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Fishery waste valorization: Sulfated ZrO₂ as a heterogeneous catalyst for chitin and chitosan depolymerization

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Chitin and chitosan are abundant unique sources of biologically-fixed nitrogen mainly derived from residues of the fishery productive chain. Their high potential as nitrogen-based highly added-value platform molecules is still largely unexploited and a catalytic way for their valorization would be strongly desirable within a biorefinery concept. Here we report our results obtained with a series of heterogeneous catalysts in the depolymerization of chitosan and chitin to acetylglucosamine. Copper catalysts supported on SiO₂, SiO₂-Al₂O₃, SiO₂-ZrO₂, ZrO₂ and the corresponding bare oxides/mixed oxides were tested, together with a sulfated zirconia system (ZrO_2 -SO₃H) that revealed to be extremely selective towards glucosamine, both for chitosan and chitin, thus giving pretty high yields with respect to the values reported so far (44% and 21%, respectively). The use of a heterogeneous catalyst alone, without the need of any additives or the combination with a mineral acid, makes these results remarkable.

KEYWORDS

chitin, chitosan, solid acids, ZrO₂, sulfated catalyst

Introduction

Chitin is the second most abundant biopolymer besides cellulose and is the first aminosaccharide based-one. It can be extracted on large scale (10¹¹ tons/year) from crustaceans shell waste, the industrial waste material of fisheries, through a well-established protocol, and it can be also obtained from insects. (Jia et al., 2014) (Hou et al., 2020) (Sagawa et al., 2019) (Mohan et al., 2022).

For these reasons, chitin and its deacetylated form, chitosan, can be considered as relevant second generation renewable raw materials, that through biorefinery processing produce added-value platform molecules, particularly when the nitrogen-containing functional group is preserved. Despite this, their applications are often limited to wastewater treatment, food industry, cosmetics, and nanomaterials and composite materials preparation. (Yan and Chen, 2015) (Gómez-López et al., 2020) (De et al., 2015) (Ahmed et al., 2020) (Jayakumar et al., 2010) (Sanandiya et al., 2020) (Deliyanni

et al., 2015) (Mäki-Arvela et al., 2007) Thus, chitin and chitosan represent abundant and unique sources of biologically-fixed nitrogen nowadays still largely un-exploited.

By applying the same approach used for cellulose and hemicellulose, chitin and chitosan can be depolymerized into their constituent monomers, namely 2-acetamido-2-deoxy-D-glucose (*N*-acetylglucosamine, GlcNAc) and 2-amino-2deoxy-D-glucose (glucosamine, GlcN), that are the starting point for the synthesis of new and sustainable renewable molecules in which the nitrogen atom is retained (Figure 1). (Pelckmans et al., 2017) (Omari et al., 2012) (Ohmi et al., 2013) (Sadiq et al., 2018) (Osada et al., 2013) (Dai et al., 2019) (Sagawa et al., 2019) (Jia et al., 2017) (Drover et al., 2012) (Xu et al., 2022).

For example, the catalytic oxidation of GlcN affords the valueadded glucosaminic acid. (Ohmi et al., 2013) GlcN can also be dehydrated into deoxyfructosazine and fructosazine, that find application as flavouring agents, but also show potential antimicrobial and pharmacological activity. (Jia et al., 2014) (Jia et al., 2017) (Bhattacherjee et al., 2016) Typically, these compounds price is around 70/mg USD and 32/mg USD by Santa Cruz Biotech-Nology, respectively. (Jia et al., 2014) (Jia et al., 2017) Analogously to 5-(hydroxymethyl)furfural (HMF) from glucose, the dehydration of GlcNAc leads to the 3-acetamido-5-acetylfuran, a promising N-containing platform molecule that provides many opportunities for accessing a wide range of products. Omari et al. (2012), Drover et al. (2012), Padovan et al. (2020), Sadiq et al. (2018), Chen et al. (2014) GlcNAc can be also converted into Chromogen I and III that recently have attracted great attention as new functional food additives and as potential therapeutics, having potent biological activities. (Osada et al., 2013) (Zheng et al., 2017) At the same time, the combination of hydrogenation and dehydration reactions leads to the 2-acetamide-2-deoxyisosorbide, an analogous of isosorbide, which is a new potential precursor to N-containing polymers. (Sagawa et al., 2019).

On the other hand, when the nitrogen atom is lost, the valorisation of chitin and chitosan produces compounds such as HMF, levulinic acid and C2–C6 polyols, similarly to what happens in the cellulose and hemicellulose deconstruction pathways (Figure 1). (Kim et al., 2018) (Hou et al., 2020) (Omari et al., 2012) (Wang et al., 2013) (Bobbink et al., 2015) (Pandit et al., 2020) (Ohmi et al., 2013) (Padovan et al., 2020) (Xu et al., 2022) (Kougioumtzis et al., 2018).

