



Heteroleptic Coordination Environments in Metal-Mediated DNA G-Quadruplexes

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The presence of metal centers with often highly conserved coordination environments is crucial for roughly half of all proteins, having structural, regulatory, or enzymatic function. To understand and mimic the function of metallo-enzymes, bioinorganic chemists pursue the challenge of synthesizing model compounds with well-defined, often heteroleptic metal sites. Recently, we reported the design of tailored homoleptic coordination environments for various transition metal cations based on unimolecular DNA G-guadruplex structures, templating the regioselective positioning of imidazole ligandosides L¹. Here, we expand this modular system to more complex, heteroleptic coordination environments by combining L^{I} with a new benzoate ligandoside L^{B} within the same oligonucleotide. The modifications still allow the correct folding of parallel tetramolecular and antiparallel unimolecular G-quadruplexes. Interestingly, the incorporation of L^B results in strong destabilization expressed in lower thermal denaturation temperatures T_m . While no transition metal cations could be bound by G-quadruplexes containing only L^B, heteroleptic derivatives containing both L^I and L^B were found to complex Cu^{II}, Ni^{II}, and Zn^{II}. Especially in case of Cu^{II} we found strong stabilizations of up to $\Delta T_m = +34^{\circ}$ C. The here shown system represents an important step toward the design of more complex coordination environments inside DNA scaffolds, promising to culminate in the preparation of functional metallo-DNAzymes.

Keywords: bioinorganic chemistry, coordination chemistry, DNA, G-quadruplex, DNAzymes

INTRODUCTION

Proteins are involved in a vast number of processes ranging from structural and regulatory functions to enzymatic reactions. Roughly half of all proteins depend on metal cations helping to maintain a desired folding or serving as catalytic centers or redox cofactors (Raven et al., 1999; Lu et al., 2009; Rubino and Franz, 2012). Which function the respective metal ion adopts is strongly dependent on its properties, including accessible spin states, oxidation potential, Lewis-acidity, and bioavailability (Holm et al., 1996; Waldron et al., 2009). These properties are further fine-tuned by a well-defined first and second coordination sphere. The former is directly involved in metal coordination are the amino acids histidine, glutamic/aspartic acid, methionine, cysteine, or the backbone amide groups (Holm et al., 1996; Degtyarenko, 2000; Shook and Borovik, 2010; Valdez et al., 2014). In contrast, the second coordination sphere is not directly involved in metal binding but regulates catalytic processes, proton or electron shuttling, substrate transport, and

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effects selectivity (Colquhoun et al., 1986; Degtyarenko, 2000; Waldron et al., 2009; Shook and Borovik, 2010; Zhao et al., 2013; Valdez et al., 2014; Cornish et al., 2016).

The design of artificial metallo-enzyme mimics with improved or novel properties is attracting increasing interest, but remains challenging. In the area of preparative bioinorganic chemistry, focus is set on small, multidentate chelate complexes, often requiring tedious multistep syntheses and only covering effects of the first coordination sphere (Samuel et al., 2010; Kanady et al., 2011; Anderson et al., 2013; Dicke et al., 2018). More biologically oriented approaches involve the replacement of natural metal cofactors with metal centers not known in nature. An example is the replacement of hemin in myoglobin with an iridium or rhodium porphyrin complex for enantioselective cyclopropanation reactions (Key et al., 2016; Litman et al., 2018). Another approach is embedding metal cofactors by covalent or non-covalent interactions into empty cavities of usually metalfree proteins. This was successfully applied in a series of examples enabling catalysis of the asymmetric transfer hydrogenation of imines (Wu et al., 2019), ring-closing metathesis (Jeschek et al., 2016), oxime (Drienovská et al., 2018), and hydrazine (Drienovská et al., 2018; Mayer et al., 2019) formation and hydration of alkenes (Drienovská et al., 2017). In contrast to the aforementioned examples, a more bottom up approach is the de novo design of new metallo-proteins by the precise arrangement of certain structural motifs to create a metal binding site (Raven et al., 1999; Lu et al., 2009; Rubino and Franz, 2012). In recent years, a more efficient alternative was developed based on small artificial peptoid structures. Due to their simple accessibility by solid phase synthesis and their capability to form well-ordered secondary structures, many examples were shown for selective metal binding and catalytic applications (Baskin and Maayan, 2016; Knight et al., 2017; Baskin et al., 2018; Ghosh et al., 2018).

Another type of biopolymers forming well-ordered secondary structures are oligonucleotides. In contrast to peptides, RNA and DNA only consist of four nucleotide building blocks, thus reducing the possibilities to create diverse coordination environments for a range of metal cations. To overcome this limitation, different strategies were developed to covalently or non-covalently anchor metal-chelating ligands inside DNA. Roelfes and co-workers pioneered the design of various oligonucleotides capable of Michael-Additions, Carbene transfer, syn-hydrations of alkenes or Diels-Alder reactions (Roelfes and Feringa, 2005; Coquière et al., 2007; Boersma et al., 2010a,b; Rioz-Martínez et al., 2016). Other groups used modified quadruplexes for sequence-specific DNA cleavage, light controlled thrombin catalysis or peroxidase mimicking DNAzymes (Xu et al., 2009; Ali et al., 2019; Wang et al., 2019). A difficulty of this approach lies in the largely unknown exact position and coordination environment of the catalytic centers. This difficulty could be overcome in the field of metal-mediated base pairing, where the hydrogen bonding interaction of canonical base pairs is replaced by metal coordination, leading to highly stabilized DNA structures (Mandal and Müller, 2017). While first examples included only the involvement of canonical bases (Katz, 1963), the field was later expanded by the incorporation of a variety of artificial nucleobases culminating in the development of

