Check for updates

OPEN ACCESS

EDITED BY Kunyu Zhang, South China University of Technology, China

REVIEWED BY Zhiwei Fang, Johns Hopkins University, United States Hani Nasser Abdelhamid, Assiut University, Egypt

*CORRESPONDENCE Dongxu Wang, wang_dong_xu@jlu.edu.cn

[†]These authors have contributed equally to this work

SPECIALTY SECTION

This article was submitted to Biomaterials, a section of the journal Frontiers in Bioengineering and Biotechnology

RECEIVED 15 June 2022 ACCEPTED 04 July 2022 PUBLISHED 22 July 2022

CITATION

Yang Z, Liu W, Liu H, Li R, Chang L, Kan S, Hao M and Wang D (2022), The applications of polysaccharides in dentistry. *Front. Bioeng. Biotechnol.* 10:970041. doi: 10.3389/fbioe.2022.970041

COPYRIGHT

© 2022 Yang, Liu, Liu, Li, Chang, Kan, Hao and Wang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or

reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

The applications of polysaccharides in dentistry

Zhijing Yang^{1,3†}, Weiwei Liu^{1,3†}, Huimin Liu^{1,3}, Rong Li², Lu Chang^{1,3}, Shaoning Kan^{1,3}, Ming Hao^{1,3} and Dongxu Wang²*

¹Department of Oral and Maxillofacial Surgery, Hospital of Stomatology, Jilin University, Changchun, China, ²Laboratory Animal Center, College of Animal Science, Jilin University, Changchun, China, ³Jilin Provincial Key Laboratory of Tooth Development and Bone Remodeling, Hospital of Stomatology, Jilin University, Changchun, China

Polysaccharides are natural polymers widely present in animals, plants, and several microorganisms. Polysaccharides have remarkable properties, including easy extractions, degradability, and renewability, and have no apparent toxicity, making them ideal for biomedical applications. Moreover, polysaccharides are suitable for repairing oral tissue defects and treating oral diseases due to their excellent biocompatibility, biosafety, anti-inflammatory, and antibacterial properties. The oral cavity is a relatively complex environment vulnerable to numerous conditions, including soft tissue diseases, hard tissue disorders, and as well as soft and hard tissue diseases, all of which are complex to treat. In this article, we reviewed different structures of natural polysaccharides with high commercial values and their applications in treating various oral disease, such as drug delivery, tissue regeneration, material modification, and tissue repair.

KEYWORDS

natural polysaccharide, oral disease, tissue repair, tissue regeneration, biomaterials

Introduction

Oral diseases are a global public health concern, often affecting the aesthetics and diet of patients, and in some cases, they cause systematic diseases (P aradowska-Stolarz et al., 2021; Wong et al., 2019). Treating oral diseases require many biological materials. Dental caries treatment requires filling the dental pulp with resin (Malkondu et al., 2014), pulp capping agent (Kunert and Lukomska-Szymanska, 2020), and gutta percha (Del Fabbro et al., 2016). Also, denture restoration, impression, and dental implant materials are required for tooth loss treatment (Roumanas, 2009). Tissue-guided regeneration and defect repair materials are necessary for oral surgery (Frassica and Grunlan, 2020; Niu et al., 2021). Toothpaste, floss, and mouthwash are needed to prevent oral diseases (Zhang et al., 2021). The common materials used in dentistry include polymethylmethacrylate (PMMA), vulcanized rubber, celluloid, phenol-formaldehyde, and polyvinyl chloride (PVC) (P aradowska-Stolarz et al., 2021). Ideal dentistry materials should possess high biocompatibility as well as good biofunction and mechanical properties (Abdelhamid and Mathew, 2022). In stomatology, natural polysaccharides are used to modify materials repairing chewing organs, and improve oral health (Vergnes and Mazevet, 2020).

Polysaccharides are natural branched or non-branched polymers, usually composed of more than 10 monosaccharides joined together *via* glycosidic bonds (Yu et al., 2018).



Polysaccharides are widely present in plants, microorganisms and animals. They possess anti-tumor, anti-oxidation, and antiinflammatory properties, favouring their application in biomedicine (Silva et al., 2021). In recent years, natural polysaccharides and their derivatives have been widely used in packaging, food and pharmaceutical industries, and dentistry, given their sustainability, renewability, biodegradability, and non-toxicity. Also, the wide application of polysaccharides in the above-mentioned industries is attributed to their structural diversity (Torres et al., 2019). Materials used in the oral cavity are in direct contact with human tissue; thus, they should be biocompatible and non-toxic. Moreover, they should promote efficient healing of the mouth to perform its main functions, including chewing, breathing and talking (Whitters et al., 1999). In addition, given the role of the mouth on the facial outlook, the materials used in the oral cavity need special consideration. Previous reports have shown that natural polysaccharides have numerous advantages in treating oral diseases. In this review, we discussed several natural polysaccharides with high commercial values and their applications in treating different oral diseases.

Classification of polysaccharides

Polysaccharides can be broadly classified into homogeneous heterogeneous polysaccharides. Homogeneous and polysaccharides derived are from joining several monosaccharide molecules of the same kind, including starch, cellulose, and chitosan. Heterogeneous polysaccharides are derivatives of different monosaccharide molecules. The common heterogeneous polysaccharides include hyaluronic acid and chondroitin sulfate. Since they are naturally available, easy to obtain, non-toxic, cheap, biodegradable, and biocompatible, polysaccharides are suitable for the oral cavity environment (Prajapati et al., 2014). The most commonly used polysaccharides are shown in Figure 1.



Chitosan

linear, semi-crystalline Chitosan is а natural polysaccharide derived from chitin, readily available in crustaceans and fungal cell walls. The U.S. Food and Drug Administration (FDA) has approved chitosan for food and drug use (Wang et al., 2020). Chitosan contains random glucosamine (deacetylation unit) and N-acetylglucosamine (acetyl unit) units linked to the primary chain through β (1-4) glycosidic bonds. Chitosan is positively charged due to the presence of the amino group (Abd El-Hack et al., 2020). The molecule has a straight-chain structure, and weighs between 10 and 1,000 kDa (Kou et al., 2021). Chitosan is widely utilized for antibacterial purposes (Muxika et al., 2017), drug release (Hudson and Margaritis, 2014), target therapy (Mushtaq et al., 2021; Narmani and Jafari, 2021), and drug delivery in stomatology (Ahsan et al., 2018). Studies have shown that it effectively inhibits the biofilm formation and the acid production capacity of Streptococcus mutans, the main causal agent of dental caries (Kawakita et al., 2019). Several delivery systems that carry drugs for anti-inflammatory purposes, tissue, and periodontitis treatment have been modeled around chitosan (Sah et al., 2019; Baranov et al., 2021). Chitosan has been used for modifying nanomaterials and hydrogel, and its derivatives for treating oral diseases have been gradually developed (Wang et al., 2020; Shivakumar et al., 2021) (Figure 2).

