

Isolation and Characterization of Human Intestinal Bacteria

Cytobacillus oceanisediminis NB2 for probiotic potential

Monika Yadav¹, Tarun Kumar¹, Akshay Kanakan², Ranjeet Maurya^{2,3}, Rajesh Pandey^{2†}, and Nar Singh Chauhan^{1*}

¹Department of Biochemistry, Maharshi Dayanand University, Rohtak, Haryana, India

²Integrative GENomics of Host-PathogEn (INGEN-HOPE) laboratory, CSIR-Institute of Genomics and Integrative Biology (CSIR-IGIB), Mall Road, Delhi-110007, India

³Academy of Scientific and Innovative Research (AcSIR), Ghaziabad-201002, India

*Corresponding author

Nar Singh Chauhan (nschauhan@mdurohtak.ac.in)

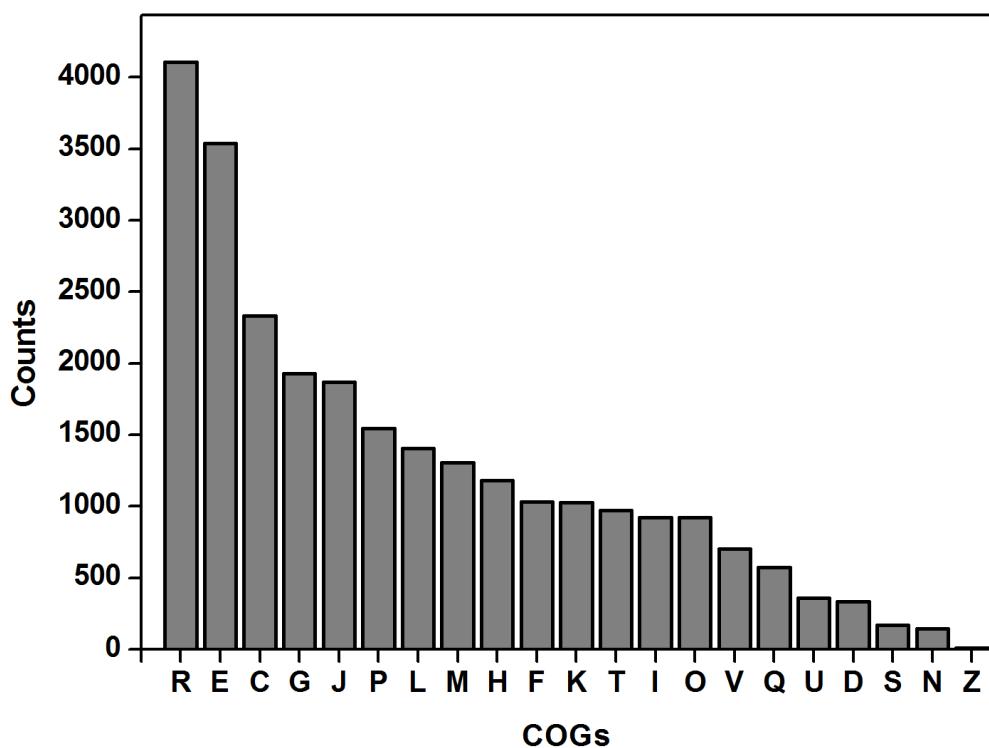
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Supplementary Figure S1. COG categories observed within the genome of *Cytobacillus oceanisediminis* NB2 where R codes for General function prediction only, E codes for Amino acid transport and metabolism, C codes for Energy production and conversion, G codes for Carbohydrate transport and metabolism, J codes for Translation, ribosomal structure and biogenesis, P codes for Inorganic ion transport and metabolism, L codes for Replication, recombination and repair, M codes for Cell wall/membrane/envelope biogenesis, H codes for Coenzyme transport and metabolism, F codes for Nucleotide transport and metabolism, K codes for Transcription, T codes for Signal transduction mechanisms, I codes for Lipid transport and metabolism, O codes for Posttranslational modification, protein turnover, chaperones, V codes for Defense mechanisms, Q codes for Secondary metabolites biosynthesis, transport and catabolism, U codes for Intracellular trafficking, secretion, and vesicular transport, D codes for Cell cycle control, cell division, chromosome partitioning, S codes for Function unknown, N codes for Cell motility, and Z codes for Cytoskeleton.



Supplementary Table S1. Genomic features of *Cytobacillus oceanisediminis* NB2 and other *Bacillus* strains used for comparative genome analysis.