programmable metal wires inside DNA duplexes (Tanaka et al., 2006; Clever et al., 2007; Mandal et al., 2016; Sandmann et al., 2019). Later, the concept was expanded from duplex to triplex DNA (Tanaka et al., 2002) and i-motifs (Abdelhamid et al., 2018), while we and others started to focus on G-quadruplexes (Miyoshi et al., 2007; Smith et al., 2012; Engelhard et al., 2013). The latter ones form from guanine-rich sequences where four Gresidues cyclize to planar G-tetrads via Hoogsteen base pairing. Multiple G-tetrads form a G-quadruplex via π - π stacking interactions. Key to their high stability is the incorporation of a central cation-typically Na⁺ or K⁺ (Hänsel-Hertsch et al., 2017; Neidle, 2017). Our group was the first to report Cu^{II}mediated tetramolecular G-quadruplexes based on pyridine and imidazole ligands (Engelhard et al., 2013, 2018b; Punt and Clever, 2019a), aimed at a range of applications. For example, dinuclear systems were employed as Cu^{II}-based EPR-rulers for accurate distance measurements (Engelhard et al., 2018a). We later expanded this concept to unimolecular G-quadruplexes, equipped with oligonucleotide loops which form cavities above the G-quadruplex stem in which the metal complexes are embedded (Engelhard et al., 2017). In a recent study, we further showed that these G-quadruplexes can act as robust templates to arrange different numbers of imidazole ligandosides, leading to fine-tuned affinities for a range of transition metal cations with respect to their preferred coordination environments (Punt and Clever, 2019b). While only homoleptic systems were investigated in that study, we herein expand the modular concept to heteroleptic systems with different donor functionalities. We introduce the design of mixed systems with imidazole and benzoate ligands, inspired by metallo-proteins, where the combination of imidazoles and carboxylate is often involved in metal coordination (e.g., in the 2-His-1-carboxylate facial triad) (Greenblatt et al., 1998; Koehntop et al., 2005). We show how this combination affects both, G-quadruplex stability and metal complexation.

RESULTS

In this study we report the incorporation of a new benzoate ligandoside L^B in combination with the known imidazole ligandoside L^{I} . Both were incorporated in the (S) configuration as GNA (glycol nucleic acid) building blocks (Zhang et al., 2005, 2006) by solid phase synthesis into tetramolecular and unimolecular G-quadruplexes. The phosphoramidite of LI was synthesized as previously described (Punt and Clever, 2019b). For the new benzoate ligand L^B, a literature procedure was adopted (Engelhard et al., 2017). Accordingly, an initial nucleophilic attack of deprotonated solketal to methyl 4-(bromomethyl)benzoate followed by acidic deprotection reaction led to protected benzoate ligandoside (R)-4. Its structure and absolute configuration were confirmed by singlecrystal X-ray diffraction (Figure 1). The primary hydroxyl group was DMT-protected (DMT = dimethoxytrityl) followed by a phosphorylation reaction yielding phosphoramidite building block (S)-6. DNA solid phase synthesis was then performed according to standard literature procedures with



FIGURE 1 Synthesis of benzoate ligandoside L^B and molecular structure of ligandoside L^1 (top). (1) NaH, CH₃CN; (2) CH₃COOH, THF/H₂O; (3) DMT-CI, DIPEA, DMAP, THF; (4) CEDIP-CI, DIPEA, CH₂Cl₂; (5) automated solid-phase DNA synthesis. The single-crystal X-ray structure of the protected ligandoside (*R*)-4 is shown. G-quadruplex formation (bottom) of htel $L^B_2 L^1_2$ creates a heteroleptic coordination environment for transition metal ions (M = Co, Ni, Cu, Zn). Gray tiles: guanosine; red: ligandoside L^B ; blue: ligandoside L^1 ; green circles: adenosine; yellow circles: thymidine. DMT = dimethoxytrityl; DIPEA = N,N-diisopropylethylamine; DMAP = N,N-dimethylaminopyridin; CEDIP-CI = 2-cyanoethyl N,N-diisopropylchlorophosphoramidite.



extended coupling times for the ligandosides L^{I} and L^{B} (see **Supplementary Material** for details). Coupling efficiencies for L^{B} and L^{I} were typically >99% per step. After solid phase synthesis, oligonucleotides were cleaved from the solid support and deprotected in 0.4 M NaOH in methanol/water (4:1) at 55°C for 16 h. Standard deprotection with concentrated ammonium hydroxide was avoided due to the risk of forming amides instead of carboxylates from the benzoate esters. After reversed-phase HPLC purification, oligonucleotides were desalted and DMT-groups removed using C18 *SepPak* cartridges and aq. TFA (2%). The oligonucleotides were then lyophilized at stored at -20° C.

