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RECEIVED 06 July 2023 ACCEPTED 07 August 2023 PUBLISHED 18 August 2023

CITATION

Peng Y, Ma Y, Luo Z, Jiang Y, Xu Z and Yu R (2023) *Lactobacillus reuteri* in digestive system diseases: focus on clinical trials and mechanisms. *Front. Cell. Infect. Microbiol.* 13:1254198. doi: 10.3389/fcimb.2023.1254198

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Lactobacillus reuteri in digestive system diseases: focus on clinical trials and mechanisms

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Objectives: Digestive system diseases have evolved into a growing global burden without sufficient therapeutic measures. *Lactobacillus reuteri* (*L. reuteri*) is considered as a new potential economical therapy for its probiotic effects in the gastrointestinal system. We have provided an overview of the researches supporting various *L. reuteri* strains' application in treating common digestive system diseases, including infantile colic, diarrhea, constipation, functional abdominal pain, *Helicobacter pylori* infection, inflammatory bowel disease, diverticulitis, colorectal cancer and liver diseases.

Methods: The summarized literature in this review was derived from databases including PubMed, Web of Science, and Google Scholar.

Results: The therapeutic effects of *L. reuteri* in digestive system diseases may depend on various direct and indirect mechanisms, including metabolite production as well as modulation of the intestinal microbiome, preservation of the gut barrier function, and regulation of the host immune system. These actions are largely strain-specific and depend on the activation or inhibition of various certain signal pathways. It is well evidenced that *L. reuteri* can be effective both as a prophylactic measure and as a preferred therapy for infantile colic, and it can also be recommended as an adjuvant strategy to diarrhea, constipation, *Helicobacter pylori* infection in therapeutic settings. While preclinical studies have shown the probiotic potential of *L. reuteri* in the management of functional abdominal pain, inflammatory bowel disease, diverticulitis, colorectal cancer and liver diseases, its application in these disease settings still needs further study.

Conclusion: This review focuses on the probiotic effects of *L. reuteri* on gut homeostasis via certain signaling pathways, and emphasizes the importance of these probiotics as a prospective treatment against several digestive system diseases.

KEYWORDS

Lactobacillus reuteri, gut microbiota, infantile colic, diarrhea, constipation, functional abdominal pain, inflammatory bowel disease, colorectal cancer

1 Introduction

Digestive system diseases arise rapidly through the interaction of genetic and environmental characteristics and are considered to cause considerable healthcare burdens and costs (Peery et al., 2022; Ricciardiello, 2022). Based on the latest data, digestive system diseases are responsible for millions of healthcare encounters and thousands and thousands of deaths that cost billions of dollars in the United States every year (Peery et al., 2022). Of these digestive system diseases, inflammatory bowel disease (IBD) is estimated to affect 790 per 100,000 people in 2025 (Kaplan, 2015). According to the most recent worldwide estimates, there were 930,000 deaths of colorectal cancer (CRC) in 2020 and over 1.9 million new cases (Morgan et al., 2023). The similar phenomenon of increasing incidence is also seen in infantile colic (IC), functional abdominal pain (FAP), and other gastrointestinal (GI) diseases (Thapar et al., 2020; Peery et al., 2022; Indrio et al., 2023). Given this situation, there is an urgent need to deploy personalized treatments for these diseases.

Nowadays public interest of probiotics applications in therapeutic settings is growing. One of the most widely used probiotics, Lactobacillus, can be detected in the GI tract of animals in varying concentrations depending on the species, age of the host, or placement inside the gut (Zhao et al., 2023). The Lactobacillus is a large heterogeneous set of facultative anaerobic bacteria that are nonsporulating, and Gram-positive comprising Lactobacillus (L.) acidophilus, L. reuteri, L. casei, L. bulgaricus, L. rhamnosus, and so on (Mu et al., 2018). Among these, L. reuteri is an extensively investigated probiotic that was first isolated in 1962, resides in tissues of numerous mammals, and provides multiple health benefits for the host including producing antimicrobial molecules and regulating the host immune system (Yu et al., 2023b). L. reuteri was discovered in various body sites, which included breast milk, skin, urinary tract, and GI tract (Yu et al., 2023b). Numerous L. reuteri strains were found to play unique roles in different diseases covering hypercholesterolemia, skin infection, allergic asthma, periodontitis, and autism spectrum disorders as essential probiotics, especially in GI diseases (Sgritta et al., 2019; Kubota et al., 2020; Dargenio et al., 2021). Mounting evidence indicates that the symbiotic gut microbiota and host immune system work together to preserve gut homeostasis (Saeed et al., 2022). Growing data has revealed that the etiology of disorders of the digestive system is strongly influenced by disrupted gut microbes (Quaglio et al., 2022). Various L. reuteri strains have been investigated in digestive system diseases like IC, diarrhea, constipation, FAP, IBD, CRC, as well as liver diseases, and the findings are generally promising (Dias et al., 2021; Saviano et al., 2021; Werlinger et al., 2022; Happy Tummy et al., 2023).

But there are still doubts about clinical application of L. reuteri, so it is important to understand the underlying mechanism of L. reuteri in gut health. The effects of L. reuteri on GI diseases may include preservation of gut barrier function, suppression of excessive immune responses as well as oxidative stress, modification of the composition of the intestinal flora, and production of metabolites. In this review, we discussed the application of *L. reuteri* in digestive system diseases and outlined the potential mechanisms.

2 Clinical application of *L. reuteri* in digestive system diseases

2.1 Infantile colic

IC is typically characterized by daily full-force crying for a minimum of 3 hrs, on a minimum of 3 days per week, for at least 3 weeks (Gordon et al., 2018). It is reported about one in four infants younger than three months develop colic (Indrio et al., 2023). Although IC often resolves by three months of age, it remains to be stressful for parents and may even cause maternal postpartum depressive symptoms (Mercuri et al., 2023). Considerable research indicates that the intestinal microbiome's composition is related to infant colic symptoms, even though the pathophysiology of this digestive condition is still inadequately comprehended (Korpela et al., 2020). Firstly, it has been hypothesized that the developing GI microbiome and the occurrence of excessive crying in infants between the ages of two weeks and three months-the normal time frame during which the gut is typically colonized by bacteria are related (Zeevenhooven et al., 2018). Additionally, the gut microbiota interacts with the gut-brain axis through multiple signaling pathways, including neural, endocrine, immune, and humoral signaling pathways (Deng et al., 2021). Given this, it may influence central and gut neural functions, such as pain perception in infants via the gut-brain axis, which may be associated with excessive crying (Socala et al., 2021). In the past decades, numerous studies illustrated that the GI microbial profiles of infants with colic differ from those without colic in microbial diversity, stability, and community composition (de Weerth et al., 2013; Savino et al., 2017). Moreover, the results showed colicky infants were less frequently colonized by Lactobacillus, Bifidobacterium and more frequently enriched with the gasforming Coliforms (mostly Escherichia, Klebsiella) or other species (Savino et al., 2009; de Weerth et al., 2013; Savino et al., 2017; Sommermeyer et al., 2022; Kozhakhmetov et al., 2023).

We concluded that L. reuteri may be useful in the improvement of breastfeeding babies with IC based on six randomized controlled trials (RCTs), shown in Table 1. Accordingly, administering 1×10^8 colony-forming units (CFU) of L. reuteri DSM 17938 per day for 21 to 30 days can significantly decrease daily crying and fussing times, and reduce the duration of crying episodes in colicky breast-fed infants, consistent with remission of maternal depression (Savino et al., 2010; Szajewska et al., 2013; Chau et al., 2015; Mi et al., 2015; Savino et al., 2018a; Savino et al., 2018b). Similar results were reported when analyzing the probiotic potential of L. reuteri ATCC 55730 in colicky infants, the superiority was still maintained compared with simethicone intervention (Savino et al., 2007). However, whether L. reuteri is useful in the treatment of formulafed colicky infants is still controversial. Data shown in the other two RCT studies demonstrated that L. reuteri did not help manage the symptoms of colicky infants (Sung et al., 2014; Turco et al., 2021).

TABLE 1 Randomized controlled trials of infantile colic.

Year	Intervention	Duration	population	Outcome	Reference
2021	partially hydrolysed formula + maltodextrins+ +DSM 17938 vs standard formula	28 days	Formula-fed colicky infants (<4 months)	significantly lower mean daily crying time in the standard formula group	(Turco et al., 2021)
2018	DSM 17938 vs placebo	1 month	breast-fed colicky infants (<4 months)	significantly shorter crying times, increased circulating FOXP3 concentration, reduced fecal calprotectin in the probiotic group	(Savino et al., 2018b)
2018	DSM 17938 vs placebo	28 days	breast-fed colicky infants (<60 days)	a significant decrease in daily crying time, increased mRNA expression of FoxP3	(Savino et al., 2018a)
2017	DSM 17938 vs placebo	42 days	breast-fed colicky infants (3 weeks to 3 months)	did not significantly change crying time	(Fatheree et al., 2017)
2015	DSM 17938 vs placebo	21 days	breast-fed colicky infants (<60 days)	significantly shorter crying and fussing times in the probiotic group	(Chau et al., 2015)
2015	DSM 17938 and Vit D ₃ vs Vit D ₃	3 months	Newborns (<10 days)	significantly reduced pediatric consultations for infantile colic	(Savino et al., 2015)
2015	DSM 17938 vs placebo	21 days	breast-fed colicky infants (<4 months)	significantly reduced daily crying time, improvement in maternal depression in the probiotic group	(Mi et al., 2015)
2014	DSM 17938 vs placebo	3 months	newborns (<1 week)	significantly reduced duration of crying time	(Indrio et al., 2014)
2014	DSM 17938 vs placebo	1 month	Breast-fed and formula- fed colicky infants (<3 months)	No benefit (increased fussing occurred only in formula fed infants)	(Sung et al., 2014)
2013	DSM 17938 vs placebo	21 days	Breast-fed colicky infants (<5 months)	significantly reduced crying time in the probiotic group	(Szajewska et al., 2013)
2010	DSM 17938 vs placebo	21 days	Breast-fed colicky infants (2 to 16 weeks)	significantly reduced crying time in the probiotic group, a significant increase in fecal lactobacilli and a reduction in fecal Escherichia coli	(Savino et al., 2010)
2007	ATCC 55730 vs simethicone	28 days	Breast-fed colicky infants (21 to 90 days)	significantly reduced daily crying times	(Savino et al., 2007)
2020	DSM 26866 vs placebo	28 days	pregnant women (last 4 weeks of pregnancy)	significantly decreased frequency of colic and lower colic severity in the intervention group	(Pourmirzaiee et al., 2020)

