



Ferroptosis, a Potential Therapeutic Target in Alzheimer's Disease

Kai Chen^{1,2†}, Xiaobing Jiang^{2†}, Moxin Wu³, Xianming Cao³, Wendai Bao^{1,4*} and Ling-Qiang Zhu^{1*}

¹ Key Lab of Neurological Disorder of Education Ministry, Department of Pathophysiology, School of Basic Medicine, Tongji Medical College, Huazhong University of Science and Technology, Wuhan, China, ² Department of Neurosurgery, Wuhan Union Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan, China, ³ Department of Jiujiang Clinical Research Center for Precision Medicine, Affiliated Hospital of Jiujiang University, Jiujiang, China, ⁴ College of Biomedicine and Health, Huazhong Agricultural University, Wuhan, China

OPEN ACCESS

Edited by:

Guangyong Peng, Saint Louis University, United States

Reviewed by:

Chunying Li, Georgia State University, United States Jun Xu, Capital Medical University, China

*Correspondence:

Wendai Bao 512483463@qq.com Ling-Qiang Zhu zhulq@mail.hust.edu.cn

[†]These authors have contributed equally to this work

Specialty section:

This article was submitted to Cell Death and Survival, a section of the journal Frontiers in Cell and Developmental Biology

> Received: 02 May 2021 Accepted: 09 July 2021 Published: 05 August 2021

Citation:

Chen K, Jiang X, Wu M, Cao X, Bao W and Zhu L-Q (2021) Ferroptosis, a Potential Therapeutic Target in Alzheimer's Disease. Front. Cell Dev. Biol. 9:704298. doi: 10.3389/fcell.2021.704298 Cell death is a common phenomenon in the progression of Alzheimer's disease (AD). However, the mechanism of triggering the death of neuronal cells remains unclear. Ferroptosis is an iron-dependent lipid peroxidation-driven cell death and emerging evidences have demonstrated the involvement of ferroptosis in the pathological process of AD. Moreover, several hallmarks of AD pathogenesis were consistent with the characteristics of ferroptosis, such as excess iron accumulation, elevated lipid peroxides, and reactive oxygen species (ROS), reduced glutathione (GSH), and glutathione peroxidase 4 (GPX4) levels. Besides, some ferroptosis inhibitors can relieve AD-related pathological symptoms in AD mice and exhibit potential clinical benefits in AD patients. Therefore, ferroptosis is gradually being considered as a distinct cell death mechanism in the pathogenesis of AD. However, direct evidence is still lacking. In this review, we summarize the features of ferroptosis in AD, its underlying mechanisms in AD pathology, and review the application of ferroptosis inhibitors in both AD clinical trials and mice/cell models, to provide valuable information for future treatment and prevention of this devastating disease.

Keywords: ferroptosis, cell death, iron, lipid peroxidation, AD

INTRODUCTION

Ferroptosis is a recently defined iron-dependent form of cell death induced by the small molecule erastin (Dixon et al., 2012). This newly described process possesses different morphological, biochemical and genetical features, which is distinct from apoptosis, necrosis, autophagy, and other forms of cell death (Weiland et al., 2019). The classic features of ferroptosis include the specifical changes of cellular morphology, the iron-dependent accumulation of lipid peroxides, and ROS, the depletion of GSH, and inactivation of GPX4, and a distinct set of regulated genes (Dixon et al., 2012; Dixon, 2017; Chen et al., 2021). Additionally, ferroptosis could be specifically inhibited by a series of inhibitors, such as iron chelators deferoxamine (DFO) and lipid peroxidation inhibitors ferrostatin-1 (Fer-1), but not sensitive to apoptotic or necroptotic inhibitors (Dixon et al., 2012). Notably, signatures of ferroptosis, such as excess iron accumulation, elevated lipid peroxides, and ROS generation have long been discovered in brains of AD patients and model mice (Zhang et al., 2012; Hambright et al., 2017; Ayton et al., 2019). Moreover, some ferroptosis inhibitors, such as desferrioxamine (DFE) and vitamin E have exhibited clinical benefits in AD

1

(Mclachlan et al., 1991; Dysken, 2014; **Figure 1**). Therefore, these findings strongly indicated the involvement of ferroptosis in the pathogenesis of AD. However, the direct evidence of ferroptosis in AD pathology is still lacking. In this article, we will systematically review the current knowledge of ferroptosis in AD, discuss the underlying mechanism, and describe the possible therapeutic strategies to treat this disease.

The clinical features of AD which were consistent with the characteristics of ferroptosis were noted here, including high concentrations of iron in AD brains, brain atrophy coincident with the iron accumulated sites, elevated lipid peroxidation products, such as MDA, isoprostanes, 4-HNE, malondialdehyde and acrolein, and clinical benefits of some ferroptosis inhibitors.

CHARACTERISTICS OF FERROPTOSIS IN AD

Excess Iron Accumulation

Ferroptosis is an iron-dependent cell death form and emerging evidences have reported that iron overload could directly induce ferroptosis in vivo and in vitro under pathological process (Wang et al., 2017; Fang et al., 2019). In the central nervous system, iron is an essential element that is involved in many important biological processes in brain, such as oxygen transportation, myelin production, and the synthesis of neurotransmitters (Ward et al., 2014). Several studies have observed the selective accumulation of iron in the $A\beta$ aggregation and neurofibrillary tangles of AD brains (Good et al., 1992; Tao et al., 2014), and the excess brain iron accumulation was associated with accelerated cognitive decline in AD patients (Ayton et al., 2019). Moreover, high concentrations of iron which was detected by immune-histochemical staining (Xian-hui et al., 2015; Sands et al., 2016) and MRI analysis (Schenck et al., 2006; Langkammer et al., 2014), are accumulated in various brain regions [including temporal cortex (Ayton et al., 2019), hippocampus (Schenck and Zimmerman, 2004; Antharam et al., 2012), caudate nucleus and the basal ganglia (Langkammer et al., 2014)] and associated

with the motor and cognitive impairments of AD pathology. Furthermore, age-related iron accumulation contributed to the tissue damage and pathologic manifestations of AD (Raven et al., 2013). Collectively, these findings indicate that excess iron accumulation is a common pathological feature in the course of AD.

Brain atrophy is a prominent feature of AD and reduced volumes of medial-temporal part of brain, especially the hippocampus was frequently reported in the brains of AD patients, which was closely correlated with the impaired global cognitive performance (de Jong et al., 2008; Uysal and Ozturk, 2020). Moreover, the diffuse cortical atrophy could also be observed in patients with abnormal cognitive scores (Ten Kate et al., 2018). Interestingly, the brain regions which suffered neuronal degeneration and brain atrophy were highly coincident with the sites of iron accumulation (Horvath et al., 2012), these findings strongly suggest that the excess iron accumulation is linked to interferonopathies of AD. On the other hand, iron is an important component of the catalytic subunit of lipoxygenase (LOX) that catalyzes the oxygenation of polyunsaturated fatty acids (PUFAs) (Kuhn et al., 2015). It could generate the ROS through the donation of electrons to oxygens, and promote the ferroptosis (Stockwell et al., 2017). Thus, the perturbed iron distribution in AD probably induced or enhanced the ferroptosis, which could contribute to neuronal death and degeneration in this disease.