Organism	Strain	Isolation source	Genome Size (Mb)	CDS	tRNA	rRNA	Genome Reference (NCBI ID)
<i>Cytobacillus oceanisediminis</i>	NB2	Human feces	5.2	5195	108	19	SUB11205683
<i>Bacillus coagulans</i>	LA204	Soil	3.37	2975	84	30	NZ_CP009709.1
<i>Bacillus coagulans</i>	strain HM-08	healthy Chicken intestine	3.37	2975	84	30	CP010525.1
<i>Bacillus subtilis</i>	strain UD1022	Root	4.02	3,933	86	30	CP011534.1
<i>Bacillus subtilis</i>	strain B-1	Oil field	3.94	3706	59	19	CP009684.1
<i>Bacillus clausii</i>	ENTPro	Soil and Water	4.44	4323	59	5	CP012475.1
<i>Cytobacillus oceanisediminis</i>	2691	Intertidal sediment	5.4	5679	106	33	CP015506.1
<i>Cytobacillus oceanisediminis</i>	strain YPW-V2	rhizosphere taken during the low tide period	5.33	5507	107	36	NZ_CP062790.1
<i>Bacillus infantis</i>	strain AcN21-9_AcN21-9_ctg006	sediment	5.84	4911	83	19	NZ_JAIVAO010000006.1
<i>Bacillus firmus</i>	NCTC10335	type strain of <i>Bacillus firmus</i>	4.8	4,436	36	108	UFTC000000000.1
<i>Bacillus subtilis</i> subsp. <i>subtilis</i>	str. 168	type strain of <i>Bacillus firmus</i>	4.22	4,237	30	86	AL009126.3
<i>Bacillus subtilis</i> subsp. <i>spizizenii</i>	str. W23	type strain of <i>Bacillus firmus</i>	4.03	3,912	24	77	CP002183.1
<i>Bacillus velezensis</i>	Strain BIM B-454D	Soil	4.2	4091	86	27	NZ_CP082262.1

Supplementary Table S2. A comparison of the growth conditions of *Cytobacillus oceanisediminis* NB2 with other *Bacillus* strains.

Sr. No.	NaCl	KCl	Organism	pH	Temperature (°C)	Arsenic	Cadmium chloride	Lead acetate	Silver nitrate
1	Up to 10%	ND	<i>Bacillus cereus</i> strain 01552	4.35-7.85	4.9-10	10µM	1.6308g/L	1.8307g/L	10µM
2	Up to 10%	ND	<i>Bacillus cereus</i> strain 5064	4.35-7.85	ND	ND	ND	ND	ND
3	Up to 15%	ND	<i>Bacillus subtilis</i> strain AS-4	6-8	17-37	ND	1000µg/L	1000µg/L	ND
4	Up to 10%	ND	<i>Bacillus cereus</i>	4.6-7.5	7-55	ND	ND	ND	ND
5	Up to 7%	ND	<i>Bacillus cereus</i>	4.9-9.3	4.5-55	ND	ND	ND	ND
6	0.5M	0.1M	<i>Bacillus subtilis</i>	4-9.5	Upto 60	10µM	1000µg/L	1000µg/L	10µM
7	No growth in 7%	ND	<i>Bacillus licheniformis</i>	5-9	Upto 135°C	ND	1.6308g/L	1.8307g/L	ND
8	No growth in 7%	ND	<i>Bacillus pumilus</i>	ND	ND	ND	ND	ND	ND
9	Up to 12%	ND	<i>Bacillus velezensis</i>	ND	15-45°C	ND	ND	ND	ND
10	Up to 10%	ND	<i>Bacillus safensis</i>	ND	ND	2mM	ND	ND	ND
11	ND	ND	<i>Bacillus subtilis</i> FNCC 0059	4-8.5	25-45°C	ND	1.6308g/L	1.8307g/L	ND
12	ND	ND	<i>Bacillus alcalophilus</i>	8-14	ND	ND	ND	ND	ND
13	Up to 7%	Up to 10%	<i>Cytobacillus oceanisediminis</i> NB2	5-9	20-40°C	10mM	ND	2mM	2mM

Supplementary Table S3. Nucleotide similarity of 16S rRNA gene of *Cytobacillus oceanisediminis* NB2 with other *Bacillus* strains.

Sr. No.	Matched organism	Accession Id	% identity	E value
1	<i>Cytobacillus oceanisediminis</i> 2691	CP015506.1	99	0.0
2	<i>Cytobacillus oceanisediminis</i> strain YPW-V2	CP062790.1	99	0.0
3	<i>Bacillus firmus</i>	AY833571.2	96	0.0
4	<i>Bacillus</i> sp. SW41	HM584798.1	99	0.0
5	<i>Bacillus firmus</i> isolate CV93b	AJ717384.1	99	0.0
6	<i>Bacillus oceanisediminis</i> strain NIOT- Ba-6	KJ575007.1	99	0.0
8	<i>Bacillus firmus</i> strain XJSL1-4	GQ903383.1	99	0.0

Supplementary Table S4. Carbohydrate utilization profile of *Cytobacillus oceanisediminis* NB2 in comparison to phylogenetically related *Bacillus* strains