In contrast to the valorization of cellulose and hemicellulose, chitin and chitosan depolymerization studies are only at early stages, despite the similar chemical structure of these natural polymers. Even if more efforts would be needed, particularly aimed at improving selectivity and at reducing waste, the transformation of cellulose and hemicellulose can inspire chitin and chitosan exploitation, which is still very limited.

The first step for the profitable use of chitin and chitosan as feedstock for biorefineries is the identification of an effective depolymerisation protocol to obtain oligomers or the corresponding monomers. Few attempts are reported in the literature. Most of the works relies on the use of toxic and dangerous homogeneous systems, based on mineral acids (HNO₃, H₂SO₄, and HCl) or free radicals, sometimes with the aid of mechanical treatments, such as ball-milling. (Harish Prashanth and Tharanathan, 2007) (Einbu and Vårum, 2008)



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(Zhang and Yan, 2017) (Yabushita et al., 2015) (Kobayashi et al., 2021) The use of heterogeneous catalysts is more convenient, since they are safer to handle, generally less toxic and, with respect to mineral acids, they entail lower purification costs and they do not require the neutralization of the reaction medium. Unfortunately, the processes reported are effective only after chitosan solubilization in a mineral acid, such as H₂SO₄ and HCl, as in the case of Amberlyts-15 (GlcN yield = 58%) (Sagawa et al., 2019), and of a glucose-derived solid acid (GlcN yield = 98.1%). (Zhang et al., 2018) On the other hand, when a solid acid catalyst is used alone (e.g., H-Mordenite), low molecular weight chitosan oligomers are obtained. (Pandit et al., 2020) Sometimes, during the reaction the loss of nitrogen atoms occurs. In these cases, the formation of the products usually derived from cellulose and hemicellulose valorisation, such as levulinic acid and HMF, is observed. (Omari et al., 2012) (Hou et al., 2020).

In this work we investigated the activity of some oxides and mixed oxides as simple heterogeneous catalysts for the selective conversion of chitosan and chitin into GlcN.

Experimental

Materials

All chemicals, in particular low molecular weight chitosan, chitin from shrimp shells and HPLC standards (formic acid, acetic acid), were obtained from Sigma-Aldrich, with the exception of HMF that was purchased from AVA Biochem.

 ZrO_2 (Melcat XZO1521), SiO_2 -Al₂O₃ 135 (Aldrich), SiO_2 A (Merk), and pyrogenic SiO_2 B (Aeroperl 300/30) are commercial materials respectively purchased from Mel-Chemicals, Sigma-Aldrich and Evonik. SiO_2 - ZrO_2 containing a 4.7% nominal amount of zirconia was kindly provided by Grace Davison (Worms, Germany) Textural properties are reported in Table 1.

Catalysts preparation

Copper catalysts were prepared according to the Chemisorption-Hydrolysis method (Zaccheria et al., 2013)

TABLE 1 Physiochemical characterization of the materials.

Sample	SSA (m ² /g)	PV (cm^3/g)
SiO ₂ A	460	0.82
SiO ₂ B	267	1.65
SiO ₂ -Al ₂ O ₃	496	0.74
SiO ₂ -ZrO ₂	304	1.62
ZrO_2	358	0.20

SSA, Specific Surface Area; PV = Pore Volume.

(Scotti et al., 2017) by adding 10 g of support (SiO₂ A, SiO₂ B, and SiO₂-Al₂O₃) to an aqueous $[Cu(NH_3)_4]^{2+}$ solution, prepared by dropping NH₄OH to a 25 mL of a Cu(NO₃)₂·3H₂O solution until pH 9 had been reached. After 20 min under stirring, the slurry, held in an ice bath at 0°C, was diluted with 3 L of water for 2 h. The solid was separated by vacuum filtration with a Büchner funnel, washed with water, dried overnight at 110°C, and calcined in air at 350°C for 4 h.

Sulphonated ZrO_2 catalyst was prepared according to Fărcașiu and Li, 1995 protocol. 5 g of the oxide were soaked in 75 mL of aq. 1 M H₂SO₄ for 2 h, filtered, and dried overnight at 110°C. Finally, the dried material was calcined at 350°C for 4 h.

Chitin and chitosan hydrolysis

Before reaction, the catalyst (400 mg) was placed in a glass reactor and dehydrated at 270°C in an oven (20 min in air +20 min under vacuum). The reactions were performed in a Hastelloy Parr autoclave. The reactor was loaded with 750 mg of chitin/chitosan, 50 mL of H₂O and the dried catalyst. The autoclave was sealed, evacuated, and filled with 4 atm of N₂. The system was heated up to the desired temperature under stirring (750 rpm). After the desired time, the autoclave was vented, and the catalyst and the insoluble residues were separated by filtration.