One study showed that a partially hydrolyzed formula added with maltodextrins and L. reuteri DSM 17938 showed no preference for the standard formula on colicky symptoms, but the unsatisfactory results may attribute to the addition of maltodextrins in the probiotics group (Turco et al., 2021). In another study included breastfed and formula-fed colicky newborns, researchers found that L. reuteri DSM 17938 had no beneficial impact on crying or fusing time, instead, more fussing occurred in formula-fed infants following probiotic treatment (Sung et al., 2014). The conflict results may be explained by the differences in gut microbiota, whose composition in early infancy seemed to be influenced by various factors, including the delivery mode, infant feeding pattern, gestational age, and the history of antibiotic use (Kapourchali and Cresci, 2020). Formula-fed babies exhibited an increased gut bacterial community richness relative to breast-fed babies, with a greater abundance of Clostridium difficile (Azad et al., 2013). Following the administration of L. reuteri DSM 17938, breast-fed colicky infants showed a substantial rise in fecal Lactobacilli and a drop in fecal Escherichia (E.) coli (Savino et al., 2010; Savino et al., 2018b). From these findings, the therapeutic effectiveness of L. reuteri could be influenced by the microbiome composition of the infant's GI tract, which would account for the differences between the effect on crying duration in breast-fed and formula-fed infants.

It is also reported that colicky infants showed increased serum concentrations of inflammatory interleukin (IL)-8, monocyte chemoattractant protein-1, and higher fecal calprotectin levels, indicating the existence of systemic and intestinal local inflammation (Partty et al., 2017). L. reuteri DSM 17938 treatment can increase circulating FoxP3 concentration and reduce fecal calprotectin, indicating the underlying mechanisms of L. reuteri to improve colic may be linked to the property of alleviating inflammation (Savino et al., 2018a; Savino et al., 2018b). According to a recent secondary analysis research based on a large observational study, L. reuteri-containing formula was superior to standard formula without probiotics in preventing infant colic, similar to breast feeding (Happy Tummy et al., 2023). Furthermore, the probiotic potential of L. reuteri DSM 17938 given at a dosage of 1×10^8 CFU per day for 90 days to prevent IC is demonstrated by two RCT studies in newborns (Indrio et al., 2014; Savino et al., 2015). This statement was also supported by one prospective cohort study, which showed maternal consumption with L. reuteri DSM 26866 in the latter four weeks of gestation was

able to protect infants from colic (Pourmirzaiee et al., 2020). In conclusion, since *L. reuteri* may ameliorate colic symptoms mainly by altering the microbial colonization pattern in colicky infants, it is a receptive therapy for colicky infants without side effects and recommendable supplements for newborns to prevent IC.

2.2 Diarrhea

Recent data has shown that acute diarrheal disease leads to 179 million outpatient visits every year in the United States, associated with various pathogen infections such as Shigella, E. coli, rotavirus, norovirus and antibiotic use (Meisenheimer Es Md et al., 2022). Diarrhea is also identified as the leading infectious cause of childhood morbidity and mortality (Ahmed et al., 2021). Previously several specific probiotic strains have demonstrated antidiarrheal effects (Skrzydlo-Radomanska et al., 2020; Lukasik et al., 2022). An increasing number of studies have been conducted to clarify the function of L. reuteri in diarrhea since Shornikova and his colleagues first analyzed the effectiveness of L. reuteri DSM 17938 in pediatric acute watery diarrhea and demonstrated that it can dosage-dependently shorten the duration of acute watery diarrhea (Shornikova et al., 1997a; Shornikova et al., 1997b). Previous nine RCTs, which demonstrated that L. reuteri DSM 17938 was capable of reducing the frequency, length of time, and incidence of diarrhea in children and adults, especially in those with lower nutritional status, supported the drug's potential benefits in treating and preventing diarrheal diseases (Agustina et al., 2012; Francavilla et al., 2012; Jones et al., 2013; Dinleyici et al., 2014; Gutierrez-Castrellon et al., 2014; Dinleyici et al., 2015; Pernica et al., 2017; Maragkoudaki et al., 2018; Kambale et al., 2023). Additionally, L. reuteri DSM 17938 could shorten a child's stay in the hospital due to acute gastroenteritis or infectious diarrhea (Dinlevici et al., 2014; Szymanski and Szajewska, 2019). However, in the situation of nosocomial diarrhea, three RCTs produced contrary findings (Wanke and Szajewska, 2012; Urbanska et al., 2016; Kolodziej and Szajewska, 2019). The investigators found that L. reuteri DSM 17938 was ineffective in preventing diarrhea in inpatient children received antibiotics, regardless of the given dosage $(1 \times 10^8$ CFU once daily, or 2×10^8 CFU twice daily, or 2×10^9 CFU once daily while in hospital) (Wanke and Szajewska, 2012; Urbanska et al., 2016; Kolodziej and Szajewska, 2019). While L. reuteri ATCC 55730 was reported to be able to prevent antibioticassociated diarrhea in hospitalized adults in another study (Cimperman et al., 2011). Different from the prior studies, this clinical trial analyzed adult patients who were mainly treated with combined antibiotic therapy, and the subjects were given L. reuteri ATCC 55730 1×10⁸ CFU twice daily for 28 days (Cimperman et al., 2011). Given these limited data, the function of L. reuteri in preventing antibiotic-associated diarrhea is ambiguous, which may be strain-specific and depend on the gut microbiota community disrupted by antibiotics. Of note, none of the mentioned clinical studies analyzed the fecal L. reuteri abundance before or after L. reuteri administration. All these RCT studies are depicted in Table 2.

Enterotoxigenic E. coli is a main pathogen of infectious diarrhea in childhood and postweaning piglets (Kotloff, 2022; Qin et al., 2023b). Gut microbiota disruption observed in diarrheal piglets was described as a reduction of Lactobacillus, enriched abundance of E. coli, and increased lipopolysaccharide (LPS) biosynthesis (Li et al., 2023b). Supplement of L. reuteri can reverse the enriched Pseudomonadota and the depleted Bacteroidota triggered by E. coli infection and reduce jejunum E. coli content, thus preserving intestinal disruption (Wang et al., 2018; Lu et al., 2023). L. reuteri stains including L. reuteri HCM2 and L. reuteri PSC102 have been reported to suppress the growth of E. coli in vitro and inhibit its adherence to intestinal epithelial cells (Wang et al., 2018; Ali et al., 2023). Moreover, L. reuteri was also reported to possess the ability to upregulate the expression of tight junction (TJ) proteins and Ecadherin, thereby maintaining intestinal permeability in infected mice (Karimi et al., 2018; Lu et al., 2023). In a rotavirus-infected murine model, L. reuteri can reduce diarrhea duration and alleviate inflammation, probably associated with accelerated intestinal epithelium turnover and restored gut microbiome diversity (Preidis et al., 2012). Particularly, the efficiency of L. reuteri to increase enterocyte proliferation and migration seemed to be influenced by host nutritional status (Preidis et al., 2012). Except these, the therapeutic application of L. reuteri in infectious diarrhea may also be related to its metabolite production such as reuterin, with the ability to inhibit an extensive range of germs (Collinson et al., 2020).

These results illustrated that the therapeutic efficiency of *L. reuteri* in diarrhea may depend on regulation of gut microbiota and barrier, which is strain-specific and reliant on age, pathogen, nutritional status, and the disease situation as well. To sum up, *L. reuteri* can be recommended as an adjuvant to rehydration treatment for diarrhea in therapeutic settings, but its function in prophylactic settings still requires more research, especially in the antibiotics-use setting.

2.3 Constipation

Constipation is an intestinal condition that could affect individuals of all ages and dramatically lower their quality of life (Saviano et al., 2021). With a prevalence spanning from 7% to 30%, constipation is also one of the most prevalent pediatric disorders (Wegner et al., 2018). The fundamental mechanism of constipation, which typically has a functional cause without an organic origin, is largely unknown (Mulhem et al., 2022). It is suggested that disruption of the intestinal flora play a role in functional constipation (FC) (Zhang et al., 2021). The results may be conflict reliant on measurements. Based on traditional microbiological culture tests, enriched Bifidobacteria and Lactobacillus were observed in children with FC, while decreased abundance of Bifidobacterium and Lactobacillus was seen in adult FC patients (Zoppi et al., 1998; Khalif et al., 2005). Nowadays though gene sequencing analysis, it seems consistent that Firmicutes relative abundance is increased while Bacteroidetes frequency is heterogeneous based on population, and the gut microbe pattern

TABLE 2 Randomized controlled trials of diarrhea and functional constipation.

