

## ***Supplementary Material***

# **Functional Characterization Of UDP-glycosyltransferases Involved in Anti-viral Lignan Glycosides Biosynthesis in *Isatis indigotica***

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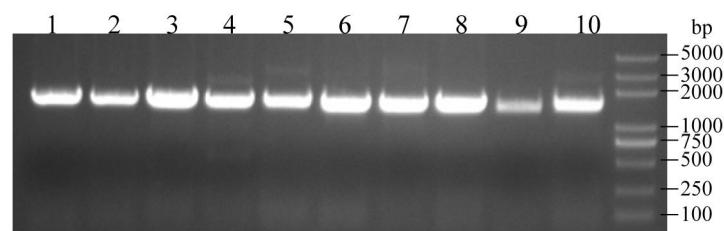
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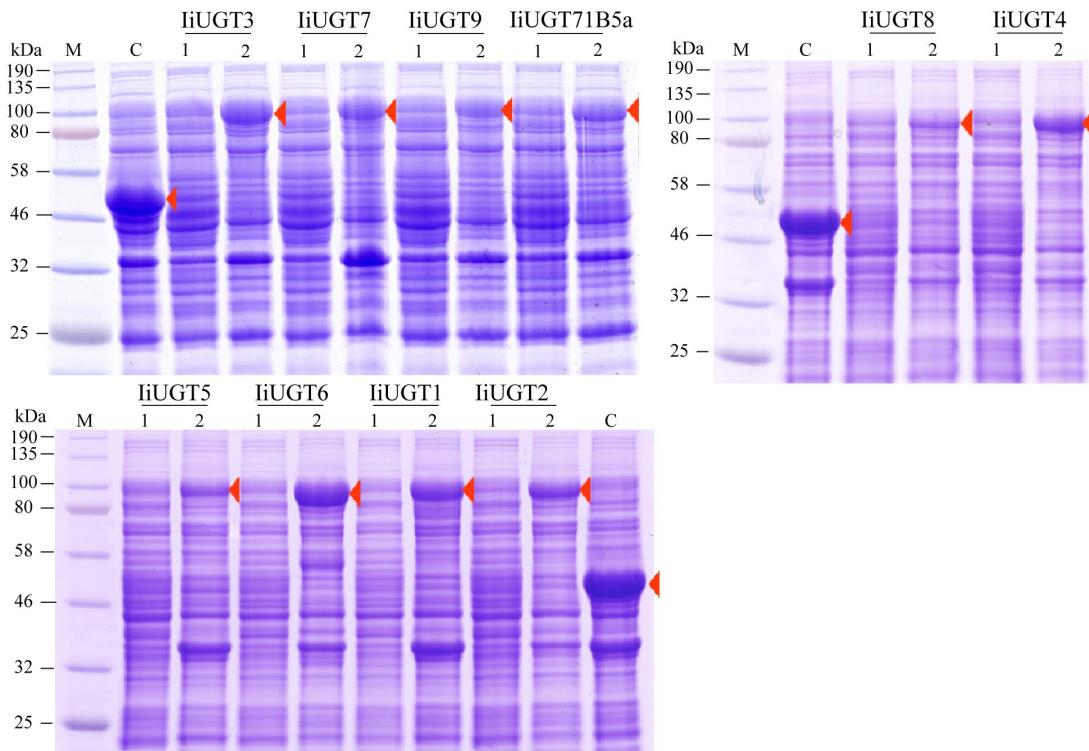
Luqi Huang  
huangluqi01@126.com

2    **1.1 Supplementary Figures**



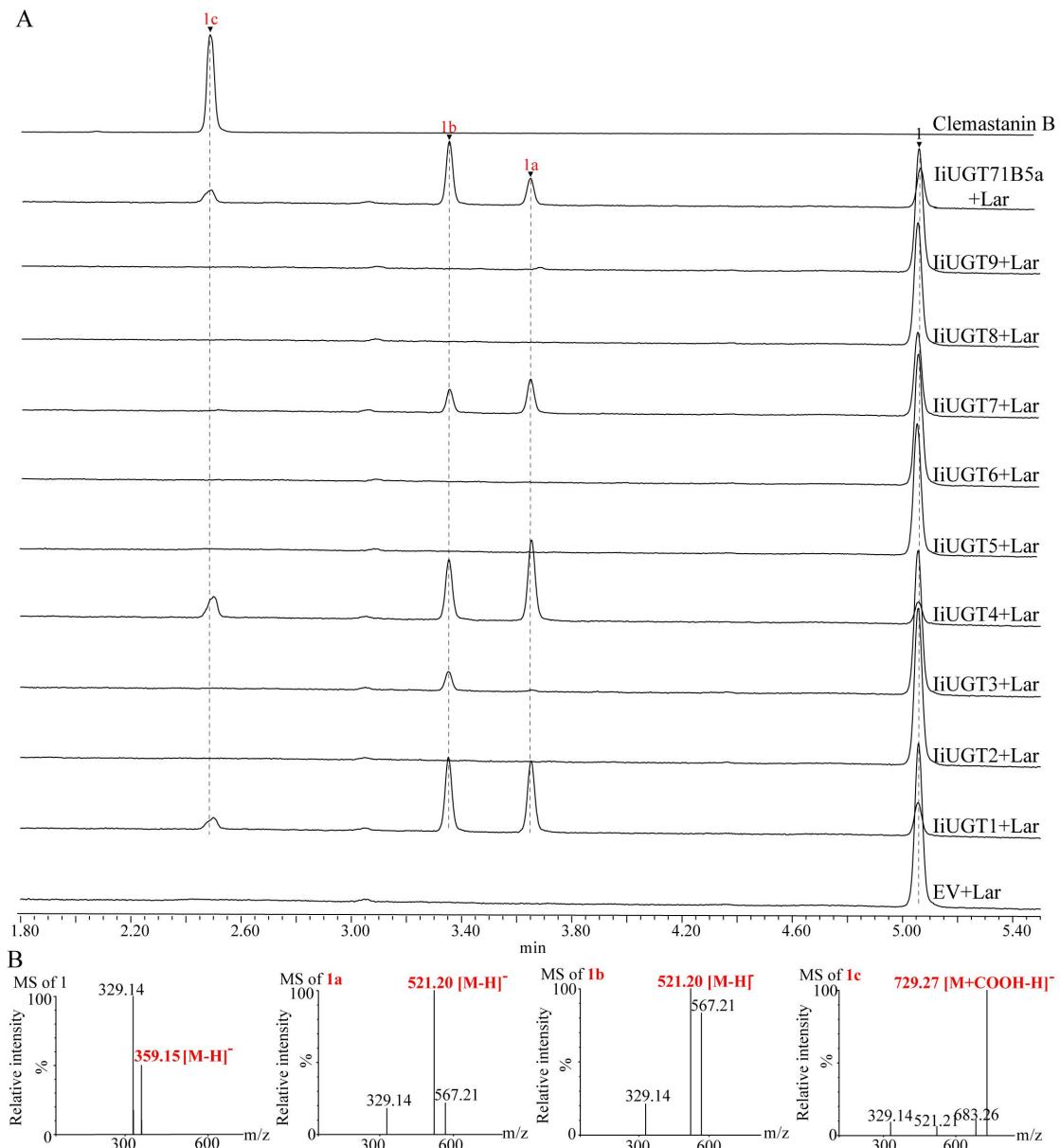
3    **Supplementary Figure 1.** PCR amplification of the ORF of ten candidate *IiUGT*  
4    genes.

5    Lane M: DNA Marker DL5000; Lane 1~Lane 10: *IiUGT1*~*IiUGT7IB5a*.



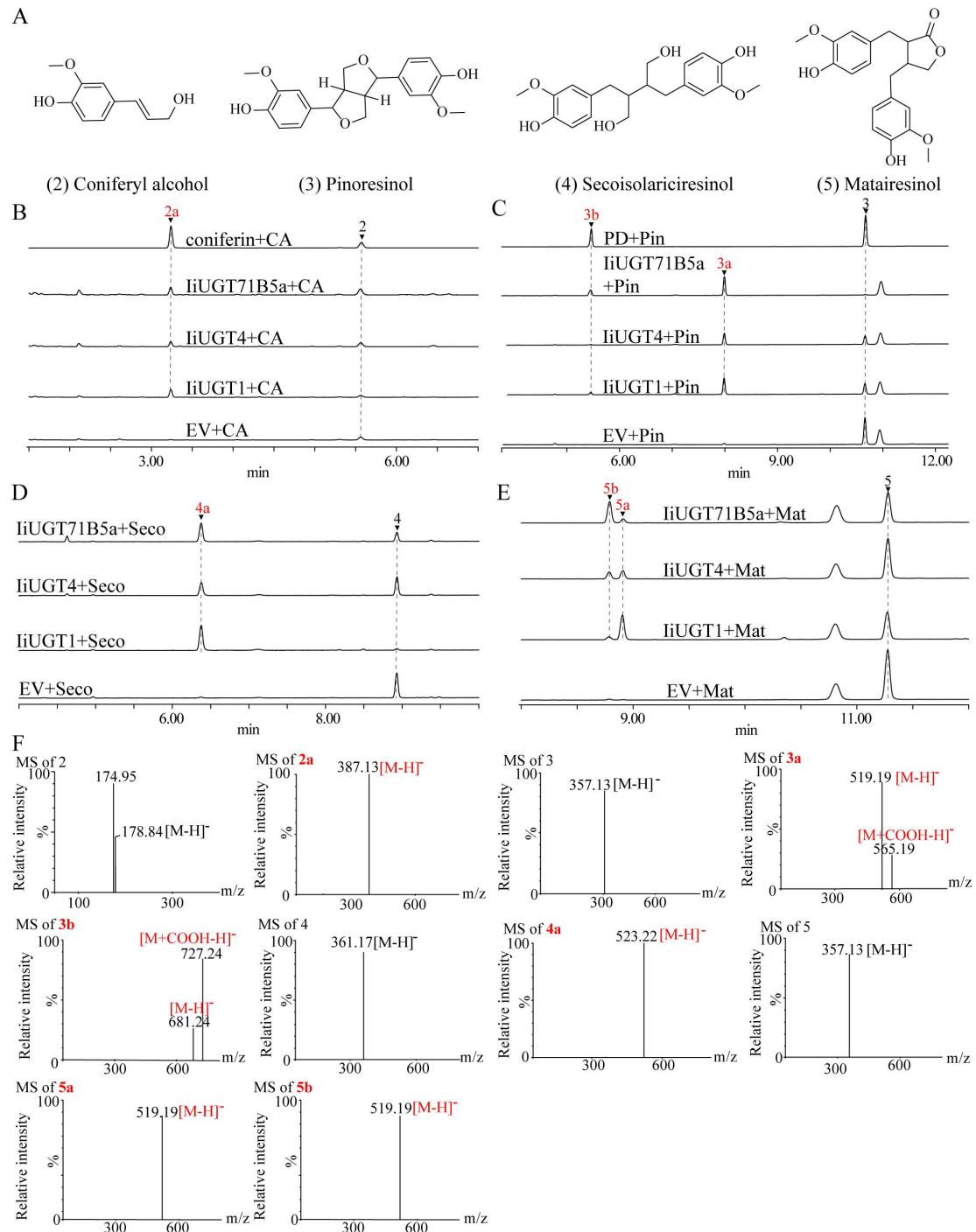
6      Supplementary Figure 2. SDS-PAGE analysis of expression of  
7      His-MBP-pET28a-*IiUGT*.