Elevated Lipid Peroxides and ROS

Accumulation of massive lipid peroxides to lethal levels was another characteristic of ferroptosis, and the biomarkers of lipid peroxides were also elevated in the AD pathology. Clinically, lipid peroxidation metabolites showed co-localization with the amyloid plaques, and were highly related to the AD progression (Benseny-Cases et al., 2014). For early and non-invasive diagnosis, various studies were done to evaluate the lipid peroxidation products, such as malondialdehyde (MDA) (Krishnan and Rani, 2014), isoprostanes (Peskind et al., 2014), 4-Hydroxynonenal (4-HNE) (Di Domenico et al., 2017),



malondialdehyde and acrolein (Williams et al., 2006; Di Domenico et al., 2017), as the diagnostic markers in the onset of AD. Among these candidates, MDA, isoprostanes, and 4-HNE were most promising and consistent between different studies (Pena-Bautista et al., 2019). These findings indicated that the accumulated lipid peroxides also contribute to the neuropathology of AD, and some lipid peroxidation products can act as the biomarkers for AD diagnosis and prognosis.

Besides the increased lipid peroxides, accumulated lipid ROS and decreased cortical GSH content also have been found in the AD pathology. Previous studies reported that excess ROS is generated under pathological AD conditions and reducing ROS accumulation could restore cognition in AD model rats (Smith et al., 2010; Wang et al., 2016). Moreover, the levels of membrane phospholipids [phosphatidylethanolamine (PE) and phosphatidylinositol (PI)]-derived total fatty acids were significantly decreased in the hippocampus of AD patients (Prasad et al., 1998), and AD-associated reductions of GSH levels also have been observed in both animal models (Resende et al., 2008; Zhang et al., 2012) and the human brains (Chiang et al., 2017). Notably, the levels of GSH showed a close relationship with the brain amyloidosis and AD pathology. Overall, all of these evidences implicated that lipid oxidative stress, the key process of ferroptosis, is also intimately involved in the pathological development of AD.

FERROPTOSIS-RELATED SIGNALING PATHWAYS IN AD

Ferroptosis in AD has been extensively studied in the past decades and there are multiple signaling pathways were reported in the regulation of ferroptotic cell death in AD, such as the perturbed iron export and transport in iron metabolism pathway, the reduced GSH and Gpx4 levels in redox homeostasis pathway, the accumulated lipid peroxidation and ROS generation in lipid metabolism pathway and some other potential signaling pathways, as summarized in **Figure 2**. In the following text, we will discuss the mechanism of each ferroptosis-related signaling pathway in AD.

Signaling pathways that may be pathologically involved in the ferroptosis in AD. (1) APP ferroxidase activity is inhibited and Ferroportin (Fpn) is reduced in the AD brain, which inhibited the iron export and resulted in the excess iron accumulation. Moreover, the levels of miR-124, which directly induced a posttranscriptional deregulation of Fpn, also increased in AD patients; (2) Iron accumulation aggravated toxic A β deposition and tau hyperphosphorylation, which disrupted GSH synthesis and reduced Gpx4 levels (3); (4) Elevated LOX catalyzed PUFAs and generated ROS via Fenton reaction. Additionally, A β deposition also led to lipid peroxidation and ROS generation. (5) Other pathways, such as p53, Nrf2, MAPK etc.

Iron Metabolism Pathway

Iron dysregulation, one of the hallmarks of ferroptosis, have been implicated in AD pathogenesis for a long time. Iron dyshomeostasis has been reported to aggravate toxic amyloid- β

(A β) deposition and hyperphosphorylated tau aggregation, the two main histopathological features of AD. Indeed, tau hyperphosphorylation and aggregation also contribute to toxic neuronal iron accumulation, which leads to a vicious cycle (Yan and Zhang, 2019). Moreover, rats fed with high iron-diet demonstrated an increased A β deposition and tau hyperphosphorylation, and iron chelator effectively attenuated A β load and tau aggregation (Sripetchwandee et al., 2014). Similarly, Tau deficiency impairs amyloid-beta precursor protein (APP)-mediated iron export and this change can be prevented by a moderate iron chelator, clioquinol (Lei et al., 2012). Taken together, these studies suggest that iron dysregulation plays a deleterious role in AD and targeting iron metabolism pathway may be an effective way for manipulating ferroptosis in AD.

Iron export and iron transport are the two aspects of maintaining iron homeostasis. However, the expression levels of two major iron-exporters, ferroportin (Fpn) and ironhomeostatic peptide hepcidin, were both reported to be reduced in the AD brain (Raha et al., 2013). Moreover, the APP ferroxidase activity, which could interact with iron-exporter Fpn and facilitate the removal of cytoplasmic iron, also was inhibited in AD postmortem neocortex (Duce et al., 2010). Interestingly, the levels of miR-124, which directly induced a posttranscriptional deregulation of Fpn is also increased in AD patients, future confirming the deregulation of Fpn in AD (Wang et al., 2018; Bao et al., 2020). Accordingly, a recent study indicated that deficiency of Fpn in principal neurons in neocortex increased iron levels and induced AD-like hippocampal atrophy and memory deficits, while restoring Fpn expression could effectively ameliorate the memory loss and ferroptosis in AD model mice (Bao et al., 2021). Besides the perturbed iron export, the expression levels of divalent metal transporter 1 (Dmt1), the main transporter of divalent metal ions which is colocalized with the $A\beta$ in AD pathology, were significantly increased in the AD transgenic model and aged murine model (Xianhui et al., 2015; Tian et al., 2018). Interestingly, silencing of endogenous Dmt1 could reduce iron accumulation and lead to reduction of Aß secretion (Zheng et al., 2009). Additionally, transferrin receptor protein 1 (Tfr1), the major vehicle of iron delivery into cells, also was significantly upregulated in AD mice. Treatment with the active compounds of herbs ameliorate impaired cognition and this beneficial effect is associated with reduced impairment of iron metabolism (Yu et al., 2019; Ma et al., 2021). Thus, enhancing iron export or inhibiting iron transport could be a promising strategy to reduce the ferroptotic cell death in this disease.