Since L^{I} had already been established in tetramolecular and unimolecular G-quadruplexes, we first investigated the

influence of \mathbf{L}^{B} in the tetramolecular G-quadruplex $(\mathbf{L}^{B}\mathbf{G}_{4})_{4}$. Clear formation of a parallel G-quadruplex was observed by CD spectroscopy with a positive Cotton effect around ~260 nm (see **Figure S25**). Thermal denaturation experiments showed a melting temperature T_{m} of 27°C which was significantly lower compared to previously reported $(\mathbf{L}^{I}\mathbf{G}_{4})_{4}$ ($T_{m} = 36^{\circ}$ C; Punt and Clever, 2019b). Since \mathbf{L}^{B} and \mathbf{L}^{I} are sharing the same backbone modification, we ascribe this destabilization to a repulsive effect between the negatively charged benzoates and phosphates (**Figure 2**). Next, the interaction of $(\mathbf{L}^{B}\mathbf{G}_{4})_{4}$ with a series of transition metal cations was investigated. In contrast to $(\mathbf{L}^{I}\mathbf{G}_{4})_{4}$ which was shown to complex \mathbf{Cu}^{II} , \mathbf{Ni}^{II} , \mathbf{Co}^{II} , and \mathbf{Zn}^{II} , no signs for metal complexation in $(\mathbf{L}^{B}\mathbf{G}_{4})_{4}$ were observed (see **Figures S3, S4**). This may be explained by the harder character



of the benzoate ligand, competing with hard ligands such as the contained chloride, cacodylate buffer or phosphate backbones. However, even for hard and oxophilic transition metal cations, including Gd^{III} and Ce^{III}, no interactions were found.

Mixing ligands in tetramolecular G-quadruplexes leads to statistical mixtures, which makes it challenging to design distinct heteroleptic coordination environments (see Supplementary Material for details). On the other hand, the folding of unimolecular G-quadruplexes into discrete topologies enables programmable ligand arrangements. Consequently, we moved forward to incorporate L^B in unimolecular Gquadruplexes. At first, L^B was incorporated four times in htel L^B_4 . Similar to $(L^BG_4)_4$, incorporation of L^B caused strong destabilization ($T_m = 12^{\circ}$ C) compared to htelL^I₄ ($T_m =$ 33° C). Successive replacement of L^B with L^I was accompanied with a linear increase in stabilization for each replacement (htelL^B₃L^I $T_m = 17^{\circ}$ C, htelL^B₂L^I₂ $T_m = 23^{\circ}$ C, htelL^BL^I₃ $T_m = 28^{\circ}$ C), highlighting the additive destabilizing effect of L^B (Figure 3). CD spectroscopy of $htelL^{B}_{4}$, $htelL^{B}_{3}L^{I}$, $htelL^{B}_{2}L^{I}_{2}$, and htelL^BL^I₃ showed clear signatures corresponding to an antiparallel G-quadruplex topology with a positive Cotton effect around ~294 nm in all cases (see Figures S26, S27). This is consistent with the previous observations for homoleptic G-quadruplexes containing only L^I. Next, the interaction with different transition metal cations was investigated. As for $(L^BG_4)_4$, for $htelL^B_4$, $htelL^B_3L^I$, and $htelL^B_2L^I_2$, thermal denaturation experiments showed no signs for interaction with the examined transition metal cations (Cu^{II}, Ni^{II}, Zn^{II}, Co^{II},

V^{IV}O). Pleasingly, this changed for htelL^BL^I₃ that showed a weak but distinct stabilization after addition of 1 equiv. of Cu^{II} ($\Delta T_m = +4^{\circ}$ C). Additional equivalents resulted in no further stabilization consistent with a specific binding of Cu^{II}. CD spectroscopy further confirmed retention of a clear antiparallel topology (see **Figures S6, S7, S11–S16**).

After we could show that at least three imidazole ligands are required to complex Cu^{II}, we moved forward to a new series of sequences with six incorporated ligands (htel $\mathbf{L}^{B}_{4}\mathbf{L}^{I}_{2}$, htel $\mathbf{L}^{B}_{3}\mathbf{L}^{I}_{3}$, htelL^B₂L^I₄). Again, the formation of G-quadruplexes with a clear antiparallel topology was observed by CD spectroscopy (see Figures S28, S29). Likewise, comparison of the thermal stabilities showed the destabilizing effect of \mathbf{L}^{B} (htel $\mathbf{L}^{\mathrm{B}}_{4}\mathbf{L}^{\mathrm{I}}_{2}$ $T_{m} = 17^{\circ}\mathrm{C}$, htel $\mathbf{L}^{\mathrm{B}}_{3}\mathbf{L}^{\mathrm{I}}_{3}$ $T_{m} = 26^{\circ}\mathrm{C}$, htel $\mathbf{L}^{\mathrm{B}}_{2}\mathbf{L}^{\mathrm{I}}_{4}$ $T_{m} =$ 26°C), however, not in the linear fashion as observed for the series $\text{htel} \mathbf{L}^{B}_{4-n} \mathbf{L}^{I}_{n}$ (n = 0-4). For the examined set of sixligand-containing sequences, however, direct T_m comparison is not appropriate due to the chosen modification pattern (see Table 1). When investigating the interaction with metal cations, for htelL^B₄L^I₂, a clear stabilization after addition of Cu^{II} ($\Delta T_m = + 6^{\circ}C$) was observed. Considering that for htelL^B₂L^I₂ almost no stabilization was observed ($\Delta T_m = +$ 1° C), we conclude that in htel $L^{B}_{4}L^{I}_{2}$ an involvement of one or two ligandosides L^B into metal coordination is very likely. When further replacing L^B with L^I as in htel $L^B_3 L^I_3$ and htelL^B₂L^I₄, the Cu^{II}-mediated thermal stabilization successively increased from $\Delta T_m = + 9^{\circ}$ C (htelL^B₃L^I₃) to $\Delta T_m = +$ 34°C (htelL^B₂L^I₄). This extremely high thermal stabilization