Cellulose

Cellulose is one of the most abundant biological polymers on Earth (Lynd et al., 2002). Plant cellulose is present in most green plants and algae. Bacterial cellulose is naturally secreted by several non-pathogenic bacteria, including acetobacter, agrobacterium, and rhizobium. Cellulose has a well-arranged three-dimensional fiber network, 3.0-3.5 µm thick (Klemm et al., 2005; Svensson et al., 2005). Compared with plant cellulose, bacterial cellulose, excluding lignin and hemicellulose, contains small fibers (100 times lower than plant cellulose) and has a highly crystalline structure. Heat, steam, ethylene oxide gas, and radiation will not destroy the inherent physicochemical properties and structural integrity of cellulose (de Oliveira Barud et al., 2020). The three-dimensional structure of cellulose is very similar to the extracellular matrix (ECM) of living tissues and, thus, suitable for preserving oxygen and nutrients, providing a favourable environment for the growth and proliferation of cells in stomatology (Halib et al., 2019). Moreover, cellulose can be used for tissue engineering as scaffolds and implants for wound healing, drug delivery, and dental materials (de Oliveira Barud et al., 2016; de Oliveira Barud et al., 2020; Klinthoopthamrong et al., 2020). Studies have shown that restorable BC could be used in GBR to improve bone generation. Moreover, the electron beam irradiation (EI) can sever the BC glucose bonds to increase biodegradation (An et al., 2017).



Hyaluronic acid

Hyaluronic acid (HA) is a glycosaminoglycan composed of repeated N-acetyl D-glucosamine and D-glucuronic acid units (Marinho et al., 2021). It is the main skin extracellular matrix (ECM) component and participates in inflammatory responses, angiogenesis, and tissue regeneration. Given its antiinflammatory property, HA is often used to treat oral ulcers (Graca et al., 2020). HA is also a good drug delivery system, given its mucosal adhesion and ease of chemical modification (Huang and Huang, 2018; Salari et al., 2021). Notably, a local administration strategy is preferred for treating oral diseases than systemic administration. HA does not affect other body organs and tissues when used for drug delivery. Moreover, the local treatment ensures better delivery of drugs to soft periodontal tissue, gingiva, periodontal ligament, and hard tissues, such as alveolar bone and cementum. Catechol (Cat)modified chitosan/hyaluronic acid nanoparticles (NPs) is a newly developed system for delivering doxorubicin (DOX) to oral squamous cell carcinoma. It displays excellent adhesion to the oral mucosa and delivery of DOX to the target tissues in a sustained manner (Pornpitchanarong et al., 2020). HA has good biocompatibility, biodegradability, and hydrophilicity and can alleviate inflammatory pain. These desired biological properties promote the use of HA in the oral cavity (Figure 3).

Marine algae polysaccharides

Seaweed polysaccharides, including alginate, agar, carrageenan, galactan, fucoidan, urfan, and laver

polysaccharides, are widely used in pharmaceutical industry (Rahmati et al., 2019; Beaumont et al., 2021). Marine algae polysaccharides possess numerous physical and chemical properties. These types of polysaccharides are soft and expansible (Bilal and Iqbal, 2019). Sodium alginate and agarose, widely used in treating oral diseases, are the common marine algae polysaccharides. Sodium alginate was included on the list of Pharmacopoeia products in the United States as early as 1938. Sodium alginate is a by-product of kelp or sargassum, β-D-mannuronic acid (β-Dmannuronic, m), and *a*- L-guluronic acid (α-L-guluronic, g), joined together via the $1 \rightarrow 4$ linkages. The compound is stable, soluble, viscous, and safe, suitable for pharmaceutical use (Sanchez-Ballester et al., 2021). Furthermore, it contains numerous -COO- units that give it its polyanionic behavior in an aqueous solution, facilitating drug attachment and adhesion to the mucosal membrane (Amin and Boateng, 2022). Some studies have suggested that sodium alginate could treat recurrent aphthous ulcers. The compound can alleviate the pain and improve the persistence of a drug in the oral mucosa (Laffleur and Kuppers, 2019; Suharyani et al., 2021). In addition, alginate is mainly used for stomatology as an impression of oral materials (Cervino et al., 2018). Obtaining an oral model is an important step before orthodontics and prosthodontics treatment. Alginate impression materials have remarkable fluidity, resilience, plasticity, and high drug delivery accuracy, promoting their clinical use.

Agarose is a linear polymer polysaccharide extracted from kelp (Zucca et al., 2016), composed of alternating $1 \rightarrow 3$ -linked β -Dgalactopyranose and $1 \rightarrow 4$ -linked 3,6 anhydro- α -



L-galactopyranose units, designated as G and A residues, respectively (Trivedi et al., 2015). Agarose is a potential hydrogel for the controlled release of bioactive substances, given its good biocompatibility and natural biodegradability (Kim et al., 2019). In the oral cavity, agarose gel regulates the size and shape of hydroxyapatite crystals (Ling et al., 2019). A new biomimetic mineralization system containing agarose gel can induce a dense hydroxyapatite layer on the surface of demineralized dentin to block dentin tubules for dentin remineralization, a potentially new method for treating dentin hypersensitivity and dental caries (Ning et al., 2012). These two algal polysaccharides are highly biocompatible and have broad dental applications, including treating oral diseases.

Applications of natural polymers in stomatology

The oral cavity is located in the lower 1/3 of the maxillofacial region, starting from the lips. It has several sections, including the buccal, the inner lining of the cheeks, the pharynx at the back, the palate on the upper side, and the

floor of the mouth on the lower side. Together, these parts form the cavitas oris propria (Tuominen and Rautava, 2021). Dental caries is an idiopathic disease of the oral cavity (Akintoye and Mupparapu, 2020). The teeth structure is different from that of bones and includes the enamel, dentin, pulp, and cementum. Enamel is the hardest tissue in the body that has neither blood vessels nor nerves and cannot regenerate. The dentin is softer than the enamel and can regenerate or repair itself to a certain degree. However, excessive teeth damage can create irreversible defects. If dental caries is not treated on time, the enamel is continually eaten up, causing bacteria infection in the pulp and periapical tissue. This eventually causes pulpitis, periapical periodontitis, jaw osteomyelitis, and other concurrent diseases, inducing severe pain and tissue damage. Moreover, dental caries not only destroy the integrity of the chewing organs but also affects the digestive function, seriously affecting an individual's overall health. Natural polysaccharides, which have antibacterial, drug delivery, and material modification capability with remarkable biocompatibility, are suitable for different dental functions to prevent oral diseases of soft and hard tissues or a combination of both (Figure 4).

Treatment of oral hard tissue diseases

Oral hard tissue diseases include dental caries, maxillofacial fractures, and bone defects (Melo et al., 2013). In dentistry, chitosan and alginate are used to improve the property of adhesive material. A combination of chitosan and triclosan supplemented with resin inhibits biofilm formation and improves the stability between dentin and adhesive interface, promoting long-term edge sealing (Machado et al., 2019). A mixture of chitosan and glass ionomer cement (GIC) can be another repair material that allows the slow-release of fluoride (Ibrahim et al., 2020). Chitosan reacts with the GIC to generate a compound with a better ion release rate, which is important for tooth structure (Mulder and Anderson-Small, 2019). Moreover, a formulation of type I collagen (Col), nanocrystalline hydroxyapatite (HAp), and alginate (Alg) is suitable for 3D printing of scaffolds with properties similar to those of natural dentin. The formulation effectively treats tooth sensitivity by blocking the dentin microtubules (Naseri et al., 2021). On the whole, polysaccharides could be modified to generate desired products suitable for treating hard tissue diseases in dentistry.