Redox Homeostasis Pathway

Redox homeostasis is a vital component of a healthy physiological steady state. GSH is the major cellular antioxidant which maintaining redox homeostasis from the following two aspects: bind to ferrous iron in the labile iron pool (LIP) to prevent the iron-dependent oxidization (Hider and Kong, 2011) and as the substrate of GPX4-mediate lipid detoxification (Friedmann Angeli et al., 2014; Yang et al., 2014). Numerous studies have shown that alterations of GSH content impaired redox



homeostasis and are associated with the occurrence of ferroptosis in AD (Pocernich and Butterfield, 2012; Hambright et al., 2017). Casley et al. (2002) reported that addition of AB could induce depletion of GSH in cultured neurons. Inversely, administration of GSH precursor gamma-glutamylcysteine ethyl ester (GCEE) increased cellular GSH levels and protected against Aβ-mediated neurotoxicity in neuronal cells (Boyd-Kimball et al., 2005). Moreover, AD-associated reductions of brain GSH levels have been observed during AD onset and progression in vitro (Ghosh et al., 2012) and in vivo (Resende et al., 2008). These findings strongly indicated that disruption in GSH homeostasis is associated with AD pathology. In addition to alterations in GSH levels, GSH-associated antioxidant enzyme glutathione S-transferases (GSTs), which catalyze the reaction between GSH and 4-HNE, was also changed in AD pathology (Aquilano et al., 2014). Both the enzyme activity and protein levels of GSTs were significantly decreased in most brain regions and cerebrospinal fluid (CSF) in AD patients (Lovell et al., 1998). Moreover, glutathione S-transferase omega-1 and 2 genes (GSTO1, GSTO2) polymorphisms have been implicated as risk factors for earlier onset of AD (Allen et al., 2012). Thus, restoring GSH levels would be an effective way of attenuating AD progression. Indeed, some researches have focused on finding potential approaches for restoring GSH levels in AD patients (Bermejo et al., 2008; Cacciatore et al., 2012).

In addition to the disruption of GSH synthesis, inhibition of Gpx4 was another vulnerability factor for ferroptosis-related AD pathogenesis (Friedmann Angeli et al., 2014). Gpx4 is a member of an antioxidant enzyme family which catalyze the reduction of hydroperoxides by GSH or other biological reductants

(Cardoso et al., 2017). This family was composed of eight isoenzymes (Gpx1-Gpx8) and Gpx4 is the most widely expressed one in brain (Wang et al., 2013). Previous study has reported that Gpx4 was identified as the key regulatory enzyme in ferroptosis of AD (Yang et al., 2014). Ablation of Gpx4 in forebrain, the frequently affected region in AD pathology, leads to cognitive impairment and hippocampal neurodegeneration in Gpx4 brain inducible knockout (Gpx4BIKO) mice. Moreover, the classical features of ferroptosis, such as elevated lipid peroxidation, extracellular signal-regulated kinase (ERK) activation and amplified neuroinflammation, were also observed and treatment with a ferroptosis inhibitor could ameliorate neurodegeneration in those mice (Hambright et al., 2017). Besides the inhibition of Gpx4 enzyme activity, the expression of guanine-rich sequencebinding factor (Grsf1), which controls the translation of Gpx4, were also downregulated in brain of AD model mice (Yoo et al., 2010). Moreover, polymorphisms in Gpx1 and Gpx4 were significantly associated with memory impairment and AD in a South Brazilian population (da Rocha et al., 2018). Collectively, these findings demonstrate that decreased GSH content and reduced Gpx4 levels lead to redox imbalance, targeting this redox homeostasis pathway could help alleviate ferroptosis-related damage in AD.

Lipid Metabolism Pathway

Science ferroptosis is a lipid peroxidation-driven cell death form, the abundance and localization of PUFAs, which determine the degree of lipid peroxidation, also was crucial for the execution of ferroptosis in AD (Stockwell et al., 2017). Previous study revealed a co-localization of lipid peroxidation products and Aß plaques in the brain of AD patients (Butterfield and Boyd-Kimball, 2018). Butterfield reported that AB peptides led to lipid peroxidation. In turn, lipid peroxidation products increase APP processing (Butterfield, 2020). Noticeably, lipoxygenase (LOX), cyclooxygenases (COXs), and cytochrome p450 (CYPs), three well-defined classes of lipid oxidation enzymes which catalyze the deoxygenation of PUFAs, have been reported were changed in AD pathology (Shintoku et al., 2017; Wu et al., 2018). Di Meco et al. (2017) reported that the enzymatic activity of 12/15-lipoxygenase (12/15-LOX) was significantly increased in the brains of AD patients, and its protein levels influence the memory and learning abilities in mouse model. Importantly, pharmacological inhibition of 12/15-LOX can reverse this AD-like phenotype (Di Meco et al., 2017). Therefore, the significant dysregulation of lipid peroxidation and perturbed lipid metabolism suggested an early involvement of this pathway in AD pathophysiology.

Sphingosine kinase1 (Sphk1), which could acetylate cyclooxygenase 2 (COX2), also were downregulated in the brains of AD patients and AD mice. Restoration of the expression of Sphk1 could promote the acetylation of COX2 and improve the AD-like pathology in APP/PS1 mice (Lee et al., 2018). Sphk2, another neuronal sphingosine kinase, also have been reported to protect against hippocampal volume loss and demyelination in male J20 mice (Lei et al., 2019). Additionally, a fatty acid synthase inhibitor, CMS121, protects against ferroptosis-induced lipid peroxidation and alleviates cognitive impairment in APP/PS1-transgenic mice (Ates et al., 2020). Taken together, these findings demonstrated that restoration of lipid peroxidation effectively rescued the AD-like symptoms and lipid metabolism pathway will be a potential therapeutic target for the prevention of this disease in the future.

Other Pathways

Besides these directly involved signal pathways, a series of molecules that mediating the cell protection and neuronal

TABLE 1 Summany of forrontosic inhibitors that have been applicated in AD

death may also regulate the occurrence of ferroptosis in AD pathogenesis. Emerging evidences implicated that p53, a critical regulator of cell cycle and cell death, directly participate in the induction of ferroptosis (Jiang et al., 2015; Gnanapradeepan et al., 2018; Zhang W. et al., 2018). Jiang et al. (2015) demonstrated that p53 repressed expression of SLC7A11, the key component of the cystine/glutamate antiporter, thus inhibiting cystine uptake and sensitizing tumor cells to ferroptosis. Interestingly, p53 were also reported to play a pivotal function in oxidative stress-triggered neuronal death in the progression of AD (Cenini et al., 2008). However, whether p53 could directly regulate the ferroptosis in AD still need to be further explored. Similar to these reports, the activation of mitogen-activated protein kinase (MAPK) signaling pathway also were observed in both ferroptosis induction (Dixon et al., 2012) and AD pathological process (Kheiri et al., 2018), probably indicating the involvement of this signaling pathway in the ferroptosis in AD.

A key transcriptional regulator, nuclear factor erythroid2related factor 2 (Nrf2), which containing the antioxidant response elements (ARE) and targeting the antioxidant effectors (etc., HO-1 and GSH), also was involved in the ferroptosis in AD (Kerr et al., 2017). Kerr et al. (2017) reported that AB inhibit Nrf2 activity in Drosophila, whereas specific inhibition of Keap1, the negative regulator of Nrf2, can rescue Nrf2 activity and ameliorate AB-induced neuronal toxicity. Many other studies also demonstrated that genetic ablation of Nrf2 in APP/PS1 mice promotes AD-like pathology (Joshi et al., 2015; Ren et al., 2020). Importantly, Nrf2 was thought to participate in the process of ferroptosis on account of its function in glutathione, iron, and lipids metabolism (Dodson et al., 2019). Although the underlying mechanism was still elusive, abundant evidence suggested the protective roles of Nrf2 in ferroptosis and neurodegeneration (Abdalkader et al., 2018; Bahn and Jo, 2019). Considering the critical role of Nrf2 in ferroptosis-related damage in AD, some Nrf2 inducers could also be efficaciously therapeutic agents for this disease.