Name	Sequence 5' 3'	No metal	Co ^{II}	Ni ^{II}	Cu ^{II}	Ζn ^{II}
L ^I G ^[a]	r, B ^u	36	63 (+27)	73 (+37)	76 (+40)	52 (+16)
L ^B G _n	L ^B G n	27	27 (0)	27 (0)	27 (0)	27 (0)
htelL ^I 4A ^[a]	agg L^{I} TT a L^{I} g gtt agg L^{I} TT a L^{I} g g	33	35 (+2)	45 (+12)	56 (+23)	36 (+3)
htelL ^B 4	agg L^B TT a L^B g gtt agg L^B Tt a L^B g g	12	12 (0)	12 (0)	12 (0)	12 (0)
htel L I ₄ B	agg L^{I} Tt t L^{I} g gtt agg L^{I} Tt t L^{I} g g	40	40 (0)	46 (+6)	60 (+20)	40 (0)
htel L^B3L I	agg L^{I} TT a L^{B} g gTT agg L^{B} TT a L^{B} g g	17	17 (0)	17 (0)	17 (0)	17 (0)
htelL ^B 2L ^I 2	agg L^{I} TT a L^{B} g gtt agg L^{I} TT a L^{B} g g	23	23 (0)	23 (+0)	24 (+1)	23 (0)
htel L^BLI₃	agg L^{I} TT a L^{I} g gtt agg L^{I} TT a L^{B} g g	28	28 (0)	28 (+0)	32 (+4)	28 (0)
htel L I ^[a]	AGG L ⁱ tl ⁱ tl ⁱ g gtt agg L ⁱ tl ⁱ tl ⁱ g g	36	44 (+8)	59 (+23)	54 (+18)	44 (+8)
htelL ^B ₄ L ¹ ₂	agg L ^b tl ⁱ tl ^b g gtt agg L ^b tl ⁱ tl ^b g g	17	17 (0)	18 (+1)	23 (+6)	18 (+1)
htel L^B3L^I3	AGG L ^b tl ⁱ tl ⁱ g gtt agg L ^b tl ^b tl ⁱ g g	26	25 (-1)	26 (+0)	35 (+9)	31 (+5)
htelL ^B 2L ^I 4	AGG L ⁱ tl ^b tl ⁱ g gtt agg L ⁱ tl ^b tl ⁱ g g	26	27 (+1)	48 (+22)	60 (+34)	32 (+6)

TABLE 1 Sequences investigated in this study and respective denaturation temperatures T_m (and ΔT_m) in absence and presence of 1 equiv. of Cu^{II}, Ni^{II}, Zn^{II}, Co^{II} (assumed to be oxidized to Co^{III} under the experimental conditions).

Marked in bold font are the incorporated ligandosides L^B and L^I. Conditions: 4 μM (tetramolecular) or 1.88 μM (unimolecular) ssDNA, 100 mM NaCl (tetramolecular) or KCl (unimolecular), 10 mM lithium cacodylate buffer (LiCaCo) pH 7.2 and, if present, 1 equiv. transition metal cation (with respect to the folded G-quadruplex). [a] Punt and Clever (2019b).



is unprecedented for unimolecular G-quadruplexes and much higher compared to the reported G-quadruplexes htel L_6^{I} ($\Delta T_m = + 18^{\circ}$ C) and htel L_4^{I} A ($\Delta T_m = + 23^{\circ}$ C) (Punt and Clever, 2019b).

The formation of 1:1 complexes for htelL^B₂L^I₄ with Cu^{II} and Ni^{II} was further confirmed by native ESI mass spectrometry. To understand whether a G-quadruplex is folded or unfolded in the

gas phase, the intrinsic property of G-quadruplexes is exploited that in their folded state they always bind n-1 potassium ions (where n = number of G-tetrads). For a folded G-quadruplex with two G-tetrads, a main signal corresponding to the adduct with one distinct potassium ion would be expected, followed by a statistical distribution of adducts with further unspecifically bound potassium cations. On the other hand, for an unfolded G-quadruplex, the main signal would correspond to the mass of the DNA strand without potassium ions. The mass spectrum shows a main signal corresponding to $[htelL^{B}_{2}L^{I}_{4}+Cu+K-7H]^{4-}$ (**Figure 4**), thus strongly indicating a folded G-quadruplex coordinating to a Cu^{II} or Ni^{II} ion in the gas phase (D'Atri et al., 2015; Lecours et al., 2017).