Product analysis

Products were analysed by HPLC using an Agilent 1,200 Infinity LC system equipped with a Quaternary gradient pump unit and a refractive index (RID) detector. MetaCarb H Plus Column 300 × 7.8 mm (Agilent Technologies, Inc.), hyphenated with MetaCarb H Plus Guard Column 50 × 4.6 mm, was used as analytical column. MilliQ water was used as the mobile phase and the separation was carried for 2 h under the following conditions: flow rate 0.4 mL/min at 60°C, wavelength λ = 195 nm, RID temperature 35°C. Before analysis, the reaction mixture was filtered through a 0.45 µm microporous membrane, diluted with milliQ water at a 0,66 pV dilution factor, and injected (20 µl) with a 50 µl glass syringe (Agilent Technologies, Inc.). To calculate the GlcN yield (wt%), a linear calibration curve (r2 > 0.99) was obtained using GlcN as standard and the following equation was used:

$$Y(wt\%) = \frac{mmol_{GlcN} \cdot MM_{GlcN}}{mg_{reagent}},$$
(1)

Where $mmol_{GlcN}$ is obtained from the HPLC analysis and $mg_{reagent}$ refers to chitin/chitosan.





The conversion of chitin and chitosan into soluble products was calculated through total organic carbon (TOC) analysis on the liquid filtrated solutions. TOC analysis was performed using a TOC-LCSH analyser purchased from Shimadzu. Samples for TOC were all prepared at a 16 dilution factor mixing 3.125 mL of the filtered reaction mixture with milliQ water to a final volume of 50 mL. The instrument performs the analysis of the TC and IC and create the TOC value given the relationship TOC = TC—IC.

Results and discussion

In a previous work, some of us reported that the selectivity in the hydrolysis of cellulose can be addressed by using different catalysts based on oxides and mixed oxides (e.g. SiO_2 , SiO_2 - ZrO_2 , and SiO_2 - Al_2O_3) and the corresponding copper-based ones. The Lewis and Brønsted acidity features of the tested materials accounted for the different distribution of the products obtained. (Mariani et al., 2014) In particular, when a CuO/ SiO₂ catalyst is used glucose is obtained in high amount. On



TABLE 2 Chitosan depolymerization (T = 200° C; N₂ = 4 atm; H₂O = 50 mL; Chitosan = 750 mg; cat = 400 mg; t = 5 h).

Catalyst	C (%)	Y _{GlcN} (%)
no cat	27	4
SiO ₂ -Al ₂ O ₃	30	4
SiO ₂ -ZrO ₂	32	5
16 CuO/SiO ₂ A	38	6
16 CuO/SiO ₂ B	57	5
16 CuO/SiO ₂ -Al ₂ O ₃	36	7
ZrO_2	33	6
ZrO ₂ -SO ₃ H	32	23
	no cat $SiO_2-Al_2O_3$ SiO_2-ZrO_2 16 CuO/SiO_2 A 16 CuO/SiO_2 B 16 CuO/SiO_2-Al_2O_3 ZrO_2	no cat 27 SiO ₂ -Al ₂ O ₃ 30 SiO ₂ -ZrO ₂ 32 16 CuO/SiO ₂ A 38 16 CuO/SiO ₂ B 57 16 CuO/SiO ₂ -Al ₂ O ₃ 36 ZrO ₂ 33

the other hand, when ion-like copper species, as those formed over a SiO_2 -Al₂O₃, were present, lactic acid is the main product. With respect to the non-catalysed reaction, the bare supports have an important effect on the selectivity, giving significantly higher amounts of glucose and HMF.

Based on the previous promising results obtained with cellulose, we applied heterogeneous catalysis to chitin and chitosan (Scheme 1), to obtain GlcN. Thus, SiO_2 A and B,

 ${\rm SiO_2\text{-}Al_2O_3}$ and the corresponding copper supported materials were tested at first in chitosan depolymerisation.

The results are reported in Table 2, entries 1–6, together with the non-catalysed reaction. Surprisingly, the use of silicas and silica-alumina did not result in significant improvements in both conversion to soluble products (30%–38%) and GlcN yield (4%–7%), nor did the supported Cu catalysts, except for CuO/SiO₂ B. CuO/SiO₂ B gave a higher conversion (57%), but the GlcN selectivity was still low (5%). Different products were found in the HPLC chromatograms (Figure 2), but, due to the complexity of the reaction, we were able to identify only GlcN, and very small amounts of acetic and formic acid. To note that neither HMF or levulinic acid were found in the reaction mixtures. Future work will be devoted to the identification of added-value products, others than GlcN, even in small amounts.