Reagent	Target	Study models	Conclusions	References
Deferoxamine (DFO)	Iron	Human cancer cells	DFO inhibited erastin-induced ROS accumulation and cell death	Dixon et al., 2012
Desferrioxamine (DFE)	Iron	Clinical patients	DFE slowed the clinical progression of the dementia associated with AD	Mclachlan et al., 1991
Clioquinol	Iron	Mouse model	Clioquinol prevented Tau deficiency impaired iron export	Lei et al., 2012
α-Lipoic acid (LA)	Iron	Mouse model	Effectively rescued tauopathy and cognitive impairment in P301S mice	Zhang Y. et al., 2018
Ferrostatin-1(Fer-1) Liproxstatin-1(Lip-1)	Lipid peroxidation	Primary neurons Mouse model	Fer-1/Lip-1 effectively ameliorate $A\beta$ induced neuronal death and memory loss	Bao et al., 2021
Vitamin E	Lipid peroxidation	Clinical patients	Vitamin E slowed cognitive decline in patients with mild to moderate AD	Dysken et al., 2014
N-acetylcysteine (NAC)	Lipid peroxidation	Mouse model	Attenuated spatial memory deficits and synaptic plasticity loss in AD mice	More et al., 2018
Coenzyme Q10 (CoQ10)	Lipid peroxidation	Mouse model	Improved AD-Type behavioral and pathological symptoms in APP/PS1 mice	Muthukumaran et al., 2018
Selenium (Se)	Gpx4	Mouse model	Improved cognitive impairment and AD -related pathological symptoms in mice	Zhang Z. et al., 2018

APPLICATION OF FERROPTOSIS INHIBITORS IN AD

Ferroptosis Inhibitors in AD Clinical Trials

Emerging evidences suggested that ferroptosis is a potential therapeutic target for AD. Some ferroptosis inhibitors have already showed possible clinical benefits in clinical trials. A two year, single-blind study which administration of iron chelator DFE or oral placebo (lecithin) in 48 patients with probable AD. The researchers found that DFE treatment led to significant reduction in the rate of deterioration in AD, and the mean rate of decline was twice as rapid for the no-treatment group. These results revealed the potential role of ferroptosis inhibitor DFE in slowing the clinical progression of the dementia associated with AD (Mclachlan et al., 1991). Moreover, in a large randomized clinical trial, another low potency anti-ferroptotic agent, vitamin E, also exhibited clinical benefits in patients with mild-moderate AD. The benefits of vitamin E in mild to moderate AD by slowing functional decline and decreasing caregiver burden (Dysken, 2014; Dysken et al., 2014). Collectively, these findings suggested the potential clinical benefits of ferroptosis inhibitors in mild to moderate AD and application of ferroptosis inhibitors may slow the progression of this disease.

Ferroptosis Inhibitors in AD Cell and Animal Models

Ferroptosis inhibitors also showed the protective roles in AD cell and animal models. A recent study found that Fer-1 and liproxstatins-1(Lip-1), two ferroptosis inhibitors, could effectively ameliorate the neuronal death and memory loss induced by AB aggregation in vitro and in vivo (Bao et al., 2021). A naturally occurring iron chelator, α-Lipoic acid (LA), effectively rescued tauopathy and cognitive impairment in P301S Tau transgenic mice. Interestingly, Tau-induced iron overload and lipid peroxidation, which are involved in ferroptosis, were significantly blocked by LA administration. They also found that LA improve these abnormalities by reducing ROS content and increasing the expression level of GPX4 (Zhang Y. et al., 2018). Futuremore, selenium (Se), the key regulator of GPX4 activity, has been considered to be related to AD pathologies (Chmatalova et al., 2017; Vinceti et al., 2017). Supplementation with organic form of Se, Se-enriched yeast (Se-yeast) or selenomethionine (Se-Met), could improve cognitive impairment and AD-related pathological symptoms in animal model of AD (Zhang Z. et al., 2018). Similarly, More et al. (2018) reported that N-acetylcysteine (NAC), an antioxidant and a glutathione precursor, attenuate the spatial memory deficits and synaptic plasticity loss by decreasing lipid oxidation and increasing GSH content in animal model of

REFERENCES

Abdalkader, M., Lampinen, R., Kanninen, K. M., Malm, T. M., and Liddell, J. R. (2018). Targeting Nrf2 to suppress ferroptosis and mitochondrial dysfunction in neurodegeneration. *Front. Neurosci.* 12:466. doi: 10.3389/fnins.2018.00466 AD (da Costa et al., 2017; More et al., 2018). Another antioxidant, a nanomicellar water-soluble formulation of coenzyme Q10 (Ubisol-Q10), drastically inhibited A β plaque formation and improved long term memory (Muthukumaran et al., 2018). Taken together, these results indicate that ferroptosis may be an important neurodegenerative mechanism in AD and ferroptosis inhibitor could provide promising approach for treatment and prevention of this disease.

CONCLUSION AND PROSPECTS

Alzheimer's disease is a progressive brain degenerative disease which is the most common form of dementia among older people. In this review, we provided comprehensive summarization about the potential involvement of ferroptosis in AD and discussed the underlying signaling pathways that were probably involved in the ferroptotic death of neuronal cells during the pathogenesis of this disease.

Furthermore, we also summarized the ferroptosis inhibitors that have been applicated in the treatment and prevention of this disease (**Table 1**). The discovery of ferroptosis and the potential regulatory signaling pathways in this article, provide not only novel insights into the neuronal death in AD, but also trackable targets for this disease. Finally, ferroptosis also has been shown to implicate in the pathological cell death of several other neurologic diseases, such as Parkinson's disease (PD), Huntington's diseases (HD), amyotrophic lateral sclerosis (ALS), ischemic and hemorrhagic stroke, traumatic brain injury (TBI), and ischemia-reperfusion injury (Guiney et al., 2017; Stockwell et al., 2017; Weiland et al., 2019; Shen et al., 2020; Yan et al., 2021). Further research needs to be done to test the clinical benefits of ferroptosis inhibitors in these neurologic diseases.

AUTHOR CONTRIBUTIONS

L-QZ and WB conceived and designed the work. KC and XJ wrote the manuscript and revised it. KC, XC, and MW designed the table and figures. All authors contributed to the article and approved the submitted version.

FUNDING

This work was supported partially by the National Natural Science Foundation of China (82030032, 32070960, 81871108, 81829002, and 81771150), National Program for Support of Top - Notch Young Professionals, and Academic Frontier Youth Team of Huazhong University of Science and Technology to L-QZ.

