

## ***Supplementary Material***

### **Polyphenolic and glucosinolate profile distribution among the aerial parts of *Nasturtium officinale* (Watercress)**

Sotiris Kyriakou<sup>1</sup>, Kyriaki Mihailidou<sup>1</sup>, Tom Amery<sup>2</sup>, Paul Kyle<sup>3</sup>, Paul G. Winyard<sup>4</sup>, Dimitrios T. Trafalis<sup>5</sup>, Franco Rodrigo<sup>5,6</sup>, Aglaia Pappa<sup>7</sup>, Mihalis I Panayiotidis<sup>1\*</sup>

<sup>1</sup>Department of Cancer Genetics, Therapeutics & Ultrastructural Pathology, The Cyprus Institute of Neurology & Genetics, Nicosia 2371, Cyprus

<sup>2</sup>The Watercress Company, Dorchester DT2 8QY, UK

<sup>3</sup>Watercress Research Limited, Devon TQ12 4AA, UK

<sup>4</sup>Laboratory of Pharmacology, Medical School, National & Kapodistrian University of Athens, 11527 Athens, Greece

<sup>5</sup>Redox Biology Centre, University of Nebraska-Lincoln, Lincoln, NE 68583, USA

<sup>6</sup>Department of Veterinary Medicine & Biomedical Sciences, University of Nebraska-Lincoln, Lincoln, NE 68583, USA

<sup>7</sup>Department of Molecular Biology & Genetics, Democritus University of Thrace, 68100 Alexandroupolis, Greece

\* Correspondence:  
Mihalis I. Panagiotidis  
[mihalisp@cing.ac.cy](mailto:mihalisp@cing.ac.cy)

**Table S1:** Multiple Reaction Monitoring conditions for glucosinolates in UPLC-MS/MS analysis

Glucosinolate	Chemical formula	Molecular weight	[M-H] <sup>-</sup> (m/z)	MS <sup>2</sup> fragments (m/z)	Cone voltage (V)	Collision energy (eV)	Retention time (R <sub>t</sub> )
Glucoiberin	C <sub>11</sub> H <sub>21</sub> NO <sub>10</sub> S <sub>3</sub>	423.48	422.1	97.2 259.15	33	26 25	1.22
Glucoraphanine	C <sub>12</sub> H <sub>23</sub> NO <sub>10</sub> S <sub>3</sub>	437.51	436.1	96.95 372.0	26	23 20	1.20
Glucocamelinin	C <sub>18</sub> H <sub>35</sub> NO <sub>10</sub> S <sub>3</sub>	521.66	520.0	97.0 456.4	26	26 19	5.88
Glucoarabin	C <sub>17</sub> H <sub>33</sub> NO <sub>10</sub> S <sub>3</sub>	547.64	546.01	331.05 96.95	35	26 32	7.42
Homoglucocamelinin	C <sub>19</sub> H <sub>37</sub> NO <sub>10</sub> S <sub>3</sub>	535.69	534.15	470.0 519.4	19	25 25	6.30
Glucoraphenin	C <sub>12</sub> H <sub>21</sub> NO <sub>10</sub> S <sub>3</sub>	435.59	434.05	96.95 259.0	32	16 18	1.21
Glucocheirolin	C <sub>11</sub> H <sub>21</sub> NO <sub>11</sub> S <sub>3</sub>	439.48	438.05	96.95 75.1	19	29 26	1.23
Glucolepidiin	C <sub>9</sub> H <sub>17</sub> NO <sub>9</sub> S <sub>2</sub>	347.36	346.05	75.1 96.95	26	26 14	1.27
Glucoerucin	C <sub>12</sub> H <sub>23</sub> NO <sub>9</sub> S <sub>3</sub>	421.51	420.01	96.9 259.0	32	19 19	3.75
Glucobreroxin	C <sub>13</sub> H <sub>24</sub> NO <sub>9</sub> S <sub>3</sub>	435.52	434.1	96.9 259.0	17	17 22	4.90
Sinigrin	C <sub>10</sub> H <sub>17</sub> NO <sub>9</sub> S <sub>2</sub>	359.37	357.8	96.95 194.93	35	17 21	1.44
Gluconapin	C <sub>11</sub> H <sub>19</sub> NO <sub>9</sub> S <sub>2</sub>	373.40	372.0	96.9	35	34	2.11
Glucobrassicinapin	C <sub>12</sub> H <sub>21</sub> NO <sub>9</sub> S <sub>2</sub>	387.43	386.0	96.95	23	12	3.65
Progoitrin	C <sub>11</sub> H <sub>19</sub> NO <sub>10</sub> S <sub>2</sub>	389.4	388.05	96.95 195.05	9	15 20	1.23
Epiprogoitrin	C <sub>11</sub> H <sub>19</sub> NO <sub>10</sub> S <sub>2</sub>	389.4	388.05	97 195.1	22	19 19	1.45

Glucotropaeolin	C <sub>14</sub> H <sub>19</sub> NO <sub>9</sub> S <sub>2</sub>	409.43	408.0	96.95 166.01	20	20 19	3.70
Gluconasturtiin	C <sub>15</sub> H <sub>21</sub> NO <sub>9</sub> S <sub>2</sub>	423.46	422.05	96.9	18	16	4.87
Sinalbin	C <sub>14</sub> H <sub>19</sub> NO <sub>10</sub> S <sub>2</sub>	425.43	424.0	75.15 96.95	20	26 21	1.71
Glucolimnanthin	C <sub>15</sub> H <sub>21</sub> NO <sub>10</sub> S <sub>2</sub>	439.46	438.05	96.95 260.0	22	22 18	4.39
Glucobrassicin	C <sub>16</sub> H <sub>20</sub> N <sub>2</sub> O <sub>9</sub> S <sub>2</sub>	448.47	446.95	97.0 259.0	22	18 22	4.21
Neoglucobrassicin	C <sub>17</sub> H <sub>22</sub> N <sub>2</sub> O <sub>10</sub> S <sub>2</sub>	478.49	477.0	97.05 446.0	22	22 11	5.64
4-methoxyglucobrassicin	C <sub>17</sub> H <sub>22</sub> N <sub>2</sub> O <sub>10</sub> S <sub>2</sub>	478.49	477.05	195.0 275.0	22	20 20	5.17
Glucomoringin	C <sub>20</sub> H <sub>29</sub> NO <sub>14</sub> S <sub>2</sub>	571.57	570.0	570.0	28	10	1.28