One of the major issues in the depolymerisation of natural polymers (cellulose, chitin, and chitosan) is their robust crystallinity due the presence of extensive inter- and intramolecular hydrogen bonds. (Margoutidis et al., 2018) These exploratory experiments shed light on the more difficult depolymerization of chitosan with respect to what observed with cellulose, and the same pool of catalysts that successfully drove the selectivity in cellulose hydrolysis revealed to be ineffective in the present case. Pursuing our effort in the search for an effective catalyst for the production of GlcN from chitosan, we therefore moved to sulfated zirconia (ZrO₂-SO₃H), known to possess strong Brønsted acidity. (Katada et al., 2000) The protocol used for the sulphation was straightforward (Fărcașiu and Li, 1995), and the effectiveness of the treatment was confirmed by FT-IR of adsorbed pyridine (Figure 3). The parent ZrO₂ shows only the presence of Lewis acid sites, with the typical bands at 1,446 cm⁻¹ and 1,607 cm⁻¹, while in the ZrO₂-SO₃H sample two signals at 1,542 cm⁻¹ and 1,638 cm⁻¹, ascribable to Brønsted acid sites, can also be detected. The sulfation does not seem to affect the structure of the oxides, as the surface area of the parent ZrO₂ and the one of the ZrO₂-SO₃H materials do not show a significant difference (358 m²/g vs. 367 m²/g).

Interestingly, ZrO_2 - SO_3H resulted to be an active material for chitosan depolymerisation (Table 2, entry 8). With this catalyst conversion reaches a value of 32%, coherent with results obtained on silica-based catalysts; however, GlcN selectivity is significantly higher, resulting in a 5-fold increase (23%). For comparison, the activity of the parent ZrO_2 is similar to the one observed in the non-catalytic reaction, thus highlighting the fundamental role of the sulfation treatment.

 ZrO_2 -SO₃H is a promising catalyst for the selective hydrolysis of chitosan to GlcN. Differently to what observed with cellulose, where Lewis acid catalysts favour the monomer (glucose) product (Mariani et al., 2014), in chitosan depolymerisation the role of strong Brønsted acidity is critical to produce its building block (GlcN). In this respect, Omari et al. reported that Lewis acid homogeneous catalysts give higher HMF/LA yield, with respect to Brønsted ones, in the depolymerisation of chitosan by using SnCl₄·5H₂O and microwave irradiation. (Omari et al., 2012) Unfortunately, no information about the GlcN yields, that would be helpful to compare the effect of the different acidity, was reported.

By virtue of the positive results shown by ZrO_2 -SO₃H, we tried to improve the yield in GlcN by optimising the reaction parameters (time and temperature, Figure 4). The results show that most of chitosan converts within the first hour, with the formation of GlcN. A small increase in the GlcN yield was observed at 175°C and, especially, at 200°C, while conversion to a soluble product has a fluctuating behaviour. The maximum GlcN yield was 27%. The higher discrepancy between conversion and selectivity observed at 175°C could be related to the formation of soluble oligomers. It worth noting that the role of soluble oligomers in addressing the product distribution has already been put in light in cellulose deconstruction. (Chambon et al., 2011) A further increase to 200°C leads to easier hydrolysis of oligomers, but at the same time humins formation is favoured.

This effect is clearly shown by observing the colour of the reaction solutions after filtration (Figure 5).

The formation of the decomposition products at higher temperature, likely due to the Maillard reaction (Wu et al., 2014), is highlighted by the browning of the solution going from 150°C to 200°C. In any case, the selectivity to GlcN is generally quite satisfying, with values that are generally higher than 65%–70%. As expected, the better selectivity was obtained at 150°C with a value of 87% at 1 h up to 99% after 12 h, and a single peak of GlcN was observed (Figure 2). With increasing temperature, the peak of GlcN is more intense, but at the same time the chromatogram reveals the formation of some by-products, such as formic acid and smaller amounts of acetic acid (Figure 2). In order to maximize the yield in GlcN the amount of the catalyst was increased up to 800 mg. This resulted in a GlcN yield of 44% at 200°C in 2.5 h. This preliminary result paves the way to further studies focused on the design of a more



Chitosan depolymerisation over ZrO_2 - SO_3H catalyst. C (%) = conversion to soluble products; Y (%) = GlcN yield; S (%) = GlcN selectivity, (chitosan = 750 mg; cat = 400 mg; H₂O = 50 mL, N₂ = 4 atm).