8      Lane M: molecular weight marker; Lane C: The crude protein induced by IPTG in *E.*  
9      *coli* Rosetta (DE3) containing the empty vector induced by IPTG; Lane 1: The crude  
10     protein in *E. coli* Rosetta (DE3) containing the recombinant vector without IPTG;  
11     Lane 2: The crude protein in *E. coli* Rosetta (DE3) containing the recombinant vector  
12     induced by IPTG.



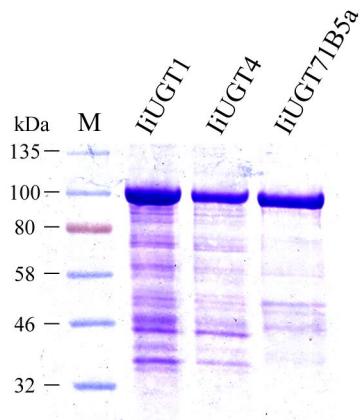
13      Supplementary Figure 3. UPLC/Q-TOF-MS analysis of the reactions of candidate  
 14      LiUGTs with UDP-glucose and (1) lariciresinol as substrates.

15      (A) The enzyme reactions of crude proteins of *E. coli* carrying empty vector (EV),  
 16      LiUGTs were assayed with UDP-glucose as the sugar donor. (1) lariciresinol; (1a)  
 17      lariciresinol-4'-O- $\beta$ -D-glucoside; (1b) lariciresinol-4-O- $\beta$ -D-glucoside; (1c)  
 18      clemastanin B. (B) MS spectra of the products in negative mode.



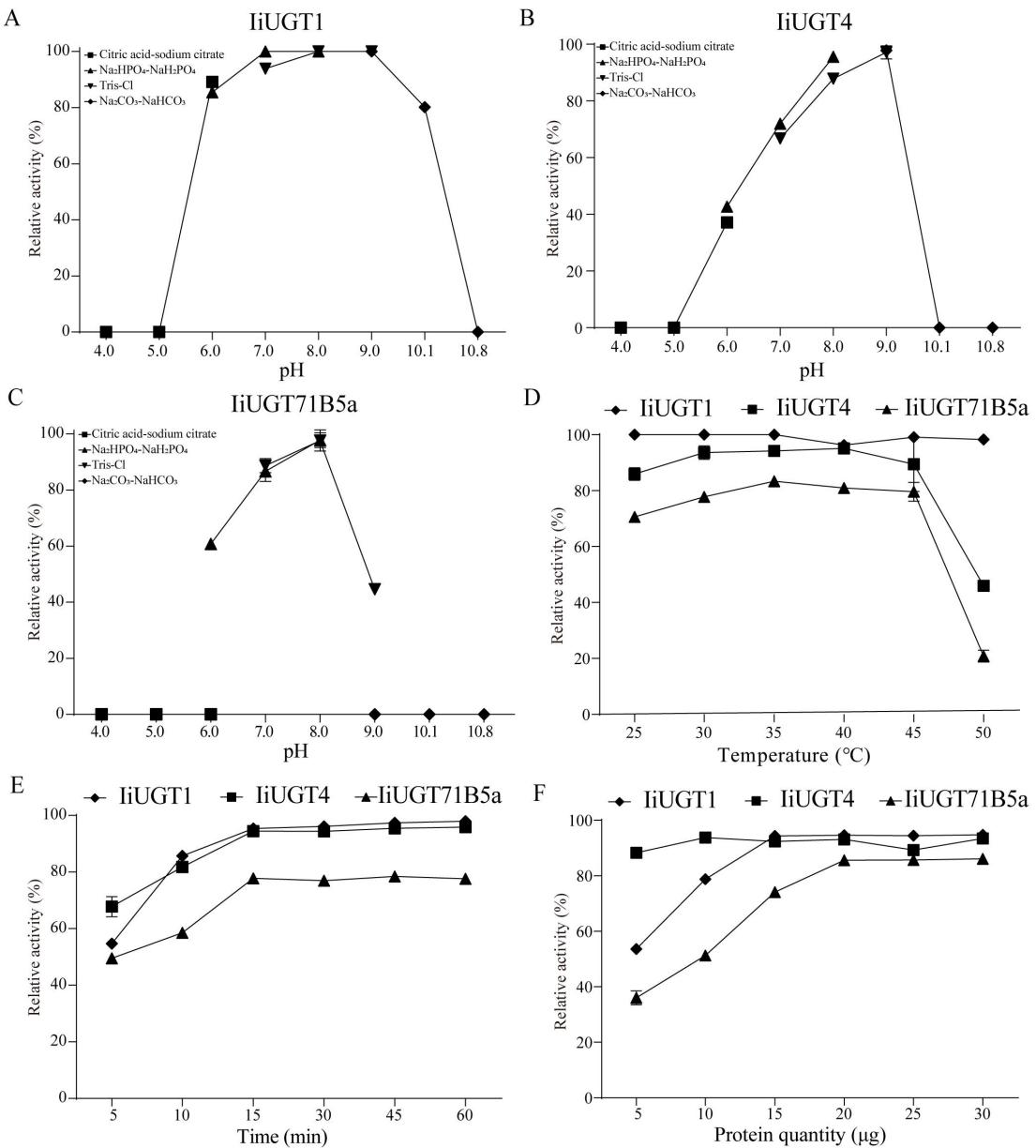
19 **Supplementary Figure 4.** Enzymatic activity of candidate LiUGT proteins.

20 (A) Chemical structures of lignans (glycosy acceptors). UPLC-ESI-MS/MS analysis  
 21 of candidate LiUGT enzymatic reaction products against coniferyl alcohol (B) and  
 22 pinoresinol (C), secoisolariciresinol (D) and matairesinol (E), respectively. The  
 23 enzyme reactions of crude proteins of *E. coli* carrying empty vector (EV), LiUGT1,  
 24 LiUGT4, LiUGT10 were assayed with UDP-glucose as the sugar donor. (B) 2. CA,  
 25 coniferyl alcohol; 2a. coniferin. (C) 3. Pin, pinoresinol; 3b. PD, pinoresinol  
 26 diglucoside. (D) 4. Seco, secoisolariciresinol. (E) 5. Mat, matairesinol. (F) MS spectra  
 27 of the products in negative mode.



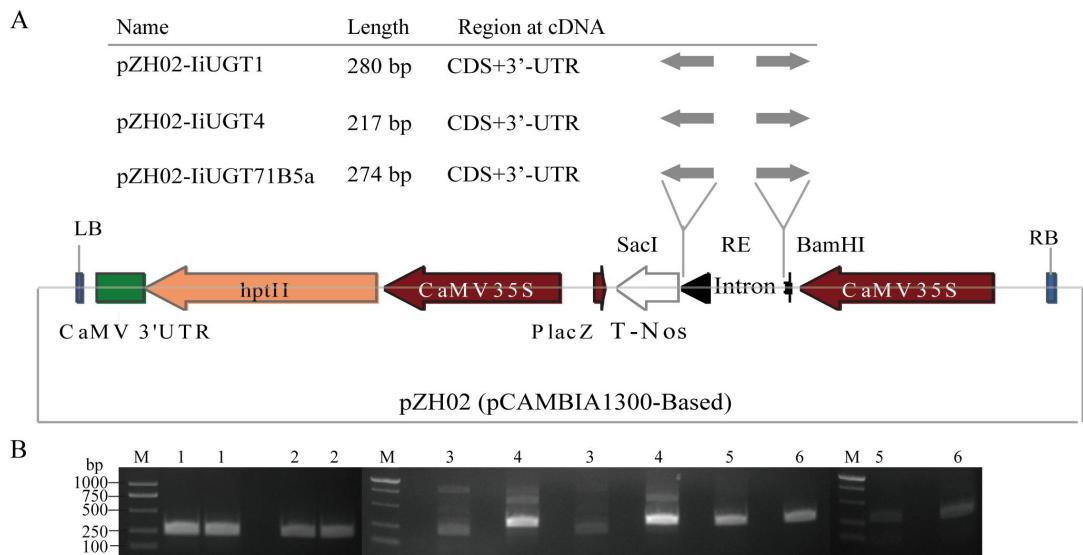
28 **Supplementary Figure 5.** SDS-PAGE analysis of purified proteins.