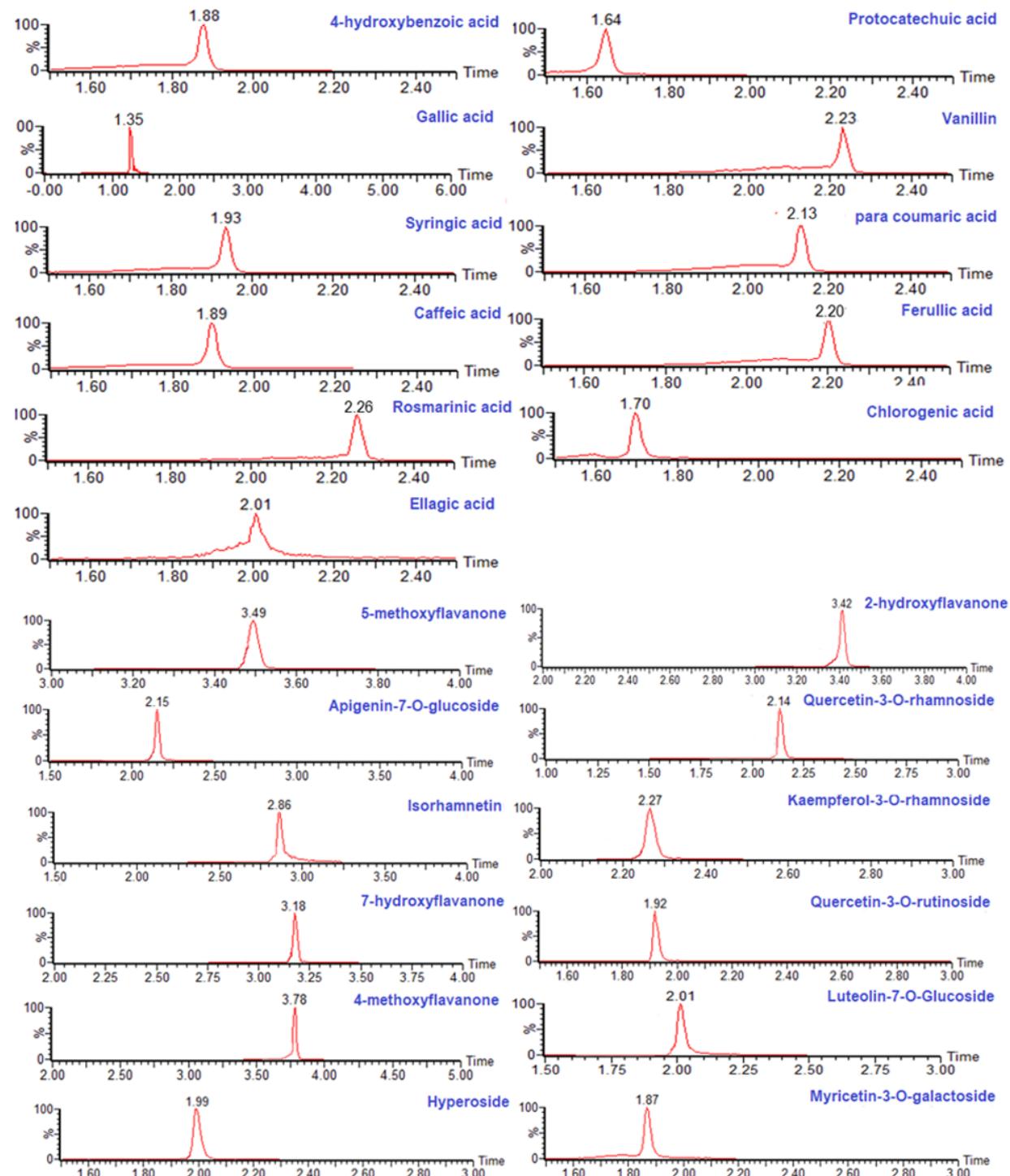
**Table S2:** Multiple Reaction Monitoring conditions for polyphenolic compounds in UPLC-MS/MS analysis

Polyphenolic compound	Chemical formula	Molecular weight	[M-H] <sup>±</sup> (m/z)	MS <sup>2</sup> fragments (m/z)	Cone voltage (V)	Collision energy (eV)	Retention time (R <sub>t</sub> )
4-hydroxybenzoic acid	C <sub>7</sub> H <sub>6</sub> O <sub>3</sub>	138.12	136.95	65.0 93.0	23	25 13	1.88
Protocatechuic acid	C <sub>7</sub> H <sub>6</sub> O <sub>4</sub>	154.12	152.95	108.95	25	13	1.64
Gallic acid	C <sub>7</sub> H <sub>6</sub> O <sub>5</sub>	170.12	168.95	78.98 124.95	23	22 15	1.37
Vanillin	C <sub>8</sub> H <sub>8</sub> O <sub>3</sub>	152.15	151.0	92.2 136.0	22	20 15	2.23
Syringic acid	C <sub>9</sub> H <sub>10</sub> O <sub>5</sub>	198.17	197.0	122.95 182.0	27	23 13	1.93
p-coumaric acid	C <sub>9</sub> H <sub>8</sub> O <sub>3</sub>	164.16	163.0	119.0	15	13	2.13
Caffeic acid	C <sub>9</sub> H <sub>8</sub> O <sub>4</sub>	180.16	178.95	134.95	25	13	1.89
Ferulic acid	C <sub>10</sub> H <sub>10</sub> O <sub>4</sub>	194.18	192.95	134.0 178.0	26	25 12	2.20
Rosmarinic acid	C <sub>18</sub> H <sub>16</sub> O	360.32	359.2	161.0 197.0	10	15 15	2.26
Chlorogenic acid	C <sub>16</sub> H <sub>18</sub> O <sub>9</sub>	354.31	353.1	84.0 191.02	22	44 14	1.70
Ellagic acid	C <sub>14</sub> H <sub>6</sub> O <sub>8</sub>	302.19	301	145.0 173.0	35	34 36	2.00
2'-hydroxyflavanone	C <sub>15</sub> H <sub>12</sub> O <sub>3</sub>	240.27	239	119.3 93.1	40	25 16	3.42
7-hydroxyflavanone	C <sub>15</sub> H <sub>12</sub> O <sub>3</sub>	240.27	239.05	135.2 91.15	41	25 23	3.42
4'-methoxyflavanone	C <sub>16</sub> H <sub>14</sub> O <sub>3</sub>	254.29	255.15	240 161.3	31	17 22	3.78
5-methoxyflavanone	C <sub>16</sub> H <sub>14</sub> O <sub>3</sub>	254.29	255.15	151.3	34	22	3.49
Apigenin-7-O-glucoside	C <sub>21</sub> H <sub>20</sub> O <sub>10</sub>	432.38	431.15	268.35	35	22	2.15
Luteolin-7-O-glucoside	C <sub>21</sub> H <sub>20</sub> O <sub>11</sub>	448.38	449.15	287.1	34	31	2.01