Year	intervention	symptom	population	Outcome	Reference
2023	DSM 17938+ GG vs placebo for 1 month	diarrhea	Children with uncomplicated severe acute malnutrition	lower number of days of diarrhea lower risk of diarrhea (>16 months)	(Kambale et al., 2023)
2019	DSM 17938 vs placebo for the duration of antibiotic treatment	diarrhea	Hospitalized children received antibiotics	not effective in the prevention of diarrhea	(Kolodziej and Szajewska, 2019)
2019	DSM 17938+ rehydration therapy vs placebo+ rehydration therapy for 5 days	diarrhea	children with acute gastroenteritis (<5 years)	did not reduce the duration of diarrhea a shorter duration of hospitalization	(Szymanski and Szajewska, 2019)
2018	DSM 17938+ oral rehydration salts+ zinc vs placebo + oral rehydration salts+ zinc	diarrhea	non-hospitalized infants with acute diarrhea	without statistical significance but a better trend of the severity and duration of diarrhea	(Maragkoudaki et al., 2018)
2017	DSM 17938 vs placebo for 60 days	diarrhea	hospitalized children with acute diarrhea (2-60 months)	lower odds of recurrent diarrhea	(Pernica et al., 2017)
2016	DSM 17938 vs Placebo	diarrhea	Hospitalized children (1-48 months)	not effective in preventing nosocomial diarrhea	(Urbanska et al., 2016)
2015	DSM 17938 +oral rehydration salts vs oral rehydration salts for 5 days	diarrhea	outpatient children with acute infectious diarrhea	reduced duration of diarrhea	(Dinleyici et al., 2015)
2014	DSM 17938 vs placebo for 5 days	diarrhea	Hospitalized children with acute gastroenteritis	reduced the duration of diarrhea, reduced mean hospital stays	(Dinleyici et al., 2014)
2014	DSM 17938 vs placebo for 3 months	diarrhea	preschool children (6-36 months)	reduced the frequency and duration of episodes of diarrhea	(Gutierrez- Castrellon et al., 2014)
2013	NCIMB 30242 vs Placebo for 9 weeks	diarrhea	healthy hypercholesterolemic subjects	Improved symptoms related to diarrhea	(Jones et al., 2013)
2012	DSM 17938 vs Placebo	diarrhea	Hospitalized children (1-48 months)	no effect on the overall incidence of nosocomial diarrhea	(Wanke and Szajewska, 2012)
2012	DSM 17938 vs Placebo	diarrhea	Healthy children (1-6 years)	reduced diarrhea incidence in children with lower nutritional status	(Agustina et al., 2012)
2012	DSM 17938+oral rehydration salts vs placebo + oral rehydration salts	diarrhea	Hospitalized children with acute diarrhea (6-36 months)	reduced the duration of watery diarrhea	(Francavilla et al., 2012)
2011	ATCC 55730 vs placebo for 4 weeks	diarrhea	hospitalized adults	lower frequency of antibiotic-associated diarrhea	(Cimperman et al., 2011)
2020	DSM 17938 vs placebo	functional constipation	Children with cerebral palsy and chronic constipation	a significant decrease in stool pH and increased defecation, improvement in the history of excessive stool retention, the presence of fecal mass in the rectum, painful defecation,	(Garcia Contreras et al., 2020)
2020	DSM 17938 + magnesium oxide vs DSM 17938+ placebo vs placebo+ magnesium oxide for 4 weeks	functional constipation	Children with functional constipation (6 months -6 years)	significant improvement in defecation frequency in DSM 17938 group	(Kubota et al., 2020)
2019	DSM 17938 vs placebo for 105 days	functional constipation	adults with functional constipation	significantly reduce the serum levels of 5-HT	(Riezzo et al., 2019)
2018	DSM 17938 + macrogol vs placebo + macrogol for 8 weeks	functional constipation	Children with functional constipation	No significant difference	(Wegner et al., 2018)

(Continued)

TABLE 2 Continued

Year	intervention	symptom	population	Outcome	Reference
2018	DSM 17938 vs placebo for 105 days	functional constipation	Adults with functional constipation	helps for defecation, improved abdominal discomfort, pain and bloating	(Riezzo et al., 2018)
2018	DSM 17938 + lactulose vs placebo + lactulose	functional constipation	Children with functional constipation	no difference in the stool frequency, stool consistency, pain	(Jadresin et al., 2018)
2014	DSM 17938 vs placebo for 4 weeks	functional constipation	Adults with functional constipation	increased bowel movements	(Ojetti et al., 2014)
2010	DSM 17938 vs placebo for 8 weeks	functional constipation	Infants with functional constipation (> 6 months)	higher frequency of bowel movements, no significant difference in stool consistency	(Coccorullo et al., 2010)

differs in different subtypes of constipation as well (Zhu et al., 2014; Mancabelli et al., 2017; Yu et al., 2023a).

A couple of RCTs have evaluated the effectiveness of *L. reuteri* DSM 17938 in treating FC, depicted in Table 2. The investigations demonstrated that *L. reuteri* DSM 17938 single-drug administration significantly alleviated constipation both in children and adults, involving improvement in defecation frequency, painful defecation, and reduced fecal mass in the rectum (Coccorullo et al., 2010; Ojetti et al., 2014; Riezzo et al., 2018; Garcia Contreras et al., 2020; Kubota et al., 2020). However, *L. reuteri* DSM 17938 did not show additional improvement in FC when regarded as adjuvant therapy for lactulose or macrogol (Jadresin et al., 2018; Wegner et al., 2018; Kubota et al., 2020). Only one study investigated the gut microbiome community alternations, in which the limited data revealed that *L. reuteri* treatment did not alter the gut microbiota composition of the pediatric subjects with FC (Kubota et al., 2020).

To note, current evidence showed that L. reuteri DSM 17938 improved bowel movements, but did not affect stool consistency (Coccorullo et al., 2010; Kubota et al., 2020). The defecation frequency was found to be negatively correlated with genus Oscillospira, Megasphaera, and Ruminococcus (Kubota et al., 2020). On one hand, L. reuteri DSM 17938 can improve intestinal motility and promote intestinal transit, partially via controlling some GI peptide pathways. According to research by Riezzo and colleagues, the therapeutic efficacy of L. reuteri DSM 17938 in FC may be associated with serum brain-derived neurotrophic factor and serotonin (5-HT) concentrations (Riezzo et al., 2019). Tryptophan metabolism may be impacted by L. reuteri, which would reduce the level of 5-HT in circulation (Montgomery et al., 2022). The connection between the intestinal flora and the enteric neural systems involves the 5-HT pathways, which have advantages for irritating the local enteric nerve responses and promoting motility (Ge et al., 2017; Agus et al., 2018). On the other hand, L. reuteri DSM 17938 is capable of producing short-chain fatty acids (SCFAs), the substances involved in boosting intestinal peristalsis and colonic myoelectric cell response, all of which are effective for treating chronic FC (Wu et al., 2013; Hurst et al., 2014; Calvigioni et al., 2023).

However, there is still no sufficient data in the field of *L. reuteri* application for constipation prevention. A prospective RCT study carried out by Indrio and colleagues revealed that oral supplementation with *L. reuteri* DSM 1793 in newborns effectively prevented constipation within the first 3 months after birth (Indrio et al., 2014). In conclusion, present findings indicate that *L. reuteri* can be recommended to be used in the management of FC as an alternative therapy for traditional drugs.

2.4 Functional abdominal pain

FAP, which is the most prevalent type of abdominal pain in children and teenagers, with a prevalence of about 14 percent, is described as chronic or recurring pain without an organic cause (Bao et al., 2023). Although most FAP is often self-managed, it can have adverse effects on life quality, suffered children's after-school activities, and school attendance, and may interfere with regular family life (Thapar et al., 2020). The disease was currently considered a disorder of the "brain-gut axis", and gut microbiome changes could be engaged in its pathogenesis, but the available data is insufficient to draw a conclusion (Zhou et al., 2022a). Given the limitations of medication in children, probiotics arise as a possibly pleasant therapeutic approach for the treatment of pediatric FAP (Trivic et al., 2021). Six RCT investigations on L. reuteri DSM 17938's medicinal potential are summarized in Table 3. The studies demonstrated that administration of L. reuteri DSM 17938 for 4 to 12 weeks reduced the pain intensity, the frequency of episodes, and increased days without pain in irritable bowel syndrome or FAPaffected children without adverse events (Romano et al., 2014; Eftekhari et al., 2015; Weizman et al., 2016; Jadresin et al., 2017; Maragkoudaki et al., 2017; Jadresin et al., 2020). However, one study proposed that L. reuteri DSM 17938 was equally effective in relieving pain in both the placebo and L. reuteri groups of children with FAP (Eftekhari et al., 2015). The authors speculated that the result may be explained by psychological effects since L. reuteri DSM 17938 showed no superiority to placebo (Eftekhari et al., 2015). None of the studies explored gut flora profiles after L.

Year	intervention	population	Outcome	References
2020	DSM 17938 vs placebo for 12 weeks	Children with functional abdominal pain (4-18 years)	significant increased days without pain, reduced intensity of pain (follow up of 4weeks)	(Jadresin et al., 2020)
2017	DSM 17938 vs placebo for 12 weeks	Children with functional abdominal pain or irritable bowel syndrome (4-18 years)	significantly reduced severity of abdominal pain, increased days without pain (follow-up of 4weeks)	(Jadresin et al., 2017)
2017	DSM 17938 vs placebo for 4 weeks	Children with functional abdominal pain (5-16 years)	reduced the frequency and intensity of abdominal pain episodes (follow-up of 8weeks)	(Maragkoudaki et al., 2017)
2016	DSM 17938 vs placebo for 4 weeks	Children with functional abdominal pain (6-15 years)	relieving frequency and intensity of functional abdominal pain (follow-up of 4weeks)	(Weizman et al., 2016)
2015	DSM 17938 vs placebo for 4 weeks	Children with functional abdominal pain (4-16 years)	the pain was significantly reduced in both groups, but no significant difference compared with placebo (follow-up of 8weeks)	(Eftekhari et al., 2015)
2014	DSM 17938 vs placebo for 4 weeks	Children with functional abdominal pain (6-16 years)	lower pain intensity (follow-up of 4 weeks)	(Romano et al., 2014)

TABLE 3 Randomized controlled trials of functional abdominal pain.

reuteri treatment, so we cannot draw a conclusion about the precise mechanism of *L. reuteri* application in pediatric FAP.