29 SDS-PAGE analysis showed that the purified IiUGTs fused with a HIS-MBP tag  
30 (41.34 kDa).



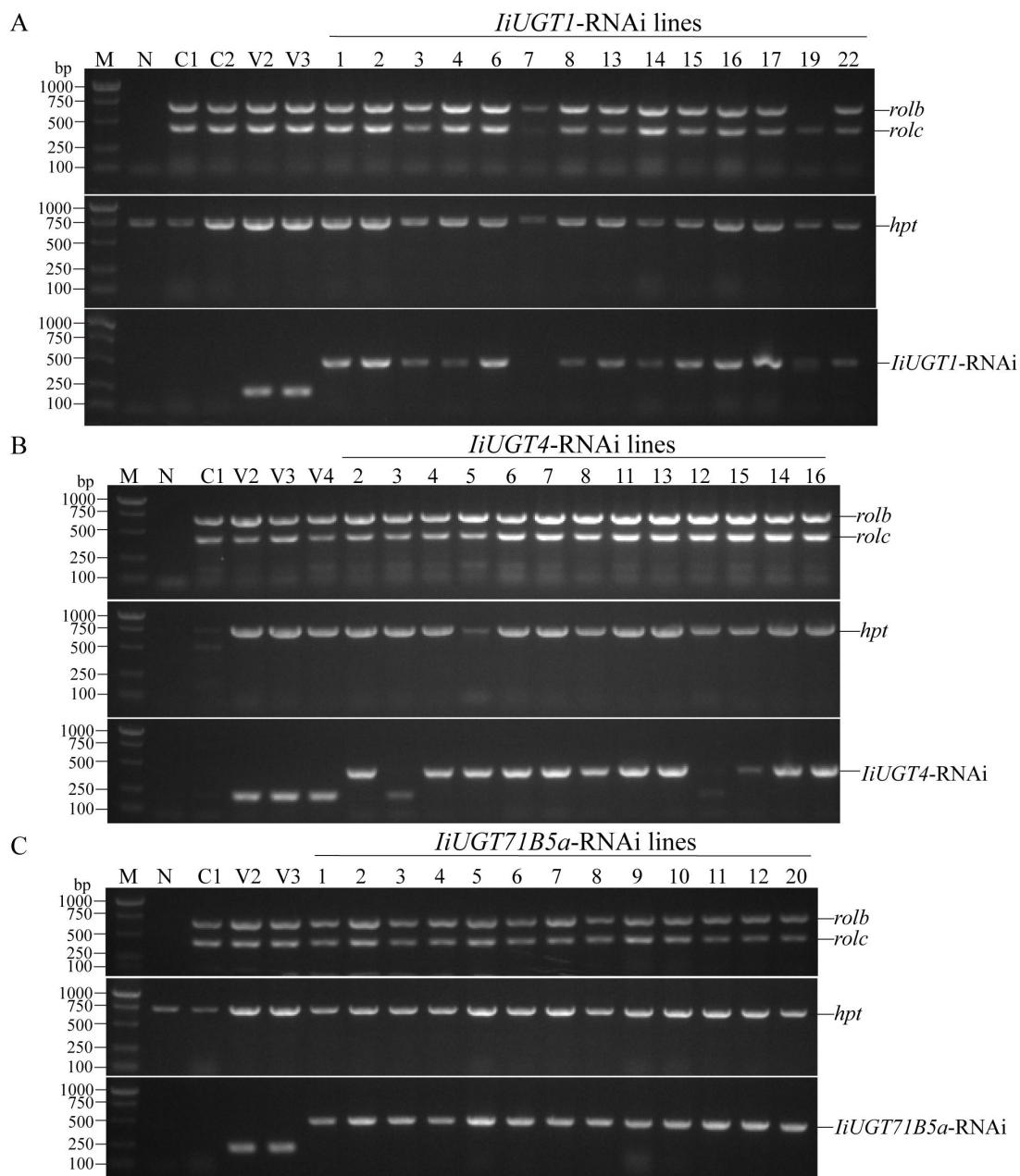
31 **Supplementary Figure 6.** Properties of recombinant LiUGTs.

32 To verify pH preference (A), (B), (C), optimal temperature (D), optimal reactive  
 33 time (E), and optimal reactive protein quantity (F), reactions were examined using  
 34 lariciresinol and UDP-glucose as substrates as described in the experimental  
 35 procedures. The squares represent Citric acid-sodium citrate Buffer (pH 4.0-6.0), the  
 36 triangles represent Sodium phosphate Buffer (pH 6.0-8.0), the inverted triangles  
 37 represent Tris-Cl Buffer (pH 7.0-9.0) and the diamonds represent Sodium carboxylate  
 38 Buffer (pH 9.0-10.8) in (A), (B), (C). Values of the relative activities are average  $\pm$   
 39 SD ( $n=3$ ), with maximum activity levels assumed to be 100%.



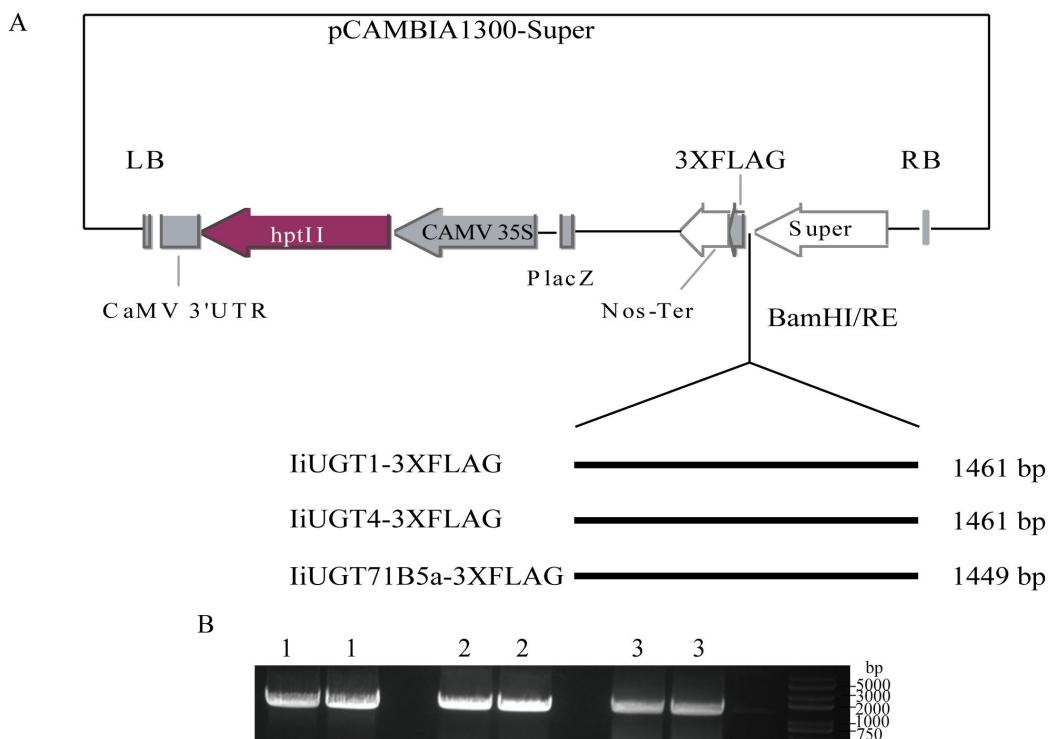
40 **Supplementary Figure 7.** RNAi silencing of *IiUGT1*, *IiUGT4* and *IiUGT10* in *I.*  
41 *indigofera* hairy roots.

42 (A) Schematic diagram of the RNAi vector. CaMV 35S, CaMV 35S promater; hpt II,  
43 hygromycin resistance gene; Intron, the Pdk intron; grey arrow, the 200-300bp  
44 fragment of the coding region of *IiUGTs*. (B) PCR amplification of the target *IiUGT*'s  
45 interference fragmnets. M: DNA Marker; Lane 1: *IiUGT4*-RNAi sence; Lane 2:  
46 *IiUGT4*-RNAi antisence; Lane 3: *IiUGT10*-RNAi sence; Lane 4: *IiUGT1*-RNAi sence;  
47 Lane 5: *IiUGT1*-RNAi antisence; Lane 6: *IiUGT10*-RNAi antisence.



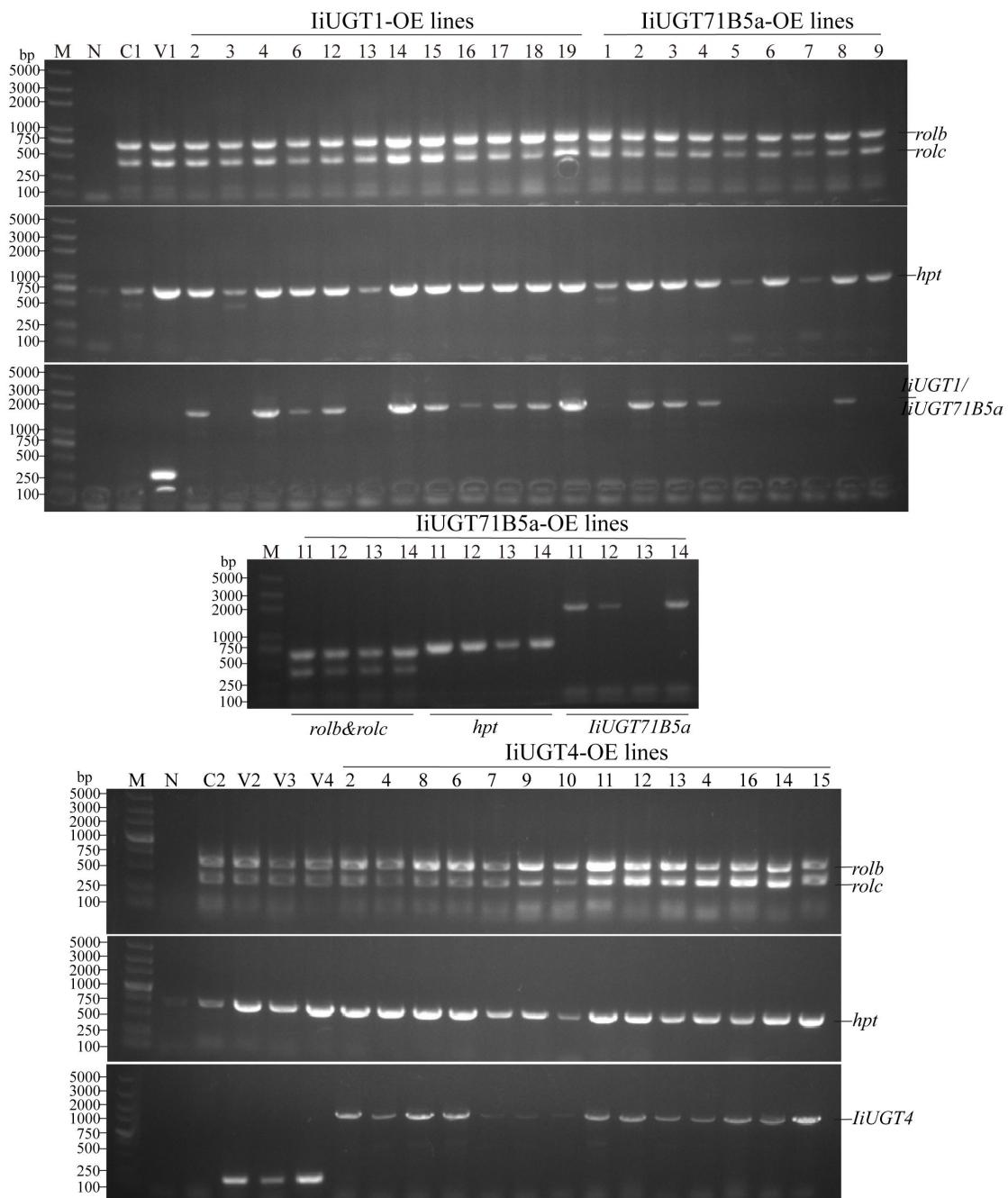
48      **Supplementary Figure 8.** PCR analysis of *rolb*, *rolc*, *hpt* and additional *IiUGT*-RNAi  
 49      fragment in *I. indigofera* hairy roots.

