Supplementary Figures

Proteomic analysis of the regulatory networks of ClpX in a model cyanobacterium *Synechocystis* sp. PCC 6803

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Fig. S1. The motif amino acid sequences and their corresponding E-values.



Fig. S2. The construction of clpX insertional mutant strain. (**A**) The schematic shows the construction of clpX insertional mutant strain by homologous recombination. Verification of the clpX insertional mutant strain by (**B**) PCR and (**C**) sequencing.



Fig. S3. Cell morphology of *Synechocystis* cells. Bright-field and fluorescent images in the RFP channel were used to determine cell morphology. Images were recorded using a fluorescent microscope (Olympus, BX53, Japan) at a magnification of 100×.



Fig. S4. Transmission electron microscopy of the *Synechocystis* cells. Stained ultrathin sections of WT (A-B) and $\Delta clpX$ (C-D). The black arrow points at the cell membrane. Images were recorded using the TEM system (Hitachi, Japan) at a magnification of $10.000 \times and 25.000 \times$.



Fig. S5. The whole cell absorption spectra of WT and $\Delta clpX$. Each spectrum is the average of three measurements.



Fig. S6. The estimation of oxidative stress levels in WT and $\Delta clpX$ strains. (A) Relative quantitative analysis of DCF fluorescence intensity in WT and $\Delta clpX$ strains. [Relative of intracellular ROS = [(fluorescence intensity of treatment group) / (fluorescence intensity of control group) × 100%]. (B) The measurement of the lipid peroxidation levels in WT and $\Delta clpX$ strains. ROS, reactive oxygen species; MDA, malondialdehyde.



Fig. S7. The protein-protein interaction (PPI) network analysis of differentially expressed proteins (DEPs) using STRING database. The 172 DEPs were input into the STRING database for PPI network analysis, and achieved a PPI network of 3.12 average nodes, with PPI enrichment p value = 3.85×10^{-08} . The six primary clusters of subnetworks were analyzed by MCL clusters in STRING.



Fig. S8. Comparison of the protein profiles of wild-type and $\Delta clpX$ strains using a quantitative proteomic strategy. Volcano plots showing *p* values ($-\log_{10}$) versus the fold change (\log_2) of DEPs. Proteins with *p* < 0.05 and fold change > 1.2 or < 0.83 are considered to be differentially expressed. Proteins with differential expression were annotated according to their functions and highlighted in different colors.



Fig. S9. The Voronoi treemap visualizes functionally organized quantitative information as area of hierarchically organized information on abundance of (**A**) annotated proteins and (**B**) hypothetical or unknown proteins. Proteins were clustered according to their functional categories as areas of different colors.