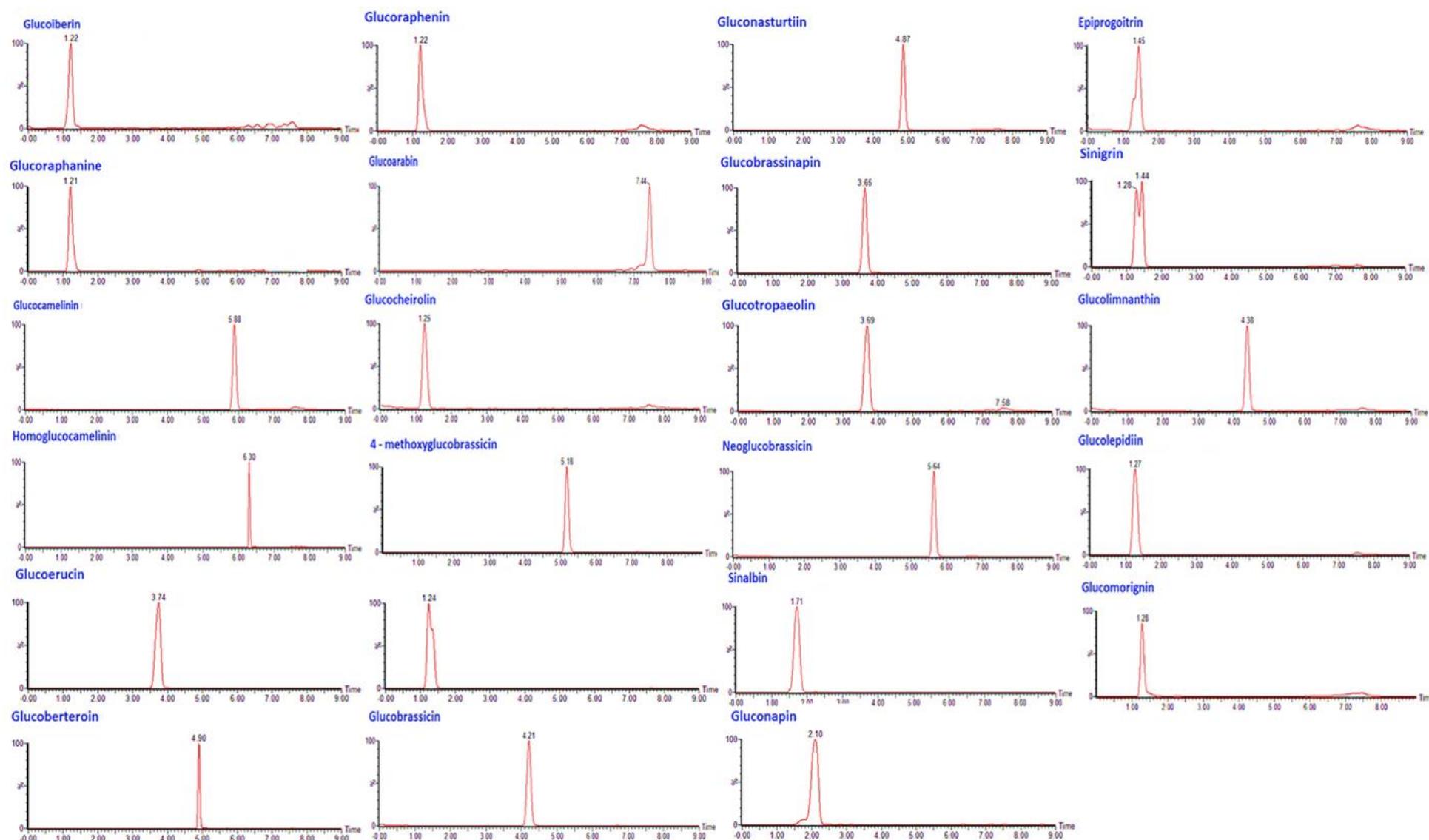
Isorhamnetin	C <sub>16</sub> H <sub>12</sub> O <sub>7</sub>	316.28	315	151.0 300.2	43	30 20	2.86
Quercetin-3-O-rhamnoside	C <sub>21</sub> H <sub>20</sub> O <sub>11</sub>	448.38	447.01	271.0 300.0	43	47 28	2.14
Quercetin-3-O-rutinoside	C <sub>27</sub> H <sub>30</sub> O <sub>16</sub>	610.53	609.1	300.0 271	47	39 65	1.92
Hyperoside	C <sub>21</sub> H <sub>20</sub> O <sub>12</sub>	464.38	463.3	300.0 271.15	47	24 44	1.99
Myricetin-3-galactoside	C <sub>21</sub> H <sub>20</sub> O <sub>13</sub>	480.38	479.05	271.1 287.1	48	39 44	1.87
Kaempferol-3-O-rutinoside	C <sub>21</sub> H <sub>20</sub> O <sub>10</sub>	432.39	431.05	255.3 284.2	45	42 28	2.27
Ipriflavone	C <sub>18</sub> H <sub>16</sub> O <sub>3</sub>	280.33	281.3	240	40	19	4.17
Naringin	C <sub>22</sub> H <sub>32</sub> O <sub>14</sub>	580.54	579.15	271.1 151.5	45	33 40	2.21