50      Lane M: DNA Marker; Lane N: Negative control; Lane C1, C2: Wild-type lines; Lane  
 51      V2, V3: Control lines with the empty construct.



**52 Supplementary Figure 9.** Overexpression of *IiUGTs* in *I. indigofera* hairy roots.

(A) The full length ORF of *IiUGTs* were inserted into the pCAMBIA1300-Super expression vector under control of the Super promoter, respectively. (B) PCR amplification of the ORF of the target *IiUGTs*. Lane M: DNA Marker; Lane 1: *IiUGT1*; Lane 2: *IiUGT4*; Lane 3: *IiUGT10*.

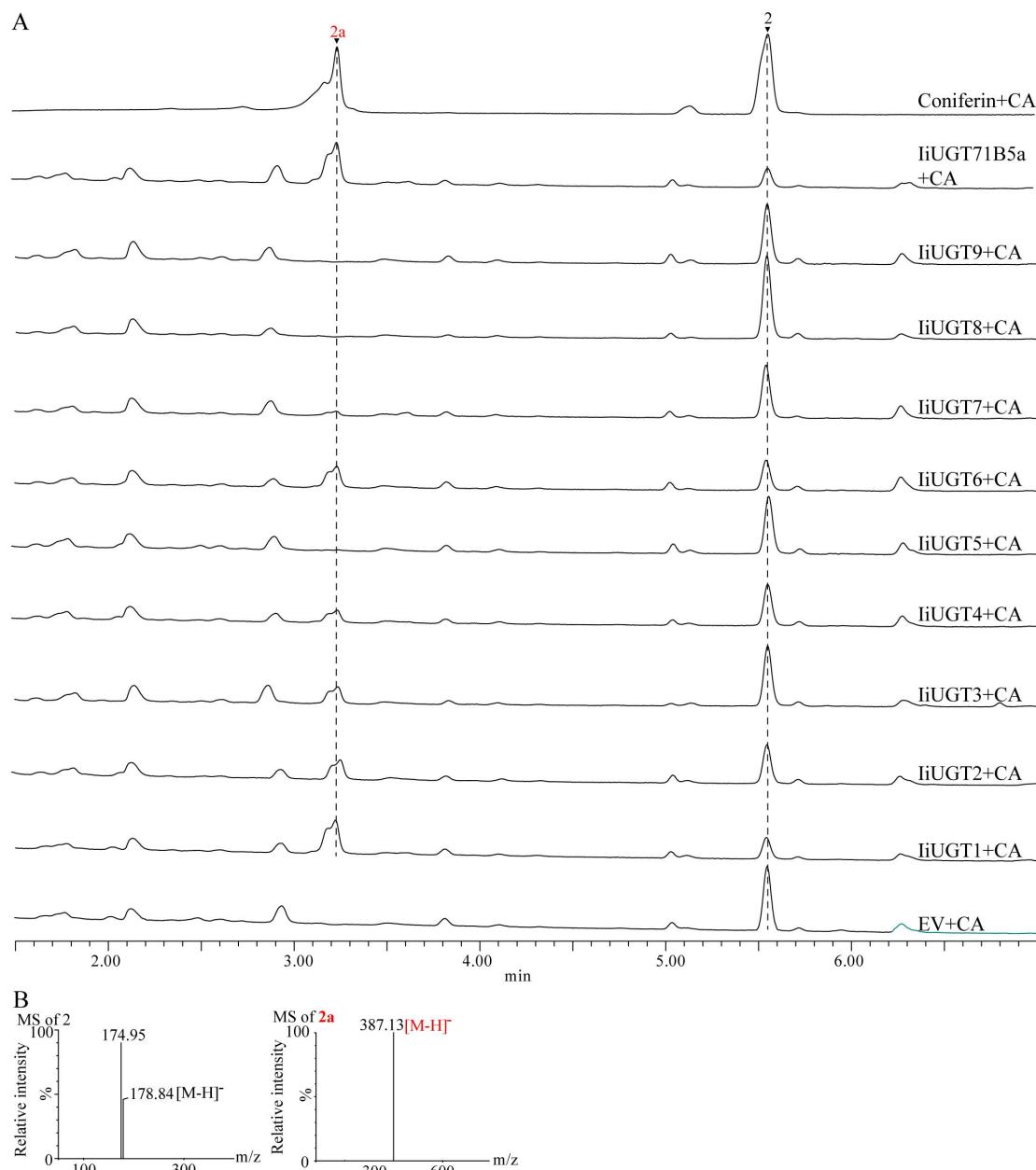


57 **Supplementary Figure 10.** PCR analysis of *rolb*, *rolc*, *hpt* and the exogenous *IiUGT*  
 58 gene with flag tag in *I. indigotica* hairy roots.

59 (A) Lane M: DNA Marker; Lane N: Negative control; Lane C1, C2: Wild-type lines;  
 60 Lane V1, V2, V3: Control lines with the empty construct.

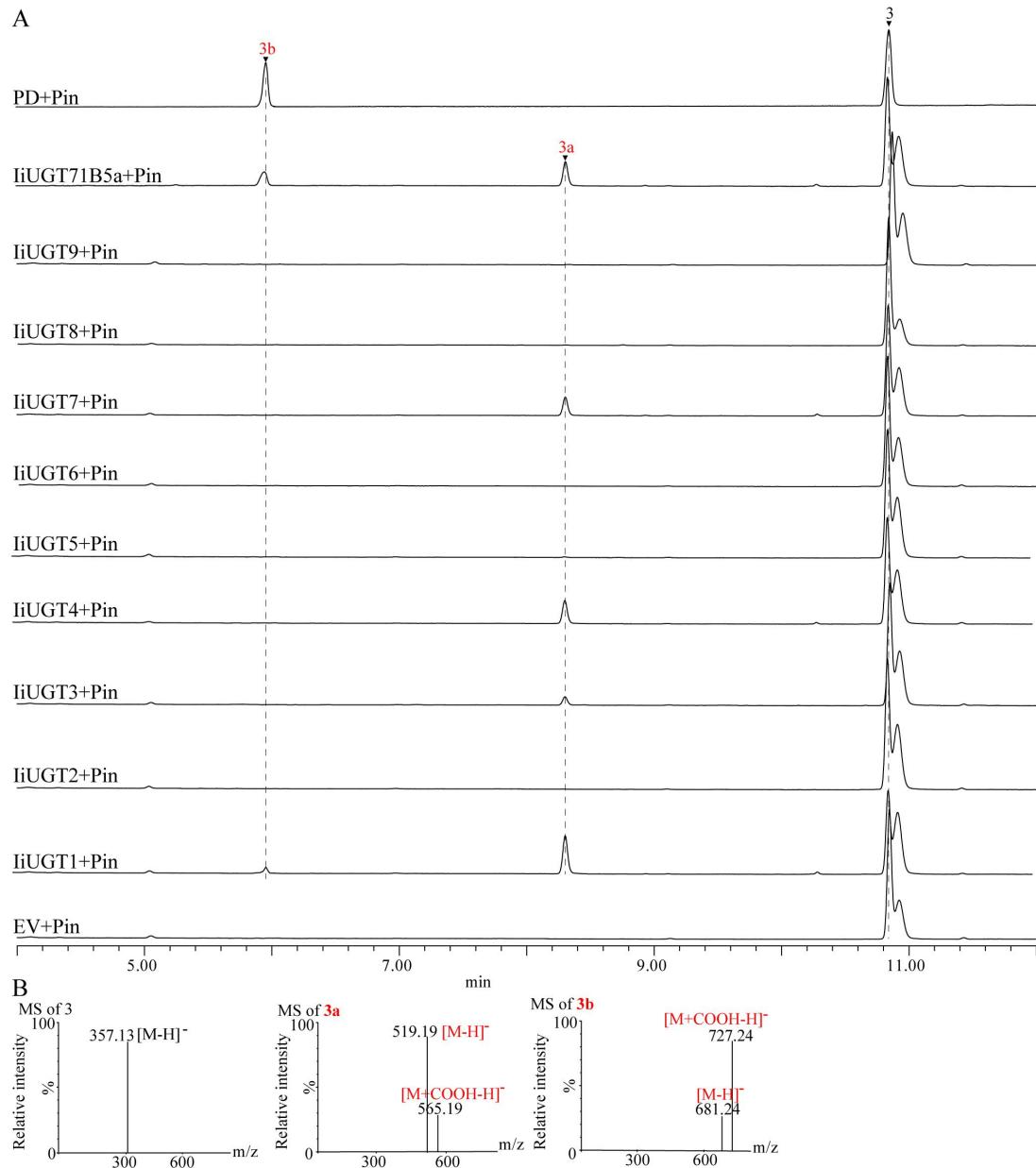
Gene ID	Position of PSPG box	Sequence of PSPG box
LiUGT1	350-393	WAPQAEILAHQAVGGFLTHCGWN <span style="background-color: #d9e1f2;">S</span> VLESVVSGVPMITWPL
LiUGT4	355-398	WAPQTAILANPAVRGFVSHCGWN <span style="background-color: #d9e1f2;">S</span> TLESLWFVGVP <span style="background-color: #d9e1f2;">I</span> ATWPL
LiUGT71B5a	349-392	WAPQVAVLAKPAIGGFVTHCGWN <span style="background-color: #d9e1f2;">S</span> MLESLWFVGVP <span style="background-color: #d9e1f2;">M</span> VTWPL
UGT71A9	348-391	WAPQMAVLSHPAVGGFVSHCGWN <span style="background-color: #d9e1f2;">S</span> VLESVWC <span style="background-color: #d9e1f2;">G</span> VPM <span style="background-color: #d9e1f2;">A</span> VWPL
UGT74S1	334-377	WCSQLQVLASGKVGC <span style="background-color: #d9e1f2;">F</span> VTHCGWN <span style="background-color: #d9e1f2;">S</span> TLEALSLGVPM <span style="background-color: #d9e1f2;">A</span> MPE

61 **Supplementary Figure 11.** Amino acid sequence alignment of the PSPG conserved  
 62 motif for LiUGTs and lignan glycosylation UGTs.