Sr.No.	Substrate	<i>Cytobacillus oceanisediminis</i> NB2	<i>Bacillus oceanisediminis</i> sp. nov. H2	<i>Bacillus</i> strain YAS1	<i>Bacillus clausii</i>	<i>Bacillus megaterium</i>	<i>Bacillus firmus</i> RAB
1	Lactose	ND	+	+	+	+	ND
2	Xylose	+	ND	ND	+	ND	ND
3	Maltose	+	+	+	+	+	ND
4	Fructose	ND	ND	+	+	ND	+
5	Dextrose	ND	ND	+	+	+	+
6	Galactose	ND	+	+	+	ND	ND
7	Raffinose	+	ND	ND	ND	ND	ND
8	Trehalose	+	+	ND	+	ND	ND
9	Melibiose	+	ND	ND	ND	ND	ND
10	Sucrose	+	ND	ND	+	+	ND
11	L-arabinose	+	ND	ND	+	ND	ND
12	Mannose	+	+	ND	+	ND	ND
13	Inulin	ND	+	ND	+	ND	ND
14	Sodium glucan	ND	ND	ND	ND	ND	ND
15	Glycerol	ND	+	+	+	ND	ND
16	Salicin	ND	ND	ND	ND	ND	ND
17	Dulcitol	ND	ND	ND	ND	ND	ND
18	Inositol	ND	+	ND	ND	ND	ND
19	Sorbitol	ND	+	ND	+	ND	ND
20	Mannitol	ND	+	-	+	+	ND
21	Adonitol	ND	ND	ND	ND	ND	ND
22	Arabitol	ND	ND	ND	ND	ND	ND
23	Erythritol	ND	ND	ND	ND	ND	ND
24	α -Methyl-D	ND	ND	ND	ND	ND	ND
25	Rhamnose	ND	ND	ND	+	ND	ND
26	Cellobiose	ND	ND	ND	ND	ND	ND

27	Melezitose	ND	ND	ND	ND	ND	ND
28	α -Methyl-D-mannoside	ND	ND	ND	ND	ND	ND
29	Xylitol	ND	+	ND	ND	ND	ND
30	ONPG	ND	ND	ND	ND	ND	ND
31	Esculin hydrolysis	+	ND	+	ND	ND	ND
32	D-arabinose	ND	ND	ND	ND	ND	ND
33	Citrate utilization	+	ND	ND	ND	ND	ND
34	Malonate utilization	ND	ND	ND	ND	ND	ND
35	Sorbose	ND	ND	ND	ND	ND	ND

Supplementary Table S5. Antibiotic susceptibility of *Cytobacillus oceanisediminis* NB2 in comparison to phylogenetically related *Bacillus* strains.

Sr. No.	Antibiotic	<i>Cytobacill us oceanisedi minis</i> <i>NB2</i>	<i>Bacillus oceanisedi minis</i>	<i>Bacillus</i> sp., strain YAS1	<i>Bacillus clausii</i>	<i>Bacillus megaterium</i>	<i>Bacillus firmus</i>
1	Amikacin	ND	ND	ND	ND	ND	ND
2	Amoxicillin	ND	ND	ND	ND	ND	ND
3	Bacitracin	ND	ND	ND	ND	ND	ND
4	Cephalothin	+	ND	ND	ND	ND	+
5	Erythromycin	ND	ND	ND	+	ND	ND
6	Novobiocin	ND	ND	ND	ND	ND	ND
7	Oxytetracycline	ND	ND	ND	ND	+	ND
8	Vancomycin	ND	ND	ND	ND	+	+
9	Ceflaxzone	+	ND	ND	ND	ND	ND
10	Ceftazidime	+	ND	ND	ND	ND	ND
11	Cefotaxime	ND	ND	ND	ND	ND	ND
12	Lincomycin	ND	ND	ND	+	ND	ND
13	Netilin	ND	ND	ND	ND	ND	ND
14	Ofloxacin	+	ND	ND	ND	ND	ND

Supplementary Table S6. BUSCO assessment of genome of *Cytobacillus oceanisediminis* NB2.

Assembly	<i>Cytobacillus oceanisediminis</i> NB2 contigs
# contigs (>= 0 bp)	203
# contigs (>= 1000 bp)	30
# contigs (>= 5000 bp)	25
# contigs (>= 10000 bp)	25
# contigs (>= 25000 bp)	23
# contigs (>= 50000 bp)	23
Total length (>= 0 bp)	5235740
Total length (>= 1000 bp)	5170386
Total length (>= 5000 bp)	5163368
Total length (>= 10000 bp)	5163368
Total length (>= 25000 bp)	5135080
Total length (>= 50000 bp)	5135080
# contigs	56
Largest contig	705313
Total length	5187457
GC (%)	41.41
N50	289521
N75	204462
L50	5
L75	10
# N's per 100 kbp	0

Supplementary Table S7. Comparison of the genome features of *Cytobacillus oceanisediminis* NB2 with reference genomes of the *Bacillus* species