Allen, M., Zou, F. G., Chai, H. S., Younkin, C. S., Miles, R., Nair, A. A., et al. (2012). Glutathione S-transferase omega genes in Alzheimer and Parkinson disease risk, age-at-diagnosis and brain gene expression: an association study with mechanistic implications. *Mol. Neurodegener.* 7:13. doi: 10.1186/1750-1326-7-13

- Antharam, V., Collingwood, J. F., Bullivant, J. P., Davidson, M. R., Chandra, S., Mikhaylova, A., et al. (2012). High field magnetic resonance microscopy of the human hippocampus in Alzheimer's disease: quantitative imaging and correlation with iron. *Neuroimage* 59, 1249–1260. doi: 10.1016/j.neuroimage. 2011.08.019
- Aquilano, K., Baldelli, S., and Ciriolo, M. R. (2014). Glutathione: new roles in redox signaling for an old antioxidant. *Front. Pharmacol.* 5:196. doi: 10.3389/fphar. 2014.00196
- Ates, G., Goldberg, J., Currais, A., and Maher, P. (2020). CMS121, a fatty acid synthase inhibitor, protects against excess lipid peroxidation and inflammation and alleviates cognitive loss in a transgenic mouse model of Alzheimer's disease. *Redox Biol.* 36:101648. doi: 10.1016/j.redox.2020.101648
- Ayton, S., Wang, Y., Diouf, I., Schneider, J. A., Brockman, J., Morris, M. C., et al. (2019). Brain iron is associated with accelerated cognitive decline in people with Alzheimer pathology. *Mol. Psychiatry* 25, 2932–2941. doi: 10.1038/s41380-019-0375-7
- Bahn, G., and Jo, D. G. (2019). Therapeutic approaches to Alzheimer's disease through modulation of NRF2. *Neuromolecular Med.* 21, 1–11. doi: 10.1007/ s12017-018-08523-5
- Bao, W., Pang, P., Zhou, X., Hu, F., Xiong, W., Chen, K., et al. (2021). Loss of ferroportin induces memory impairment by promoting ferroptosis in Alzheimer's disease. *Cell Death Differ*. 28, 1548–1562. doi: 10.1038/s41418-020-00685-9
- Bao, W., Zhou, X., Zhou, L., Wang, F., Yin, X., Lu, Y., et al. (2020). Targeting miR-124/Ferroportin signaling ameliorated neuronal cell death through inhibiting apoptosis and ferroptosis in aged intracerebral hemorrhage murine model. *Aging Cell* 19:e13235.
- Benseny-Cases, N., Klementieva, O., Cotte, M., Ferrer, I., and Cladera, J. (2014). Microspectroscopy (muFTIR) reveals co-localization of lipid oxidation and amyloid plaques in human Alzheimer disease brains. *Anal. Chem.* 86, 12047– 12054. doi: 10.1021/ac502667b
- Bermejo, P., Martín-Aragón, S., Benedí, J., Susín, C., Felici, E., Gil, P., et al. (2008). Peripheral levels of glutathione and protein oxidation as markers in the development of Alzheimer's disease from mild cognitive impairment. *Free Radic. Res.* 42, 162–170. doi: 10.1080/10715760701861373
- Boyd-Kimball, D., Sultana, R., Abdul, H., and Butterfield, D. (2005). Gammaglutamylcysteine ethyl ester-induced up-regulation of glutathione protects neurons against Abeta(1-42)-mediated oxidative stress and neurotoxicity: implications for Alzheimer's disease. J. Neurosci. Res. 79, 700–706. doi: 10.1002/ jnr.20394
- Butterfield, D. (2020). Brain lipid peroxidation and alzheimer disease: synergy between the butterfield and mattson laboratories. *Ageing Res. Rev.* 64:101049. doi: 10.1016/j.arr.2020.101049
- Butterfield, D., and Boyd-Kimball, D. (2018). Oxidative stress, amyloid-β peptide, and altered key molecular pathways in the pathogenesis and progression of Alzheimer's disease. *J. Alzheimers Dis.* 62, 1345–1367. doi: 10.3233/jad-17 0543
- Cacciatore, I., Baldassarre, L., Fornasari, E., Mollica, A., and Pinnen, F. (2012). Recent advances in the treatment of neurodegenerative diseases based on GSH delivery systems. Oxid. Med. Cell. Longev. 2012:240146.
- Cardoso, B. R., Hare, D. J., Bush, A. I., and Roberts, B. R. (2017). Glutathione peroxidase 4: a new player in neurodegeneration? *Mol. Psychiatry* 22, 328–335. doi: 10.1038/mp.2016.196
- Casley, C. S., Land, J. M., Sharpe, M. A., Clark, J. B., Duchen, M. R., and Canevari, L. (2002). beta-amyloid fragment 25-35 causes mitochondrial dysfunction in primary cortical neurons. *Neurobiol. Dis.* 10, 258–267. doi: 10.1006/nbdi.2002. 0516
- Cenini, G., Sultana, R., Memo, M., and Butterfield, D. A. (2008). Elevated levels of pro-apoptotic p53 and its oxidative modification by the lipid peroxidation product, HNE, in brain from subjects with amnestic mild cognitive impairment and Alzheimer's disease. *J. Cell Mol. Med.* 12, 987–994. doi: 10.1111/j.1582-4934.2008.00163.x
- Chen, X., Comish, P., Tang, D., and Kang, R. (2021). Characteristics and biomarkers of ferroptosis. *Front. Cell Dev. Biol.* 9:637162. doi: 10.3389/fcell. 2021.637162
- Chiang, G. C., Mao, X., Kang, G., Chang, E., Pandya, S., Vallabhajosula, S., et al. (2017). Relationships among cortical glutathione levels, brain amyloidosis, and memory in healthy older adults investigated in vivo with H-1-MRS and

pittsburgh compound-B PET. Am. J. Neuroradiol. 38, 1130–1137. doi: 10.3174/ ajnr.a5143