FIGURE 5

Filtered product solution collected at different temperature (chitosan = 750 mg; $ZrO_2\text{-}SO_3H$ = 400 mg; H_2O = 50 mL; N_2 = 4 atm, t = 1 h).

main product (Scheme 2), while its acetylated form, the GlcNAc, was observed only in small amounts (Figure 7).

With respect to chitosan, the activity of the Zr-based catalyst decreases and a maximum GlcN yield of 21% (200° C, 12 h) was obtained.

By combining these results, with those obtained by some of us with cellulose, under similar experimental conditions, the following depolymerization tendency was observed: cellulose (Mariani et al., 2014)> chitosan > chitin. Copperbased catalysts and mixed oxides, such as SiO2-Al2O3 or SiO₂-ZrO₂, can be considered as promising materials for cellulose valorization, whilst fail with aminosaccharide polymers. With these substrates stronger acidity and sulfated systems are required. It has already been highlighted into the literature that the reactivity of chitin dramatically differs from those of cellulose and chitosan. (Deringer et al., 2016) (Yabushita et al., 2015) (Kaczmarek et al., 2019) Indeed, chitin has a higher crystallinity due to the presence of acetamido groups that enables the formation of numerous inter- and intra-molecular hydrogen bonds between linear chains.

Our results are very significant, if compared to those reported in the literature. Indeed, as above mentioned, the depolymerisation of chitin and chitosan are complicated and



performant ZrO_2 -based material, in order to the decrease the catalyst amount and the reaction temperature (thus limiting humins formation). In particular, the development of an optimized sulfation protocol would lead to an increase of the Brønsted acid sites density on the catalyst surface and in turn to an improved activity. The catalytic depolymerization of chitin over the ZrO_2 -SO₃H catalyst was also studied and the results are reported in Figure 6. The reaction produces directly GlcN as the

poorly studied reactions. Enzymatic routes are mainly pursued to obtain oligosaccharides or the deacetylation of chitin to chitosan. Moreover, they suffer from high cost and long reaction time, thus their use is often limited to the laboratory scale. (Kaczmarek et al., 2019) (Mohan et al., 2022) (Zhang et al., 2022) The use of toxic and dangerous homogeneous systems, such as mineral acids (HNO₃, H₂SO₄, and HCl) or free radicals, sometimes with the aid of





mechanical treatments, is reported so far. (Harish Prashanth and Tharanathan, 2007) (Einbu and Vårum, 2008) (Zhang and Yan, 2017) (Yabushita et al., 2015) Some processes end up in the loss of the N atom from the resulting product. This is the case of ionic liquids (Hou et al., 2020) and of a combination of $SnCl_4$ · $5H_2O$ with microwave irradiation (Omari et al., 2012), where HMF and levulinic acid were obtained. When a heterogeneous catalyst is used, its combination with a homogeneous one is always requested. For example, Amberlyts-15 and H_2SO_4 gave a GlcN yield of 58%. (Dai et al., 2019), while a glucose-derived solid acid catalyst resulted in a GlcN yield of 98.1%, but only after complete dissolution of chitosan in a hydrochloric acid solution. (Zhang et al., 2018) If the solid acid (H-Mordenite) is not assisted by a homogeneous one only oligomers were obtained, as results of an incomplete deconstruction of chitosan into the monomer. (Pandit et al., 2020) In this paper, we presented a new approach, in which the heterogeneous catalyst is used alone. The fruitful marriage between a high surface area zirconia and a simple sulfation protocol, gave us an active, selective, and easy-to-prepare material. Future work will be focused on the use of different zirconia-based materials (Zaccheria et al., 2020), and on the investigation of different sulfation procedures, to design a more performant system.

Conclusion

A series of heterogeneous catalysts, based on oxides and mixed oxides have been studied in the reaction of chitosan and chitin depolymerization. They revealed to be much more recalcitrant to deconstruction with respect to cellulose. On the other hand, a material having strong Brønsted acidity, namely ZrO₂-SO₃H, resulted in a good selectivity towards GlcN and a yield of 21% from chitin and 44% from chitosan. To the best of our knowledge, our protocol is the first example relying on the use of a heterogeneous catalyst alone, active without the addition of any additives or mineral acids. This represents a step ahead, compared to homogeneouscatalysed processes that require massive neutralisation and purification steps, or the enzymatic ones that suffer for high costs and long reaction times. (Kaczmarek et al., 2019).

Data availability statement

The raw data supporting the conclusion of this article will be made available by the authors, without undue reservation.