Genome feature	<i>Cytobacillus oceanisediminis</i> NB2	<i>Bacillus clausii</i> strain ENTPro	<i>Bacillus coagulans</i> strain HM-08	<i>Bacillus infantis</i> NRRL B-14911	<i>Bacillus subtilis</i> strain B-1	<i>Bacillus velezensis</i> strain BIM B-454D	<i>Cytobacillus firmus</i> strain NCTC10335	<i>Cytobacillus oceanisediminis</i> 2691
Bases (Mb)	5.23	4.44	3.37	4.88	4.22	4.01	4.80	5.63
CDS	5195	4446	3057	4707	4237	3761	4775	5297
rRNA	19	5	30	27	30	21	36	18
tRNA	108	59	84	86	86	81	107	77

Supplementary Table S8. Subsystems related protein features for the resistance to metal/metalloid resistance identified in the genome of *Cytobacillus oceanisediminis* NB2.

Nature of Resistance	Identified Protein Feature	No. of gene features
Arsenic resistance	Arsenate reductase (EC 1.20.4.1)	18
Arsenic resistance	Arsenic efflux pump protein	33
Arsenic resistance	Arsenical resistance operon repressor	4
Arsenic resistance	Arsenical-resistance protein ACR3	4
Cadmium resistance	Cadmium efflux system accessory protein	6
Cadmium resistance	Cadmium-transporting ATPase (EC 3.6.3.3)	65
Resistance to chromium compounds	Chromate transport protein ChrA	8
Cobalt-zinc-cadmium resistance	Cobalt-zinc-cadmium resistance protein	66
Cobalt-zinc-cadmium resistance	Cobalt-zinc-cadmium resistance protein CzcD	10
Copper homeostasis:copper tolerance	Cytoplasmic copper homeostasis protein cutC	2
Copper homeostasis: copper tolerance	Magnesium and cobalt efflux protein CorC	5
Zinc resistance	Response regulator of zinc sigma-54-dependent two-component system	2
Cobalt-zinc-cadmium resistance	Transcriptional regulator, MerR family	31

Supplementary Table S9. List of CAZzymes determined within *Cytobacillus oceanisediminis* NB2 genome.

Sr No.	Enzyme family	Activities in Family	Hits
1	Carbohydrate Esterase Family 4	Acetyl xylan esterase; Chitin deacetylase; Chitooligosaccharide-deacetylase; Peptidoglycan GlcNAc-deacetylase; Peptidoglycan N-acetylmuramic acid deacetylase	9
2	Glycoside Hydrolase Family 43 / Subf 11	β -1,3-xylosidase; Xylan 1,4- β -xylosidase; α -L-arabinofuranosidase	1
3	Glycosyl Transferase Family 27	Polypeptide α -N-acetylgalactosaminyltransferase	1
4	Glycosyl Transferase Family 4	Sucrose synthase; Sucrose-phosphate synthase; α -glucosyltransferase; Lipopolysaccharide N-acetylglucosaminyltransferase; Phosphatidylinositol α -mannosyltransferase; GDP-Man: Man1GlcNAc2-PP-dolichol α -1,3-mannosyltransferase; GDP-Man: Man3GlcNAc2-PP-dolichol/Man4GlcNAc2-PP-dolichol α -1,2-mannosyltransferase; Digalactosyldiacylglycerol synthase; 1,2-diacylglycerol 3-glucosyltransferase; Dиглucosyldиacylglycerol synthase; Trehalosephosphorylase; NDP-Glc: α -glucose α -glucosyltransferase / α , α -trehalose synthase; GDP-Man: Man2GlcNAc2-PP-dolichol α -1,6-mannosyltransferase; UDP-GlcNAc: 2-deoxystreptamine α -N-acetylglucosaminyltransferase; UDP-GlcNAc: ribostamycin α -N-acetylglucosaminyltransferase; UDP-Gal α -galactosyltransferase; UDP-Xyl α -xylosyltransferase; UDP-GlcA α -glucuronyltransferase; UDP-Glc α -glucosyltransferase; UDP-GalNAc: GalNAc-PP-Und α -1,3-N-acetylgalactosaminyltransferase; UDP-GalNAc: N,N'-diacetylbacillosaminyl-PP-Und α -1,3-N-acetylgalactosaminyltransferase; ADP-dependent α -maltose-1-phosphate synthase; UDP-GlcNAc: polypeptide α -N-acetylglucosaminyltransferase; UDP-GlcNAc: α -N-acetylglucosaminyltransferase	11
5	Carbohydrate-Binding Module Family 50	Modules of approx. 50 residues found attached to various enzymes from families GH18, GH19, GH23, GH24, GH25 and GH73, i.e. enzymes	6