**Table S3:** Multiple Reaction Monitoring conditions for derivatized (into thioureas) isothiocyanates in UPLC-MS/MS analysis

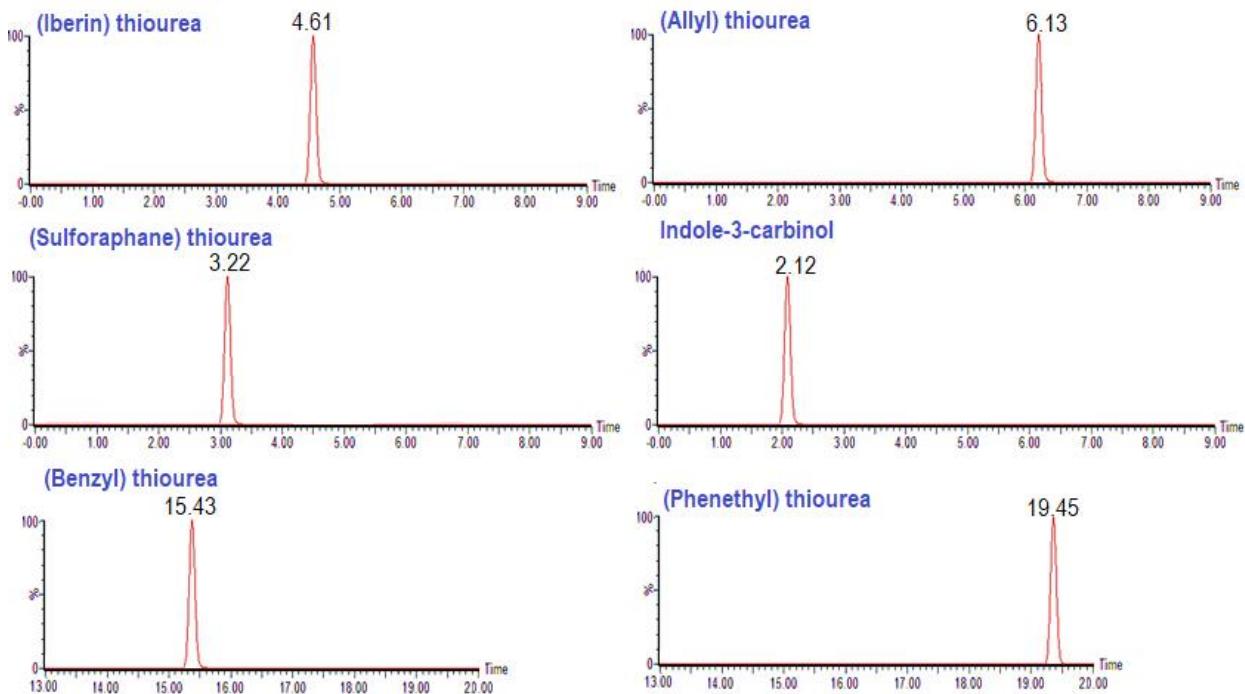
(Isothiocyanate) derivatized thiourea	Chemical formula	Molecular weight	[M-H] <sup>+</sup> (m/z)	MS <sup>2</sup> fragments (m/z)	Cone voltage (V)	Collision energy (eV)	Retention time (R <sub>t</sub> )
(Iberin) thiourea	C <sub>5</sub> H <sub>12</sub> N <sub>2</sub> OS <sub>2</sub>	180.29	181.3	163.4	22	15	4.61
(Allyl) thiourea	C <sub>4</sub> H <sub>7</sub> N <sub>2</sub> S	115.18	116.18	99.8	19	13	6.13
(Sulforaphane) thiourea	C <sub>6</sub> H <sub>14</sub> N <sub>2</sub> OS <sub>2</sub>	194.31	195.4	117.2	12	11	3.22
(Benzyl) thiourea	C <sub>8</sub> H <sub>10</sub> N <sub>2</sub> S	166.24	167.4	148.9	18	9	15.43
(Phenethyl) thiourea	C <sub>9</sub> H <sub>12</sub> N <sub>2</sub> S	180.27	181.0	104.99	20	17	19.45
Indole-3-carbinol	C <sub>9</sub> H <sub>9</sub> NO	147.17	148.0	117	17	9	2.12



**Figure S1:** Extracted UPLC- ESI-MS/MS chromatograms of polyphenolic compounds (including phenolic acids and flavonoids) in watercress sample extracts.



**Figure S2:** Extracted UPLC- ESI-MS/MS chromatograms of glucosinolates in watercress sample extracts.



**Figure S3:** Extracted UPLC- ESI-MS/MS chromatograms of thioureas (derivatized isothiocyanates) in watercress sample extracts.

**Table S4:** The limit of detection (LOD), quantification (LOQ), linearity, precision and accuracy results for the screened intact glucosinolates. The calibration equations represent the peak area as a function of concentration in ppb. The intra- and inter- day experimental data concern data that have been collected from a six days experiment, whereas the %recovery data are the means of three indepent experiments.