Jahn-Teller-distorted Cu^{II} usually favors the coordination of four strongly associated ligands in a square planar geometry, with two additional ligands more loosely bound in axial positions (Halcrow, 2012). After proving a 1:1 complex for $htel L^{B}_{2}L^{I}_{4}$ and Cu^{II}, the question was if all six ligands are participating in metal coordination or if only L^I is involved. Therefore, a new sequence htelL^I₄B was synthesized where L^B was replaced with thymidines. Addition of Cu^{II} led to a thermal stabilization of $\Delta T_m = +20^{\circ}$ C, much lower compared to $\text{htel} \mathbf{L}^{\text{B}}_{2} \mathbf{L}^{\text{I}}_{4}$ ($\Delta T_{m} = +34^{\circ}\text{C}$). However, when looking at the absolute melting temperature T_m in presence of Cu^{II}, one notices that they are the same for both sequences (htelL^B₂L^I₄ $T_m = 60^{\circ}$ C, htelL^I₄B $T_m = 60^{\circ}$ C). This could mean that Cu^{II} coordination by $htel L^{B}_{2}L^{I}_{4}$ simply compensates the destabilizing effect of L^B and no benzoate ligand was involved in Cu^{II} coordination. Further studies are required to shed light on this question.

Besides Cu^{II}, the addition of Zn^{II} and Ni^{II} to htelL^B₂L^I₄ and htelL^B₃L^I₃ led to thermal stabilizations. These results were highly intriguing for two reasons. Quadruplex htelL^B₂L^I₄ was significantly more stabilized with Ni^{II} ($\Delta T_m = + 22^{\circ}$ C) compared to Zn^{II} ($\Delta T_m = + 6^{\circ}$ C). However, in htelL^B₃L^I₃, the opposite effect was observed, showing a higher stabilization after Zn^{II} addition ($\Delta T_m = +5^{\circ}C$), while for Ni^{II} no complexation was observed. This adds to the established variation of ligand number and position a third laver to our system to fine-tune metal affinities by the introduction of heteroleptic systems. As last question, we were interested whether ZnII in htelLB3LI3 is coordinated by one or more benzoates. Interestingly, other sequences shown to complex Zn^{II} (htelL^I₄A $\Delta T_m = + 3^{\circ}C$, htelL^I₆ $\Delta T_m = + 8^{\circ}$ C) always contain at least four counts of L^I. Since in $htel L^B_3 L^I_3$ only three L^I were available, we conclude that an involvement of L^B in coordination to the Zn^{II} cation is likely.

CONCLUSION

A new benzoate-based ligandoside L^B was established in tetramolecular and unimolecular G-quadruplex structures. Homoleptic G-quadruplex $(L^BG_4)_4$ was found to form a clear parallel topology. Its thermal stability indicated a strongly destabilizing effect of L^B compared to L^I which was attributed to an accumulation of negative charges. Also, no interactions between a series of transition metal cations and $(L^BG_4)_4$ were found. Similarly, for the unimolecular G-quadruplex htel L^B_4 , a destabilizing effect of L^B and no interactions with transition metal cations were observed. The successive replacement of L^B with L^I in htel $L^B_3 L^I$, htel $L^B_2 L^I_2$, htel $L^B L^I_3$, and htel L^I_4 resulted in a linear increase of the thermal stability. In addition, for htel $L^B L^I_3$, a weak thermal stabilization after addition of 1 equiv. Cu^{II} indicated specific binding.

When moving to systems with six incorporated ligands, a tremendously high thermal stabilization was observed after addition of Cu^{II} to htelL^B₂L^I₄ ($\Delta T_m = + 34^{\circ}$ C). In comparison, for htelL^I₄B, addition of Cu^{II} resulted in a stabilization of only $\Delta T_m = +20^{\circ}$ C. However, the absolute melting temperatures T_m of htelL^B₂L^I₄ ($T_m = 60^{\circ}$ C) and htelL^I₄B ($T_m = 60^{\circ}$ C) are the same, indicating that Cu^{II} complexation is rather compensating the destabilizing effect of L^{B} . More interesting were the results for htelL^B₂L^I₄ and htelL^B₃L^I₃ after addition of Zn^{II} and Ni^{II}, respectively. HtelL^B₂L^I₄ was significantly more stabilized by Ni^{II} $(\Delta T_m = + 22^{\circ}\text{C})$ compared to Zn^{II} ($\Delta T_m = + 6^{\circ}\text{C}$). However, in htelL^B₃L^I₃, the opposite effect was observed, showing a higher stabilization after Zn^{II} addition ($\Delta T_m = + 5^{\circ}C$) while for Ni^{II} no complexation was found. This expands our toolbox to design tailored binding sites for various transition metal cations. Previously, we had shown to fine-tune coordination environments by varying position and number of ligands. Here, we expand this approach by combining two ligandosides, L^B and L^I, which we regard as an important step for the design of metal-selective G-quadruplexes with application in diagnostics, selective catalysis, and DNA nanotechnology.

DATA AVAILABILITY STATEMENT

The datasets generated for this study can be found in the Cambridge Crystallographic Data Center under the CCDC identifier 1961648.