		cleaving either chitin or peptidoglycan	
6	Glycoside Hydrolase Family 109	α -N-acetylgalactosaminidase; β -N-acetylhexosaminidase	7
7	Glycoside Hydrolase Family 13 / Subf 31	Hexosyltransferases; Oligosaccharide a-4-glucosyltransferase; Palatinase; α -amylase; Oligo-a-1,6-glucosidase; α -glucosidase; Glucodextranase; Isomaltulose synthase /sucrose isomerase /sucrose glucosylmutase	3
8	Glycosyl Transferase Family 28	1,2-diacylglycerol 3- β -galactosyltransferase; 1,2-diacylglycerol 3- β -glucosyltransferase; UDP-GlcNAc: Und-PP-MurAc-pentapeptide β -N-acetylglucosaminyltransferase; Digalactosyldiacylglycerol synthase	3
9	Glycoside Hydrolase Family 32	Invertase; Endo-inulinase; β -2,6-fructan 6-levanbiohydrolase; Endo-levanase; exo-inulinase; Fructan β -(2,1)-fructosidase/1-exohydrolase; Fructan β -(2,6)-fructosidase/6-exohydrolase; Sucrose:sucrose 1-fructosyltransferase; Fructan:fructan 1-fructosyltransferase; Sucrose:fructan 6-fructosyltransferase; Fructan:fructan 6G-fructosyltransferase; Levanfructosyltransferase; Sucrose:sucrose 6-fructosyltransferase (6-SST); Cycloinulo-oligosaccharide fructanotransferase	1
10	Carbohydrate Esterase Family 14	N-acetyl-1-D-myo-inositol-2-amino-2-deoxy- α -D-glucopyranosidedeacetylase; Diacetylchitobiosedeacetylase; Mycothiol S-conjugate amidase	3
11	Carbohydrate-Binding Module Family 41; Glycoside Hydrolase Family 13 / Subf 41	Modules of approx. 100 residues found in primarily in bacterial pullulanases; [retaining] α -amylase	2
12	Glycosyl Transferase Family 51	Murein polymerase	6
13	Glycoside Hydrolase Family 31	α -glucosidase; α -galactosidase; α -mannosidase; α -1,3-glucosidase; Sucrase-isomaltase; α -xylosidase; α -glucanlyase; isomaltosyltransferase; Oligosaccharide α -1,4-glucosyltransferase; α -N-acetylgalactosaminidase; Sulfoquinovosidase; α -6-glucosyltransferase	2
14	Carbohydrate-Binding Module Family 32	Binding to galactose and lactose	1

15	Carbohydrate Esterase Family 9	N-acetylglucosamine 6-phosphate deacetylase; N-acetylgalactosamine 6-phosphate deacetylase	2
16	Auxiliary Activity Family 4	Vanillyl-alcohol oxidase	5
17	Auxiliary Activity Family 3	Cellbiose dehydrogenase; Glucose 1-oxidase; aryl alcohol oxidase; Alcohol oxidase; pyranose oxidase; Glucose 1-dehydrogenase (FAD, quinone); Pyranose dehydrogenase; Oligosaccharide dehydrogenase (FAD)	1
18	Glycoside Hydrolase Family 18	Chitinase; Lysozyme; Endo- β -N-acetylglucosaminidase; Peptidoglycan hydrolase with endo- β -N-acetylglucosaminidase specificity; Nod factor hydrolase; Xylanase inhibitor; Concanavalin B; Narbonin	5
19	Glycoside Hydrolase Family 13 / Subf 9	α -1,4-glucan branching enzyme	1
20	Glycosyl Transferase Family 5	UDP-Glc: glycogen glucosyltransferase; ADP-Glc: starch glucosyltransferase; NDP-Glc: starch glucosyltransferase; UDP-Glc: α -1,3-glucan synthase UDP-Glc: α -1,4-glucan synthase	1
21	Glycosyl Transferase Family 35	Glycogen or starch phosphorylase	1
22	Carbohydrate-Binding Module Family 68; Glycoside Hydrolase Family 13 / Subf 14	Binding to maltotriose and maltotetraose; α -glycosidase; α -amylase; Pullulanase; Isoamylase	1
23	Glycoside Hydrolase Family 170	6-phospho-N-acetylmuramidase	1
24	Carbohydrate Esterase Family 9	N-acetylglucosamine 6-phosphate deacetylase; N-acetylgalactosamine 6-phosphate deacetylase	2
25	Glycoside Hydrolase Family 73	Lysozyme; Mannosyl-glycoprotein endo- β -N-acetylglucosaminidase; Peptidoglycan hydrolase with endo- β -N-acetylglucosaminidase specificity	1
26	Glycoside Hydrolase Family 4	Maltose-6-phosphate glucosidase; α -glucosidase; α -galactosidase; 6-phospho- β -glucosidase; α -glucuronidase; α -galacturonase; Palatinase	1
27	Auxiliary Activity Family 6	1,4-benzoquinone reductase	1
28	Glycoside Hydrolase Family 171	Peptidoglycan β -N-acetylmuramidase	1