References

Ahmed, M. J., Hameed, B. H., and Hummadi, E. H. (2020). Review on recent progress in chitosan/chitin-carbonaceous material composites for the adsorption of water pollutants. *Carbohydr. Polym.* 247, 116690. doi:10.1016/j.carbpol.2020. 116690

Bhattacherjee, A., Hrynets, Y., and Betti, M. (2016). Fructosazine, a polyhydroxyalkylpyrazine with antimicrobial activity: Mechanism of inhibition against extremely heat resistant *Escherichia coli. J. Agric. Food Chem.* 64, 8530–8539. doi:10.1021/acs.jafc.6b03755

Bobbink, F. D., Zhang, J., Pierson, Y., Chen, X., and Yan, N. (2015). Conversion of chitin derived N-acetyl-d-glucosamine (NAG) into polyols over transition metal catalysts and hydrogen in water. *Green Chem.* 17, 1024–1031. doi:10.1039/c4gc01631a

Chambon, F., Rataboul, F., Pinel, C., Cabiac, A., Guillon, E., and Essayem, N. (2011). Cellulose hydrothermal conversion promoted by heterogeneous Brønsted and Lewis acids: Remarkable efficiency of solid Lewis acids to produce lactic acid. *Appl. Catal. B Environ.* 105, 171–181. doi:10.1016/j.apcatb.2011.04.009

Chen, X., Chew, S. L., Kerton, F. M., and Yan, N. (2014). Direct conversion of chitin into a N-containing furan derivative. *Green Chem.* 16, 2204–2212. doi:10.1039/c3gc42436g

Dai, J., Gözaydln, G., Hu, C., and Yan, N. (2019). Catalytic conversion of chitosan to glucosaminic acid by tandem hydrolysis and oxidation. *ACS Sustain. Chem. Eng.* 7, .9b01912–12407. doi:10.1021/acssuschemeng.9b01912

De, S., Balu, A. M., Van Der Waal, J. C., and Luque, R. (2015). Biomass-derived porous carbon materials: Synthesis and catalytic applications. *ChemCatChem* 7, 1608–1629. doi:10.1002/cctc.201500081

Deliyanni, E. A., Kyzas, G. Z., Triantafyllidis, K. S., and Matis, K. A. (2015). Activated carbons for the removal of heavy metal ions : A systematic review of recent literature focused on lead and arsenic ions. Beijing, 699–708. doi:10.1515/chem-2015-0087

Deringer, V. L., Englert, U., and Dronskowski, R. (2016). Nature, strength, and cooperativity of the hydrogen-bonding network in α-chitin. *Biomacromolecules* 17, 996–1003. doi:10.1021/acs.biomac.5b01653

Drover, M. W., Omari, K. W., Murphy, J. N., and Kerton, F. M. (2012). Formation of a renewable amide, 3-acetamido-5-acetylfuran, via direct conversion of N-acetyld-glucosamine. *RSC Adv.* 2, 4642–4644. doi:10.1039/c2ra20578e

Author contributions

NS formulated the ideas and conceptualized the work; YR and VP prepared the catalysts and performed the catalytic tests; NS characterized the materials; NS, FZ, and VP wrote the original draft and prepared the figures; NR, LC, FZ, and NS revised and edited the manuscript.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Einbu, A., and Vårum, K. M. (2008). Characterization of chitin and its hydrolysis to GlcNAc and GlcN. *Biomacromolecules* 9, 1870–1875. doi:10.1021/bm8001123

Fărcașiu, D., and Li, J. Q. (1995). Preparation of sulfated zirconia catalysts with improved control of sulfur content. *Appl. Catal. A General* 128, 97–105. doi:10. 1016/0926-860X(95)00077-1

Gómez-López, P., Puente-Santiago, A., Castro-Beltrán, A., Santos do Nascimento, L. A., Balu, A. M., Luque, R., et al. (2020). Nanomaterials and catalysis for green chemistry. *Curr. Opin. Green Sustain. Chem.* 24, 48–55. doi:10.1016/j.cogsc.2020. 03.001

Harish Prashanth, K. V., and Tharanathan, R. N. (2007). Chitin/chitosan: Modifications and their unlimited application potential-an overview. *Trends Food Sci. Technol.* 18, 117–131. doi:10.1016/j.tifs.2006.10.022

Hou, W., Zhao, Q., and Liu, L. (2020). Selective conversion of chitin to levulinic acid catalyzed by ionic liquids: Distinctive effect of: N-Acetyl groups. *Green Chem.* 22, 62–70. doi:10.1039/c9gc02669j

Jayakumar, R., Menon, D., Manzoor, K., Nair, S. V., and Tamura, H. (2010). Biomedical applications of chitin and chitosan based nanomaterials - a short review. *Carbohydr. Polym.* 82, 227–232. doi:10.1016/j.carbpol.2010.04.074

Jia, L., Liu, X., Qiao, Y., Pedersen, C. M., Zhang, Z., Ge, H., et al. (2017). Mechanism of the self-condensation of GlcNH2: Insights from *in situ* NMR spectroscopy and DFT study. *Appl. Catal. B Environ.* 202, 420–429. doi:10.1016/ j.apcatb.2016.09.058

