

### Supplementary Material

#### **Supplementary Figures**



## Supplementary Figure 1. Expression of CD244 and CD160 on CD4<sup>+</sup> T cells of healthy individuals from different age groups

(A-B) Box plots of the percentage of CD244<sup>+</sup> and CD160<sup>+</sup> cells on CD4<sup>+</sup> T cells from healthy donors in different age groups (n = 52-84 each group). *P* values were obtained by Kruskal-Wallis test followed by Dunn's multiple comparisons test. (C-D) Correlation analysis of age and CD244 (C), CD160 (D) expression on CD4<sup>+</sup> T cells from all healthy individuals. Spearman's non-parametric test was used to test for correlations. \**P* < 0.05.



Supplementary Figure 2. Distribution of CD8<sup>+</sup> T-cell subsets and CD244, CD160 expression on CD8<sup>+</sup> T-cell subsets from different age groups

(A-B) Gating strategy for CD244 and CD160 expression on each subset ( $T_N$ ,  $T_{SCM}$ ,  $T_{CM}$ ,  $T_{TM}$ ,  $T_{EM}$  and  $T_{TE}$ ) of CD8<sup>+</sup> T cells. (C) Distribution of  $T_N$ ,  $T_{SCM}$ ,  $T_{CM}$ ,  $T_{TM}$ ,  $T_{EM}$  and  $T_{TE}$  in CD8<sup>+</sup> T cells from different age groups (n = 17-34 each group). *P* values were obtained by Kruskal-Wallis test followed by Dunn's multiple comparisons test ( $T_N$ ,  $T_{SCM}$ ,  $T_{CM}$ ,  $T_{TM}$  and  $T_{EM}$ ) or one-way ANOVA followed by Tukey's multiple comparisons test ( $T_{TE}$ ). (D-E) Expression of CD244 on each subset ( $T_N$ ,  $T_{SCM}$ ,  $T_{CM}$ ,  $T_{TM}$ ,  $T_{EM}$  and  $T_{TE}$ ) of CD8<sup>+</sup> T cells (n = 17-34 each group). *P* values were obtained by Kruskal-Wallis test followed by Dunn's multiple comparisons test ( $T_N$ ,  $T_{SCM}$ ,  $T_{TM}$ ,  $T_{EM}$  and  $T_{TE}$  in D,  $T_N$ ,  $T_{SCM}$ ,  $T_{TM}$ , and  $T_{EM}$  in E) or one-way ANOVA followed by Tukey's multiple ach group the comparisons test ( $T_{CM}$ ,  $T_{TM}$ ,  $T_{EM}$  and  $T_{TE}$  in D,  $T_N$ ,  $T_{SCM}$ ,  $T_{TM}$ , and  $T_{EM}$  in E) or one-way ANOVA followed by Tukey's multiple comparisons test ( $T_{CM}$  in D,  $T_{CM}$  and  $T_{TE}$  in E). (F-G) Expression of CD160 on each subset ( $T_N$ ,  $T_{SCM}$ ,  $T_{CM}$ ,  $T_{TM}$ ,  $T_{EM}$  and  $T_{TE}$  in E). (T-G) Expression of CD160 on each subset ( $T_N$ ,  $T_{SCM}$ ,  $T_{CM}$ ,  $T_{TM}$ ,  $T_{EM}$  and  $T_{TE}$  in D,  $T_N$ ,  $T_{SCM}$ ,  $T_{CM}$ ,  $T_{TM}$ ,  $T_{EM}$  and  $T_{TE}$  in E). (F-G) Expression of CD160 on each subset ( $T_N$ ,  $T_{SCM}$ ,  $T_{CM}$ ,  $T_{TM}$ ,  $T_{EM}$  and  $T_{TE}$  of CD8<sup>+</sup> T cells (n = 17-34 each group).

*P* values were obtained by Kruskal-Wallis test followed by Dunn's multiple comparisons test ( $T_N$ ,  $T_{SCM}$ ,  $T_{CM}$  and  $T_{TM}$  in F,  $T_{SCM}$ ,  $T_{TM}$ ,  $T_{EM}$  and  $T_{TE}$  in G) or one-way ANOVA followed by Tukey's multiple comparisons test ( $T_{EM}$  and  $T_{TE}$  in F,  $T_N$  and  $T_{CM}$  in G). \**P* < 0.05, \*\**P* < 0.01, \*\*\**P* < 0.001.