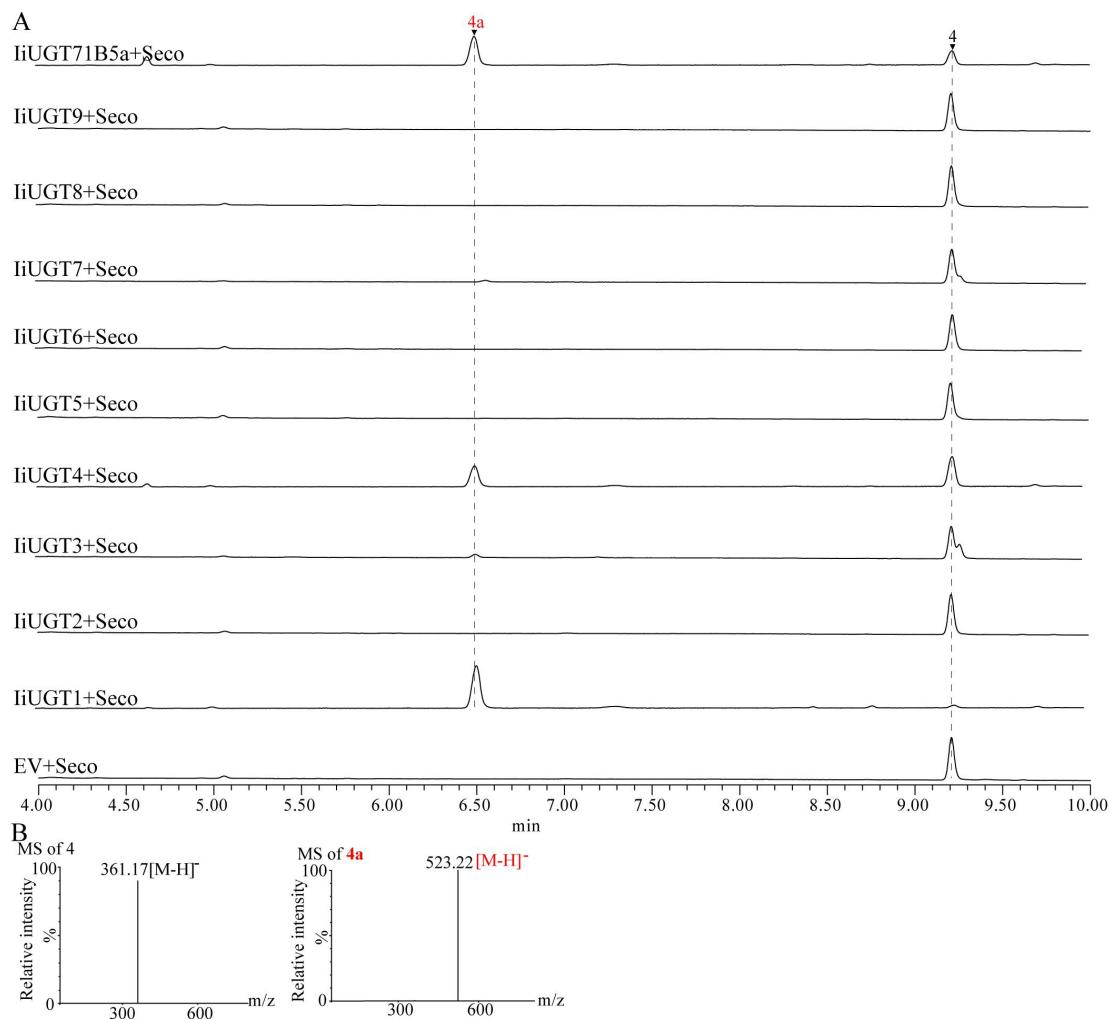
63 **Supplementary Figure 12.** UPLC/Q-TOF-MS analysis of the reactions of candidate  
 64 IiUGTs with UDP-glucose and (2) coniferaldehyde as substrates.

65 (A) The enzyme reactions of crude proteins of *E. coli* carrying empty vector (EV),  
 66 IiUGTs were assayed with UDP-glucose as the sugar donor. CA, coniferyl alcohol. (B)  
 67 MS spectra of the products in negative mode.



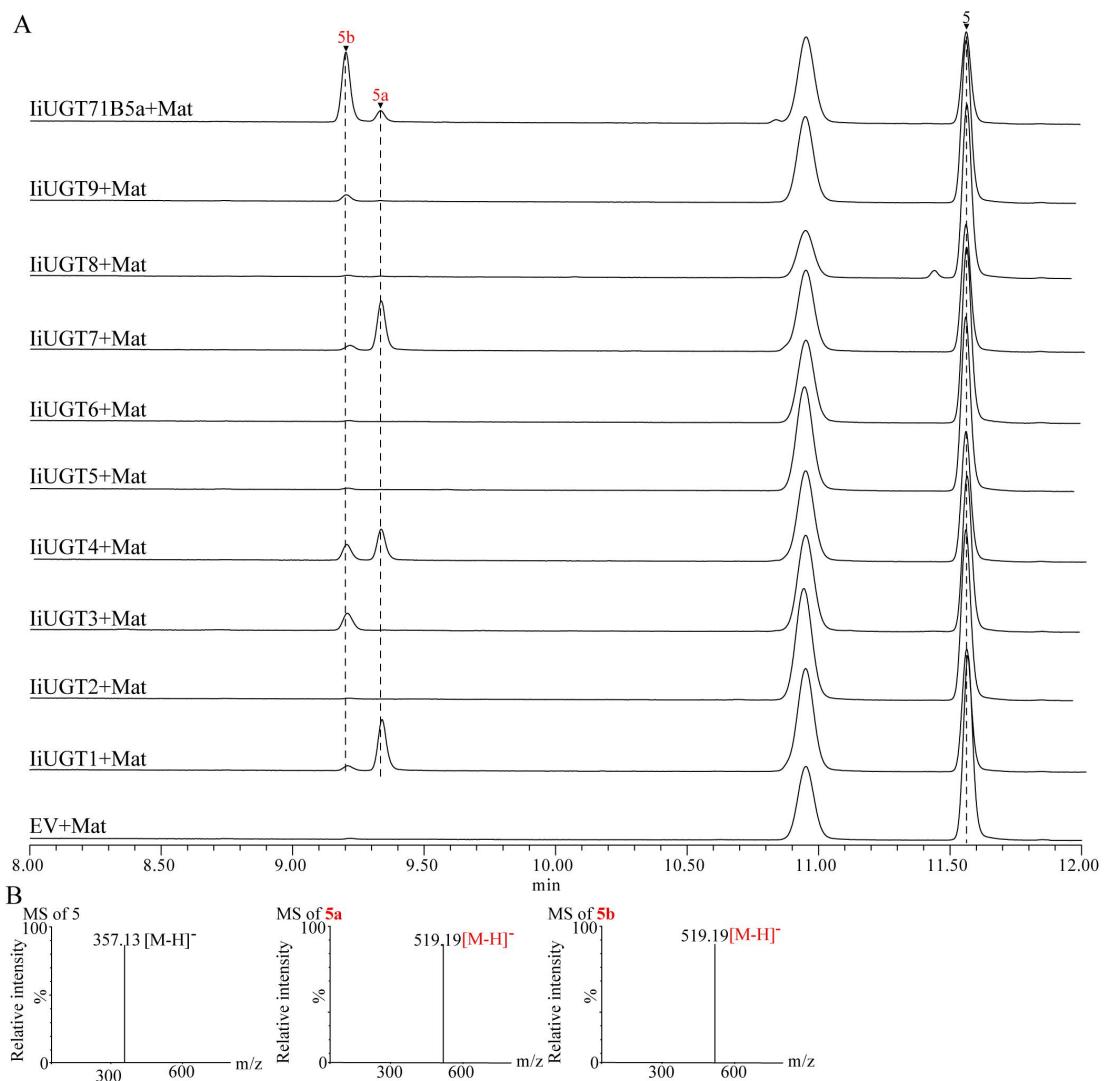
68 **Supplementary Figure 13.** UPLC/Q-TOF-MS analysis of the reactions of candidate  
69 LiUGTs with UDP-glucose and (3) pinoresinol as substrates.

70 The enzyme reactions of crude proteins of *E. coli* carrying empty vector (EV), LiUGTs  
71 were assayed with UDP-glucose as the sugar donor. Pin, pinoresinol; PD, pinoresinol  
72 diglucoside.



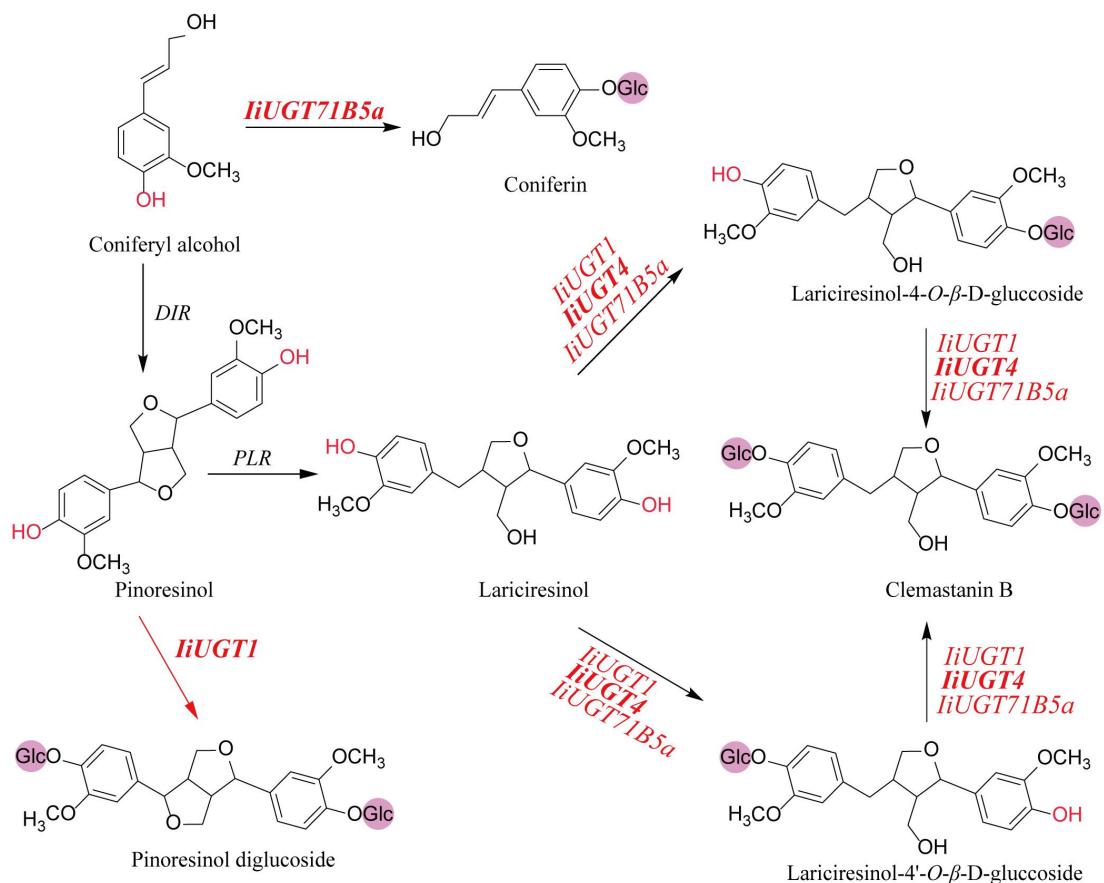
73 **Supplementary Figure 14.** UPLC/Q-TOF-MS analysis of the reactions of candidate  
 74 IiUGTs with UDP-glucose and (4) secoisolariciresinol as substrates.