Compound	Linear range (ppb)	LOD (ppb)	LOQ (ppb)	Calibration equation <sup>a</sup>	Correlation coefficient ( $r^2$ )	%RSD		
						(intra-day) <sup>b</sup>	(inter-day) <sup>c</sup>	%REC <sup>d</sup>
<b>GLUCOSINOLATES</b>								
Glucoiberin	0.27-249.80	0.27	0.79	y=0.21x-0.57	0.9998	0.98	2.28	98.3
Glucoraphanine	1.10-250.1	1.10	3.65	y=3.19x-4.6	0.9990	0.57	4.31	89.6
Glucocamelinin	1.69-250.10	1.69	5.64	y=4.54x-5.24	0.9991	0.65	2.65	84.0
Homoglucocamelinin	3.70-248.30	3.70	12.36	y=5.82x+1.04	0.9995	0.69	2.32	96.3
Glucoarabin	3.36-249.80	3.36	11.21	y=7.63x+2.22	0.9989	0.47	4.10	99.9
Glucoraphenine	5.22-246.60	5.22	17.40	y=2.41x-1.05	0.9989	0.58	1.95	100.2
Glucocheirolin	6.36-251.20	6.36	21.20	y=2.33x-5.00	0.9997	1.12	2.1	93.2
Glucolepidiin	4.26-248.60	4.26	14.20	y=1.13x-7.17	0.9954	1.32	2.53	85.0
Glucobreroxin	3.66-2.49	3.66	12.20	y=3.38x-6.27	0.9999	1.54	3.0	96.4
Glucoerucin	1.68-377.20	1.68	5.60	y=3.56x-2.64	0.9991	2.6	1.99	97.8
Sinigrin	1.96-255.40	1.96	6.54	y=1.46x-3.32	0.9981	2.2	1.1	98.4
Gluconapin	2.69-243.8	2.69	8.98	y=1.84x-1.48	0.9987	2.2	1.2	93.1
Glucobrassicinapin	4.37-246.5	4.37	14.58	y=1.16x-1.60	0.9992	2.3	1.03	87.6
Epiprogoitrin	6.79-237.40	6.79	22.65	y=0.98x-7.07	0.9995	1.89	1.04	99.5
Progoitrin	4.77-248.0	4.77	15.90	y=1.27x-0.58	0.9961	2.02	2.02	99.5
Glucolimnanthin	4.89-247.90	4.89	16.32	y=5.24x+0.93	0.9989	2.21	0.5	94.6
Sinalbin	5.36-262.30	5.36	17.89	y=2.25x-6.36	0.9932	1.87	1.37	89.9

Supplementary Material

Gluconasturtiin	2.99-258.20	2.99	9.98	y=2.08x-1.60	0.9977	2.25	4.2	95.5	<sup>a</sup>
Glucotropaeolin	2.69-250.50	2.69	8.97	y=7.07x+11.65	0.9983	1.14	0.78	92.2	
Glucobrassicin	2.99-250.50	2.99	9.99	y=2.15x-2.44	0.9994	1.6	0.96	98.7	
Glucomoringin	3.53-254.40	3.53	11.78	y=4.64x+3.73	0.9992	3.2	1.11	100.5	
4-methoxyglucobrassicin	4.29-251.90	4.29	14.30	y=2.55x-1.21	0.9984	2.89	1.63	101.2	
Neoglucobrassicin	3.75-233.90	3.75	12.50	y=8.014x-1.95	0.9993	1.21	2.42	93.7	

Chromatographic peak area (y) as a function of ppb concentration (x)

<sup>b</sup> Values are means of intra-day assays (n=6)

<sup>c</sup> Values are means of inter-day assays (n=6)

<sup>d</sup> (n=3)

**Table S5:** The limit of detection (LOD), quantification (LOQ), linearity, precision and accuracy results for the screened polyphenolic compounds. The calibration equations represent the peak area as a function of concentration in ppb. The intra- and inter- day experimental data concern data that have been collected from a six days experiment, whereas the %recovery data are the means of three indepentend experiments.

Compound	Linear range (ppb)	LOD (ppb)	LOQ (ppb)	Calibration equation <sup>a</sup>	Correlation coefficient ( $r^2$ )	%RSD			%REC <sup>d</sup>
						(intra-day) <sup>b</sup>	(inter-day) <sup>c</sup>		
<b>POLYPHENOLIC COMPOUNDS</b>									
4-hydroxybenzoic acid	3.01-499.50	3.01	14.20	y=36.87x-62.07	0.9991	1.15	2.21	98.8	
Protocatechuic acid	0.66-504.50	0.66	14.70	y=34.24x-69.4	0.9995	1.25	2.65	86.3	
Gallic acid	53.20-513.20	53.20	105.20	y=0.67x-1.5	0.9996	0.46	0.21	99.9	
Vanillin	2.87-335.00	2.87	5.62	y=0.67x-0.1	0.9999	0.98	0.95	100.4	
Syringic acid	2.01-501.60	2.01	2.86	y=7.28x-2.7	0.9996	1.36	1.01	96.6	
p-coumaric acid	0.65-497.30	0.65	1.55	y=52.84x+36.9	0.9997	1.7	1.94	93.2	
Caffeic acid	1.21-500	1.21	1.25	y=92.95x-344.4	0.9995	1.01	2.21	100.1	
Ferulic acid	2.10-505.60	2.10	12. 17	y=19.02x-68.4	0.9992	0.7	2.45	102.6	
Rosmarinic acid	2.32-499.50	2.32	2.56	y=7.03x+12.34	0.9996	1.3	3.02	86.9	
Chlorogenic acid	3.48-495.60	3.48	4.76	y=25.02x+60.3	0.9991	1.35	1.98	87.4	
Ellagic acid	5.53-499.10	5.53	75.60	y=2.18x+7.4	0.9995	1.32	3.05	89.9	
2'-hydroxyflavanone	19.50-250.00	19.50	20.12	y=38.69x+22.5	0.9998	2.7	4.32	99.5	