- Chmatalova, Z., Vyhnalek, M., Laczo, J., Hort, J., Pospisilova, R., Pechova, M., et al. (2017). Relation of plasma selenium and lipid peroxidation end products in patients with Alzheimer's disease. *Physiol. Res.* 66, 1049–1056. doi: 10.33549/ physiolres.933601
- da Costa, M., Bernardi, J., Costa, L., Fiuza, T., Brandão, R., Ribeiro, M., et al. (2017). N-acetylcysteine treatment attenuates the cognitive impairment and synaptic plasticity loss induced by streptozotocin. *Chem. Biol. Interact.* 272, 37–46. doi: 10.1016/j.cbi.2017.05.008
- da Rocha, T. J., Alves, M. S., Guisso, C. C., De Andrade, F. M., Camozzato, A., De Oliveira, A. A., et al. (2018). Association of GPX1 and GPX4 polymorphisms with episodic memory and Alzheimer's disease. *Neurosci. Lett.* 666, 32–37. doi: 10.1016/j.neulet.2017.12.026
- de Jong, L. W., Van Der Hiele, K., Veer, I. M., Houwing, J. J., Westendorp, R. G., Bollen, E. L., et al. (2008). Strongly reduced volumes of putamen and thalamus in Alzheimer's disease: an MRI study. *Brain* 131, 3277–3285. doi: 10.1093/ brain/awn278
- Di Domenico, F., Tramutola, A., and Butterfield, D. A. (2017). Role of 4-hydroxy-2-nonenal (HNE) in the pathogenesis of alzheimer disease and other selected age-related neurodegenerative disorders. *Free Radic. Biol. Med.* 111, 253–261. doi: 10.1016/j.freeradbiomed.2016.10.490
- Di Meco, A., Li, J. G., Blass, B. E., Abou-Gharbia, M., Lauretti, E., and Pratico, D. (2017). 12/15-lipoxygenase inhibition reverses cognitive impairment, brain amyloidosis, and tau pathology by stimulating autophagy in aged triple transgenic mice. *Biol. Psychiatry* 81, 92–100. doi: 10.1016/j.biopsych.2016.05. 023
- Dixon, S. (2017). Ferroptosis: bug or feature? *Immunol. Rev.* 277, 150–157. doi: 10.1111/imr.12533
- Dixon, S. J., Lemberg, K. M., Lamprecht, M. R., Skouta, R., Zaitsev, E. M., Gleason, C. E., et al. (2012). Ferroptosis: an iron-dependent form of nonapoptotic cell death. *Cell* 149, 1060–1072. doi: 10.1016/j.cell.2012.03.042
- Dodson, M., Castro-Portuguez, R., and Zhang, D. D. (2019). NRF2 plays a critical role in mitigating lipid peroxidation and ferroptosis. *Redox Biol.* 23:101107. doi: 10.1016/j.redox.2019.101107
- Duce, J. A., Tsatsanis, A., Cater, M. A., James, S. A., Robb, E., Wikhe, K., et al. (2010). Iron-export ferroxidase activity of beta-amyloid precursor protein is inhibited by zinc in Alzheimer's disease. *Cell* 142, 857–867. doi: 10.1016/j.cell.2010.08.014
- Dysken, M. W. (2014). Effect of vitamin E and memantine on functional decline in Alzheimer disease: the TEAM-AD VA cooperative randomized trial. *JAMA* 311, 1161–1161.
- Dysken, M. W., Guarino, P. D., Vertrees, J. E., Asthana, S., Sano, M., Llorente, M., et al. (2014). Vitamin E and memantine in Alzheimer's disease: clinical trial methods and baseline data. *Alzheimers Dement.* 10, 36–44.
- Fang, X. X., Wang, H., Han, D., Xie, E. J., Yang, X., Wei, J. Y., et al. (2019). Ferroptosis as a target for protection against cardiomyopathy. *Proc. Natl. Acad. Sci. U. S. A.* 116, 2672–2680. doi: 10.1073/pnas.1821022116
- Friedmann Angeli, J. P., Schneider, M., Proneth, B., Tyurina, Y. Y., Tyurin, V. A., Hammond, V. J., et al. (2014). Inactivation of the ferroptosis regulator Gpx4 triggers acute renal failure in mice. *Nat. Cell Biol.* 16, 1180–1191. doi: 10.1038/ ncb3064
- Ghosh, D., Levault, K. R., Barnett, A. J., and Brewer, G. J. (2012). A reversible early oxidized redox state that precedes macromolecular ROS damage in aging nontransgenic and 3xTg-AD mouse neurons. J. Neurosci. 32, 5821–5832. doi: 10.1523/jneurosci.6192-11.2012
- Gnanapradeepan, K., Basu, S., Barnoud, T., Budina-Kolomets, A., Kung, C. P., and Murphy, M. E. (2018). The p53 tumor suppressor in the control of metabolism and ferroptosis. *Front. Endocrinol. (Lausanne)* 9:124. doi: 10.3389/fendo.2018. 00124
- Good, P. F., Perl, D. P., Bierer, L. M., and Schmeidler, J. (1992). Selective accumulation of aluminum and iron in the neurofibrillary tangles of Alzheimer's disease: a laser microprobe (LAMMA) study. Ann. Neurol. 31, 286–292. doi: 10.1002/ana.410310310
- Guiney, S., Adlard, P., Bush, A., Finkelstein, D., and Ayton, S. (2017). Ferroptosis and cell death mechanisms in Parkinson's disease. *Neurochem. Int.* 104, 34–48. doi: 10.1016/j.neuint.2017.01.004
- Hambright, W. S., Fonseca, R. S., Chen, L., Na, R., and Ran, Q. (2017). Ablation of ferroptosis regulator glutathione peroxidase 4 in forebrain neurons promotes

cognitive impairment and neurodegeneration. *Redox Biol.* 12, 8–17. doi: 10. 1016/j.redox.2017.01.021