Treatment of oral soft tissue diseases

Diseases of the soft mouth tissues include pulpitis, periapical periodontitis, gingivitis, and other oral mucosal diseases. Infected dental pulp is very painful, significantly affecting the normal life of patients. The infection can spread to periapical tissues, causing periapical periodontitis. Gingivitis is an inflammation of teeth gums caused by accumulated dental plaque and some physical stimulation in the papillary. The disease can cause periodontitis and loss of the alveolar bone. Therefore, removing the root canal infection and dental plaque is important in treating oral soft tissue diseases. Given the unique anatomy, histology, and microbial environment of the oral cavity, materials for oral cavity uses should possess anti-bacterial properties, adhere on the oral cavity, be easy to apply, and should allow slow release of important molecules (Karolewicz, 2016; Timur et al., 2019). Natural polysaccharides can be modified to possess good adhesion, repair, and mechanical properties suitable for treating oral soft tissue diseases.

Chitosan inhibits the growth of *Porphyromonas gingivalis* and *Actinomyces aggregatum* and can modulate inflammation in human gingival fibroblasts by regulating the level of PGE2 through the JNK pathway. Chitosan is well tolerated by gingival fibroblasts and can stimulate cell proliferation via the ERK1/2 signaling pathway. Furthermore, the synergistic effect of chitosan and growth factors such as PDGF-BB stimulates the proliferation of gingival fibroblasts and inhibits the growth of *Porphyromonas gingivalis* and *Actinomyces aggregatum* (Silva et al., 2013). Bacterial cellulose fastens the solidification of diatomite and enhances the biological activity of the mineral.

Moreover, it has good biocompatibility and promotes the proliferation and adhesion of mesenchymal stem cells (Voicu et al., 2017). In addition, it does not generate oxidative stress and is, therefore, an excellent material in endodontics. Compared with the ordinary absorbent paper tips, bacterial cellulose has a higher moisture absorption rate, expansion rate, tensile strength, and biocompatibility. Moreover, it is hard and causes no obvious allergic reactions (Yoshino et al., 2013). HA and platelet lysates' complex increase the metabolism of dental pulp mesenchymal stem cells and repair damaged dental pulp/dentin tissue by stimulating the deposition of mineralized matrix (Almeida et al., 2018; Schmidt et al., 2020). A combination of bacterial cellulose (BC) and photoactivated carbene-based biological adhesive (PDZ) forms a flexible film platform that can repair soft tissue in the ever-wet mouth environment. The shear strength and adhesion of the composite have been significantly improved, making it suitable for treating oral mucosal wounds (Singh et al., 2021). Therefore, polysaccharides can be used as antibacterial, root canal therapy materials and tissue treating oral patches, underlining the novel application prospects of these materials.

Treatment of a combination of oral soft and hard tissue diseases

Diseases involving soft and hard tissues in the mouth include cancer and periodontitis. Both diseases can cause lesions, gingival, buccal and lingual mucosa recession, and even alveolar bone and jaw bone loss. Gingival recession and the alveolar bone defect caused by periodontitis are irreversible and greatly affect a person's facial appearance. In serious cases, it causes tooth loss and facial deformity, affecting eating and talking. Therefore, periodontitis should be promptly treated. The periodontium repair involves using multiple materials to prevent further damage and restore bone and periodontal losses. Pure chitosan scaffolds promote the proliferation of cementoblasts (CB) and periodontal ligament cells (PDLCs), the alkaline phosphatase activity, and mineralization level. The CS scaffolds, combined with other polymer biomaterials and bioceramics, promote rapid periodontal regeneration (Lauritano et al., 2020). Mesoporous hydroxyapatite/chitosan (MHA/CS) composite scaffolds promote periodontal tissue regeneration, inhibit the growth of Clostridium nucleatum and Porphyromonas gingivalis, promote the differentiation of periodontal ligament stem cells into osteoblasts, and upregulate the ALP activity, and the expression of RUNX-2, OPN, and DLX-5 in vitro. Moreover, MHA/CS composite induces the cementum-like tissue formation in vivo, demonstrating its potential for periodontal tissue regeneration (Liao et al., 2020). Mice models with bone defects comparing bacterial cellulose and collagen biofilms regarding guided bone regeneration during periodontal tissue repair revealed that bacterial cellulose biofilms only promote soft tissue repair in skulls but do not induce bone regeneration (Farnezi Bassi et al., 2020). However, hydrolyzed cellulose biofilm through strontium apatite modulates inflammation at the wound site and promotes the formation of connective tissue and the increase of calcium and magnesium, important elements that promote the bone generation and calcification (Luz et al., 2020). In general, the natural polysaccharides can effectively induce periodontal tissue regeneration, supporting periodontitis treatment.

Oral cancer is another disease of oral soft and hard tissues. Oral cancer is a life-threatening disease, whether local or metastatic. Natural polysaccharides have remarkable adhesion property, which is suitable for the wet oral environment. The polysaccharide sticks to the oral tissues, ensuring precise and sustained delivery of antibiotics. Combining bacterial cellulose, alginate, gelatin, and curcumin forms a multifunctional biopolymer film material that can release curcumin in saliva and has no obvious toxicity to human keratinocytes and human gingival fibroblasts. However, the biopolymer inhibits the growth of oral cancer cells and has good antibacterial activity against Escherichia coli and Staphylococcus aureus and, thus, is suitable for topical wound care and periodontitis and oral cancer treatment (Chiaoprakobkij et al., 2020). TQ/Ca-alg-PVA, a product of loading with calcium alginate and polyvinyl alcohol onto thymoquinone (TQ), inhibits early-stage oral cancer in 7,12-Dimethylbenz [a]anthracene (DMBA) painted hamster by downregulating the expression of NF-KB p50/p65, and PI3K/ AKT/mTOR mRNA (Pu et al., 2021). In summary, natural polysaccharides can achieve precise treatment and reduce the drug resistance of oral cancer.

Oral care

Caries and periodontal disease, the most common disease in the oral cavity, is primarily prevented through oral hygiene. Mouthwash containing natural polysaccharides such as chitosan is a safe and effective natural product for reducing harmful oral microorganisms (Farias et al., 2019). Chitosan mouthwash has been proven safe, and its cytotoxicity is lower than that of commercial mouthwash, and effectively inhibits Streptococcus spp. And Enterococcus spp, preventing oral diseases (Costa et al., 2014). Paper-like nanofiber materials made from bacterial cellulose and chitosan inhibit the growth of bacteria and yeast, biofilm formation, and oxidation (Cabanas-Romero et al., 2020). Given that polysaccharides are biodegradable, they are environmentally friendly biomaterials. Chitosan-based toothpaste prevents tooth enamel erosion and wear. Toothpaste supplemented with chitosan and Sn2+ prevents corrosion and abrasion of teeth gums (Schluet er et al., 2014). In addition, chitosan-containing chewing gum reduces enamel demineralization and maintains bacteriostatic levels in saliva

(Hayashi et al., 2007). Thus, the polysaccharides can be supplemented in the oral care products to prevent oral diseases.