7-hydroxyflavanone	1.97- 249.90	1.97	2.21	y=51.17x-73.6	1	2.63	1.42	98.9
4'-methoxyflavanone	2.21- 250.00	2.21	3.89	y=83.54x+60.3	0.9999	2.89	1.87	93.6
5-methoxyflavanone	6.47- 248.50	6.47	8.52	y=195.14x-493.9	0.9992	3.21	2.69	94.7
Apigenin-7-O-glucoside	1.87- 125.30	1.87	4.42	y=6.17x+3.8	0.9998	3.48	2.54	95.8
Luteolin-7-O-glucoside	2.21- 250.10	2.21	2.22	y=51.52x-89.9	0.9998	3.64	3.22	89.2
Isorhamnetin	14.01- 251.1	14.01	2.31	y=6.08x-15.4	0.9992	2.48	1.18	100.1
Quercetin-3-O-rhamnoside	1.02- 250.60	1.02	4.21	y=60.83x-38.6	0.9999	2.21	3.01	99.8
Quercetin-3-O-rutinoside	1.40- 251.30	1.40	4.32	y=97.74x+109.7	0.9999	1.35	1.89	87.4
Hyperoside	6.32- 249.90	6.32	3.21	y=3.97x+0.5	0.9998	2.14	1.37	96.3
Myricetin-3-galactoside	0.85- 251.20	0.85	2.12	y=26.38x-31.8	0.9997	1.78	1.65	100.2
Kaempferol-3-O-rutinoside	0.76- 250.00	0.76	1.21	y=25.73x+73.7	0.9997	1.36	2.21	91.2
Ipriflavone	109.90- 250.00	109.90	13.21	y=0.62x+2.2	0.9994	1.69	1.11	93.6
Naringin	3.01- 250.60	3.01	1.21	y=22.88x-43.3	0.9997	2.22	4.02	95.4

<sup>a</sup> Chromatographic peak area (y) as a function of ppb concentration (x)<sup>b</sup> Values are means of intra-day assays (n=6)<sup>c</sup> Values are means of inter-day assays (n=6)<sup>d</sup> (n=3)

**Table S6:** The limit of detection (LOD), quantification (LOQ), linearity, precision and accuracy results for the screened derivatized isothiocyanates compounds. The calibration equations represent the peak area as a function of concentration in ppb. The intra- and inter- day experimental data concern data that have been collected from a six days experiment, whereas the %recovery data are the means of three indepenteds experiments.

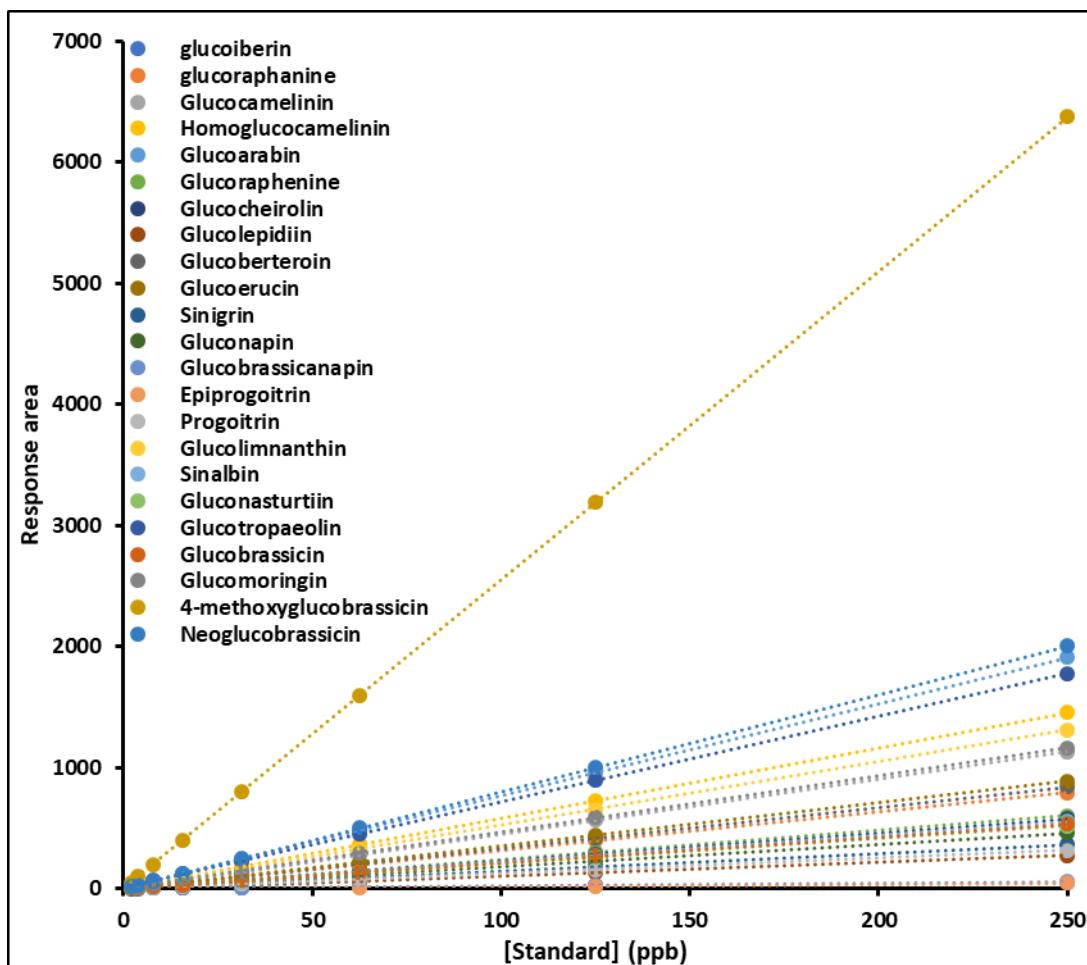
Compound	Linear range (ppb)	LOD (ppb)	LOQ (ppb)	Calibration equation <sup>a</sup>	Correlation coefficient ( $r^2$ )	%RSD		%REC <sup>d</sup>
						(intra-day) <sup>b</sup>	(inter-day) <sup>c</sup>	
<b>(ISOTHIOCYANATE) DERIVATIZED THIOUREA COMPOUNDS</b>								
(Iberin) thiourea	21.69-250.50	21.69	72.32	y=3.12x+15	0.9999	1.18	4.02	84.2
(Allyl) thiourea	24.43-151.42	24.43	81.45	y=0.67x+1.1	0.9997	1.32	2.21	80.2
(Sulforaphane) thiourea	6.96-236.12	6.96	23.14	y=29.41x-36.9	0.9996	2.15	2.21	95.6
(Benzyl) thiourea	4.68-247.92	4.68	15.36	y=25.48x-65.1	0.9982	2.13	2.43	89.7
(Phenethyl) thiourea	3.06-250	3.06	10.21	y=48.12x-1.2	0.9997	2.22	2.6	96.7
Indole-3-carbinol	2.73-251.10	2.73	9.12	y=22.88x-43.3	0.9959	1.04	2.0	93.6