29	Glycoside Hydrolase Family 3	β -glucosidase; Xylan 1,4- β -xylosidase; β -glucosylceramidase; β -N-acetylhexosaminidase; α -L-arabinofuranosidase; Glucan 1,4- β -glucosidase; Isopimeloverose-producing oligoxyloglucan hydrolase; Coniferin β -glucosidase; Exo-1,3-1,4-glucanase; β -N-acetylglucosaminidephosphorylases; β -1,2-glucosidase; β -1,3-glucosidase; Xyloglucan-specific exo- β -1,4-glucanase / exo-xyloglucanase; Stevioside- β -1,2-glucosidase; Lichenase / endo- β -1,3-1,4-glucanase; Protodioscin 26-O- β -D-glucosidase	1
30	Glycoside Hydrolase Family 1	β -glucosidase; β -galactosidase; β -mannosidase; β -glucuronidase; β -xylosidase; β -D-fucosidase; Phlorizin hydrolase; Exo- β -1,4-glucanase; 6-phospho- β -galactosidase; 6-phospho- β -glucosidase; Strictosidine β -glucosidase; Lactase; Amygdalin β -glucosidase; Prunasin β -glucosidase; Vicianin hydrolase; Raucaffricine β -glucosidase; Thioglucosidase; β -primeverosidase; Isoflavonoid 7-O- β -apiosyl- β -glucosidase; ABA-specific β -glucosidase; DIMBOA β -glucosidase; β -glycosidase; Hydroxyisourate hydrolase; β -rutinosidase / α -L-rhamnose-(1,6)- β -D-glucosidase; Protodioscin 26-O- $\hat{\beta}$ -D-glucosidase	1
31	Glycoside Hydrolase Family 13 / Subf 1	Hexosyltransferases; 4-a-glucanotransferase / amylo maltase; α -amylase; Maltotriose-producing a-amylase	1
32	Carbohydrate-Binding Module Family 34; Glycoside Hydrolase Family 13 / Subf 20	Modules of approx. 120 residues; Cyclic a-1,6-maltosyl-maltose hydrolase; α -glycosidase hydrolyzing pullulan, starch and g-cyclodextrin; α -amylase; Maltogenic α -amylase; Neopullulanase; Pullulanase; Cyclomaltodextrinase	1
33	Glycoside Hydrolase Family 13 / Subf 31	Hexosyltransferases; Oligosaccharide α -4-glucosyltransferase; Palatinase; α -amylase; Oligo-a-1,6-glucosidase; α -glucosidase; Glucodextranase; Isomaltulose synthase / sucrose isomerase / sucrose glucosylmutase	1
34	Glycoside Hydrolase Family 13 / Subf 29	α -glucosidase; α , α -trehalose-6-phosphate hydrolase	1
35	Glycoside Hydrolase Family 23	Lysozyme type G; Peptidoglycan lyase also known in the literature as peptidoglycan lytic transglycosylase; Chitinase	1

Supplementary Table S10. Tetra correlation among *Cytobacillus oceanisediminis* NB2 and other *Bacillus* highlighted by a wide distribution of z-scores

Organism name	Z-score
<i>Cytobacillus oceanisediminis</i> 2691	0.99806
<i>Cytobacillus firmus</i> NBRC 15306	0.99669
<i>Cytobacillus firmus</i> LK28	0.99604
<i>Cytobacillus firmus</i> DS1	0.99583
<i>Cytobacillus firmus</i> NCTC10335	0.99486
<i>Sporosarcina globispora</i> DSM 4	0.99381
<i>Bacillus saganii</i> V47-23a	0.96384
<i>Bacillus acidicola</i> FJAT-2406	0.95647
<i>Mesobacillus foraminis</i> CV53	0.95066
<i>Bacillus</i> sp. SG-1	0.94954
<i>Bacillus</i> sp. NRRL B-14911	0.94889
<i>Bacillus infantis</i> NRRL B-14911	0.94763
<i>Bacillus massiliiglaciei</i> Marseille-P2600	0.94738
<i>Bacillus glennii</i> V44-8	0.9438
<i>Bacillus salacetis</i> SKP7-4	0.94353
<i>Cytobacillus gottheilii</i> FJAT-2394	0.94319
<i>Fictibacillus solisalsi</i> CGMCC 1.6854	0.94182
<i>Peribacillus cavernae</i> L5	0.93871
<i>Fictibacillus enclensis</i> NIO-1003	0.93859
<i>Bacillus</i> sp. FJAT-27986	0.93546
<i>Bacillus</i> sp. SJS	0.93284
<i>Bacillus oleivorans</i> JC228	0.93039
<i>Peribacillus kribbensis</i> DSM 17871	0.92906
<i>Bacillus badius</i> DSM 5610	0.92874
<i>Peribacillus simplex</i> P558	0.92856
<i>Bacillus capparidis</i> DSM 103394	0.92848
<i>Peribacillus simplex</i> SH-B26	0.92846
<i>Bacillus gobiensis</i> FJAT-4402	0.92785
<i>Bacillus badius</i> DSM 30822	0.92777
<i>Bacillus badius</i> SGD-V-25	0.92774
<i>Bacillus badius</i> SGD-V-25	0.92774