75 The enzyme reactions of crude proteins of *E. coli* carrying empty vector (EV), IiUGTs  
 76 were assayed with UDP-glucose as the sugar donor. Seco, secoisolariciresinol.



77 **Supplementary Figure 15.** UPLC/Q-TOF-MS analysis of the reactions of candidate  
78 LiUGTs with UDP-glucose and (5) matairesinol as substrates.

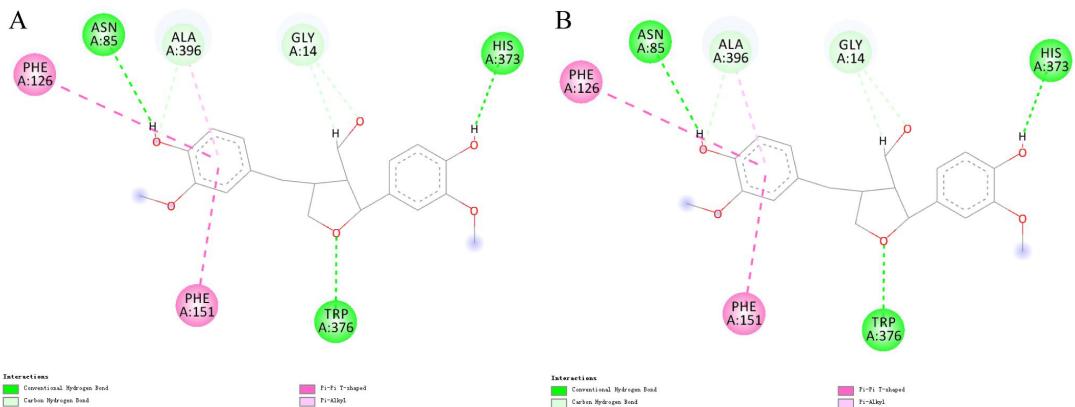
79 The enzyme reactions of crude proteins of *E. coli* carrying empty vector (EV), LiUGTs  
80 were assayed with UDP-glucose as the sugar donor. Mat, matairesinol.



81

82 **Supplementary Figure 16.** The deductive pathway of lignan glycosides in *I.*  
83 *indigotica*.

84 *IiUGT* genes playing a dominant role in the biosynthesis were in red bold.



85      **Supplementary Figure 17.** Molecular docking of IiUGT4 with (+)-lariciresinol (A),  
 86      (-)-lariciresinol (B).

87 **1.2 Supplementary Tables**88 **Supplementary Table 1.** Unigenes generated from *I. indigofera* transcriptome.

Range of Length (bp)	Total Number	Percentage (%)
200~300	19,709	33.43
300~500	13,458	23.58
500~1000	10,032	17.58
1000~2000	8,646	15.15
>2000	5,857	10.26
Total	57,072	100
Total Length	47,238,106--	
N50	1,546--	
Mean	827.69--	

**Supplementary Table 2.** The 10 *IiUGT* genes cloned in the study.

Gene Name	Subfamily	Protein Sequences
<i>IiUGT1</i>	<i>UGT72E</i>	mkitrphavmfaspghvipvielgkrlvgshgfqvtifvleadaasaqsq flnstgcdatlidlividclptidisglvdpsaffaiklltmmreiptirskeemqh kptalivdlfgldalrlggefmltyvfiasnarfvallyfptlekdaeehiikk kplampcepvrfedtlepfldptdqiyrifvpfgyvyptadgiivntwddme pkltkslqdpgkllgriarvpvypigplsrvdpsktnhpvldwlnkqpeesvl yisfgsggslsakqltelawglelsqqrffvvrvppvdssacseylsansgev qdgtlpdklfkqfisrtqerglvvpswapqaeilahqavgflthcgwnsvles vvsgvpmitwplfadqkmnatllneelgvairsrklpseevtlrveieslvrrl mvedegremrekvkkldtaemsrlcdggssheslsrvanechrllearma rga
<i>IiUGT2</i>	<i>UGT71C</i>	maketelifipvpstghllvniefakrlinlehriqtitiqmdspinphasvfars lvasqpqirlhslpvldpppsdlykrapayivqlvkktplvk davssivesr gsdsrvraglvldffcnslkdvgnelnlptyiflcnarylsmmkyipdrhrk maskldwssgdeelpipgfanpiptkfmppglfnqegyeayvelaprfaha kgilvnsiaelepfpfyfsqqhnyppvypvgpilslkdraspneeaadrdr ilrwledqessvvflcfgskgs vdepqvkeiaqalevvgerflwsirmsleei kpsdvlpegfmgrvagrglvvcgwapqevlahkaiggfvshcgwnstlesl wfgvpvatwpmyaeqqlnaftlvkelgla vdlrm dyvsgrglvcdeiara vrs lmdg gegkrvkv kemadaarkammdggssylatarfigelddgss mqitkphaamfsspgmg hlipvielakrlsanhgfrtvfvlesdaasaqskf lnstgvdvvnlp spdisdlvdpadhvvt kigimreapalrskiaemnqkpt aliidlfgtdalclaaefkmltyvliasnarylgvamyptldkhvqeehtvqrk plevpgcepvr fedtmdaylvpdeplyrdfvrhslaypkadgilvntw dem epkslkslqdpgkllgrvarvpvypgplcrpveqsktghp vldwln eqpdes vlyisfgsggsltakqltelawgleqsqqrfvwwvrvppvdgsscc eyfsanga ekkd stpeylpegfvtrtcdr glvvqswapqaevlah ravggflthcgwnstl egvvsgvpmiawplfaeqnmna allsdelgia vradnlkeavtrfeieavrk vmteegeemrmkvkkldkaemllssdgggsaheslckvt vecerflerd mdlarga
<i>IiUGT3</i>	<i>UGT72E</i>	mkqelvfipspgdghirplqvakllvdrdehisitiliipqmhgf gssnsy iaslstasedrlhy nvlsvade pnsddakpnflshinsfkpqvkatveklitspa rpdsp srlagivdmfc tdmidvanefd vpsymf tsnatfl gllshv qhly ddknydvs dldsevteleipcltcplpvkclpsvmlnkewlpialsqvrryke t kglvntfae lep qamkffs gednllptv ypgp ilnk tngp npaddkq sei lrwldeqpretvvflcfgsmggfredqakeiaialersghrfvws lrrar pegtr gppgeftnleeilpegflrtakigkvigwapqtailanpavrgfvshcgwnst leslwfgvpiatwplyaeqqvnafemveel glaveirnsfrad fmaa eselm taeeiergirclmeqdnvrdrvkemsekshvslmeg gssha allkfiedvtsn is
<i>IiUGT4</i>	<i>UGT71B</i>	mivv lvntrkvrggnrsyqflemrna elifipaptvghl vpsle larri lidq dd riritylv mklqgqshldtyvksigssl pfvr fidv peledkptfgstq saea fvy
<i>IiUGT5</i>	<i>UGT71D</i>	