Alveolar ridge preservation after tooth extraction remains an oral implant challenge. A mixture of DBBM-C and HA covered in a collagen membrane prevents dimensional shrinkage and increases bone formation after tooth extraction (Lee et al., 2021). Colonization of the bacteria is a major cause of implant failure (Jiang et al., 2020). Studies have shown that chitosan reduces colonization of *Fusobacterium nucleatum* on the implant surface, plaque biofilm formation, and decrease periimplantitis, increasing the success of implanting (Vaz et al., 2018). Therefore, chitosan is a remarkable antibacterial material that can improve the success of oral implants. Generally, the natural polysaccharide has good biocompatibility and is non-toxic and, thus, an attractive oral care material in stomatology.

Discussion

In recent years, natural polysaccharides extracted from animals, plants, and microorganisms have attracted the attention of researchers because of their good degradability, non-toxicity, and renewability (Zhao et al., 2020). At present, the antibacterial, anti-inflammatory, modifiable, tissue regeneration, and drug carrier potential of polysaccharides have been investigated (Cui et al., 2018; Serrano-Sevilla et al., 2019; Hou et al., 2020; Layek and Mandal, 2020; Zhai et al., 2020). Different polysaccharides are used for treating different oral diseases (Table 1). Natural polysaccharides have attracted greater attention than synthetic materials because they are biocompatible, biodegradable, and ecological.

Despite the advantages of natural polysaccharides in dentistry, these molecules have certain disadvantages in biomedical applications; 1) They have poor mechanical properties. The inferior adhesiveness and the short-term *in vivo* stability of natural polysaccharides limit their therapeutic efficacy (Kim et al., 2021). 2) The quality of polysaccharides is limited to their original material. The method of extraction and purification affects controlled the products' reliability (Luo et al., 2021). 3) Natural polysaccharides are highly moisture sensitive. They undergo hydrolysis during processing and are unstable in the oral cavity (Imre et al., 2019).

Natural polysaccharides are generated through natural processes. However, researches are needed to explore strategies for modifying the chemical structure of these molecules to broaden their biomedical applications. Moreover, the natural polysaccharides are made to the namomaterials (Georgouvelas et al., 2021), which might improve their inherent properties to enhance drug delivery efficacy (Ahmed and Aljaeid, 2016) and applied in wound healing (Yang et al., 2020). In future, natural material could be brought to clinical practice and their effect to environment should be valued. It is believed that advances in the development of biomaterials and

TABLE 1 The applications of polysaccharide in oral diseases.

Polysaccharides	Dental Specialties	Models	Biological Activity/Application	References
Chitosan dispersion	Dental caries	In vitro	Exert antimicrobial effect against mature S. mutans biofilms	Kawakita et al. (2019)
Bacterial cellulose	Bone regeneration	<i>In vivo</i> rat calvarial defect models	The BC biofilms exhibited significantly larger new bone area <i>in vivo</i>	Coelho et al. (2019)
Hyaluronic acid	Oral squamous cell carcinoma	<i>Ex vivo</i> porcine oral mucosal tissues	Deliver DOX to HN22 with a low IC50	Pornpitchanarong et al (2020)
Hyaluronic acid	Oral candidiasis	<i>In vivo</i> sheep buccal mucosa	Hyaluronic acid hydrogel delivers a nanotransfersome with fluconazole entrapped, which exert enhanced antifungal efficacy	Alkhalidi et al. (2020)
Alginate	Recurrent aphthous stomatitis	In vitro	Adhesion time was improved and the AL. Ambroxol was controlled release	Laffleur and Kuppers (2019)
Alginate	Tooth sensitivity	In vitro	The 3D printing dentin mimics is of good cytocompability and could block the dentinal tubule effectively	Naseri et al. (2021)
Bacterial cellulose	Pulpitis	<i>In vivo</i> Sprague–Dawley rat	BC showed higher absorption and expansion than paper points, and maintained a high tensile strength even wet.	Yoshino et al. (2013)
Chitosan	Periodontitis	In vitro	Chitosan induces the proliferation of human gingival fibroblasts by activating of the ERK1/2 signaling pathway	Silva et al. (2013)
Agarose	Dentin hypersensitivity and dental caries	In vitro	Induced a layer of tightly packed hydroxyapatite on the surface of demineralized dentine and the dentinal tubules was occluded	Ning et al. (2012)
Chitosan	Mouthwash	In vitro	The chitosan mouthwash inhibits the streptococci and enterococci and cause no major reductions to the normal oral microflora viability	Costa et al. (2014)
Chitosan	Dental caries filling materials	In vitro	Triclosan-loaded chitosan showed antibacterial activity and induced dentin/adhesive interface stability	Machado et al. (2019)
Chitosan	Modify glass ionomer restorative cements	In vitro	Chitosan modifications increase the ion release of aluminium, sodium, silicon and strontium for materials	Mulder and Anderson- Small (2019)
Chitosan	The nano hydroxyapatite/chitosan composite scaffold for periodontal regeneration	In vivo	mHA/CS could promote periodontal regeneration	Liao et al. (2020)
Bacterial cellulose	The guided bone regeneration (GBR) membranes	BALB/c mice	Promoting soft tissue repair in rat skulls	Farnezi Bassi et al. (2020)
Bacterial cellulose	The guided bone regeneration (GBR) membranes	In vivo	Modulates inflammation, promotes the formation of connective tissue and the increase of calcium and magnesium	Luz et al. (2020)
Bacterial cellulose/ Alginate	Oral mucoadhesive patches for periodontitis or oral cancer treatment	Rats	Showing anticancer activity against oral cancer cells (CAL-27), but non-cytotoxicity to HaCaT and GF cells	Chiaoprakobkij et al. (2020)
Alginate	Oral cancer	In vivo	Inhibits early-stage oral cancer	Pu et al. (2021)
Chitosan	Mouthwash	Swiss albino mice	Antimicrobial effectiveness and toxicological safety	Farias et al. (2019)
Chitosan	Toothpaste	In vitro porcine mucosa	Chitosan enhanced the efficacy of the Sn ²⁺ - containing toothpaste as an anti-erosive/anti- abrasive agent	Schlueter et al. (2014)
Chitosan	Chewing gum	In vivo hamster buccal	Chitosan-containing gum chewing has a better antibacterial effect and increases salivary secretion	Hayashi et al. (2007)
Hyaluronic Acid	Ridge preservation	In vitro	Prevents dimensional shrinkage and increases bone formation after tooth extraction	Lee et al. (2021)

molecular biology-related technologies will enhance the application of natural polysaccharides in stomatology.