AUTHOR CONTRIBUTIONS

PP and LS conducted all syntheses and DNA experiments. SS contributed to the tetramolecular systems. LK and CS contributed the X-ray structure of compound 4. PP, LS, and GC designed the study, conceived the experiments, analyzed the data, and authored the manuscript.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fchem. 2020.00026/full#supplementary-material

REFERENCES

- Abdelhamid, M. A., Fábián, L., MacDonald, C. J., Cheesman, M. R., Gates, A. J., and Waller, Z. A. (2018). Redox-dependent control of i-Motif DNA structure using copper cations. *Nucleic Acids Res.* 46, 5886–5893. doi: 10.1093/nar/gky390
- Ali, A., Bullen, G. A., Cross, B., Dafforn, T. R., Little, H. A., Manchester, J., et al. (2019). Light-controlled thrombin catalysis and clot formation using a photoswitchable G-quadruplex DNA aptamer. *Chem. Commun.* 55, 5627–5630. doi: 10.1039/C9CC01540J
- Anderson, J. S., Rittle, J., and Peters, J. C. (2013). Catalytic conversion of nitrogen to ammonia by an iron model complex. *Nature* 501, 84–87. doi: 10.1038/nature12435
- Baskin, M., and Maayan, G. (2016). A rationally designed metal-binding helical peptoid for selective recognition processes. *Chem. Sci.* 7, 2809–2820. doi: 10.1039/C5SC04358A
- Baskin, M., Zhu, H., Qu, Z.-W., Chill, J. H., Grimme, S., and Maayan, G. (2018). Folding of unstructured peptoids and formation of hetero-bimetallic peptoid complexes upon side-chain-to-metal coordination. *Chem. Sci.* 10, 620–632. doi: 10.1039/C8SC03616K
- Boersma, A. J., Coquière, D., Geerdink, D., Rosati, F., Feringa, B. L., and Roelfes, G. (2010a). Catalytic enantioselective syn hydration of enones in water using a DNA-based catalyst. *Nat. Chem.* 2, 991–995. doi: 10.1038/nchem.819
- Boersma, A. J., Megens, R. P., Feringa, B. L., and Roelfes, G. (2010b). DNA -based asymmetric catalysis. *Chem. Soc. Rev.* 39, 2083–2092. doi: 10.1039/ b811349c
- Clever, G. H., Kaul, C., and Carell, T. (2007). DNA-metal base pairs. Angew. Chem. Int. Ed. 46, 6226–6236. doi: 10.1002/anie.200701185
- Colquhoun, H. M., Stoddart, F. J., and Williams, D. J. (1986). Second-sphere coordination – a novel role for molecular receptors. *Angew. Chem. Int. Ed.* 25, 487–507. doi: 10.1002/anie.198604873
- Coquière, D., Feringa, B. L., and Roelfes, G. (2007). DNA-based catalytic enantioselective michael reactions in water. *Angew. Chem. Int. Ed.* 46, 9308–9311. doi: 10.1002/anie.200703459
- Cornish, A. J., Ginovska, B., Thelen, A., da Silva, J. C., Soares, T. A., Raugei, S., et al. (2016). Single-amino acid modifications reveal additional controls on the proton pathway of [FeFe]-hydrogenase. *Biochemistry* 55, 3165–3173. doi: 10.1021/acs.biochem.5b01044
- D'Atri, V., Porrini, M., Rosu, F., and Gabelica, V. (2015). Linking molecular models with ion mobility experiments. Illustration with a rigid nucleic acid structure. *J. Mass. Spectrom.* 50, 711–726. doi: 10.1002/jms.3590
- Degtyarenko, K. (2000). Bioinorganic motifs: towards functional classification of metalloproteins. *Bioinformatics* 16, 851–864. doi: 10.1093/bioinformatics/16.10.851
- Dicke, B., Hoffmann, A., Stanek, J., Rampp, M., Grimm-Lebsanft, B., Biebl, F., et al. (2018). Transferring the entatic-state principle to copper photochemistry. *Nat. Chem.* 10, 355–362. doi: 10.1038/nchem.2916
- Drienovská, I., Alonso-Cotchico, L., Vidossich, P., Lledós, A., Maréchal, J.-D., and Roelfes, G. (2017). Design of an enantioselective artificial metallo-hydratase enzyme containing an unnatural metal-binding amino acid. *Chem. Sci.* 8, 7228–7235. doi: 10.1039/C7SC03477F
- Drienovská, I., Mayer, C., Dulson, C., and Roelfes, G. (2018). A designer enzyme for hydrazone and oxime formation featuring an unnatural catalytic aniline residue. *Nat. Chem.* 10, 946–952. doi: 10.1038/s41557-018-0082-z
- Engelhard, D. M., Meyer, A., Berndhäuser, A., Schiemann, O., and Clever, G. H. (2018a). Di-copper(ii) DNA G-quadruplexes as EPR distance rulers. *Chem. Commun.* 54, 7455–7458. doi: 10.1039/C8CC04053B
- Engelhard, D. M., Nowack, J., and Clever, G. H. (2017). Copper-induced topology switching and thrombin inhibition with telomeric DNA G-quadruplexes. *Angew. Chem. Int. Ed.* 56, 11640–11644. doi: 10.1002/anie.201705724
- Engelhard, D. M., Pievo, R., and Clever, G. H. (2013). Reversible stabilization of transition-metal-binding DNA G-quadruplexes. *Angew. Chem. Int. Ed.* 52, 12843–12847. doi: 10.1002/anie.201307594
- Engelhard, D. M., Stratmann, L. M., and Clever, G. H. (2018b). Structure–property relationships in CuII-binding tetramolecular G-quadruplex DNA. *Chem. Eur. J.* 24, 2117–2125. doi: 10.1002/chem.201703409
- Ghosh, T., Ghosh, P., and Maayan, G. (2018). A copper-peptoid as a highly stable, efficient, and reusable homogeneous water oxidation electrocatalyst. ACS Catal. 8, 10631–10640. doi: 10.1021/acscatal.8b03661