By enhancing opioid receptor activity in the afflicted tissues, Hegde and his colleagues reported that the dorsal root ganglia neurons projecting to the dilated colon exhibited attenuated hyperexcitability after *L. reuteri* recolonization, thus preventing visceral hypersensitivity in lumen distension in rats (Hegde et al., 2020). Additionally, *L. reuteri* may exhibit the ability to reduce gut pain by reducing signaling from the pain receptor transient receptor potential vanilloid 1 channel in the intestine (Pang et al., 2022). Collectively, although *L. reuteri* could be an attractive choice in treating FAP-related diseases due to its safety feature, there is currently no powerful proof to recommend *L. reuteri* application in FAP and further long-run researches are still needed to explore the definite mechanism.

2.5 Helicobacter pylori infection

Helicobacter pylori (H. pylori) is the key pathogen of chronic active gastritis, peptic and duodenal ulcers, with the global prevalence of infection exceeding 50%, also considered a high-risk factor for gastric cancer (de Brito et al., 2019). Currently, the effectiveness of treatment has been greatly impaired by the fast global emergence of antibiotic resistance to H. pylori, and the quadruple approach for H. pylori elimination could further exacerbate GI disease (Chen et al., 2021). It is suggested that eradication of H. pylori may lead to disruption of gut microbiota, featured by reduced Actinobacteria as well as Bacteroidetes abundances and enriched Proteobacteria (Sitkin et al., 2022). Specific probiotics may be helpful in reducing side effects triggered by first-line antimicrobial therapy and enhancing the efficacy of H. pylori eradication therapy (Liang et al., 2022). In previous RCT studies, L. reuteri was shown to reduce H. pylori load but seemed to be unable to increase eradication rates of H. pylori as an adjuvant for first-line therapy (Imase et al., 2007; Francavilla et al., 2008; Mehling and Busjahn, 2013; Francavilla et al., 2014; Holz et al., 2015; Dore et al., 2019; Poonvam et al., 2019; Yang et al., 2021; Dore et al., 2022; Moreno Marquez et al., 2022; Naghibzadeh et al., 2022), shown in Table 4. Notably, several studies showed that L. reuteri can reduce abdominal pain, distension, and other eradication treatment-related adverse events, consistent with decreased GI symptom rating scale scores in H. pylori-positive subjects, thus governing patient compliance (Lionetti et al., 2006; Francavilla et al., 2008; Francavilla et al., 2014; Poonyam et al., 2019; Yang et al., 2021; Moreno Marquez et al., 2022). Two studies analyzed the alternations of the gut flora after combined therapy of this probiotics and eradication treatment, which revealed that L. reuteri can change the gut microbiota composition in H. pylori-positive patients but cannot fully counteract gut dysbiosis induced by antibiotics (Yang et al., 2021; Dore et al., 2022). It means that the benefits brought by L. reuteri supplement cannot be attributed to restored gut microbiota balance. Particularly, L. reuteri can survive the gastric acid environment and colonize the gastric mucosa (Dargenio et al., 2021). It is reported that L. reuteri can inhibit the early colonization stages of H. pylori in the human GI tract, associated with the suppression of the binding of H. pylori to putative glycolipid receptor molecules (Holz et al., 2015). Besides, L. reuteri produces reuterin, an antibiotic that targets H. pylori, thus reducing H. pylori load (Urrutia-Baca et al., 2018). Meanwhile, a most recent survey also revealed that L. reuteri 2892 attenuated H. pylori-induced gastritis by its anti-inflammatory and anti-oxidative stress characteristics as well as suppressing the gene expression of the virulence factor CagA (Forooghi Nia et al., 2023). Recent studies also focus on the relationship between L. reuteri and cancer diseases, it has been demonstrated that L. reuteri exerts an anti-tumor effect in gastric cancer MKN1 cells, but data is still limited about L. reuteri application in gastric cancers related to H. pylori infection (Kim et al., 2022a).

Overall, *L. reuteri* has shown superiority as a salvageable treatment for side effects brought by eradication therapy, yet we still need further studies to delve deeper into the research gap between *L. reuteri* application and *H. pylori*-associated cancer.

Year	intervention	population	Outcome	References
2022	quadruple therapy+ S. boulardii vs quadruple therapy+ DSMZ 17648 vs quadruple therapy	H. pylori- positive adults	no significant difference in the eradication of H. pylori in <i>L. reuteri</i> group	(Naghibzadeh et al., 2022)
2022	quadruple therapy + DSM 17938 and ATCC PTA 6475 vs quadruple therapy	H. pylori- positive adults	no significant difference in the eradication of H. pylori	(Dore et al., 2022)
2022	eradication regimen+ DSM 17938 and ATCC 6475 vs eradication regimen	H. pylori- positive adults	no differences in eradication therapy, reduced abdominal pain and distension	(Moreno Marquez et al., 2022)
2021	non-viable DSM17648+triple therapy vs placebo + triple therapy	H. pylori- positive adults	did not improve the eradication rate of H. pylori, reduced abdominal distention, diarrhea, and the Gastrointestinal Symptom Rating Scale score.	(Yang et al., 2021)
2019	quadruple therapy+ Biogaia [®] vs quadruple H. pylori- therapy H. pylori- positive adults did not increase eradication rates, reduced treatment-related adverse events and improve the patients' compliance		(Poonyam et al., 2019)	
2019	quadruple therapy + DSM 17938 and ATC 6475 vs quadruple therapy	H. pylori- positive adults		
2015	non-viable DSM17648 vs placebo	H. pylori- positive adults	reduce the load of H. pylori	(Holz et al., 2015)
2014	eradication regimen+ DSM 17938 and ATCC 6475 vs eradication regimen	H. pylori- positive adults	reduced load of H. pylori, fewer side effects, decreased Gastrointestinal Symptom Rating Scale scores, no difference in the H. pylori-eradication rate	(Francavilla et al., 2014)
2013	non-viable DSMZ17648 vs placebo	H. pylori- positive asymptomatic adults	reduced load of H. pylori	(Mehling and Busjahn, 2013)
2008	ATCC 55730+ sequential conventional treatment vs sequential conventional treatment	H. pylori- positive adults	reduced H. pylori load, decreased Gastrointestinal Symptom Rating Scale scores, no difference in eradication rates	(Francavilla et al., 2008)
2007	SD2112 vs placebo	H. pylori- positive adults	reduced load of H. pylori	(Imase et al., 2007)
2006	sequential therapy + ATCC 55730 vs sequential therapy	H. pylori- positive children	reduced frequency and intensity of antibiotic-associated side effects, decreased Gastrointestinal Symptom Rating Scale scores	(Lionetti et al., 2006)

TABLE 4 Randomized controlled trials of H. pylori infection.

2.6 Inflammatory bowel disease and diverticulitis

Crohn's disease and ulcerative colitis are categorized as chronic IBD, which have evolved into a worldwide burden with increasing incidence (Nishikawa et al., 2021). It is reported that about one in five children needing a colectomy during childhood while the intestinal damage and complications brought by Crohn's disease are still a difficulty for disease management (Cheng et al., 2021; Gordon et al., 2022). Multiple factors, such as the interaction between genetic background and environment, especially including a potential association between impaired gut microbial homeostasis and inflammation, and gut barrier disruption as well, have been suggested to be engaged in IBD pathogenesis (Larabi et al., 2020). It was observed in IBD subjects that the relative abundance of Proteobacteria, especially E. coli, was increased while the content of Bacteroidetes and Firmicutes was depleted, along with the reduced gut microbiota diversity (Nishino et al., 2018; Vester-Andersen et al., 2019). It has been revealed in a previous clinical trial, whereby, rectal infusion of L. reuteri ATCC 55730 effectively improved inflamed mucous membranes and reduced mucosal expression levels of inflammatory markers in active distal ulcerative colitis in children (Oliva et al., 2012), shown in Table 5.

According to Liu and his colleagues, peroral treatment with L. reuteri ATCC PTA 4659 improved colitis severity clinically and morphologically in mouse colitis caused by dextran sulfate sodium (Liu et al., 2022a). Other studies investigated the efficiency of L. reuteri R2LC, ATCC PTA 4659, F-9-35, and 5454, these strains also can alleviate inflammation in mice colitis, characterized by downregulated pro-inflammatory tumor necrosis factor-a (TNF- α), IL-1 β , and interferon- γ , the decisive cytokines in colitis development (Ahl et al., 2016; Sun et al., 2018; Hrdy et al., 2020). This may be explained by the boosted expression of the TJ proteins along with cytoprotective heat shock proteins after L. reuteri therapy, thereby conferring protection against defects in gut barrier function (Ahl et al., 2016; Liu et al., 2022a). Meanwhile, L. reuteri supplement can also reverse gut microbiota dysbiosis induced by colitis. It was reported that after L. reuteri FYNDL13 treatment, enriched beneficial bacteria (Bifidobacterium, Akkermansia, Blautia and Oscillospira) together with reduced harmful bacteria (Bacteroides and Sutterella) content was observed (Lin et al., 2023). Moreover, L. reuteri therapy can offset