<i>Bacillus badius</i> MTCC 1458	0.92756
<i>Bacillaceae bacterium</i> MTCC 10057	0.92742
<i>Quasibacillus thermotolerans</i> MTCC 8252	0.92714
<i>Mesobacillus campialis</i> SA2-6	0.92703
<i>Bacillus</i> sp. FJAT-22058	0.92697
<i>Halobacillus massiliensis</i> Marseille-P3554	0.92628
<i>Peribacillus simplex</i> NBRC 15720 = DSM 1321 NBRC 15720	0.92616
<i>Quasibacillus thermotolerans</i> SGZ-8	0.92612
<i>Bacillus</i> sp. Soil745	0.92599
<i>Bacillus badius</i> NBRC 15713	0.92578
<i>Domibacillus antri</i> XD80	0.92567
<i>Domibacillus epiphyticus</i> SAB 38	0.92535
<i>Mesobacillus zae</i> JJ-247	0.92533
<i>[Brevibacterium] frigoritolerans</i> FJAT-2396	0.9253
<i>Bacillus</i> sp. FJAT-21352	0.9252
<i>Bacillus methanolicus</i> PB1	0.92423
<i>Lysinibacillus odysseyi</i> 34hs-1 = NBRC 100172	0.92342
<i>Priestia abyssalis</i> DSM 25875	0.92321
<i>Siminovitchia fordii</i> DSM 16014 = CIP 108821 DSM 16014	0.92234
<i>Falsibacillus pallidus</i> DSM 25281	0.92197
<i>Jeotgalibacillus campialis</i> SF-57	0.92121
<i>Bacillus</i> sp. FJAT-27225	0.92111
<i>Cytobacillus praedii</i> FJAT-25547	0.92024
<i>Bacillus badius</i> DSM 30822	0.92777
<i>Bacillus badius</i> SGD-V-25	0.92774
<i>Bacillus badius</i> MTCC 1458	0.92756
<i>Bacillaceae bacterium</i> MTCC 10057	0.92742
<i>Quasibacillus thermotolerans</i> MTCC 8252	0.92714
<i>Mesobacillus campialis</i> SA2-6	0.92703
<i>[Brevibacterium] frigoritolerans</i> FJAT-2396	0.9253
<i>Bacillus</i> sp. FJAT-21352	0.9252
<i>Bacillus methanolicus</i> PB1	0.92423
<i>Lysinibacillus odysseyi</i> 34hs-1 = NBRC 100172	0.92342
<i>Priestia abyssalis</i> DSM 25875	0.92321
<i>Lysinibacillus odysseyi</i> 34hs-1 = NBRC 100172	0.92319
<i>Siminovitchia fordii</i> DSM 16014 = CIP 108821 DSM 16014	0.92234
<i>Falsibacillus pallidus</i> DSM 25281	0.92197

<i>Jeotgalibacillus campialis</i> SF-57	0.92121
<i>Bacillus</i> sp. FJAT-27225	0.92111
<i>Cytobacillus praedii</i> FJAT-25547	0.92024
<i>[Brevibacterium] frigoritolerans</i> FJAT-2396	0.9253
<i>Bacillus</i> sp. FJAT-21352	0.9252
<i>Bacillus methanolicus</i> PB1	0.92423
<i>Salipaludibacillus aurantiacus</i> S9	0.91951
<i>Bacillus methanolicus</i> MGA3	0.91884
<i>Cytobacillus eiseniae</i> FJAT-2352	0.91864
<i>Neobacillus fumarioli</i> NBRC 102428	0.91848
<i>Bacillus methanolicus</i> MGA3	0.9183
<i>Bacillus</i> sp. FJAT-27231	0.9171
<i>Bacillus wudalianchiensis</i> FJAT-27215	0.91518
<i>Bacillus massiliigabonensis</i> Marseille-P2639	0.91464
<i>Bacillus</i> sp. Leaf13	0.91381
<i>Bacillus</i> sp. FJAT-18017	0.91373
<i>Bacillus</i> sp. FJAT-21945	0.91335
<i>Peribacillus butanolivorans</i> DSM 18926	0.91331
<i>Bacillus</i> sp. Soil768D1	0.91316
<i>Jeotgalibacillus proteolyticus</i> 22-7	0.91296
<i>Jeotgalibacillus soli</i> P9	0.91272
<i>Neobacillus mesonae</i> FJAT-13985	0.91262
<i>Virgibacillus indicus</i> P2-C2	0.9125
<i>Domibacillus iocasae</i> DSM 29979	0.91111
<i>Oceanobacillus damuensis</i> PT-20	0.91098
<i>Cytobacillus solani</i> FJAT-18043	0.91086
<i>Bacillus</i> sp. SA1-12	0.91066
<i>Peribacillus psychrosaccharolyticus</i> ATCC 23296	0.90911
<i>Mesobacillus subterraneus</i> DSM 13966	0.90885
<i>Halobacillus karajensis</i> DSM 14948	0.90842
<i>Mesobacillus selenatarsenatis</i> SF-1	0.90834
<i>Bacillus dielmonensis</i> null	0.90825
<i>Lysinibacillus yapensis</i> YLB-03	0.90802
<i>Metabacillus indicus</i> LMG 22858	0.90801
<i>Salipaludibacillus aurantiacus</i> S9	0.91951
<i>Bacillus methanolicus</i> MGA3	0.91884
<i>Cytobacillus eiseniae</i> FJAT-2352	0.91864
<i>Neobacillus fumarioli</i> NBRC 102428	0.91848
<i>Bacillus methanolicus</i> MGA3	0.9183
<i>Bacillus</i> sp. FJAT-27231	0.9171
<i>Bacillus wudalianchiensis</i> FJAT-27215	0.91518
<i>Bacillus massiliigabonensis</i> Marseille-P2639	0.91464
<i>Bacillus</i> sp. Leaf13	0.91381
<i>Bacillus</i> sp. FJAT-18017	0.91373
<i>Bacillus</i> sp. FJAT-21945	0.91335
<i>Peribacillus butanolivorans</i> DSM 18926	0.91331