- Greenblatt, H. M., Feinberg, H., Tucker, P. A., and Shoham, G. (1998). Carboxypeptidase a: native, zinc-removed and mercury-replaced forms. *Acta Cryst.* D54, 289–305. doi: 10.1107/S0907444997010445
- Halcrow, M. A. (2012). Jahn–Teller distortions in transition metal compounds, and their importance in functional molecular and inorganic materials. *Chem. Soc. Rev.* 42, 1784–1795. doi: 10.1039/C2CS35253B
- Hänsel-Hertsch, R., Antonio, M., and Balasubramanian, S. (2017). DNA Gquadruplexes in the human genome: detection, functions and therapeutic potential. *Nat. Rev. Mol. Cell. Bio.* 18, 279–284. doi: 10.1038/nrm.2017.3
- Holm, R. H., Kennepohl, P., and Solomon, E. I. (1996). Structural and functional aspects of metal sites in biology. *Chem. Rev.* 96, 2239–2314. doi: 10.1021/cr9500390
- Jeschek, M., Reuter, R., Heinisch, T., Trindler, C., Klehr, J., Panke, S., et al. (2016). Directed evolution of artificial metalloenzymes for *in vivo* metathesis. *Nature* 537, 661–665. doi: 10.1038/nature19114
- Kanady, J. S., Tsui, E. Y., Day, M. W., and Agapie, T. (2011). A synthetic model of the Mn3Ca subsite of the oxygen-evolving complex in photosystem II. *Science* 333, 733–736. doi: 10.1126/science.1206036
- Katz, S. (1963). The reversible reaction of Hg (II) and double-stranded polynucleotides a step-function theory and its significance. *Biochim. Biophys. Acta* 68, 240–253. doi: 10.1016/0926-6550(63)90435-3
- Key, H. M., Dydio, P., Clark, D. S., and Hartwig, J. F. (2016). Abiological catalysis by artificial haem proteins containing noble metals in place of iron. *Nature* 534, 534–537. doi: 10.1038/nature17968
- Knight, A. S., Kulkarni, R. U., Zhou, E. Y., Franke, J. M., Miller, E. W., and Francis, M. B. (2017). A modular platform to develop peptoidbased selective fluorescent metal sensors. *Chem. Commun.* 53, 3477–3480. doi: 10.1039/C7CC00931C
- Koehntop, K. D., Emerson, J. P., and Que, L. (2005). The 2-His-1carboxylate facial triad: a versatile platform for dioxygen activation by mononuclear non-heme iron(II) enzymes. J. Biol. Inorg. Chem. 10, 87–93. doi: 10.1007/s00775-005-0624-x
- Lecours, M. J., Marchand, A., Anwar, A., Guetta, C., Hopkins, S. W., and Gabelica, V. (2017). What stoichiometries determined by mass spectrometry reveal about the ligand binding mode to G-quadruplex nucleic acids. *Biochim. Biophys. Acta* 1861, 1353–1361. doi: 10.1016/j.bbagen.2017.01.010
- Litman, Z. C., Wang, Y., Zhao, H., and Hartwig, J. F. (2018). Cooperative asymmetric reactions combining photocatalysis and enzymatic catalysis. *Nature* 560, 355–359. doi: 10.1038/s41586-018-0413-7
- Lu, Y., Yeung, N., Sieracki, N., and Marshall, N. M. (2009). Design of functional metalloproteins. *Nature* 460, 855–862. doi: 10.1038/nature08304
- Mandal, S., Hebenbrock, M., and Müller, J. (2016). A dinuclear mercury(II)mediated base pair in DNA. *Angew. Chem. Int. Ed.* 55, 15520–15523. doi: 10.1002/anie.201608354
- Mandal, S., and Müller, J. (2017). Metal-mediated DNA assembly with ligand-based nucleosides. *Curr. Opin. Chem. Biol.* 37, 71–79. doi: 10.1016/j.cbpa.2017.01.019
- Mayer, C., Dulson, C., Reddem, E., Thunnissen, A. W., and Roelfes, G. (2019). Directed evolution of a designer enzyme featuring an unnatural catalytic amino acid. *Angew. Chem. Int. Ed.* 58, 2083–2087. doi: 10.1002/anie.201813499
- Miyoshi, D., Karimata, H., Wang, Z.-M., Koumoto, K., and Sugimoto, N. (2007). Artificial G-wire switch with 2,2'-bipyridine units responsive to divalent metal ions. J. Am. Chem. Soc. 129, 5919–5925. doi: 10.1021/ja068707u
- Neidle, S. (2017). Quadruplex nucleic acids as targets for anticancer therapeutics. *Nat. Rev. Chem.* 1:0041. doi: 10.1038/s41570-017-0041
- Punt, P. M., and Clever, G. H. (2019a). Imidazole-modified G-quadruplex DNA as metal-triggered peroxidase. *Chem. Sci.* 10, 2513–2518. doi: 10.1039/C8SC05020A
- Punt, P. M., and Clever, G. H. (2019b). Tailored transition-metal coordination environments in imidazole-modified DNA G-quadruplexes. *Chem. Eur. J.* 25, 13987–13993. doi: 10.1002/chem.201903445
- Raven, J. A., Evans, M. C., and Korb, R. E. (1999). The role of trace metals in photosynthetic electron transport in O₂-evolving organisms. *Photosynth. Res.* 60, 111–150. doi: 10.1023/A:1006282714942
- Rioz-Martínez, A., Oelerich, J., Ségaud, N., and Roelfes, G. (2016). DNAaccelerated catalysis of carbene-transfer reactions by a DNA/cationic iron porphyrin hybrid. *Angew. Chem. Int. Ed.* 55, 14136–14140. doi: 10.1002/anie.201608121