TABLE 5	Randomized	controlled	trials of	colitis	and	diverticulitis.
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Year	intervention	symptom	population	Outcome	References
2022	ATCC PTA 4659+ fluids+bowel rest vs placebo+ fluids+bowel rest	abdominal pain, inflammatory markers and reduction of hours of hospitalization.	patients affected by acute uncomplicated diverticulitis	Decreased inflammatory markers	(Ojetti et al., 2022)
2019	ATCC PTA 4659+ ciprofloxacin +metronidazole vs placebo+ciprofloxacin +metronidazole	abdominal pain, inflammatory markers and reduction of hours of hospitalization.	patients affected by acute uncomplicated diverticulitis	reduced abdominal pain and inflammatory markers	(Petruzziello et al., 2019)
2012	ATCC 55730+mesalazine vs placebo +mesalazine	inflammation and cytokine expression of rectal mucosa	in children with active distal ulcerative colitis	decreased mucosal inflammation	(Oliva et al., 2012)

bacterial translocation to the mesenteric lymph nodes caused by colitis, leading to the amelioration of excessive immune reactions (Liu et al., 2022a). Secretory immunoglobulin (Ig) A is responsible for bacterial translocation and bacterial toxins neutralization in the intestinal mucosa (Tezuka and Ohteki, 2019). Recently, it was discovered that orally administered L. reuteri R2LC stimulated the proliferation of the B lymphocytes in Peyer's patches and induced IgA production and secretion to the intestinal lumen via the PD-1 pathway (Liu et al., 2021a). Consequently, treatment based on L. reuteri R2LC regulated alterations in the gut microbiome and shielded against colitis caused by dextransulfate-sodium (Liu et al., 2021a). Dendritic cells (DCs) are responsible for handling pathogens when the gut barriers are disrupted in IBD (Wei et al., 2022). L. reuteri was proven to possess the potential to promote DCs differentiation and maturation and boost regulatory T cells (Tregs) induction (Haileselassie et al., 2016; Hrdy et al., 2020). In addition, it is demonstrated that L. reuteri may reduce intestine inflammation by the mechanisms dependent on aryl hydrocarbon receptor activation, an immune and bacteria sensor receptor, correlated to inflammatory responses in IBD pathogenesis (Pernomian et al., 2020). The tryptophan metabolites from L. reuteri can activate aryl hydrocarbon receptors, inducing IL-22 production to maintain mucosal protection (Zelante et al., 2013). And the microbial histamine produced by L. reuteri has been proven to ameliorate intestinal inflammation in mice via activating H2R signaling (Gao et al., 2015). It was also reported that LPxTG-motif surface protein derived from L. reuteri SH 23 can alleviate inflammation in a mice colitis model via the inhibition of the MAPK-dependent NF-KB transduction way (Zong et al., 2023).

Colon mucosa and submucosa may inflame and herniate through the muscular layer in cases of diverticulitis (Barbaro et al., 2022). Diet and lifestyle factors may induce changes in the gut microbiota community that contributes to mucosal inflammation and diverticulitis (Piccioni et al., 2021). Probiotics could be beneficial in diverticular disease because they can prevent harmful bacteria from adhering to the intestinal mucosa and from producing inflammatory markers (Piccioni et al., 2021). Two previous RCTs examined *L. reuteri*'s function in managing acute uncomplicated diverticulitis (AUD) in light of recent recommendations that the condition be treated without the use of antibiotics (Petruzziello et al., 2019; Ojetti et al., 2022). They revealed that the treatment with *L. reuteri* ATCC PTA 4659 can alleviate abdominal pain and reduce inflammatory markers as well as the duration of hospitalization in patients with AUD (Petruzziello et al., 2019; Ojetti et al., 2022), shown in Table 5.

Therefore, the above preclinical studies validate the therapeutic potential of specific strains of *L. reuteri* to enhance the natural defense of gut epitheliums, alleviate hyperimmune response and restore gut microecology balance in inflammatory intestinal diseases. Although clinical data are limited, *L. reuteri* can be an attractive adjuvant therapy for IBD and diverticulitis.

2.7 Colorectal cancer

As per global statistics, CRC is ranked third as the most prevalent type of malignancy and the fourth major contributor to deaths related to cancer worldwide (Baidoun et al., 2021). As is known to all, IBD is linked to the onset and progression of CRC (Mathey et al., 2021). Dysplasia, clonal proliferation, and malignant progression might all be caused by gut dysbacteriosis, which may also have dramatic effects on genetics and epigenetics (Eslami et al., 2019). Previously, researchers identified gut flora dysbiosis in patients with CRC, including the depletion of *L. gallinarum*, *Streptococcus thermophilus* and enriched *Firmicutes* and *Bacteroidetes*, along with decreased bacterial diversity (Dai et al., 2018). Reduced abundance of *Lactobacillus*, *Roseburia* and *Bifidobacterium* in gut microbiota composition was observed in early precursors of CRC including adenoma and serrated polyp (Rezasoltani et al., 2018).

Although the prognosis of CRC patients without local or distant metastatic disease is generally favorable, there are currently inadequate viable therapies for individuals with metastatic cancer (Bell et al., 2022). Accordingly, probiotic therapy has recently gained popularity as a promising CRC therapeutic approach. Gao and colleagues demonstrated that treatment with (histidine decarboxylase) HDC⁺ L. reuteri can induce luminal histamine production, consequently, decreased colon tumor frequency and size were observed in HDC^{-/-} mice (Gao et al., 2017). Moreover, they discovered that an L. reuteri mutant with isogenic HDC deficiency that cannot generate histamine did not inhibit tumorigenesis, highlighting the essential function of histamine in preventing inflammatory responses and the development of colorectal tumors (Gao et al., 2017). Reuterin, another major compound produced by L. reuteri, was found to be downregulated in mice and human CRC (Bell et al., 2022). Consistently, reuterin has been shown to have an inhibitory

impact on the proliferative rate of CRC cell lines such as RKO at a concentration of 25µM, but not at a higher dosage of 100µM on normal colonic epithelium (Yu et al., 2023b). Indole-3-lactic acid, a tryptophan catabolite that inhibits the IL-17 signaling pathway, was shown in another recent study to be a key player in the suppression of colorectal carcinogenesis following L. reuteri treatment (Han et al., 2023). RAR-related orphan receptor γt, a nuclear receptor, was the target of this microbial metabolite product, which prevented Th17 cell development (Han et al., 2023). The potential underlying mechanism of L. reuteri's pro-apoptotic actions was investigated but still needs further investigation. L. reuteri MG5346 was reported to inhibit tumor growth by inducing cell apoptosis though upregulated caspase-9 activity (Kim et al., 2022b). Additionally, it has been demonstrated that sirtuin-3-L. reuteri interaction is essential for the development of gut tumors (Zhang et al., 2018). Sirtuin-3 is a tumor-suppressing gene and L. reuteri can inhibit the downregulation of Sirt3 expression during colorectal tumorigenesis (Zhang et al., 2018). A previous meta-analysis analyzed the relationship between intestine E-cadherin and the prognosis of CRC, they concluded that low expression of Ecadherin is a risk factor for poor prognosis in CRC sufferers (Chang et al., 2022). It has been evidenced that L. reuteri can boost E-cadherin expression, indicating a possible mechanism for treating CRC (Karimi et al., 2018).

In summary, considering these evidence, *L. reuteri* owns the ability to suppress intestinal tumor progression in direct and indirect ways, thereby it may develop into a potential adjuvant treatment, but it is premature to recommend widespread use of *L. reuteri* in CRC due to insufficient clinical data.

2.8 Liver diseases

The total cirrhosis incidence is growing and estimated to reach 112.1/100,000 person-years in 2040, largely attributed to the increase in non-alcoholic fatty liver disease and alcohol-related liver disease (Flemming et al., 2021). The liver is a central immunological organ, which is frequently exposed to metabolites derived from the intestinal microbiome, thus it is not surprising that the GI microbial community also plays a part in the pathologic process of liver diseases (Jones and Neish, 2021; Wang et al., 2021a). Particularly, gut microbiota dysbiosis may already exist at the early period of liver damage. Studies have shown alcohol-dependent subjects with liver disorder show compositional changes in the fecal microbiota, presented as a reduction in bacterial diversity and decreased relative abundance of Lactobacillus, Bifidobacterium, as well as Akkermansia muciniphila (Leclercq et al., 2014; Grander et al., 2018). The phenotype of ethanolelicited hepatitis was reversed by L. reuteri, partially by regulating fatty acid metabolic pathways in mice alcoholic liver disease models (Zheng et al., 2020). It was also reported that L. reuteri MJM60668 supplementation to mice fed a high-fat diet ameliorated hepatic steatosis by inhibiting lipogenesis, which may be attributed to the improvement of the gut microbiome dysbiosis (Werlinger et al., 2022). Chen and colleagues reported that gut microbiota imbalance promoted liver tumorigenesis and administration with L. reuteri can improve tryptophan metabolism and reduce hepatic sterol regulatory

element-binding protein 2 expression, thus suppressing tumorigenesis (Chen et al., 2022). L. reuteri was also reported to reduce IL-17A secretion of group 3 innate lymphoid cells (ILC3s) in liver by upregulating acetate levels, thus exerting anti-tumor effects in mice with hepatocellular carcinoma (Hu et al., 2023). These evidences revealed the interaction between gut microbiome and hepatic immune response as well as metabolite signal pathways. Hence the use of L. reuteri in managing liver disorders has drawn a lot of attention from researchers, although RCT studies on the bacteria's protective effects for patients with hepatic ailments are still inadequate. In one RCT research, Ferolla et al. discovered that a 3-months supplementation of L. reuteri with guar gum as well as inulin reduced hepatic steatosis in subjects with nonalcoholic steatohepatitis (Ferolla et al., 2016). However, another RCT study carried out in 2017 showed that in obese patients with type 2 diabetes, oral dosing of L. reuteri DSM 17938 for 12 weeks seemed to have no impact on the microbiota composition, adiposity, or liver steatosis, although, in some subjects with higher gut microbiota diversity before intervention, it increased insulin sensitivity (Mobini et al., 2017). The contrary outcomes may be attributed to the of microbial diversity in complicated metabolic disease conditions and the crosstalk caused by obesity. Taken together, these preclinical literatures provide evidence that L. reuteri may relieve liver injury in several signaling pathways probably by reversing gut microbiota dysbiosis. Given this, it may act as a potential probiotic for the prevention or therapeutic strategy for liver diseases, but further long-term well-designed clinical trials are still needed

3 Potential *L. reuteri* mechanisms in digestive system diseases

As illustrated in Figure 1, growing preclinical investigations revealed that *L. reuteri* restores gut microbiota balance, products antimicrobial metabolites, regulates intestinal immunity and mediates mucosal homeostasis, thereby exhibiting protective effects on digestive system diseases.