<sup>a</sup> Chromatographic peak area (y) as a function of ppb concentration (x)

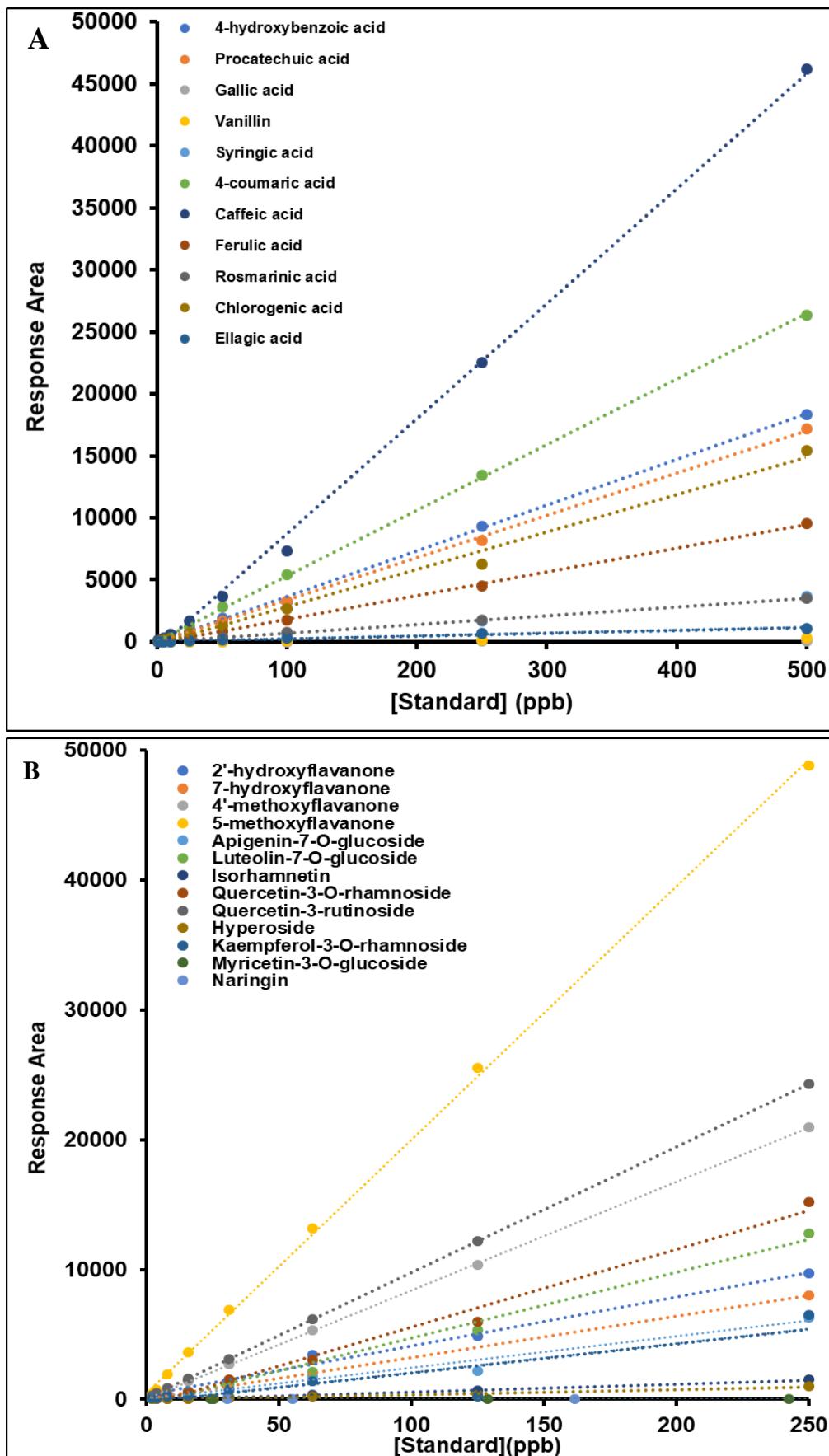
<sup>b</sup> Values are means of intra-day assays (n=6)

<sup>c</sup> Values are means of inter-day assays (n=6)

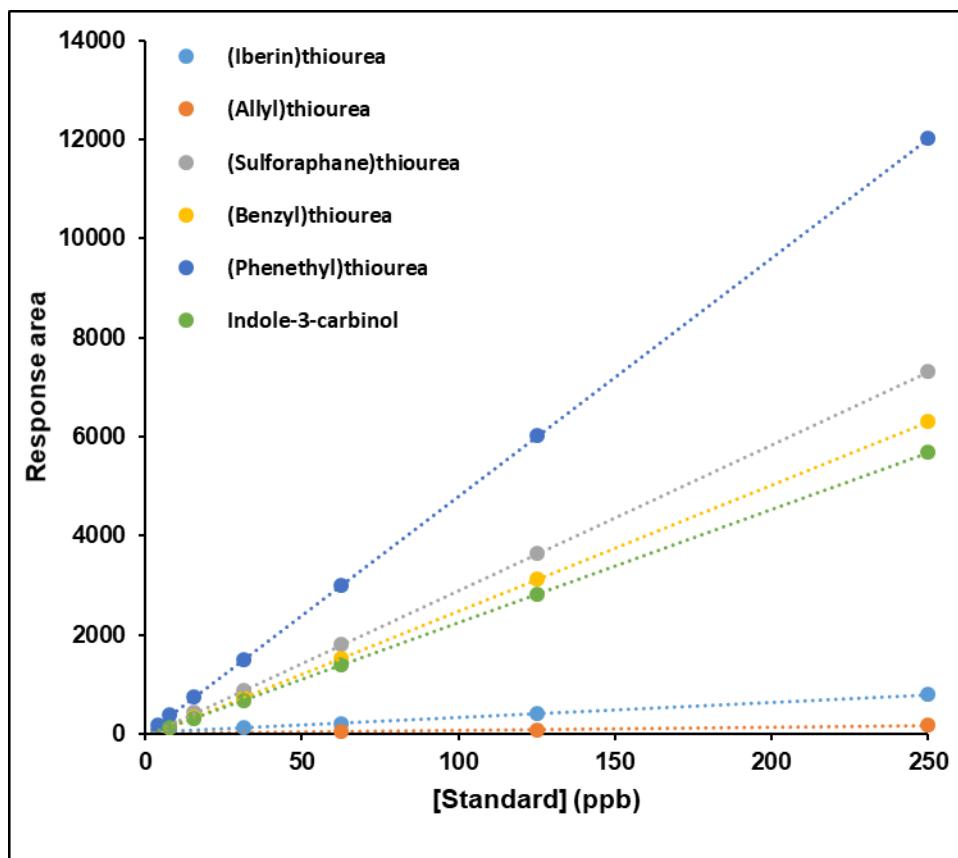
<sup>d</sup> (n=3)