Author Contributions

ZY and DW wrote the manuscript. ZY, HL, LC, RL, MH, SK, and WL collected the references and prepared figures. All authors reviewed the manuscript.

Funding

This work was supported by the Fundamental Research Funds of the Jilin Province, Department of Finance (Grant No. jcsz202189313), the Jilin Scientific and Technological Development Program (Grant Nos. 81602377), the Changchun Scientific and Technological Development Program (Grant Nos. 21ZY26), the Jilin Province Scientific

References

Abd El-Hack, M. E., El-Saadony, M. T., Shafi, M. E., Zabermawi, N. M., Arif, M., Batiha, G. E., et al. (2020). Antimicrobial and antioxidant properties of chitosan and its derivatives and their applications: a review. *Int. J. Biol. Macromol.* 164, 2726–2744. doi:10.1016/j.ijbiomac.2020.08.153

Abdelhamid, H. N., and Mathew, A. P. (2022). Cellulose-based nanomaterials advance biomedicine: a review. Int. J. Mol. Sci. 23, 5405. doi:10.3390/ijms23105405

Ahmed, T. A., and Aljaeid, B. M. (2016). Preparation, characterization, and potential application of chitosan, chitosan derivatives, and chitosan metal nanoparticles in pharmaceutical drug delivery. *Drug Des. devel. Ther.* 10, 483–507. doi:10.2147/DDDT.S99651

Ahsan, S. M., Thomas, M., Reddy, K. K., Sooraparaju, S. G., Asthana, A., and Bhatnagar, I. (2018). Chitosan as biomaterial in drug delivery and tissue engineering. *Int. J. Biol. Macromol.* 110, 97–109. doi:10.1016/j.ijbiomac.2017.08.140

Akintoye, S. O., and Mupparapu, M. (2020). Clinical evaluation and anatomic variation of the oral cavity. *Dermatol. Clin.* 38, 399–411. doi:10.1016/j.det.2020. 05.001

Alkhalidi, H. M., Hosny, K. M., and Rizg, W. Y. (2020). Oral gel loaded by fluconazolesesame oil nanotransfersomes: Development, optimization, and assessment of antifungal activity. *Pharmaceutics* 13 (1). doi:10.3390/pharmaceutics13010027

Almeida, L. D. F., Babo, P. S., Silva, C. R., Rodrigues, M. T., Hebling, J., Reis, R. L., et al. (2018). Hyaluronic acid hydrogels incorporating platelet lysate enhance human pulp cell proliferation and differentiation. *J. Mat. Sci. Mat. Med.* 29, 88. doi:10.1007/s10856-018-6088-7

Amin, M. K., and Boateng, J. S. (2022). Enhancing stability and mucoadhesive properties of chitosan nanoparticles by surface modification with sodium alginate and polyethylene glycol for potential oral mucosa vaccine delivery. *Mar. Drugs* 20, 156. doi:10.3390/md20030156

An, S. J., Lee, S. H., Huh, J. B., Jeong, S., Park, J. S., Gwon, H. J., et al. (2017). Preparation and characterization of resorbable bacterial cellulose membranes treated by electron beam irradiation for guided bone regeneration. *Int. J. Mol. Sci.* 18, 2236. doi:10.3390/ijms18112236

Baranov, N., Popa, M., Atanase, L. I., and Ichim, D. L. (2021). Polysaccharidebased drug delivery systems for the treatment of periodontitis. *Molecules* 26, 2735. doi:10.3390/molecules26092735

Beaumont, M., Tran, R., Vera, G., Niedrist, D., Rousset, A., Pierre, R., et al. (2021). Hydrogel-forming algae polysaccharides: from seaweed to biomedical applications. *Biomacromolecules* 22, 1027–1052. doi:10.1021/acs.biomac.0c01406

Bilal, M., and Iqbal, H. M. N. (2019). Marine seaweed polysaccharides-based engineered cues for the modern biomedical sector. *Mar. Drugs* 18, 7. doi:10.3390/md18010007

and Technological Development Program (Grant Nos. 20200801077GH).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Cabanas-Romero, L. V., Valls, C., Valenzuela, S. V., Roncero, M. B., Pastor, F. I. J., Diaz, P., et al. (2020). Bacterial cellulose-chitosan paper with antimicrobial and antioxidant activities. *Biomacromolecules* 21, 1568–1577. doi:10.1021/acs.biomac. 0c00127

Cervino, G., Fiorillo, L., Herford, A., Laino, L., Troiano, G., Amoroso, G., et al. (2018). Alginate materials and dental impression technique: a current state of the art and application to dental practice. *Mar. Drugs* 17, 18. doi:10.3390/md17010018

Chiaoprakobkij, N., Suwanmajo, T., Sanchavanakit, N., and Phisalaphong, M. (2020). Curcumin-Loaded bacterial cellulose/alginate/gelatin as a multifunctional biopolymer composite film. *Molecules* 25, 3800. doi:10. 3390/molecules25173800

Coelho, F., Cavicchioli, M., Specian, S. S., Scarel-Caminaga, R. M., Penteado, L. A., Medeiros, A. I., et al. (2019). Bacterial cellulose membrane functionalized with hydroxiapatite and anti-bone morphogenetic protein 2: A promising material for bone regeneration. *PLoS One* 14 (8), e0221286. doi:10.1371/journal.pone. 0221286

Costa, E. M., Silva, S., Costa, M., Pereira, M., Campos, D., Odila, J., et al. (2014). Chitosan mouthwash: toxicity and *in vivo* validation. *Carbohydr. Polym.* 111, 385–392. doi:10.1016/j.carbpol.2014.04.046

Cui, X., Wang, S., Cao, H., Guo, H., Li, Y., Xu, F., et al. (2018). A review: the bioactivities and pharmacological applications of polygonatum sibiricum polysaccharides. *Molecules* 23, 1170. doi:10.3390/molecules23051170

de Oliveira Barud, H. G., da Silva, R. R., Borges, M. A. C., Castro, G. R., Ribeiro, S. J. L., and da Silva Barud, H. (2020). Bacterial nanocellulose in dentistry: perspectives and challenges. *Molecules* 26, 49. doi:10.3390/molecules26010049

de Oliveira Barud, H. G., da Silva, R. R., da Silva Barud, H., Tercjak, A., Gutierrez, J., Lustri, W. R., et al. (2016). A multipurpose natural and renewable polymer in medical applications: bacterial cellulose. *Carbohydr. Polym.* 153, 406–420. doi:10. 1016/j.carbpol.2016.07.059

Del Fabbro, M., Corbella, S., Sequeira-Byron, P., Tsesis, I., Rosen, E., Lolato, A., et al. (2016). Endodontic procedures for retreatment of periapical lesions. *Cochrane Database Syst. Rev.* 10, CD005511. doi:10.1002/14651858.CD005511.pub3

Farias, J. M., Stamford, T. C. M., Resende, A. H. M., Aguiar, J. S., Rufino, R. D., Luna, J. M., et al. (2019). Mouthwash containing a biosurfactant and chitosan: an eco-sustainable option for the control of cariogenic microorganisms. *Int. J. Biol. Macromol.* 129, 853–860. doi:10.1016/j.ijbiomac.2019.02.090

