.55 ± 0.66 | |
| 19 | 44-1 | 88 | 14 | 151 | 59.3 | 51 | 96 | 15.8 | 44.50 | 14.78 | 33.21 | - |
| 20 | 44-2 | 85 | 13 | 151 | 55.3 | 58 | 110 | 14.7 | 46.54 | 15.40 | 33.09 | |
| 21 | 44-3 | 86 | 15 | 150 | 56.5 | 52 | 102 | 15.2 | 44.56 | 14.34 | 32.18 | |
| | Mean \pm SD | 86.33 ± 1.53 | 14.00 ± 1.00 | 150.67 ± 0.58 | 57.03 ± 2.05 | 53.67 ± 3.78 | 102.67 ± 7.02 | 15.23 ± 0.55 | 45.20 ± 1.16 | 14.84 ± 0.53 | 32.83 ± 0.56 | |
| 22 | BS 100E-1 | 81 | 9 | 147 | 49.4 | 17 | 29 | 13.6 | 24.55 | 4.55 | 18.53 | - |
| 23 | BS 100E-2 | 83 | 7 | 146 | 45.3 | 13 | 22 | 12.9 | 22.14 | 4.12 | 18.61 | |
| 24 | BS 100E-3 | 83 | 9 | 148 | 44.8 | 14 | 25 | 13.7 | 21.45 | 4.22 | 19.67 | |
| | Mean + SD | 82.33 + 1.15 | 8.33 + 1.15 | 147.00 ± 1.00 | 46.50 + 2.52 | 14.67 ± 2.08 | 25.33 + 3.51 | 13.40 ± 0.43 | 22.71 ± 1.63 | 4.29 ± 0.22 | 18.94 ± 0.64 | |
| 25 | PBG7-1 | 88 | 16 | 153 | 59.3 | 58 | 114 | 15.9 | 51 44 | 16.5 | 32.08 | - |
| 25 | PBG7-2 | 89 | 17 | 154 | 54.5 | 54 | 107 | 16.2 | 47.62 | 15.4 | 32.00 | |
| 20 | PBG7-3 | 88 | 16 | 155 | 57.6 | 57 | 100 | 15.2 | 49.55 | 15.8 | 31 89 | |
| 21 | Mean + SD | 88.33 + 0.58 | 16.33 ± 0.58 | 154.00 ± 1.00 | 57.13 + 2.43 | 54.67 + 3.05 | 107.00 + 7.00 | 15.93 ± 0.25 | 49.54 + 1.91 | 15.9 ± 0.56 | 32.10 ± 0.22 | |
| | | | 10.00 = 0.00 | | | 2 = 2.00 | | | · · · · · · · · · · · · · · · · · · · | | 22.10 2 0.22 | |

SUPPLEMENTARY TABLE 5 Agronomic traits of BC_2F_2 and BC_2F_3 populations derived from Cross C (PBG7 × BS 100E).

Data on BC_2F_2 population are presented for plants analyzed for recurrent parent genome recovery; data on BC_2F_3 population are based on three plants phenotypically similar to PBG7 and presented as mean \pm SD; figures in parentheses are recurrent parent recovery percentages for agronomic traits calculated as plant trait value/value of PBG 7 for that trait \times 100; ⁶ Harvest Index = Seed yield per plant/Biological yield \times 100.