3.1 Remodeling the gut microbiota

It is well-documented that *L. reuteri* can alter the bacterial diversity and abundance of various microorganisms, largely strain-specific. For instance, administration with *L. reuteri* LR6 and KT260178 both resulted in higher counts of total *Lactobacillus*, *Bifidobacterium* (Garg et al., 2020; Yang et al., 2020). While *L. reuteri* DSM 17938 supplement improved bacterial diversity along with enriched *Firmicutes* and reduced abundance of *Bacteroidetes* (Liu et al., 2019). Similar results have been observed in disease conditions. After supplement with *L. reuteri* DSM 17938, investigators observed enriched fecal *Lactobacilli* and a decrease in the colicky infants' fecal *E. coli* counts (Savino et al., 2010; Savino et al., 2018b). Consistently, *L. reuteri* ATCC PTA 4659 peroral therapy alleviated mice colitis by maintaining the diversity of the colon's microbiome (Liu et al., 2022a).



The beneficial properties of L. reuteri on gut microflora dysbiosis may be partially explained by the competition of L. reuteri and these harmful genera for the intestinal epithelium binding sites (Walsham et al., 2016). Walsham et al. reported that L. reuteri can bind to the epithelial cell surface by mucus-binding proteins CmbA and MUB, thus inhibiting enteropathogenic E. coli adherence to the gut epithelium (Walsham et al., 2016). Bioinformatics analysis further revealed that the inhibition mechanism may be associated with PI3K-Akt and MAPK signaling pathways (Qin et al., 2023b). Furthermore, L. reuteri may inhibit harmful bacteria by the production of antimicrobial metabolites like reuterin (Asare et al., 2020). The probiotic management of the gastrointestinal microbiota may also be mediated by regulating the intestinal pH through lactic acid production as well as some other antimicrobial substances like acetic acid and ethanol of some L. reuteri strains (Vitale et al., 2023). L. reuteri is demonstrated to be effective against a range of GI microbial infections, including Salmonella, Enterobacteriaceae, and others, partially depending on the synthesis of these compounds

(Kao et al., 2020; Laosee et al., 2022; Qin et al., 2023a). In mice with colitis caused by dextran-sulfate-sodium, dosing of *L. reuteri* R2LC may remodel the makeup of the gut microbiome via enhanced intestinal IgA production (Liu et al., 2021a). It has been demonstrated that IgA is responsible for the selection of intestinal colonized bacteria and tends to resist pathogenic bacteria (Schnupf et al., 2018).

Overall, *L. reuteri* is shown to modulate the gut microbial community, characterized by enriched beneficial genera and diminished pathogenetic genera. Nonetheless, further research is required to determine the interaction between GI health and the modulated gut microbiota.

3.2 Production of antimicrobial metabolites

With the development of metabolomics and bacterial genetics strategy, small compounds produced by *L. reuteri* were identified. It has been proposed that the metabolite products of *L. reuteri* strains

are responsible for their antibacterial and immunoregulatory properties. We covered a few well-known metabolites associated with *L. reuteri's* probiotic potential in this section.

3.2.1 Reuterin

Reuterin is a broad-spectrum antimicrobial compound generated by specific strains of L. reuteri during the anaerobic metabolism of glycerol (Asare et al., 2020). This antimicrobial compound, which is a blend of different 3-hydroxypropionaldehyde (3-HPA) forms, may suppress the proliferation of both Gram-positive and -negative bacteria, protozoa, and fungi (Asare et al., 2020; Saviano et al., 2021). Apart from being used as a food preservative product, reuterin was also identified as the most inhibitory compound in CRC cell lines (Bell et al., 2022). Recently, it was shown that reuterin can suppress tumor growth in the mouse model implanted with CRC xenograft tumors by inducing oxidative stress (Bell et al., 2022). The authors showed that reuterin can inhibit the growth of CRC cell lines with a dose of 25µM but the concentration of reuterin at a dose of 100µM seemed not cytotoxic to non-cancerous colon cell lines (Bell et al., 2022). The molecular mechanism might be explained by that reuterin can increase reactive oxygen species and thus reduce ribosomal biogenesis by selectively binding to cysteine residues, accompanied by increased gene expression downstream of nuclear factor erythroid-derived 2 (Nrf-2) (Bell et al., 2022). However, it was also revealed in vitro experiments that reuterin treatment at a dose of 250µM significantly suppressed LPS-induced oxidative stress by enhancing Nrf-2 expression and reducing reactive oxygen species production in HD11 macrophage cells (Xu et al., 2022). These data revealed that the effects of reuterin depend on certain cell types, reuterin concentration, and stimulation time.

3.2.2 Histamine

Biogenic amines, especially histamine, were reported to be able to modulate the host immune system (Dvornikova et al., 2023). Several strains of L. reuteri such as L. reuteri DSM 20016 are capable of producing histamine from the dietary component L-histidine, an amino acid (Smythe and Efthimiou, 2022). According to Lin and his colleagues, L. reuteri ATCC PTA 6475 inhibited the MAPK signaling pathway, reducing the generation of TNF in monocytederived macrophages isolated from children with Crohn's disease (Lin et al., 2008). By using mass spectrometry, investigators further proved that the key substance of L. reuteri ATCC PTA 6475-mediated TNF inhibition was histamine (Thomas et al., 2012). Histamine's pleiotropic actions are achieved by activating the H1R, H2R, H3R, and H4R histamine receptors present in mammalian cells (Shulpekova et al., 2021). Via activation of H2R on human monocytoid cell line THP-1 cells, histamine stimulated increased levels of cyclic adenosine monophosphate (cAMP) and improved protein kinase A activity, thus inhibiting MEK/ERK MAPK signaling and alleviating inflammation (Thomas et al., 2012). Consistently, Gao and his colleagues discovered that oral administration of HDC+ L. reuteri capable of converting Lhistidine to histamine might successfully alleviate inflammatory reactions in a murine model of colitis (Gao et al., 2015). Subsequently, they found that by causing immature myeloid cells to aggregate, HDC deficiency can accelerate the development of CRC that is linked to inflammation, hinting that histamine may have antitumorigenic properties (Gao et al., 2017). H1R, H2R, and, to a less extent, H4R are expressed by myeloid cells in the GI tract (Sander et al., 2006). Consequently, histamine may be involved in intestinal immunomodulation and inflammation by mediating myeloid cells (Sander et al., 2006). Besides, it is reported that improved overall survival outcomes in patients with CRC were correlated with elevated gene expression of H2R and decreased gene expression of H1R in the intestinal mucosa (Shi et al., 2019). The study further demonstrated that histamine-producing *L. reuteri* can suppress tumorigenesis in an established CRC mouse model but compared to H1R antagonists, H2R antagonists dramatically increased both the size and number of tumors (Shi et al., 2019). Given these data, different receptors of histamine may play distinct roles in the gut.

3.2.3 Exopolysaccharide

Exopolysaccharide (EPS), a substance generated by *L. reuteri*, is essential for the development of biofilms and *L. reuteri*'s adhesion to surfaces of the epithelium (Wang et al., 2021b). And *in vitro E. coli* adherence to porcine epithelial cells was also inhibited by EPS synthesized by *L. reuteri* (Ksonzekova et al., 2016). Consequently, the gene expression of proinflammatory cytokines like TNF- α and IL-6 brought on by *E. coli* or *Salmonella Typhimurium* infection was suppressed by EPS derived from *L. reuteri* (Ksonzekova et al., 2016; Kissova et al., 2022).

3.2.4 Short-chain fatty acids

SCFAs are a set of metabolite products derived from L. reuteri and engaged in regulating immune response and inflammatory reactions, thereby, promoting intestinal health (Smith et al., 2013; Oh et al., 2021). Intestinal SCFAs mainly consist of acetate, propionate, and butyrate, which can be generated by L. reuteri in different situations (Liu et al., 2022c; Calvigioni et al., 2023; Van den Abbeele et al., 2023). SCFAs are also suggested to be essential for ameliorating gut microbiota dysbiosis in mice colitis treated with L. reuteri (Lee et al., 2022). Particularly, butyrate is the typical metabolite which can enhance gastrointestinal motility by mediating the enteric neurons and maintaining the proliferation of Cajal cells though the AKT-NF-KB pathway (He et al., 2020). Besides, L. reuteri was reported to exert anti-tumor effects in mice with hepatocellular carcinoma by upregulating acetate levels (Hu et al., 2023). Recently propionate and butyrate were also reported to suppress the adhesion and invasion ability of Salmonella Typhimurium in vitro experiments (Liu et al., 2022b).

Together, these observations provide substantial evidence that metabolites generated by *L. reuteri* are involved in the regulation of gut immune response and remodeling of the commensal microbiota composition via the activation of various signal pathways.

3.3 Enhancement of intestinal epithelial barrier

Numerous studies showed that intestinal epithelial barrier damage disrupts immune homeostasis and contributes to many intestinal disorders, especially IBD and CRC (Leibovitzh et al., 2023). The capacity of L. reuteri to boost gut barrier function has been demonstrated in several investigations (Yi et al., 2018; Gao et al., 2022). On one hand, L. reuteri owns the ability to maintain the intestinal epithelial repair function. Wu and colleagues found that L. reuteri reconstructed the epithelial integrity by activating the Wnt/βcatenin pathway, thereby promoting the intestine epithelial cell proliferation and inducing intestinal stem cell differentiation to Paneth cells, even under the condition of TNF-induced intestinal inflammation (Wu et al., 2020). On the other hand, L. reuteri was also reported to maintain the integrity of the gut barrier by upregulating the expression of TJ proteins, evidenced in in preclinical studies (Gao et al., 2022; Liu et al., 2022a; Li et al., 2023a). TJ proteins are primary for epithelial cell proliferation and gut permeability (Kuo et al., 2021). Bacterial extracellular membrane vesicles derived from L. reuteri contain lipoteichoic acid, identified as a Toll-like receptor (TLR) 2 activator (Pang et al., 2022). According to research, TLR2 activation strengthened the intestinal barrier by increased expression of TJ proteins via the PI3K-Akt-signaling pathway (Gu et al., 2016). L. reuteri was also reported to induce TJ proteins expression via the activated PKC-Nrf-2/HO-1pathway and inhibited NF-KB pathway, along with reduced apoptosis of intestinal epithelial cells (Zhou et al., 2022b). In conclusion, these results indicated the therapeutic benefits of L. reuteri in gastrointestinal diseases by repairing the gut barrier function though different pathways.