Farnezi Bassi, A. P., Bizelli, V. F., Brasil, L. F. d. M., Pereira, J. C., Al-Sharani, H. M., Momesso, G. A. C., et al. (2020). Is the bacterial cellulose membrane feasible for osteopromotive property? *Membr.* (*Basel*) 10, 230. doi:10.3390/membranes10090230

Frassica, M. T., and Grunlan, M. A. (2020). Perspectives on synthetic materials to guide tissue regeneration for osteochondral defect repair. *ACS Biomater. Sci. Eng.* 6, 4324–4336. doi:10.1021/acsbiomaterials.0c00753

Georgouvelas, D., Abdelhamid, H. N., Li, J., Edlund, U., and Mathew, A. P. (2021). All-cellulose functional membranes for water treatment: adsorption of metal ions and catalytic decolorization of dyes. *Carbohydr. Polym.* 264, 118044. doi:10.1016/j.carbpol.2021.118044

Graca, M. F. P., Miguel, S. P., Cabral, C. S. D., and Correia, I. J. (2020). Hyaluronic acid-based wound dressings: a review. *Carbohydr. Polym.* 241, 116364. doi:10.1016/j.carbpol.2020.116364

Halib, N., Ahmad, I., Grassi, M., and Grassi, G. (2019). The remarkable threedimensional network structure of bacterial cellulose for tissue engineering applications. *Int. J. Pharm. X.* 566, 631–640. doi:10.1016/j.ijpharm.2019.06.017

Hayashi, Y., Ohara, N., Ganno, T., Ishizaki, H., and Yanagiguchi, K. (2007). Chitosan-containing gum chewing accelerates antibacterial effect with an increase in salivary secretion. J. Dent. (Shiraz). 35, 871–874. doi:10.1016/j.jdent.2007.08.004

Hou, C., Chen, L., Yang, L., and Ji, X. (2020). An insight into anti-inflammatory effects of natural polysaccharides. *Int. J. Biol. Macromol.* 153, 248–255. doi:10.1016/j.ijbiomac.2020.02.315

Huang, G., and Huang, H. (2018). Application of hyaluronic acid as carriers in drug delivery. *Drug Deliv. (Lond).* 25, 766–772. doi:10.1080/10717544.2018. 1450910

Hudson, D., and Margaritis, A. (2014). Biopolymer nanoparticle production for controlled release of biopharmaceuticals. *Crit. Rev. Biotechnol.* 34, 161–179. doi:10. 3109/07388551.2012.743503

Ibrahim, M. S., Garcia, I. M., Kensara, A., Balhaddad, A. A., Collares, F. M., Williams, M. A., et al. (2020). How we are assessing the developing antibacterial resin-based dental materials? a scoping review. *J. Dent. (Shiraz).* 99, 103369. doi:10. 1016/j.jdent.2020.103369

Imre, B., Garcia, L., Puglia, D., and Vilaplana, F. (2019). Reactive compatibilization of plant polysaccharides and biobased polymers: review on current strategies, expectations and reality. *Carbohydr. Polym.* 209, 20–37. doi:10.1016/j.carbpol.2018.12.082

Jiang, X., Yao, Y., Tang, W., Han, D., Zhang, L., Zhao, K., et al. (2020). Design of dental implants at materials level: an overview. *J. Biomed. Mat. Res. A* 108, 1634–1661. doi:10.1002/jbm.a.36931

Karolewicz, B. (2016). A review of polymers as multifunctional excipients in drug dosage form technology. *Saudi Pharm. J.* 24, 525–536. doi:10.1016/j.jsps.2015. 02.025

Kawakita, E. R. H., Re, A. C. S., Peixoto, M. P. G., Ferreira, M. P., Ricomini-Filho, A. P., Freitas, O., et al. (2019). Effect of chitosan dispersion and microparticles on older Streptococcus mutans biofilms. *Molecules* 24, 1808. doi:10.3390/molecules24091808

Kim, C., Jeong, D., Kim, S., Kim, Y., and Jung, S. (2019). Cyclodextrin functionalized agarose gel with low gelling temperature for controlled drug delivery systems. *Carbohydr. Polym.* 222, 115011. doi:10.1016/j.carbpol.2019. 115011

Kim, J., Kim, S., Son, D., and Shin, M. (2021). Phenol-hyaluronic acid conjugates: correlation of oxidative crosslinking pathway and adhesiveness. *Polym. (Basel)* 13, 3130. doi:10.3390/polym13183130

Klemm, D., Heublein, B., Fink, H. P., and Bohn, A. (2005). Cellulose: fascinating biopolymer and sustainable raw material. *Angew. Chem. Int. Ed.* 44, 3358–3393. doi:10.1002/anie.200460587

Klinthoopthamrong, N., Chaikiawkeaw, D., Phoolcharoen, W., Rattanapisit, K., Kaewpungsup, P., Pavasant, P., et al. (2020). Bacterial cellulose membrane conjugated with plant-derived osteopontin: preparation and its potential for bone tissue regeneration. *Int. J. Biol. Macromol.* 149, 51–59. doi:10.1016/j. ijbiomac.2020.01.158

Kou, S. G., Peters, L. M., and Mucalo, M. R. (2021). Chitosan: a review of sources and preparation methods. *Int. J. Biol. Macromol.* 169, 85–94. doi:10.1016/j.ijbiomac. 2020.12.005

Kunert, M., and Lukomska-Szymanska, M. (2020). Bio-inductive materials in direct and indirect pulp capping-a review article. *Mater. (Basel)* 13, 1204. doi:10. 3390/ma13051204

Laffleur, F., and Kuppers, P. (2019). Adhesive alginate for buccal delivery in aphthous stomatitis. *Carbohydr. Res.* 477, 51–57. doi:10.1016/j.carres.2019.03.009

Lauritano, D., Limongelli, L., Moreo, G., Favia, G., and Carinci, F. (2020). Nanomaterials for periodontal tissue engineering: chitosan-based scaffolds. A systematic review. *Nanomater. (Basel)* 10, 605. doi:10.3390/nano10040605

Layek, B., and Mandal, S. (2020). Natural polysaccharides for controlled delivery of oral therapeutics: a recent update. *Carbohydr. Polym.* 230, 115617. doi:10.1016/j. carbpol.2019.115617

Lee, J. B., Chu, S., Ben Amara, H., Song, H. Y., Son, M. J., Lee, J., et al. (2021). Effects of hyaluronic acid and deproteinized bovine bone mineral with 10% collagen for ridge preservation in compromised extraction sockets. *J. Periodontol.* 92, 1564–1575. doi:10.1002/JPER.20-0832

Liao, Y., Li, H., Shu, R., Chen, H., Zhao, L., Song, Z., et al. (2020). Mesoporous hydroxyapatite/chitosan loaded with recombinant-human amelogenin could enhance antibacterial effect and promote periodontal regeneration. *Front. Cell. Infect. Microbiol.* 10, 180. doi:10.3389/fcimb.2020.00180