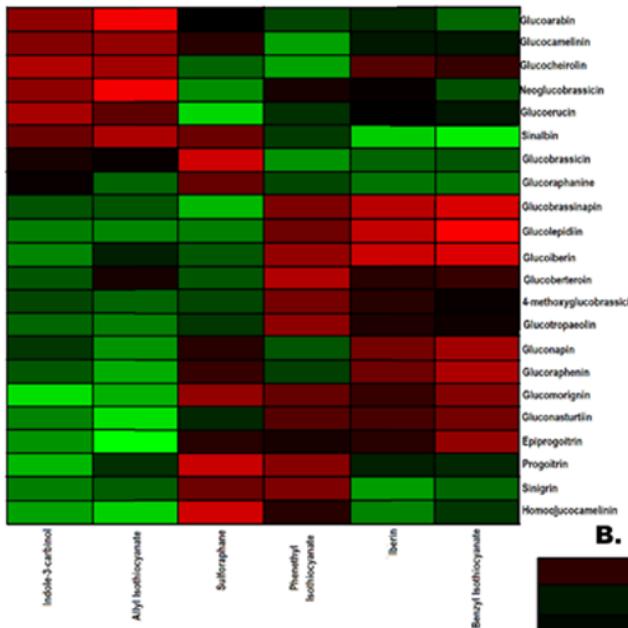
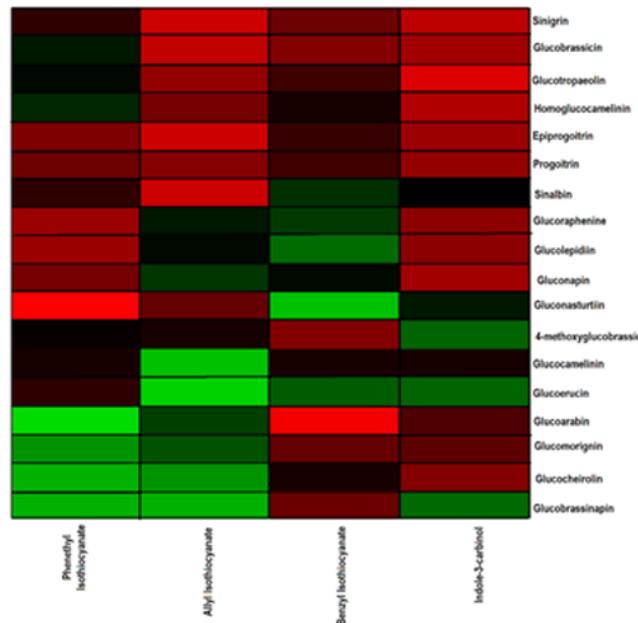
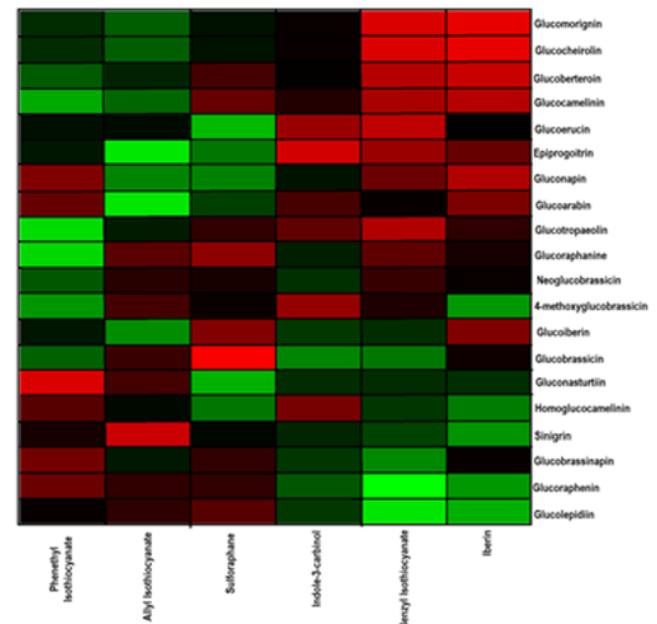
**Figure S4:** Calibration curve of glucosinolate standards at various concentrations (1.95–250 ppb) used for the determination of intact glucosinolates in the aerial parts (flowers, leaves and stems) of watercress.



**Figure S5:** Calibration curve of (A) phenolic acid and (B) flavonoid standards at various concentrations (1.95-500 ppb) used for the determination of polyphenolic compounds in the aerial parts (flowers, leaves and stems) of watercress.



**Figure S6:** Calibration curve of derivatised isothiocyanates into the thiourea standards at various concentrations (1.95-250 ppb) used for the determination of isothiocyanates in the aerial parts (flowers, leaves).

**A. WATERCRESS FLOWERS**

**B. WATERCRESS LEAVES**

**C. WATERCRESS STEMS**


**Figure S7:** Spearman's coefficient analysis between intact GL and ITC contents on watercress (A) flowers, (B) stems and (C) leaves. Red indicates positive (+1.0) correlation while green indicate a negative correlation. Black suggests no correlation.