<i>Bacillus</i> sp. Soil768D1	0.91316
<i>Jeotgalibacillus proteolyticus</i> 22-7	0.91296
<i>Jeotgalibacillus soli</i> P9	0.91272
<i>Neobacillus mesonae</i> FJAT-13985	0.91262
<i>Virgibacillus indicus</i> P2-C2	0.9125
<i>Domibacillus iocasae</i> DSM 29979	0.91111
<i>Oceanobacillus damuensis</i> PT-20	0.91098
<i>Cytobacillus solani</i> FJAT-18043	0.91086
<i>Bacillus</i> sp. SA1-12	0.91066
<i>Peribacillus psychrosaccharolyticus</i> ATCC 23296	0.90911
<i>Mesobacillus subterraneus</i> DSM 13966	0.90885
<i>Halobacillus karajensis</i> DSM 14948	0.90842
<i>Mesobacillus selenatarsenatis</i> SF-1	0.90834
<i>Bacillus dielmonensis</i> null	0.90825
<i>Lysinibacillus yapensis</i> YLB-03	0.90802
<i>Metabacillus indicus</i> LMG 22858	0.90801
<i>Salipaludibacillus aurantiacus</i> S9	0.91951
<i>Bacillus methanolicus</i> MGA3	0.91884
<i>Cytobacillus eiseniae</i> FJAT-2352	0.91864
<i>Neobacillus fumarioli</i> NBRC 102428	0.91848
<i>Bacillus methanolicus</i> MGA3	0.9183
<i>Bacillus</i> sp. FJAT-27231	0.9171
<i>Bacillus wudalianchiensis</i> FJAT-27215	0.91518
<i>Bacillus massiliigabonensis</i> Marseille-P2639	0.91464
<i>Bacillus</i> sp. Leaf13	0.91381
<i>Bacillus</i> sp. FJAT-18017	0.91373
<i>Bacillus</i> sp. FJAT-21945	0.91335
<i>Peribacillus butanolivorans</i> DSM 18926	0.91331
<i>Bacillus</i> sp. Soil768D1	0.91316
<i>Jeotgalibacillus proteolyticus</i> 22-7	0.91296
<i>Jeotgalibacillus soli</i> P9	0.91272
<i>Neobacillus mesonae</i> FJAT-13985	0.91262
<i>Virgibacillus indicus</i> P2-C2	0.9125
<i>Domibacillus iocasae</i> DSM 29979	0.91111
<i>Oceanobacillus damuensis</i> PT-20	0.91098
<i>Cytobacillus solani</i> FJAT-18043	0.91086
<i>Bacillus</i> sp. SA1-12	0.91066
<i>Peribacillus psychrosaccharolyticus</i> ATCC 23296	0.90911
<i>Mesobacillus boroniphilus</i> JCM 21738	0.90737
<i>Metabacillus indicus</i> DSM 16189	0.90715
<i>Bacillus</i> sp. 17376	0.90669
<i>Lentibacillus halodurans</i> CGMCC 1.3702	0.90615
<i>Halobacillus karajensis</i> HD-02	0.90612
<i>Alteribacillus bidgolensis</i> P4B	0.90589
<i>Sediminibacillus albus</i> CGMCC 1.6502	0.90577
<i>Halobacillus karajensis</i> HD-03	0.90542

<i>Domibacillus tundrae</i> PAMC 80007	0.90521
<i>Terribacillus saccharophilus</i> DSM 21619	0.90479
<i>Alteribacillus bidgolensis</i> DSM 25260	0.90479