Ling, Z., He, Y., Huang, H., Xie, X., Li, Q. I., and Cao, C. Y. (2019). Effects of oligopeptide simulating DMP-1/mineral trioxide aggregate/agarose hydrogel biomimetic mineralisation model for the treatment of dentine hypersensitivity. *J. Mat. Chem. B* 7, 5825–5833. doi:10.1039/c9tb01684h

Luo, M., Zhang, X., Wu, J., and Zhao, J. (2021). Modifications of polysaccharide-based biomaterials under structure-property relationship for biomedical applications. *Carbohydr. Polym.* 266, 118097. doi:10.1016/j. carbpol.2021.118097

Luz, E., das Chagas, B. S., de Almeida, N. T., de Fatima Borges, M., Andrade, F. K., Muniz, C. R., et al. (2020). Resorbable bacterial cellulose membranes with strontium release for guided bone regeneration. *Mater. Sci. Eng. C* 116, 111175. doi:10.1016/j. msec.2020.111175

Lynd, L. R., Weimer, P. J., van Zyl, W. H., and Pretorius, I. S. (2002). Microbial cellulose utilization: fundamentals and biotechnology. *Microbiol. Mol. Biol. Rev.* 66, 506–577. doi:10.1128/MMBR.66.3.506-577.2002

Machado, A. H. S., Garcia, I. M., Motta, A. S. D., Leitune, V. C. B., and Collares, F. M. (2019). Triclosan-loaded chitosan as antibacterial agent for adhesive resin. *J. Dent. (Shiraz).* 83, 33–39. doi:10.1016/j.jdent.2019.02.002

Malkondu, O., Karapinar Kazandag, M., and Kazazoglu, E. (2014). A review on biodentine, a contemporary dentine replacement and repair material. *Biomed. Res. Int.* 2014, 160951. doi:10.1155/2014/160951

Marinho, A., Nunes, C., and Reis, S. (2021). Hyaluronic acid: a key ingredient in the therapy of inflammation. *Biomolecules* 11, 1518. doi:10.3390/biom11101518

Melo, M. A., Guedes, S. F., Xu, H. H., and Rodrigues, L. K. (2013). Nanotechnology-based restorative materials for dental caries management. *Trends Biotechnol.* 31, 459–467. doi:10.1016/j.tibtech.2013.05.010

Mulder, R., and Anderson-Small, C. (2019). Ion release of chitosan and nanodiamond modified glass ionomer restorative cements. *Clin. Cosmet. Investig. Dent.* 11, 313–320. doi:10.2147/CCIDE.S220089

Mushtaq, A., Li, L., and Grondahl, L. (2021). Chitosan nanomedicine in cancer therapy: targeted delivery and cellular uptake. *Macromol. Biosci.* 21, e2100005. doi:10.1002/mabi.202100005

Muxika, A., Etxabide, A., Uranga, J., Guerrero, P., and de la Caba, K. (2017). Chitosan as a bioactive polymer: processing, properties and applications. *Int. J. Biol. Macromol.* 105, 1358–1368. doi:10.1016/j.ijbiomac.2017.07.087

Narmani, A., and Jafari, S. M. (2021). Chitosan-based nanodelivery systems for cancer therapy: recent advances. *Carbohydr. Polym.* 272, 118464. doi:10.1016/j. carbpol.2021.118464

Naseri, S., Cooke, M. E., Rosenzweig, D. H., and Tabrizian, M. (2021). 3D printed *in vitro* dentin model to investigate occlusive agents against tooth sensitivity. *Mater.* (*Basel*) 14, 7255. doi:10.3390/ma14237255

Ning, T. Y., Xu, X. H., Zhu, L. F., Zhu, X. P., Chu, C. H., Liu, L. K., et al. (2012). Biomimetic mineralization of dentin induced by agarose gel loaded with calcium phosphate. *J. Biomed. Mat. Res.* 100, 138–144. doi:10.1002/jbm.b.31931

Niu, X., Wang, L., Xu, M., Qin, M., Zhao, L., Wei, Y., et al. (2021). Electrospun polyamide-6/chitosan nanofibers reinforced nano-hydroxyapatite/polyamide-6 composite bilayered membranes for guided bone regeneration. *Carbohydr. Polym.* 260, 117769. doi:10.1016/j.carbpol.2021.117769

Paradowska-Stolarz, A., Wieckiewicz, M., Owczarek, A., and Wezgowiec, J. (2021). Natural polymers for the maintenance of oral health: review of recent advances and perspectives. *Int. J. Mol. Sci.* 22, 10337. doi:10.3390/ijms221910337

Pornpitchanarong, C., Rojanarata, T., Opanasopit, P., Ngawhirunpat, T., and Patrojanasophon, P. (2020). Catechol-modified chitosan/hyaluronic acid nanoparticles as a new avenue for local delivery of doxorubicin to oral cancer cells. *Colloids Surfaces B Biointerfaces* 196, 111279. doi:10.1016/j.colsurfb.2020. 111279

Prajapati, V. D., Maheriya, P. M., Jani, G. K., and Solanki, H. K. (2014). Retracted: Carrageenan: a natural seaweed polysaccharide and its applications. *Carbohydr. Polym.* 105, 97–112. doi:10.1016/j.carbpol.2014.01.067

Pu, Y., Hu, S., Chen, Y., Zhang, Q., Xia, C., Deng, H., et al. (2021). Thymoquinone loaded calcium alginate and polyvinyl alcohol carrier inhibits the 7, 12dimethylbenz[a]anthracene-induced hamster oral cancer via the downregulation of P13K/AKT/mTOR signaling pathways. *Environ. Toxicol.* 36, 339–351. doi:10.1002/tox.23040 Rahmati, M., Alipanahi, Z., and Mozafari, M. (2019). Emerging biomedical applications of algal polysaccharides. *Curr. Pharm. Des.* 25, 1335–1344. doi:10. 2174/1381612825666190423160357

Roumanas, E. D. (2009). The social solution-denture esthetics, phonetics, and function. J. Prosthodont. 18, 112-115. doi:10.1111/j.1532-849X.2009.00440.x

Sah, A. K., Dewangan, M., and Suresh, P. K. (2019). Potential of chitosan-based carrier for periodontal drug delivery. *Colloids Surfaces B Biointerfaces* 178, 185–198. doi:10.1016/j.colsurfb.2019.02.044

Salari, N., Mansouri, K., Valipour, E., Abam, F., Jaymand, M., Rasoulpoor, S., et al. (2021). Hyaluronic acid-based drug nanocarriers as a novel drug delivery system for cancer chemotherapy: a systematic review. *DARU J. Pharm. Sci.* 29, 439–447. doi:10. 1007/s40199-021-00416-6

Sanchez-Ballester, N. M., Bataille, B., and Soulairol, I. (2021). Sodium alginate and alginic acid as pharmaceutical excipients for tablet formulation: Structure-function relationship. *Carbohydr. Polym.* 270, 118399. doi:10.1016/j.carbpol. 2021.118399