Jia, L., Wang, Y., Qiao, Y., Qi, Y., and Hou, X. (2014). Efficient one-pot synthesis of deoxyfructosazine and fructosazine from d-glucosamine hydrochloride using a basic ionic liquid as a dual solvent-catalyst. *RSC Adv.* 4, 44253–44260. doi:10.1039/ c4ra06832g

Kaczmarek, M. B., Struszczyk-Swita, K., Li, X., Szczęsna-Antczak, M., and Daroch, M. (2019). Enzymatic modifications of chitin, chitosan, and chitooligosaccharides. *Front. Bioeng. Biotechnol.* 7, 243. doi:10.3389/fbioe.2019. 00243

Katada, N., Endo, J. I., Notsu, K. I., Yasunobu, N., Naito, N., and Niwa, M. (2000). Superacidity and catalytic activity of sulfated zirconia. *J. Phys. Chem. B* 104, 10321–10328. doi:10.1021/jp0022120 Kim, H. S., Kim, S. K., and Jeong, G. T. (2018). Efficient conversion of glucosamine to levulinic acid in a sulfamic acid-catalyzed hydrothermal reaction. *RSC Adv.* 8, 3198–3205. doi:10.1039/c7ra12980g

Kobayashi, H., Suzuki, Y., Sagawa, T., Kuroki, K., Hasegawa, J. Y., and Fukuoka, A. (2021). Impact of tensile and compressive forces on the hydrolysis of cellulose and chitin. *Phys. Chem. Chem. Phys.* 23, 15908–15916. doi:10.1039/d1cp01650d

Kougioumtzis, M. A., Marianou, A., Atsonios, K., Michailof, C., Nikolopoulos, N., Koukouzas, N., et al. (2018). Production of 5-HMF from cellulosic biomass: Experimental results and integrated process simulation. *Waste Biomass Valorization* 9, 2433–2445. doi:10.1007/s12649-018-0267-0

Mäki-Arvela, P., Holmbom, B., Salmi, T., and Murzin, D. Y. (2007). Recent progress in synthesis of fine and specialty chemicals from wood and other biomass by heterogeneous catalytic processes. *Catal. Rev.* 49, 197–340. doi:10.1080/01614940701313127

Margoutidis, G., Parsons, V. H., Bottaro, C. S., Yan, N., and Kerton, F. M. (2018). Mechanochemical amorphization of α -chitin and conversion into oligomers of N-acetyl- d -glucosamine. ACS Sustain. Chem. Eng. 6, 1662–1669. doi:10.1021/ acssuschemeng.7b02870

Mariani, M., Zaccheria, F., Psaro, R., and Ravasio, N. (2014). Some insight into the role of different copper species as acids in cellulose deconstruction. *Catal. Commun.* 44, 19–23. doi:10.1016/j.catcom.2013.07.010

Mohan, K., Ganesan, A. R., Ezhilarasi, P. N., Kondamareddy, K. K., Rajan, D. K., Sathishkumar, P., et al. (2022). Green and eco-friendly approaches for the extraction of chitin and chitosan: A review. *Carbohydr. Polym.* 287, 119349. doi:10.1016/j. carbpol.2022.119349

Ohmi, Y., Nishimura, S., and Ebitani, K. (2013). Synthesis of α-amino acids from glucosamine-HCl and its derivatives by aerobic oxidation in water catalyzed by au nanoparticles on basic supports. *ChemSusChem* 6, 2259–2262. doi:10.1002/cssc.201300303

Omari, K. W., Besaw, J. E., and Kerton, F. M. (2012). Hydrolysis of chitosan to yield levulinic acid and 5-hydroxymethylfurfural in water under microwave irradiation. *Green Chem.* 14, 1480–1487. doi:10.1039/c2gc35048c

Osada, M., Kikuta, K., Yoshida, K., Totani, K., Ogata, M., and Usui, T. (2013). Non-catalytic synthesis of Chromogen i and III from N-acetyl-d-glucosamine in high-temperature water. *Green Chem.* 15, 2960–2966. doi:10.1039/c3gc41161c

Padovan, D., Kobayashi, H., and Fukuoka, A. (2020). Facile preparation of 3-Acetamido-5-acetylfuran from N-Acetyl-d-glucosamine by using commercially available aluminum salts. *ChemSusChem* 13, 3594–3598. doi:10.1002/cssc. 202001068

Pandit, A., Deshpande, C., Patil, S., Jain, R., and Dandekar, P. (2020). Mechanistic insights into controlled depolymerization of Chitosan using H- Mordenite. *Carbohydr. Polym.* 230, 115600. doi:10.1016/j.carbpol.2019.115600