		dfierniplvrnivmdilsspaldgvtvkgivadffclpmvdvakdvslpfhvfdlttsvflammqyladrhskdtsfvrnsgemlsipgfvpnvpanvlptalmedgyeayvkilfakakgivilntyfdlepislnhfhneqnypsvyavgpfnpnaphpdqdlarrdelkvwlldqpeasvvflcfgsmgrlsgplvkeiahglelcqyrlwsrteevthddlfpegfldrvsrgmicgwspqveilahkavgsfvshcgwnslveslwfgvpivtwpmyaeqqlnaflmvkelnlavemkldyrvrsdlvnaneietaircvmnednnlrvkrvidisqmarkatlngssylatekfiqdvigikpmktselivplpetghllstiegkrllddrismitilsmklyaphadaslasltasepgirlislpeiqdpppiklldtssetyildfveknipflktirdlvssgedsnhvaglildffcvdldigrevnlpsyifmtnsfngflgflqylperhrsissedessgdeelpipafvnrvpakvlppgvfdklsygtlvkigerlneakgifvnsfsevepyaaehfsrggdypypraypvgpvlntgrtnpglasaqyaemmkwlldqpdssvlfcfgsmgvfsaaqiteiahaielvgfrfiwairtnmegdgdpheplpegfvdrtmrgivcswapqvdlahkatggfvshcgwnsiqeslwyygvpiatwpmyaeqqlnafevmkelglaveirldyvadgdrvtleivsadeiaairslmdgdnlinkkvrevsaarkavsdggssmvatgdfirdilgdhfmeqphallvasplglghlipilegnrlssvlnihvtvlavpsgsssspteteairaavargtceiaelpsvdiehlvedpatvatrifekmratrpaqvdaatkamnrkptvmividffatglmsvaddvgvtakyvvpshawflavmvlpvldkvvegeyidikepmkipgcrsvgpdelmdtmfdrsdrqyrecvrcgeeipmsdgilvntweelqgntlaalredgelsrvmkvpvyai gpfvrsngpiekpksifewldkqrdrsvvyvclgsggilsleqtmelawglelgsgqsflwvllrptsylearssdddqvsaglpegfldrtrgvglvvtqwapqveilshgsiggflshcgwssvlesltkgvpivawplyaeqwmmatllteeigavrtselpskkvigreevaslvrkivaeedeegrkvraakaeevkatseaawaqggsshgsllewakrcrlvcdsqiimgnqeiiifvpypipghllvtielakylikrdnrihtitilhwtlplaphadlfakslvaseprirrlftlpdpvnpppfelflrateayvleftkctvplvrealstivssrdgsdpvwvaglvldffcvplievgnefnlpsyifltnagflgilkylperhrriaselelsehhpiqfvssvpskvlpsqfqvresyeawieiaqkfpkakgivilvnsftcleqnaftyfaclpenfpvypvgpvlssledrspdldtsqcrvmtwlddqpessvvylcfgsfgvlgепqieeiaraeleisshrflwsirtekatpydllpegfmdrtvskglvcgwapqevlahkavggfvshcgwnsvleslwfvdvpiatwplyaeqqlnaytmvkelglsvlerldyvsakkvivkadeiagairslmdgedtprrvkkmaeaarmalmeeggssfvavkrfiddlvgedfmqntkphaamltspgmghvipviqlgkrlagfhgfhtifvleadaasaqsqflnspgcdattlvdiglpspdisglvepsasfgtklltmmreavpsirskaemqhkptalivdlsldalrlggefmltyfiasnarfvalmmyfptldrveeehiikkplaipgceplrfedtfeifldpssqmyqecvplglvyatvdgiivntwdmepktilkslqdpkllgriarvpiyigplcrpvdpksktnhpvldwlnkqpdesvlyisfgsggslsakqltelawglelsqqr fvwwvrppvdgsacsayfsntgqvrdgtpdylpeefvsrtlerglvvp swapqaeilahtavgflthcgwnsilesvvigvpmiawplfaeqmtnatllneelgiavrsrrlpseglilreeiealvrrimvdeegcvmrkkvkklsdaekslscetlsrvaeecerrlehdrsmarga
<i>IiUGT6</i>	<i>UGT71C</i>	
<i>IiUGT7</i>	<i>UGT72D</i>	
<i>IiUGT8</i>	<i>UGT71C</i>	
<i>IiUGT9</i>	<i>UGT72E</i>	

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*IiUGT71B5a    UGT71B*

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mkielvfipspgighrstvelakqlvngddrlsitviiprsggdatdsaqissl  
faasqdrlyetisvadeptadrlptqlyiknqkpqvrdavakildparvdsspr  
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eldesvnelefpcltrypveclpylfiskewlppmdqarsfrkmkgilvntv  
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gvpmvtwplyaeqkvnavfmveelglaveirrslkgdilmaggmetvaaed  
ierairrvmeqdsdvrnrvkemaekchvaltdggssqvalrkfiqdvienvv

v

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**Supplementary Table 3.** GenBank accession numbers of UGT proteins in Figure 3.

Gene	Accession	Species
UGT71A5	BAF96584.1	<i>Antirrhinum majus</i>
UGT88D3	Q33DV3.1	<i>A. majus</i>
At3Rt	NP_564357.1	<i>Arabidopsis thaliana</i>
UGT71B1	NP_188812.1	<i>A. thaliana</i>
UGT71B6	NP_188815.2	<i>A. thaliana</i>
UGT71C1	NP_180536.1	<i>A. thaliana</i>
UGT73B1	NP_567955.1	<i>A. thaliana</i>
UGT73C6	NP_181217.1	<i>A. thaliana</i>
UGT74F2	OAP07463.1	<i>A. thaliana</i>
UGT75C1	AAL69494.1	<i>A. thaliana</i>
UGT71B2	NP_188813.1	<i>A. thaliana</i>
UGT78D2	NP_197207.1	<i>A. thaliana</i>
UGT89C1	Q9LNE6.1	<i>A. thaliana</i>
UGT74AN1	AXF50399.1	<i>Asclepias curassavica</i>
UGT94B1	Q5NTH0.1	<i>Bellis perennis</i>
UGT71F1	AAS94330.1	<i>Beta vulgaris</i>
UGT708G1	BBA18062.1	<i>Citrus japonica</i>
CmF7G2"RT	Q8GVE3.2	<i>Citrus maxima</i>
UGT708G2	BBA18063.1	<i>Citrus unshiu</i>
UGT78K6	4REL_A	<i>Clitoria ternatea</i>
UGT709G1	APU54677.1	<i>Crocus sativus</i>
F7GAT	ANC70234.1	<i>Erigeron breviscapus</i>
UGT708C1	BAP90360.1	<i>Fagopyrum esculentum</i>
FiF3GT	AAD21086.1	<i>Forsythia x intermedia</i>
UGT71A18	BAI65912.1	<i>F. x intermedia</i>
UGT73B4	NP_001354361.1	<i>Glycine max</i>
GmF3G2"GT	BAR88077.1	<i>G. max</i>
GmF3G6"RT	BAN91401.1	<i>G. max</i>
UGT78K1	ADC96620.1	<i>G. max</i>
UGT88E3.	NP_001235161.1	<i>G. max</i>
UGT73P12	BBN60804.1	<i>G. uralensis</i>
VhA5GT	Q9ZR25.1	<i>Glandularia x hybrida</i>
GeIF7GT	BAC78438.1	<i>Glycyrrhiza echinata</i>
UGT73F17	AXS75258.1	<i>Glycyrrhiza uralensis</i>
UGT72B11	ACB56923.1	<i>Hieracium pilosella</i>
HvF3GT	P14726.1	<i>Hordeum vulgare</i>
UGT79G16.	Q53UH5.1	<i>Ipomoea purpurea</i>
UGT71B5b	QTI0875.1	<i>Isatis indigotica</i>
UGT88D2	BAE48240.1	<i>Linaria vulgaris</i>
UGT74S1	AGD95005	<i>Linum usitatissimum</i>
UGT73A10	BAG80536.1	<i>Lycium barbarum</i>

UGT71A12	BAF96585.1	<i>Lycium chinense</i>
UGT71A13	ABL85473.1	<i>Maclura pomifera</i>
UGT75L4	ABL85474.1	<i>M. pomifera</i>
UGT88A4	ABL85471.2	<i>M. pomifera</i>
UGT71G1	XP_003615613.1	<i>Medicago truncatula</i>
UGT78G1	A6XNC6.1	<i>M. truncatula</i>
UGT88F1	NP_001315652.1	<i>Malus domestica</i>
MiCGTb	AMM73095.1	<i>Mangifera indica</i>
Ns3RT	BAC10994.1	<i>Nierembergia sp. NB17</i>
OcUGT1	AWD73588.1	<i>Ornithogalum longebracteatum</i>
UGT71A16	ACZ44836.1	<i>Pyrus communis</i>
UGT71K2	ACZ44837.1	<i>P. communis</i>
UGT88F2	ACZ44838.1	<i>P. communis</i>
Pff3GT	BAA19659.1	<i>P. frutescens</i>
PhA5GT	BAA89009.1	<i>P. x hybrida</i>
PhF3GT	BAA89008.1	<i>P. x hybrida</i>
PfA5GT	Q9ZR27.1	<i>Perilla frutescens</i>
PhA3G6"RT	CAA50376.1	<i>Petunia x hybrida</i>
PtUGT1	BBK15460.1	<i>Polygala tenuifolia</i>
RhA53GT	Q4R1I9.1	<i>Rosa hybrid cultivar</i>
SbB7GAT	Q76MR7.1	<i>Scutellaria baicalensis</i>
SbF7GT	BAA83484.1	<i>S. baicalensis</i>
UGT78B4	QLB54224.1	<i>S. baicalensis</i>
UGT94D1	BAF99027.1	<i>Sesamum indicum</i>
UGT71A9	XP_011100453.1	<i>S. indicum</i>
UGT71E1	Q6VAB2.1	<i>Stevia rebaudiana</i>
UGT74G1	AAR06920.1	<i>S. rebaudiana</i>
UGT71A8	BAF96581.1	<i>Sesamum alatum</i>
UGT71A10	BAF96583.1	<i>Sesamum radiatum</i>
TcCGT1	QCZ42162.1	<i>Trollius chinensis</i>
UGT74M1	ABK76266.1	<i>Vaccaria hispanica</i>
UGT73A16	ACO44747.1	<i>Withania somnifera</i>

**Supplementary Table 4.**  $^1\text{H}$ -NMR,  $^{13}\text{C}$ -NMR spectra of monoglycoside products.

	(+)-Lariciresinol-4- <i>O</i> - $\beta$ -D-Glucoside		(+)-Lariciresinol-4'- <i>O</i> - $\beta$ -D-Glucoside	
	$\delta\text{H}$	$\delta\text{C}$	$\delta\text{H}$	$\delta\text{C}$
1		134.3		132.1
2	6.89 (d, $J = 1.8$ Hz)	109.2	6.79 (d, $J = 1.8$ Hz)	112
3		149.5		149.4
4		145.7		145.9
5	7.09 (d, $J = 8.2$ Hz)	114.6	6.71 (d, $J = 8.0$ Hz)	114.8
6	6.74 (brs)	120.9	6.64 (dd, $J = 8.0, 1.8$ Hz)	120.7
7	2.98 (dd, $J = 13.4, 4.8$ Hz) 2.55 (dd, $J = 13.4, 11.4$ Hz)	32.3	2.90 (dd, $J = 13.5, 5.0$ Hz) 2.50 (dd, $J = 13.4, 11.2$ Hz)	32.2
8	2.72 - 2.77 (m)	42.4	2.68 - 2.74 (m)	42.4
9	3.99 (dd, $J = 8.3, 6.5$ Hz) 3.70 (dd, $J = 8.4, 6.0$ Hz)	72	4.00 (dd, $J = 8.3, 6.7$ Hz) 3.74 (dd, $J = 8.3, 6.4$ Hz)	72.3
3-OCH <sub>3</sub>	3.84 (s)	55	3.83 (s)	55
1'		135.7		138.1
2'	6.90 (d, $J = 1.8$ Hz)	112.9	6.98 (d, $J = 2.0$ Hz)	109.9
3'		147.6		147.6
4'		145		144.4
5'	6.77 - 6.78 (m)	116.9	7.13 (d, $J = 8.4$ Hz)	116.5
6'	6.75 - 6.76 (m)	118.4	6.88 (dd, $J = 8.4, 2.0$ Hz)	118.2
7'	4.75 (d, $J = 6.9$ Hz)	82.6	4.83 (d, $J = 6.4$ Hz)	82.4
8'	2.35 - 2.40 (m)	52.7	2.32 - 2.37 (m)	52.7
9'	3.81 - 3.86 (m) 3.62 - 3.65 (m)	59	3.84 - 3.88 (m) 3.66 - 3.68 (m)	59.1
3'-OCH <sub>3</sub>	3.85 (s)	55.3	3.86 (s)	55.3
Glc-1	4.87 (d, $J = 7.6$ Hz)	101.6	4.88 (d, $J = 7.6$ Hz)	101.5
Glc-2	3.43 - 3.49 (m)	73.5	3.44 - 3.48 (m)	73.5
Glc-3	3.57 - 3.60 (m)	76.4	3.57 - 3.60 (m)	76.4
Glc-4	3.38 - 3.40 (m)	70	3.36 - 3.40 (m)	70
Glc-5	3.66 - 3.68 (m)	76.8	3.63 - 3.66 (m)	76.8
Glc-6	3.85 - 3.88 (m) 3.69 - 3.71 (m)	61.1	3.84 - 3.88 (m) 3.68 - 3.70 (m)	61.1

92 **Supplementary Table 5.** List of primers used in the study.