Schlueter, N., Klimek, J., and Ganss, C. (2014). Effect of a chitosan additive to a Sn2+-containing toothpaste on its anti-erosive/anti-abrasive efficacy--a controlled randomised *in situ* trial. *Clin. Oral Investig.* 18, 107–115. doi:10.1007/s00784-013-0941-3

Schmidt, J., Pilbauerova, N., Soukup, T., Suchankova-Kleplova, T., and Suchanek, J. (2020). Low molecular weight hyaluronic acid effect on dental pulp stem cells *in vitro. Biomolecules* 11, 22. doi:10.3390/biom11010022

Serrano-Sevilla, I., Artiga, A., Mitchell, S. G., De Matteis, L., and de la Fuente, J. M. (2019). Natural polysaccharides for siRNA delivery: nanocarriers based on chitosan, hyaluronic acid, and their derivatives. *Molecules* 24, 2570. doi:10.3390/molecules24142570

Shivakumar, P., Gupta, M. S., Jayakumar, R., and Gowda, D. V. (2021). Prospection of chitosan and its derivatives in wound healing: proof of patent analysis (2010-2020). *Int. J. Biol. Macromol.* 184, 701–712. doi:10.1016/j.ijbiomac. 2021.06.086

Silva, A. C. Q., Silvestre, A. J. D., Vilela, C., and Freire, C. S. R. (2021). Natural polymers-based materials: a contribution to a greener future. *Molecules* 27, 94. doi:10.3390/molecules27010094

Silva, D., Arancibia, R., Tapia, C., Acuna-Rougier, C., Diaz-Dosque, M., Caceres, M., et al. (2013). Chitosan and platelet-derived growth factor synergistically stimulate cell proliferation in gingival fibroblasts. *J. Periodontal Res.* 48, 677–686. doi:10.1111/jre.12053

Singh, J., Tan, N. C., Mahadevaswamy, U. R., Chanchareonsook, N., Steele, T. W., and Lim, S. (2021). Bacterial cellulose adhesive composites for oral cavity applications. *Carbohydr. Polym.* 274, 118403. doi:10.1016/j.carbpol.2021.118403

Suharyani, I., Fouad Abdelwahab Mohammed, A., Muchtaridi, M., Wathoni, N., and Abdassah, M. (2021). Evolution of drug delivery systems for recurrent aphthous stomatitis. *Drug Des. devel. Ther.* 15, 4071–4089. doi:10.2147/DDDT. S328371

Svensson, A., Nicklasson, E., Harrah, T., Panilaitis, B., Kaplan, D., Brittberg, M., et al. (2005). Bacterial cellulose as a potential scaffold for tissue engineering of cartilage. *Biomaterials* 26, 419–431. doi:10.1016/j.biomaterials.2004.02.049

Timur, S. S., Yuksel, S., Akca, G., and Senel, S. (2019). Localized drug delivery with mono and bilayered mucoadhesive films and wafers for oral mucosal infections. *Int. J. Pharm.* X. 559, 102–112. doi:10.1016/j.ijpharm.2019.01.029

Torres, F. G., Troncoso, O. P., Pisani, A., Gatto, F., and Bardi, G. (2019). Natural polysaccharide nanomaterials: an overview of their immunological properties. *Int. J. Mol. Sci.* 20, 5092. doi:10.3390/ijms20205092

Trivedi, T. J., Bhattacharjya, D., Yu, J. S., and Kumar, A. (2015). Functionalized agarose self-healing ionogels suitable for supercapacitors. *ChemSusChem* 8, 3294–3303. doi:10.1002/cssc.201500648

Tuominen, H., and Rautava, J. (2021). Oral microbiota and cancer development. *Pathobiology* 88, 116–126. doi:10.1159/000510979

Vaz, J. M., Pezzoli, D., Chevallier, P., Campelo, C. S., Candiani, G., and Mantovani, D. (2018). Antibacterial coatings based on chitosan for pharmaceutical and biomedical applications. *Curr. Pharm. Des.* 24, 866-885. doi:10.2174/1381612824666180219143900

Vergnes, J. N., and Mazevet, M. (2020). Oral diseases: a global public health challenge. Lancet 395, 186. doi:10.1016/S0140-6736(19)33015-6

Voicu, G., Jinga, S. I., Drosu, B. G., and Busuioc, C. (2017). Improvement of silicate cement properties with bacterial cellulose powder addition for applications in dentistry. *Carbohydr. Polym.* 174, 160–170. doi:10.1016/j. carbpol.2017.06.062

Wang, W., Meng, Q., Li, Q., Liu, J., Zhou, M., Jin, Z., et al. (2020). Chitosan derivatives and their application in biomedicine. *Int. J. Mol. Sci.* 21, 487. doi:10. 3390/ijms21020487

Whitters, C. J., Strang, R., Brown, D., Clarke, R., Curtis, R., Hatton, P., et al. (1999). Dental materials: 1997 literature review. *J. Dent. (Shiraz)*. 27, 401–435. doi:10.1016/s0300-5712(99)00007-x

Wong, F. M. F., Ng, Y. T. Y., and Leung, W. K. (2019). Oral health and its associated factors among older institutionalized residents-a systematic review. *Int. J. Environ. Res. Public Health* 16, 4132. doi:10.3390/ ijerph16214132

Yang, M., Ward, J., and Choy, K. L. (2020). Nature-inspired bacterial cellulose/methylglyoxal (BC/MGO) nanocomposite for broad-spectrum antimicrobial wound dressing. *Macromol. Biosci.* 20, e2000070. doi:10. 1002/mabi.202000070

Yoshino, A., Tabuchi, M., Uo, M., Tatsumi, H., Hideshima, K., Kondo, S., et al. (2013). Applicability of bacterial cellulose as an alternative to paper points in endodontic treatment. *Acta Biomater*. 9, 6116–6122. doi:10.1016/j. actbio.2012.12.022

Yu, Y., Shen, M., Song, Q., and Xie, J. (2018). Biological activities and pharmaceutical applications of polysaccharide from natural resources: a review. *Carbohydr. Polym.* 183, 91–101. doi:10.1016/j.carbpol.2017.12.009

Zhai, P., Peng, X., Li, B., Liu, Y., Sun, H., and Li, X. (2020). The application of hyaluronic acid in bone regeneration. *Int. J. Biol. Macromol.* 151, 1224–1239. doi:10.1016/j.ijbiomac.2019.10.169

Zhang, C., Hui, D., Du, C., Sun, H., Peng, W., Pu, X., et al. (2021). Preparation and application of chitosan biomaterials in dentistry. *Int. J. Biol. Macromol.* 167, 1198-1210. doi:10.1016/j.jibiomac.2020.11.073

Zhao, Y., Yan, B., Wang, Z., Li, M., and Zhao, W. (2020). Natural polysaccharides with immunomodulatory activities. *Mini Rev. Med. Chem.* 20, 96-106. doi:10.2174/1389557519666190913151632

Zucca, P., Fernandez-Lafuente, R., and Sanjust, E. (2016). Agarose and its derivatives as supports for enzyme immobilization. *Molecules* 21, 1577. doi:10.3390/molecules21111577