- Hider, R. C., and Kong, X. L. (2011). Glutathione: a key component of the cytoplasmic labile iron pool. *Biometals* 24, 1179–1187. doi: 10.1007/s10534-011-9476-8
- Horvath, R., Holinski-Feder, E., Neeve, V., Pyle, A., Griffin, H., Ashok, D., et al. (2012). A new phenotype of brain iron accumulation with dystonia, optic atrophy, and peripheral neuropathy. *Mov. Disord.* 27, 789–793. doi: 10.1002/ mds.24980
- Jiang, L., Kon, N., Li, T. Y., Wang, S. J., Su, T., Hibshoosh, H., et al. (2015). Ferroptosis as a p53-mediated activity during tumour suppression. *Nature* 520, 57–62. doi: 10.1038/nature14344
- Joshi, G., Gan, K., Johnson, D., and Johnson, J. (2015). Increased Alzheimer's disease-like pathology in the APP/ PS1∆E9 mouse model lacking Nrf2 through modulation of autophagy. *Neurobiol. Aging* 36, 664–679. doi: 10.1016/j. neurobiolaging.2014.09.004
- Kerr, F., Sofola-Adesakin, O., Ivanov, D. K., Gatliff, J., Perez-Nievas, B. G., Bertrand, H. C., et al. (2017). Direct Keap 1-Nrf2 disruption as a potential therapeutic target for Alzheimer's disease. *PLoS Genet*. 13:e1006593. doi: 10. 1371/journal.pgen.1006593
- Kheiri, G., Dolatshahi, M., Rahmani, F., and Rezaei, N. (2018). Role of p38/MAPKs in Alzheimer's disease: implications for amyloid beta toxicity targeted therapy. *Rev. Neurosci.* 30, 9–30. doi: 10.1515/revneuro-2018-0008
- Krishnan, S., and Rani, P. (2014). Evaluation of selenium, redox status and their association with plasma Amyloid/Tau in Alzheimer's disease. *Biol. Trace Elem. Res.* 158, 158–165. doi: 10.1007/s12011-014-9930-x
- Kuhn, H., Banthiya, S., and Van Leyen, K. (2015). Mammalian lipoxygenases and their biological relevance. *Biochim. Biophys. Acta* 1851, 308–330. doi: 10.1016/ j.bbalip.2014.10.002
- Langkammer, C., Ropele, S., Pirpamer, L., Fazekas, F., and Schmidt, R. (2014). MRI for iron mapping in Alzheimer's disease. *Neurodegener. Dis.* 13, 189–191.
- Lee, J. Y., Han, S. H., Park, M. H., Baek, B., Song, I. S., Choi, M. K., et al. (2018). Neuronal SphK1 acetylates COX2 and contributes to pathogenesis in a model of Alzheimer's disease. *Nat. Commun.* 9:1479.
- Lei, M., Teo, J., Song, H., Mcewen, H., Yup Lee, J., Couttas, T., et al. (2019). Sphingosine Kinase 2 potentiates amyloid deposition but protects against hippocampal volume loss and demyelination in a mouse model of Alzheimer's disease. J. Neurosci. 39, 9645–9659. doi: 10.1523/jneurosci.0524-19.2019
- Lei, P., Ayton, S., Finkelstein, D. I., Spoerri, L., Ciccotosto, G. D., Wright, D. K., et al. (2012). Tau deficiency induces parkinsonism with dementia by impairing APP-mediated iron export. *Nat. Med.* 18, 291–295. doi: 10.1038/nm.2613
- Lovell, M. A., Xie, C., and Markesbery, W. R. (1998). Decreased glutathione transferase activity in brain and ventricular fluid in Alzheimer's disease. *Neurology* 51, 1562–1566. doi: 10.1212/wnl.51.6.1562
- Ma, J., Qian, C., Bao, Y., Liu, M., Ma, H., Shen, M., et al. (2021). Apolipoprotein E deficiency induces a progressive increase in tissue iron contents with age in mice. *Redox Biol.* 40:101865. doi: 10.1016/j.redox.2021.101865
- Mclachlan, D. R. C., Dalton, A. J., Kruck, T. P. A., Bell, M. Y., Smith, W. L., Kalow, W., et al. (1991). Intramuscular desferrioxamine in patients with Alzheimersdisease. *Lancet* 337, 1304–1308. doi: 10.1016/0140-6736(91)92978-b
- More, J., Galusso, N., Veloso, P., Montecinos, L., Finkelstein, J., Sanchez, G., et al. (2018). N-acetylcysteine prevents the spatial memory deficits and the redoxdependent RyR2 decrease displayed by an Alzheimer's disease rat model. *Front. Aging Neurosci.* 10:399. doi: 10.3389/fnagi.2018.00399
- Muthukumaran, K., Kanwar, A., Vegh, C., Marginean, A., Elliott, A., Guilbeault, N., et al. (2018). Ubisol-Q10 (a nanomicellar water-soluble formulation of CoQ10) treatment inhibits alzheimer-type behavioral and pathological symptoms in a double transgenic mouse (TgAPEswe, PSEN1dE9) model of Alzheimer's disease. J. Alzheimers Dis. 61, 221–236. doi: 10.3233/jad-170275
- Pena-Bautista, C., Baquero, M., Vento, M., and Chafer-Pericas, C. (2019). Free radicals in Alzheimer's disease: lipid peroxidation biomarkers. *Clin. Chim. Acta* 491, 85–90. doi: 10.1016/j.cca.2019.01.021
- Peskind, E. R., Li, G., Shofer, J. B., Millard, S. P., Leverenz, J. B., Yu, C. E., et al. (2014). Influence of lifestyle modifications on age-related free radical injury to brain. *JAMA Neurol.* 71, 1150–1154. doi: 10.1001/jamaneurol.2014.1428
- Pocernich, C. B., and Butterfield, D. A. (2012). Elevation of glutathione as a therapeutic strategy in Alzheimer disease. *Biochim. Biophys. Acta* 1822, 625– 630. doi: 10.1016/j.bbadis.2011.10.003

- Prasad, M. R., Lovell, M. A., Yatin, M., Dhillon, H., and Markesbery, W. R. (1998). Regional membrane phospholipid alterations in Alzheimer's disease. *Neurochem. Res.* 23, 81–88.
- Raha, A. A., Vaishnav, R. A., Friedland, R. P., Bomford, A., and Raha-Chowdhury, R. (2013). The systemic iron-regulatory proteins hepcidin and ferroportin are reduced in the brain in Alzheimer's disease. *Acta Neuropathol. Commun.* 1:55. doi: 10.1186/2051-5960-1-55
- Raven, E., Lu, P., Tishler, T., Heydari, P., and Bartzokis, G. (2013). Increased iron levels and decreased tissue integrity in hippocampus of Alzheimer's disease detected in vivo with magnetic resonance imaging. J. Alzheimers Dis. 37, 127–136. doi: 10.3233/jad-130209
- Ren, P., Chen, J., Li, B., Zhang, M., Yang, B., Guo, X., et al. (2020). Nrf2 ablation promotes Alzheimer's disease-like pathology in APP/PS1 transgenic mice: the role of neuroinflammation and oxidative stress. Oxid. Med. Cell. Longev. 2020:3050971.
- Resende, R., Moreira, P. I., Proenca, T., Deshpande, A., Busciglio, J., Pereira, C., et al. (2008). Brain oxidative stress in a triple-transgenic mouse model of Alzheimer disease. *Free Radic. Biol. Med.* 44, 2051–2057. doi: 10.1016/j. freeradbiomed.2008.03.012
- Sands, S. A., Leung-Toung, R., Wang, Y., Connelly, J., and Levine, S. M. (2016). Enhanced histochemical detection of iron in paraffin sections of mouse central nervous system tissue: application in the APP/PS1 mouse model of Alzheimer's disease. ASN Neuro 8:1759091416670978.
- Schenck, J. F., and Zimmerman, E. A. (2004). High-field magnetic resonance imaging of brain iron: birth of a biomarker? NMR Biomed. 17, 433–445.
- Schenck, J. F., Zimmerman, E. A., Li, Z., Adak, S., Saha, A., Tandon, R., et al. (2006). High-field magnetic resonance imaging of brain iron in Alzheimer disease. *Top. Magn. Reson. Imaging* 17, 41–50. doi: 10.1097/01.rmr.0000245455.59912.40
- Shen, L., Lin, D., Li, X., Wu, H., Lenahan, C., Pan, Y., et al. (2020). Ferroptosis in acute central nervous system injuries: the future direction? *Front. Cell Dev. Biol.* 8:594. doi: 10.3389/fcell.2020.00594
- Shintoku, R., Takigawa, Y., Yamada, K., Kubota, C., Yoshimoto, Y., Takeuchi, T., et al. (2017). Lipoxygenase-mediated generation of lipid peroxides enhances ferroptosis induced by erastin and RSL3. *Cancer Sci.* 108, 2187–2194. doi: 10.1111/cas.13380
- Smith, M., Zhu, X., Tabaton, M., Liu, G., Mckeel, D., Cohen, M., et al. (2010). Increased iron and free radical generation in preclinical Alzheimer disease and mild cognitive impairment. J. Alzheimers Dis. 19, 363–372. doi: 10.3233/jad-2010-1239
- Sripetchwandee, J., Pipatpiboon, N., Chattipakorn, N., and Chattipakorn, S. (2014). Combined therapy of iron chelator and antioxidant completely restores brain dysfunction induced by iron toxicity. *PLoS One* 9:e85115. doi: 10.1371/journal. pone.0085115
- Stockwell, B. R., Friedmann Angeli, J. P., Bayir, H., Bush, A. I., Conrad, M., Dixon, S. J., et al. (2017). Ferroptosis: a regulated cell death nexus linking metabolism, redox biology, and disease. *Cell* 171, 273–285. doi: 10.1016/j.cell.2017.09.021
- Tao, Y., Wang, Y., Rogers, J. T., and Wang, F. (2014). Perturbed iron distribution in Alzheimer's disease serum, cerebrospinal fluid, and selected brain regions: a systematic review and meta-analysis. J. Alzheimers Dis. 42, 679–690. doi: 10.3233/jad-140396
- Ten Kate, M., Dicks, E., Visser, P. J., Van Der Flier, W. M., Teunissen, C. E., Barkhof, F., et al. (2018). Atrophy subtypes in prodromal Alzheimer's disease are associated with cognitive decline. *Brain* 141, 3443–3456. doi: 10.1093/brain/ awy264
- Tian, J., Zheng, W., Li, X. L., Cui, Y. H., and Wang, Z. Y. (2018). Lower expression of Ndfip1 is associated with Alzheimer disease pathogenesis through decreasing DMT1 degradation and increasing iron influx. *Front. Aging Neurosci.* 10:165. doi: 10.3389/fnagi.2018.00165
- Uysal, G., and Ozturk, M. (2020). Hippocampal atrophy based Alzheimer's disease diagnosis via machine learning methods. J. Neurosci. Methods 337:108669. doi: 10.1016/j.jneumeth.2020.108669
- Vinceti, M., Chiari, A., Eichmüller, M., Rothman, K., Filippini, T., Malagoli, C., et al. (2017). A selenium species in cerebrospinal fluid predicts conversion to Alzheimer's dementia in persons with mild cognitive impairment. *Alzheimers Res. Ther.* 9:100.
- Wang, G., Wu, Y., Zhou, T., Guo, Y., Zheng, B., Wang, J., et al. (2013). Mapping of the N-linked glycoproteome of human spermatozoa. J. Proteome Res. 12, 5750–5759. doi: 10.1021/pr400753f