Primers	Squences
LiUGT1F	GAGGGATCCATGAAGATTACAAGACCACA
LiUGT1R	GAGGCGGCCGCCTAGGCACCACGTGCCATT
LiUGT2F	GAGCATATGGCGAAAGAAACAGAGCTCA
LiUGT2R	GAGGTCGACTCAGCTGAACCATCGTCCA
LiUGT3F	GAGGGATCCATGCAAATCACAAAACCACA
LiUGT3R	GAGGCGGCCGCCTAAGCACCACGTGCCAAGT
LiUGT4F	GAGGGATCCATGAAGACATCGGAGCTAAT
LiUGT4R	GAGGCGGCCGCTAAAAGTGTATCCCCAAGAA
LiUGT5F	GAGGGATCCATGGAGCAGCCTCACGCGCT
LiUGT5R	GAGGTCGACCTATATTATTGCGAATCAC
LiUGT6F	GAGGGATCCATGGAAAAGCAAAACGCAAT
LiUGT6R	GAGGTCGACTCATTTGGGCTCCACGATT
LiUGT7F	GAGGCTAGCATGGGAATCAAGAGATCATC
LiUGT7R	GAGGTCGACCTAAAAGTCCTCGCCGACCA
LiUGT8F	GAGGGATCCATGCAGAATACAAAACCTCA
LiUGT8R	GAGGCGGCCGCTTAGGCACCACGTGCCATGC
LiUGT9F	GAGGGATCCATGGAAAATCAAGAAGCTAT
LiUGT9R	GAGGTCGACTCATTTGAGCTCCACGACT
LiUGT71B5aF	GAGGGATCCATGAAGATCGAGCTCGTGT
LiUGT71B5aR	GAGGCGGCCGCCTAGACCACAACTCTCGAT
138Actin-F	CCAGTGGTCGTACAACCGGTA
138Actin-R	TAGTTCTTCGATGGAGGAGCTG
LiUGT1RT-F	ATCAAAGGGTAAATGGACTG
LiUGT1RT-R	TCAGGTCTTGTCAAATGC
LiUGT4RT-4F	TGCTCTTCTCAAGTTATTG
LiUGT4RT-4R	TGTACCTATATGCAATGTTCTT
LiUGT71B5aRT-F	TTATCCTGTTGAATGCCTTC
LiUGT71B5aRT-R	AATTATCCGATCCGCTTTCA
LiUGT1I-1F	gatacttagttctagagagcttgagggatacggcagagatg
LiUGT1I-1R	cggggaaattcgagctgtgactgcaagtggcgctg
LiUGT1I-2F	tcgagggtacccgggtgagggatacggcagagatg
LiUGT1I-2R	acggggacttagaggtaactgcaagtggcgctg
LiUGT4I2-1F	gatacttagttctagagagctgcacgctgttcaagt
LiUGT4I2-1R	cggggaaattcgagctgcctaaaacatgtaccttatatgcaatg
LiUGT4I2-2F	tcgagggtacccgggacgctgttcaag
LiUGT4I2-2R	acggggacttagaggctaaacatgtaccttatatg
LiUGT71B5aI-1F	gatacttagttctagagagctgacgtgatecgagaatgttg
LiUGT71B5aI-1R	cggggaaattcgagctgttcaag
LiUGT71B5aI-2F	tcgagggtacccgggacgtgtcgagaatgttg
LiUGT71B5aI-2R	acggggacttagaggctgaaccatctcaag

LiUGT1SCF	GCTTCTGCAGGGGCCGGGATGAAGATTACAA GACCACA
LiUGT1SCR	GGATCCACTAGTATTAAATGGGCACCACGTGC CATTCTG
LiUGT4SCF	GCTTCTGCAGGGGCCGGGATGAAACAGGAGC TGGTTTC
LiUGT4SCR	GGATCCACTAGTATTAAATGAGAGATATTGA AGTGACATC
LiUGT71B5aSCF	GCTTCTGCAGGGGCCGGGATGAAGATCGAGC TCGTGTT
LiUGT71B5aSCR	GGATCCACTAGTATTAAATGGACCACAAACATT CTCGATCAC
RolB-F	CGAGGGGATCCGATTGCTT
RolB-R	GACGCCCTCCTCGCCTTCCT
RolC-F	TCGCCATGCCTCACCAACTCAC
RolC-R	CCTTGATCGAGCCGGGTGAGAA
HPT-F	TACACAGCCATCGGTCCAGA
HPT-R	TTAGCGAGAGCCTGACCTATTG
224-35SF	GACGCACAATCCCACATCC
NOS Ter SeqR	ATCATCGCAAGACCGGCAACAG
pZH02-F	GATAAAGAGTACCCACTGTATA
pZH02-R	caaccatgaacattaaagtg
SAIL_LB1	ggataaatagccttgcttcc
NOS Ter SeqR	ATCATCGCAAGACCGGCAACAG
LiUGT4F151A_F	cacctccaacgcgtacgGCTctcggtttg
LiUGT4F151A_R	GCcgtagcggtggagggtgtaaaacatgt
LiUGT4G14A_F	accatcacctggtgacGCCacatcaga
LiUGT4G14A_R	GCgtcaccaggtgatggatgaaaac
LiUGT4H373A_F	ccgaggatttgtcgGCCtgtggttga
LiUGT4H373A_R	GCcgacacaaatcctcgacagcagggt
LiUGT4A207F_F	agtggctaccgattgcgGCTtcacaagtg
LiUGT4A207F_R	GCcgcaatcggtagccactcctgttaa
LiUGT4D124A_F	ctggaatcggtggacGCgtctgcacg
LiUGT4D124A_R	GCgtccaccacgattccagcgagtcgttg
LiUGT4N85D_F	tgtacgccaaaccGACttcctctc
LiUGT4N85D_R	Cgggttggcgcatcagagttgggt
LiUGT4N377A-F	gtcgcaactgtggtggGCCtcgacactgg

LiUGT4N377A-R	GCccaaccacagtgcgacacaaaatcctcg
LiUGT4Q398A_F	gccactctatgccgagGCAcaagttaacg
LiUGT4Q398A_R	GCctcggcatagagtggccacgtggcta
LiUGT4S196A_F	gttaagtgttacccGCTgtgatgtt
LiUGT4S196A_R	Cgggtaaacacttaaccggcaaagga
LiUGT4S289A_F	gttcctctgtttggaGCCatgggaggtt
LiUGT4S289A_R	GCtccaaaacagaggaacacaacggtt
LiUGT4S289W_F	gttcctctgtttggaTGGatgggaggtt
LiUGT4S289W_R	CCAtccaaaacagaggaacacaacggtt
LiUGT4S378A-F	gcactgtggttgaaacGCGacactggag
LiUGT4S378A-R	Cgttccaaccacagtgcgacacaaaatc
LiUGT4W376A_F	gtgtcgcaactgtggtGCGaactcgacac
LiUGT4W376A_R	GCaccacagtgcgacacaaaatcctcg
LiUGT4Y395F-F	agccacgtggccactcTTTgccgagcaac
LiUGT4Y395F-R	Aagagtggccacgtggctatggaacacc

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93 **Supplementary Table 6.** Mass spectrometry condition parameters of multi-target  
 94 ingredients in *I. indigotica* Fort.

Compounds	Molecular formulas	Rt (min)	[M-H] <sup>-</sup>	Negative ion mode Major Fragments
Coniferyl alcohol	C <sub>10</sub> H <sub>12</sub> O <sub>3</sub>	1.95	179.07	179.07,146.21,164.04
Coniferin	C <sub>16</sub> H <sub>22</sub> O <sub>8</sub>	1.29	341.34	341.34,179.01
Pinoresinol	C <sub>20</sub> H <sub>22</sub> O <sub>6</sub>	3.32	357.38	357.38,357.38,342.10,150.81
Pinoresinol diglucoside	C <sub>32</sub> H <sub>42</sub> O <sub>16</sub>	1.78	682.66	680.90,519.02
Lariciresinol	C <sub>20</sub> H <sub>24</sub> O <sub>6</sub>	2.84	360.43	359.10,328.90
(+)-Lariciresinol-4- <i>O</i> - $\beta$ -D-glucoside	C <sub>26</sub> H <sub>34</sub> O <sub>11</sub>	2.07	522.54	521.20,359.00,329.10
(+)-Lariciresinol-4'- <i>O</i> - $\beta$ -D-glucoside	C <sub>26</sub> H <sub>34</sub> O <sub>11</sub>	2.17	523.54	521.20,359.00,329.10
Clemastanin B	C <sub>32</sub> H <sub>44</sub> O <sub>16</sub>	1.58	684.68	683.20,521.10,359.00
Secoisolariciresinol	C <sub>20</sub> H <sub>26</sub> O <sub>6</sub>	2.71	362.41	361.14,179.91,164.92
Secoisolariciresinol diglucoside	C <sub>32</sub> H <sub>46</sub> O <sub>16</sub>	2.11	686.69	684.94,523.24
Matairesinol	C <sub>20</sub> H <sub>22</sub> O <sub>6</sub>	3.58	358.38	342.12,137.02,122.01