- Roelfes, G., and Feringa, B. L. (2005). DNA-based asymmetric catalysis. Angew. Chem. Int. Ed. 44, 3230–3232. doi: 10.1002/anie.200500298
- Rubino, J. T., and Franz, K. J. (2012). Coordination chemistry of copper proteins: how nature handles a toxic cargo for essential function. J. Inorg. Biochem. 107, 129–143. doi: 10.1016/j.jinorgbio.2011.11.024
- Samuel, A. P., Co, D. T., Stern, C. L., and Wasielewski, M. R. (2010). Ultrafast photodriven intramolecular electron transfer from a zinc porphyrin to a readily reduced diiron hydrogenase model complex. *J. Am. Chem. Soc.* 132, 8813–8815. doi: 10.1021/ja100016v
- Sandmann, N., Bachmann, J., Hepp, A., Doltsinis, N. L., and Müller, J. (2019). Copper(ii)-mediated base pairing involving the artificial nucleobase 3 H -imidazo[4,5- f]quinolin-5-ol. *Dalton Trans.* 48, 10505–10515. doi: 10.1039/C9DT02043H
- Shook, R. L., and Borovik, A. (2010). Role of the secondary coordination sphere in metal-mediated dioxygen activation. *Inorg. Chem.* 49, 3646–3660. doi: 10.1021/ic901550k
- Smith, N. M., Amrane, S., Rosu, F., Gabelica, V., and Mergny, J.-L. (2012). Mercury-thymine interaction with a chair type G-quadruplex architecture. *Chem. Commun.* 48, 11464–11466. doi: 10.1039/c2cc36481f
- Tanaka, K., Clever, G. H., Takezawa, Y., Yamada, Y., Kaul, C., Shionoya, M., et al. (2006). Programmable self-assembly of metal ions inside artificial DNA duplexes. *Nat. Nanotechnol.* 1, 190–194. doi: 10.1038/nnano.2006.141
- Tanaka, K., Yamada, Y., and Shionoya, M. (2002). Formation of silver(I)mediated DNA duplex and triplex through an alternative base pair of pyridine nucleobases. J. Am. Chem. Soc. 124, 8802–8803. doi: 10.1021/ja0205100
- Valdez, C. E., Smith, Q. A., Nechay, M. R., and Alexandrova, A. N. (2014). Mysteries of metals in metalloenzymes. Acc. Chem. Res. 47, 3110–3117. doi: 10.1021/ar500227u
- Waldron, K. J., Rutherford, J. C., Ford, D., and Robinson, N. J. (2009). Metalloproteins and metal sensing. *Nature* 460, 823–830. doi: 10.1038/nature08300

- Wang, J., Yue, L., Li, Z., Zhang, J., Tian, H., and Willner, I. (2019). Active generation of nanoholes in DNA origami scaffolds for programmed catalysis in nanocavities. *Nat. Commun.* 10:4963. doi: 10.1038/s41467-019-12933-9
- Wu, S., Zhou, Y., Rebelein, J. G., Kuhn, M., Mallin, H., Zhao, J., et al. (2019). Breaking symmetry: engineering single-chain dimeric streptavidin as host for artificial metalloenzymes. J. Am. Chem. Soc. 141, 15869–15878. doi: 10.1021/jacs.9b06923
- Xu, Y., Suzuki, Y., Lönnberg, T., and Komiyama, M. (2009). Human telomeric DNA sequence-specific cleaving by G-quadruplex formation. J. Am. Chem. Soc. 131, 2871–2874. doi: 10.1021/ja807313x
- Zhang, L., Peritz, A., and Meggers, E. (2005). A simple glycol nucleic acid. J. Am. Chem. Soc. 127, 4174–4175. doi: 10.1021/ja042564z
- Zhang, L., Peritz, A. E., Carroll, P. J., and Meggers, E. (2006). Synthesis of glycol nucleic acids. Synthesis 2006, 645–653. doi: 10.1055/s-2006-926313
- Zhao, M., Wang, H.-B., Ji, L.-N., and Mao, Z.-W. (2013). Insights into metalloenzyme microenvironments: biomimetic metal complexes with a functional second coordination sphere. *Chem. Soc. Rev.* 42, 8360–8375. doi: 10.1039/c3cs60162e

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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