- Wang, H., An, P., Xie, E., Wu, Q., Fang, X., Gao, H., et al. (2017). Characterization of ferroptosis in murine models of hemochromatosis. *Hepatology* 66, 449–465. doi: 10.1002/hep.29117
- Wang, X., Hu, X., Yang, Y., Takata, T., and Sakurai, T. (2016). Nicotinamide mononucleotide protects against β-amyloid oligomer-induced cognitive impairment and neuronal death. *Brain Res.* 1643, 1–9. doi: 10.1016/j.brainres. 2016.04.060
- Wang, X., Liu, D., Huang, H., Wang, Z., Hou, T., Yang, X., et al. (2018). A novel MicroRNA-124/PTPN1 signal pathway mediates synaptic and memory deficits in Alzheimer's disease. *Biol. Psychiatry* 83, 395–405. doi: 10.1016/j.biopsych. 2017.07.023
- Ward, R., Zucca, F. A., Duyn, J. H., Crichton, R. R., and Zecca, L. (2014). The role of iron in brain ageing and neurodegenerative disorders. *Lancet Neurol.* 13, 1045–1060. doi: 10.1016/s1474-4422(14)70117-6
- Weiland, A., Wang, Y., Wu, W., Lan, X., Han, X., Li, Q., et al. (2019). Ferroptosis and its role in diverse brain diseases. *Mol. Neurobiol.* 56, 4880–4893. doi: 10.1007/s12035-018-1403-3
- Williams, T. I., Lynn, B. C., Markesbery, W. R., and Lovell, M. A. (2006). Increased levels of 4-hydroxynonenal and acrolein, neurotoxic markers of lipid peroxidation, in the brain in mild cognitive impairment and early Alzheimer's disease. *Neurobiol. Aging* 27, 1094–1099. doi: 10.1016/j.neurobiolaging.2005. 06.004
- Wu, J. R., Tuo, Q. Z., and Lei, P. (2018). Ferroptosis, a recent defined form of critical cell death in neurological disorders. J. Mol. Neurosci. 66, 197–206. doi: 10.1007/s12031-018-1155-6
- Xian-hui, D., Wei-Juan, G., Tie-Mei, S., Hong-Lin, X., Jiang-Tao, B., Jing-Yi, Z., et al. (2015). Age-related changes of brain iron load changes in the frontal cortex in APPswe/PS1DeltaE9 transgenic mouse model of Alzheimer's disease. J. Trace Elem. Med. Biol. 30, 118–123. doi: 10.1016/j.jtemb.2014.11.009
- Yan, H., Zou, T., Tuo, Q., Xu, S., Li, H., Belaidi, A., et al. (2021). Ferroptosis: mechanisms and links with diseases. *Signal Transduct. Target. Ther.* 6:49.
- Yan, N., and Zhang, J. (2019). Iron metabolism, ferroptosis, and the links with Alzheimer's disease. Front. Neurosci. 13:1443. doi: 10.3389/fnins.2019.01443
- Yang, W. S., Sriramaratnam, R., Welsch, M. E., Shimada, K., Skouta, R., Viswanathan, V. S., et al. (2014). Regulation of ferroptotic cancer cell death by GPX4. *Cell* 156, 317–331. doi: 10.1016/j.cell.2013.12.010
- Yoo, M. H., Gu, X., Xu, X. M., Kim, J. Y., Carlson, B. A., Patterson, A. D., et al. (2010). Delineating the role of glutathione peroxidase 4 in protecting cells against lipid hydroperoxide damage and in Alzheimer's disease. *Antioxid. Redox Signal.* 12, 819–827. doi: 10.1089/ars.2009.2891

- Yu, W., An, S., Shao, T., Xu, H., Chen, H., Ning, J., et al. (2019). Active compounds of herbs ameliorate impaired cognition in APP/PS1 mouse model of Alzheimer's disease. *Aging* 11, 11186–11201. doi: 10.18632/aging.102 522
- Zhang, C., Rodriguez, C., Spaulding, J., Aw, T. Y., and Feng, J. (2012). Agedependent and tissue-related glutathione redox status in a mouse model of Alzheimer's disease. J. Alzheimers Dis. 28, 655–666. doi: 10.3233/jad-2011-111244
- Zhang, W., Gai, C., Ding, D., Wang, F., and Li, W. (2018). Targeted p53 on smallmolecules-induced ferroptosis in cancers. *Front. Oncol.* 8:507. doi: 10.3389/ fonc.2018.00507
- Zhang, Y., Wang, D., Xu, S., Zhang, S., Fan, Y., Yang, Y., et al. (2018). α-Lipoic acid improves abnormal behavior by mitigation of oxidative stress, inflammation, ferroptosis, and tauopathy in P301S Tau transgenic mice. *Redox Biol.* 14, 535–548. doi: 10.1016/j.redox.2017.11.001
- Zhang, Z., Wu, Q., Chen, C., Zheng, R., Chen, Y., Ni, J., et al. (2018). Comparison of the effects of selenomethionine and selenium-enriched yeast in the tripletransgenic mouse model of Alzheimer's disease. *Food Funct.* 9, 3965–3973. doi: 10.1039/c7fo02063e
- Zheng, W., Xin, N., Chi, Z. H., Zhao, B. L., Zhang, J., Li, J. Y., et al. (2009). Divalent metal transporter 1 is involved in amyloid precursor protein processing and Abeta generation. *FASEB J.* 23, 4207–4217. doi: 10.1096/fj.09-13 5749

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2021 Chen, Jiang, Wu, Cao, Bao and Zhu. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.