3.4 Regulation of immune cell differentiation and function

L. reuteri may exhibit beneficial effects in digestive system diseases via modulation of the immune system. DCs are crucial in the interaction between microorganisms and gut immune responses because of their advantageous location and capacity to present luminal antigens as "gatekeepers" (Saez et al., 2023). Mature DCs promote the polarization of naive T cells toward T helper (Th) 1, Th2, Th17, or Treg responses (Tiberio et al., 2018). L. reuteri 5454 treatment alleviated colitis inflammation in mice, the potential molecular mechanism might be explained by the efficiency of DCs primed with L. reuteri 5454 to promote Tregs differentiation and trigger IL-22 secretion (Hrdy et al., 2020). Another study revealed that L. reuteri 17938 may activate DCs via TLR2 and lead to Tregs expansion in the intestinal mucosa, thereby alleviating experimental necrotizing enterocolitis (Hoang et al., 2018). Further evidenced by in vitro experiments, L. reuteri cell-free supernatants promoted DCs differentiation with up-regulated surface markers (CCR7, CD83, CD86, HLA-DR) and enhanced cytokine synthesis (IL-6, IL-10, and IL-23), nonetheless, these DCs exhibited diminished phagocytic activity (Haileselassie et al., 2016). The similar results were also reported by Lasaviciute and his colleagues who showed that a mixed secondary response profile was produced in DCs after priming human monocytes with L. reuteri DSM17938 secretions, secreting low levels of TNF-α, and IL-27 and high levels of IL-1 β and IL-6 (Lasaviciute et al., 2022). The particular manner by which L. reuteri modulates the phenotypic and operation of mucosal-like DC is yet fully understood. Notably,

the involvement of surface proteins was examined recently in the interaction between *L. reuteri* and DCs. The study demonstrated that *L. reuteri* can promote the release of the immune-regulating IL-10 in monocyte-derived DCs independent of its mucus adhesins while the production of pro-inflammatory cytokines (IL-6, TNF- α , IL-1 β) was enhanced by mucus adhesins (Bene et al., 2017).

Meanwhile, numerous preclinical studies showed L. reuteri also had the property to promote the proliferation of Tregs while suppressing the production of Th cells, specifically Th1 and Th2, accompanied by decreased mRNA expression of NF-KB, TLR4, TNF- α , IL-6, and upregulated IL-10 mRNA expression in the intestine (Hoang et al., 2018; Liu et al., 2019; Liu et al., 2023b). The forkhead box P3 (FoxP3) transcription factor is crucial in generating Tregs (Harada et al., 2022). And the FoxP3 mRNA expression level was shown to be elevated in peripheral blood after L. reuteri DSM 17938 administration in colicky infants (Savino et al., 2018b). In addition, the mechanism of Tregs induction may be associated with reprogrammed gut microbiota composition and SCFAs production by L. reuteri (Liu et al., 2019; Wen et al., 2021). However, the ability of L. reuteri to induce Tregs is generally strain-dependent, and certain L. reuteri strains have anti-inflammatory properties that are not always reliant on Tregs (Liu et al., 2021b; Liu et al., 2023a). Orally feeding L. reuteri can suppress Th1/Th2 responses in mesenteric lymph nodes in Treg-deficient mice via activation of adenosine A2A receptors (Liu et al., 2023a). This phenomenon may be explained by the specific property of L. reuteri to increase the level of plasma adenosine metabolites such as inosine (Liu et al., 2021b). Proteomics studies using mass spectrometry confirmed that bacterial extracellular membrane vesicles derived from L. reuteri carried numerous bacterial cell surface proteins, including 5'-nucleotidase, which initiate the process by which AMP is transformed into the signal molecule adenosine (Pang et al., 2022).

Besides, L. reuteri seemed to be able to promote M2 macrophage polarization. Pro-inflammatory M1-like macrophage indicators may be suppressed by the stress protein GroEL isolated from L. reuteri, whereas M2-like markers are favored, consequently suppressing intestinal production of pro-inflammatory markers (TNF- α , IL-1 β , interferon- γ) and increasing anti-inflammatory IL-10 and IL-13 by the activation of a TLR4 pathway (Dias et al., 2021). Except for these, Wang and colleagues reported that oral therapy of L. reuteri effectively inhibited the progression of mice colitis induced by immune checkpoint blockade treatment, and they highlighted that the therapeutic impact of L. reuteri ATCC PTA 6475 was related to a decrease of ILC3s (Wang et al., 2019). ILC3s are mainly found in the intestinal mucosa and can drive proinflammatory responses and cause immunopathological damage (Jarade et al., 2021). The mechanism is not fully clear. It is possible that L. reuteri's capacity to metabolize tryptophan into indole derivatives, which subsequently activate the aryl hydrocarbon receptor in ILC3s, is responsible for the underlying mechanism (Zelante et al., 2013; Schiering et al., 2017). Furthermore, a previous study investigated two strains extracted from piglets, L. reuteri ZJ617 and L. reuteri ZJ615, characterized by high and low adhesive ability respectively (Gao et al., 2016). The study showed that L. reuteri ZJ615 reduced phosphorylation levels

of ERK1/2 while *L. reuteri* ZJ617 had no significant effects on the ERK1/2 signaling pathway in the ilea of the LPS-stimulated mice model (Gao et al., 2016). Given this, *L. reuteri's* capacity to modulate the immune system may be somewhat dependent on its adhesive property.

Overall, these studies demonstrated that metabolites and bacterial components of *L. reuteri* as well as the restored balance of the gut microbiota are both engaged in the suppression of excessive immune response and protection the intestine from injury.

4 Conclusions and future perspectives

In the past decades, research regarding the link between digestive diseases and the gut microbiome is growing quickly, accompanied by increased interest in probiotics in gut health. *L. reuteri* may serve as a viable candidate for the treatment of digestive system disorders owing to its potent antimicrobial, immunomodulatory, and anti-inflammatory activities with nearly no safety risks.

In this review, we systematically interpreted its application in different digestive disorders, including IC, diarrhea and constipation, FAP, *H. pylori* infection, IBD, and liver diseases. The results discussed here showed that *L. reuteri* may alleviate the symptoms of digestive problems through a variety of mechanisms, such as manipulation of the intestinal microbial population, barrier function of the epithelium, regulation of immunity, and modulation of numerous metabolites. Most of the *in vitro* and *in vivo* experiments revealed beneficial impacts of *L. reuteri* but the results of some clinical trials may be controversial. This phenomenon may be explained, in part, by specific and different functions of multiple stains and the high diversity of the human gut microbiota affected by sex, population, diet as well as other factors.

According to well-established studies, *L. reuteri* treatment is effective in improving IC, diarrhea, constipation, and *H. pylori* infection, whether to be served as a monotherapy or adjuvant strategy. While given the limited clinical data, there is no sufficient support data to recommend *L. reuteri* application in FAP, IBD, diverticulitis, CRC and liver diseases. In conclusion, it is important to conduct more research on *L. reuteri*'s potential benefits for treating digestive system diseases so that it may be used in clinical settings and as a therapeutic strategy.

References

Agus, A., Planchais, J., and Sokol, H. (2018). Gut microbiota regulation of tryptophan metabolism in health and disease. *Cell Host Microbe* 23 (6), 716–724. doi: 10.1016/j.chom.2018.05.003

Agustina, R., Kok, F. J., van de Rest, O., Fahmida, U., Firmansyah, A., Lukito, W., et al. (2012). Randomized trial of probiotics and calcium on diarrhea and respiratory tract infections in Indonesian children. *Pediatrics* 129 (5), e1155–e1164. doi: 10.1542/ peds.2011-1379

Ahl, D., Liu, H., Schreiber, O., Roos, S., Phillipson, M., and Holm, L. (2016). Lactobacillus reuteri increases mucus thickness and ameliorates dextran sulphate sodium-induced colitis in mice. *Acta Physiol. (Oxf)* 217 (4), 300–310. doi: 10.1111/ apha.12695

Ahmed, T., Chisti, M. J., Rahman, M. W., Alam, T., Ahmed, D., Parvin, I., et al. (2021). Effect of 3 days of oral azithromycin on young children with acute diarrhea in

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RY: Conceptualization, Funding acquisition, Methodology, Project administration, Resources, Supervision, Visualization, Writing – review & editing. YP: Writing – original draft. YM: Funding acquisition, Writing – original draft. ZL: Conceptualization, Writing – review & editing. YJ: Software, Writing – review & editing. ZX: Methodology, Supervision, Validation, Writing – review & editing.

Funding

This work was supported by the Jiangsu Provincial Department of Science and Technology (No. BE2022698), the Wuxi Science and Technology Bureau (No.Y20222003), the Wuxi Municipal Medical Innovation Team (No.CXTD2021013), the Wuxi Commission of Health and Family Planning (No. FYKY202109).