# Supplementary Figure 3. Expression of CD244 and CD160 on CD8<sup>+</sup> T cell subsets from young (20s) and elderly (60s)

(A-B) Scatter dot plots of the expression of CD244 and CD160 cells on indicated  $CD8^+$ T cell subsets from healthy donors in 20s and 60s (n = 9-30 each group). *P* values were obtained by Mann-Whitney test or Unpaired t test.

### Supplementary Tables

Supplementary	Table 1. The seq	uence of primers f	for metabolic-related genes.
Suppremental		active of primers i	or metabolie related genesi

Genes	Forward primer $(5' \rightarrow 3')$	Reverse primer $(5' \rightarrow 3')$	
GLUT1	5'-TTGGCTCCGGTATCGTCAAC-3'	5'-GCCAGGACCCACTTCAAAGA-3'	
HK2	5'-GATTTCACCAAGCGTGGACT-3'	5'-ACAGGTGCTCTCAAGCCCTA-3'	
ENO1	5'-TCATCAATGGCGGTTCTCA-3'	5'-TTCCCAATAGCAGTCTTCAGC-3'	
PDK1	5'-CTGTGATACGGATCAGAAACCG-3'	5'-TCCACCAAACAATAAAGAGTGCT-3'	
ATP5G1	5'-ACAGCAACTTCCCACTCCAG-3'	5'-GCCAAGAATGGCATAGGAGA-3'	
mtNd1	5'-GGCTATATACAACTACGCAAAGGC-3'	5'-GGTAGATGTGGCGGGTTTTAGG-3'	
β-actin	5'-GAGCTACGAGCTGCCTGACG-3'	5'-GTAGTTTCGTGGATGCCACAG-3'	

### Supplementary Table 2. Antibody List.

Reagent	Source	Reference number (Cat#)
Anti-human CD3-BV786 (SK7)	BD Pharmingen	563800
Anti-human CD4-APC-Fire750 (SK3)	BD Pharmingen	344638
Anti-human CD8-BV421 (SK1)	BD Pharmingen	740093
Anti-human CD244-PE-D594 (2- 69)	BD Pharmingen	564881
Anti-human CD160-AF488 (BY55)	BD Pharmingen	562351
Anti-human CD98-BUV395 (UM7F8)	BD Pharmingen	744508
Anti-human CD45RA-AF700 (HI100)	BD Pharmingen	560673
Anti-human CD57-BV421 (NK-1)	BD Pharmingen	563896
Anti-human PD-1-BV711 (EH12.1)	BD Pharmingen	564017
Anti-human TIM-3-BV650 (7D3)	BD Pharmingen	565564
Anti-human Granzyme B-AF700 (GB11)	BD Pharmingen	560213
Anti-human T-bet-BV421 (O4-46)	BD Pharmingen	563318
Anti-human TNF-α BV711(MAb11)	BioLegend	502943
Anti-human IL-2-BV650 (MQ1- 17H12)	BioLegend	500334
Anti-human CCR7-BV421 (G043H7)	BioLegend	353208

Anti-human HLA-DR-AF700	BioLegend	307626
(L243)	DioLegend	507020
Anti-human CD38-BV421 (HB-7)	BioLegend	356618
Anti-human CD28-BV711	BioLegend	302948
(CD28.2)	DioLegena	
Anti-human CD27-BV650 (O323)	BioLegend	302828
Anti-human KLRG-1-APC-Fire750	BioLegend	367718
(SA231A2)	DioLegend	
Anti-human Annexin V-APC	BioLegend	640920
Anti-human Ki67-BV711 (Ki-67)	BioLegend	350516
Anti-human perforin-APC (B-D48)	BioLegend	353312
Anti-human CD71-FITC (CY1G4)	BioLegend	334104
Anti-human CD107a BV421	BioLegend	328626
(H4A3)		
Anti-human Eomes-PE-CY7	Ebioscience	25-4877-42
(WD1928)	Loioscience	
Fixable Viability Dye eFluor™ 506	Ebioscience	65-0866-14
Anti-human TIGIT-PE-Cy7	Ebioscience	25-9500-42
(MBSA43)		
Anti-human LAG-3-APC	Ebioscience	17-2239-42
(3DS223H)	Loioscience	
Anti-human 2-NBDG	Ebioscience	N13195
Anti-human IFN-γ-AF700 (4S.B3)	Ebioscience	56-7319-42
Human CD244 (999602)	R&D systems	MAB10393-100
Human CD160 (688327)	R&D systems	MAB6700
Mouse IgG 2B isotype control	R&D systems	MAB004
(688327)	Red systems	
Cell Event <sup>TM</sup> Senescence Green	Thermo Fisher Scientific	C10841