Pelckmans, M., Renders, T., Van De Vyver, S., and Sels, B. F. (2017). Bio-based amines through sustainable heterogeneous catalysis. *Green Chem.* 19, 5303–5331. doi:10.1039/c7gc02299a

Sadiq, A. D., Chen, X., Yan, N., and Sperry, J. (2018). Towards the shell biorefinery: Sustainable synthesis of the anticancer alkaloid proximicin A from chitin. *ChemSusChem* 11, 532–535. doi:10.1002/cssc.201702356

Sagawa, T., Kobayashi, H., Murata, C., Shichibu, Y., Konishi, K., and Fukuoka, A. (2019). Catalytic conversion of a chitin-derived sugar alcohol to an amidecontaining isosorbide analog. *ACS Sustain. Chem. Eng.* 7, 14883–14888. doi:10. 1021/acssuschemeng.9b02985

Sanandiya, N. D., Ottenheim, C., Phua, J. W., Caligiani, A., Dritsas, S., and Fernandez, J. G. (2020). Circular manufacturing of chitinous bio-composites via bioconversion of urban refuse. *Sci. Rep.* 10, 4632–4638. doi:10.1038/s41598-020-61664-1

Scotti, N., Zaccheria, F., Evangelisti, C., Psaro, R., and Ravasio, N. (2017). Dehydrogenative coupling promoted by copper catalysts: A way to optimise and upgrade bio-alcohols. *Catal. Sci. Technol.* 7, 1386–1393. doi:10.1039/ c6cy02670b

Wang, Y., Pedersen, C. M., Deng, T., Qiao, Y., and Hou, X. (2013). Direct conversion of chitin biomass to 5-hydroxymethylfurfural in concentrated ZnCl2 aqueous solution. *Bioresour. Technol.* 143, 384–390. doi:10.1016/j.biortech.2013.06.024

Wu, S., Hu, J., Wei, L., Du, Y., Shi, X., and Zhang, L. (2014). Antioxidant and antimicrobial activity of Maillard reaction products from xylan with chitosan/ chitooligomer/glucosamine hydrochloride/taurine model systems. *Food Chem. x.* 148, 196–203. doi:10.1016/j.foodchem.2013.10.044

Xu, B., Du, Z., Dai, J., Yang, R., Yang, D., Gu, X., et al. (2022). Progress in catalytic conversion of renewable chitin biomass to furan-derived platform compounds. *Catalysts* 12, 653. doi:10.3390/catal12060653

Yabushita, M., Kobayashi, H., Kuroki, K., Ito, S., and Fukuoka, A. (2015). Catalytic depolymerization of chitin with retention of N-acetyl group. *ChemSusChem* 8, 3760–3763. doi:10.1002/cssc.201501224

Yan, N., and Chen, X. (2015). Sustainability: Don't waste seafood waste. *Nature* 524, 155–157. doi:10.1038/524155a

Zaccheria, F., Bossola, F., Scotti, N., Evangelisti, C., Dal Santo, V., and Ravasio, N. (2020). On demand production of ethers or alcohols from furfural and HMF by selecting the composition of a Zr/Si catalyst. *Catal. Sci. Technol.* 10, 7502–7511. doi:10.1039/d0cy01427c

Zaccheria, F., Scotti, N., Marelli, M., Psaro, R., and Ravasio, N. (2013). Unravelling the properties of supported copper oxide: Can the particle size induce acidic behaviour? *Dalton Trans.* 42, 1319–1328. doi:10.1039/c2dt32454g

Zhang, H., Lu, Y., Wang, Y., Zhang, X., and Wang, T. (2018). D-Glucosamine production from chitosan hydrolyzation over a glucose-derived solid acid catalyst. *RSC Adv.* 8, 5608–5613. doi:10.1039/c7ra12490b

Zhang, J., and Yan, N. (2017). Production of glucosamine from chitin by Cosolvent promoted hydrolysis and deacetylation. *ChemCatChem* 9, 2790–2796. doi:10.1002/cctc.201601715

Zhang, R., Zhao, Q., Yi, Z., Zhang, K., Shi, J., Zhu, L., et al. (2022). Chitin oligosaccharides for the food industry: Production and applications. *Syst. Microbiol. Biomanuft*. doi:10.1007/s43393-022-00127-2

Zheng, X. Y., Peng, J. B., Livera, M. M. V. S., Luo, Y., Wang, Y. Y., Kong, X. J., et al. (2017). Selective formation of chromogen i from N-Acetyl-D-glucosamine upon lanthanide coordination. *Inorg. Chem.* 56, 110–113. doi:10.1021/acs.inorgchem. 6b02589