Acknowledgments

We would like to thank Bullet Edits for English language editing.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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low-resource settings: a randomized clinical trial. JAMA Netw. Open 4 (12), e2136726. doi: 10.1001/jamanetworkopen.2021.36726

Ali, M. S., Lee, E. B., Lim, S. K., Suk, K., and Park, S. C. (2023). Isolation and identification of Limosilactobacillus reuteri PSC102 and evaluation of its potential probiotic, antioxidant, and antibacterial properties. *Antioxid. (Basel)* 12 (2). doi: 10.3390/antiox12020238

Asare, P. T., Zurfluh, K., Greppi, A., Lynch, D., Schwab, C., Stephan, R., et al. (2020). Reuterin demonstrates potent antimicrobial activity against a broad panel of human and poultry meat campylobacter spp. isolates. *Microorganisms* 8 (1). doi: 10.3390/ microorganisms8010078

Azad, M. B., Konya, T., Maughan, H., Guttman, D. S., Field, C. J., Chari, R. S., et al. (2013). Gut microbiota of healthy Canadian infants: profiles by mode of delivery and infant diet at 4 months. *CMAJ* 185 (5), 385–394. doi: 10.1503/cmaj.121189

Baidoun, F., Elshiwy, K., Elkeraie, Y., Merjaneh, Z., Khoudari, G., Sarmini, M. T., et al. (2021). Colorectal cancer epidemiology: recent trends and impact on outcomes. *Curr. Drug Targets* 22 (9), 998–1009. doi: 10.2174/1389450121999201117115717

Bao, X., Yu, W., Chu, Z., Gao, J., Zhou, M., and Gu, Y. (2023). Functional abdominal pain disorders in children in southern Anhui province, China are related to academic stress rather than academic performance. *BMC Pediatr.* 23 (1), 333. doi: 10.1186/s12887-023-04154-3

Barbaro, M. R., Cremon, C., Fuschi, D., Marasco, G., Palombo, M., Stanghellini, V., et al. (2022). Pathophysiology of diverticular disease: from diverticula formation to symptom generation. *Int. J. Mol. Sci.* 23 (12). doi: 10.3390/ijms23126698

Bell, H. N., Rebernick, R. J., Goyert, J., Singhal, R., Kuljanin, M., Kerk, S. A., et al. (2022). Reuterin in the healthy gut microbiome suppresses colorectal cancer growth through altering redox balance. *Cancer Cell* 40 (2), 185–200.e186. doi: 10.1016/j.ccell.2021.12.001

Bene, K. P., Kavanaugh, D. W., Leclaire, C., Gunning, A. P., MacKenzie, D. A., Wittmann, A., et al. (2017). Lactobacillus reuteri surface mucus adhesins upregulate inflammatory responses through interactions with innate C-type lectin receptors. *Front. Microbiol.* 8. doi: 10.3389/fmicb.2017.00321

Calvigioni, M., Bertolini, A., Codini, S., Mazzantini, D., Panattoni, A., Massimino, M., et al. (2023). HPLC-MS-MS quantification of short-chain fatty acids actively secreted by probiotic strains. *Front. Microbiol.* 14. doi: 10.3389/fmicb.2023.1124144

Chang, K., Jiang, L., Sun, Y., and Li, H. (2022). Effect of E-cadherin on prognosis of colorectal cancer: a meta-analysis update. *Mol. Diagn. Ther.* 26 (4), 397–409. doi: 10.1007/s40291-022-00593-3

Chau, K., Lau, E., Greenberg, S., Jacobson, S., Yazdani-Brojeni, P., Verma, N., et al. (2015). Probiotics for infantile colic: a randomized, double-blind, placebo-controlled trial investigating Lactobacillus reuteri DSM 17938. *J. Pediatr.* 166 (1), 74–78. doi: 10.1016/j.jpeds.2014.09.020

Chen, C. C., Liou, J. M., Lee, Y. C., Hong, T. C., El-Omar, E. M., and Wu, M. S. (2021). The interplay between Helicobacter pylori and gastrointestinal microbiota. *Gut Microbes* 13 (1), 1–22. doi: 10.1080/19490976.2021.1909459

Chen, W., Wen, L., Bao, Y., Tang, Z., Zhao, J., Zhang, X., et al. (2022). Gut flora disequilibrium promotes the initiation of liver cancer by modulating tryptophan metabolism and up-regulating SREBP2. *Proc. Natl. Acad. Sci. U.S.A.* 119 (52), e2203894119. doi: 10.1073/pnas.2203894119

Cheng, W. X., Ren, Y., Lu, M. M., Xu, L. L., Gao, J. G., Chen, D., et al. (2021). Palmitoylation in Crohn's disease: current status and future directions. *World J. Gastroenterol.* 27 (48), 8201–8215. doi: 10.3748/wjg.v27.i48.8201

Cimperman, L., Bayless, G., Best, K., Diligente, A., Mordarski, B., Oster, M., et al. (2011). A randomized, double-blind, placebo-controlled pilot study of Lactobacillus reuteri ATCC 55730 for the prevention of antibiotic-associated diarrhea in hospitalized adults. *J. Clin. Gastroenterol.* 45 (9), 785–789. doi: 10.1097/MCG.0b013e3182166a42

Coccorullo, P., Strisciuglio, C., Martinelli, M., Miele, E., Greco, L., and Staiano, A. (2010). Lactobacillus reuteri (DSM 17938) in infants with functional chronic constipation: a double-blind, randomized, placebo-controlled study. *J. Pediatr.* 157 (4), 598–602. doi: 10.1016/j.jpeds.2010.04.066

Collinson, S., Deans, A., Padua-Zamora, A., Gregorio, G. V., Li, C., Dans, L. F., et al. (2020). Probiotics for treating acute infectious diarrhoea. *Cochrane Database Syst. Rev.* 12 (12), CD003048. doi: 10.1002/14651858.CD003048.pub4

Dai, Z., Coker, O. O., Nakatsu, G., Wu, W. K. K., Zhao, L., Chen, Z., et al. (2018). Multi-cohort analysis of colorectal cancer metagenome identified altered bacteria across populations and universal bacterial markers. *Microbiome* 6 (1), 70. doi: 10.1186/s40168-018-0451-2

Dargenio, C., Dargenio, V. N., Bizzoco, F., Indrio, F., Francavilla, R., and Cristofori, F. (2021). Limosilactobacillus reuteri strains as adjuvants in the management of Helicobacter pylori infection. *Medicina (Kaunas)* 57 (7). doi: 10.3390/medicina57070733

de Brito, B. B., da Silva, F. A. F., Soares, A. S., Pereira, V. A., Santos, M. L. C., Sampaio, M. M., et al. (2019). Pathogenesis and clinical management of Helicobacter pylori gastric infection. *World J. Gastroenterol.* 25 (37), 5578–5589. doi: 10.3748/wjg.v25.i37.5578

Deng, Y., Zhou, M., Wang, J., Yao, J., Yu, J., Liu, W., et al. (2021). Involvement of the microbiota-gut-brain axis in chronic restraint stress: disturbances of the kynurenine metabolic pathway in both the gut and brain. *Gut Microbes* 13 (1), 1–16. doi: 10.1080/19490976.2020.1869501

de Weerth, C., Fuentes, S., Puylaert, P., and de Vos, W. M. (2013). Intestinal microbiota of infants with colic: development and specific signatures. *Pediatrics* 131 (2), e550–e558. doi: 10.1542/peds.2012-1449

Dias, A. M. M., Douhard, R., Hermetet, F., Regimbeau, M., Lopez, T. E., Gonzalez, D., et al. (2021). Lactobacillus stress protein GroEL prevents colonic inflammation. *J. Gastroenterol.* 56 (5), 442–455. doi: 10.1007/s00535-021-01774-3

Dinleyici, E. C., Dalgic, N., Guven, S., Metin, O., Yasa, O., Kurugol, Z., et al. (2015). Lactobacillus reuteri DSM 17938 shortens acute infectious diarrhea in a pediatric outpatient setting. *J. Pediatr. (Rio J)* 91 (4), 392–396. doi: 10.1016/j.jped.2014.10.009

Dinleyici, E. C., Group, P. S., and Vandenplas, Y. (2014). Lactobacillus reuteri DSM 17938 effectively reduces the duration of acute diarrhoea in hospitalised children. *Acta Paediatr.* 103 (7), e300–e305. doi: 10.1111/apa.12617

Dore, M. P., Bibbo, S., Loria, M., Salis, R., Manca, A., Pes, G. M., et al. (2019). Twicea-day PPI, tetracycline, metronidazole quadruple therapy with Pylera(R) or Lactobacillus reuteri for treatment naive or for retreatment of Helicobacter pylori. Two randomized pilot studies. *Helicobacter.* 24 (6), e12659. doi: 10.1111/ hel.12659

Dore, M. P., Sau, R., Niolu, C., Abbondio, M., Tanca, A., Bibbo, S., et al. (2022). Metagenomic changes of gut microbiota following treatment of Helicobacter pylori Infection with a simplified low-dose quadruple therapy with bismuth or Lactobacillus reuteri. *Nutrients* 14 (14). doi: 10.3390/nu14142789

Dvornikova, K. A., Platonova, O. N., and Bystrova, E. Y. (2023). Inflammatory bowel disease: crosstalk between histamine, immunity, and disease. *Int. J. Mol. Sci.* 24 (12). doi: 10.3390/ijms24129937

Eftekhari, K., Vahedi, Z., KaMali Aghdam, M., and Noemi Diaz, D. (2015). A randomized double-blind placebo-controlled trial of Lactobacillus reuteri for chronic functional abdominal pain in children. *Iran J. Pediatr.* 25 (6), e2616. doi: 10.5812/ ijp.2616

Eslami, M., Yousefi, B., Kokhaei, P., Hemati, M., Nejad, Z. R., Arabkari, V., et al. (2019). Importance of probiotics in the prevention and treatment of colorectal cancer. *J. Cell Physiol.* 234 (10), 17127–17143. doi: 10.1002/jcp.28473

Fatheree, N. Y., Liu, Y., Taylor, C. M., Hoang, T. K., Cai, C., Rahbar, M. H., et al. (2017). Lactobacillus reuteri for infants with colic: a double-blind, placebo-controlled, randomized clinical trial. *J. Pediatr.* 191, 170–178.e172. doi: 10.1016/j.jpeds.2017.07.036

Ferolla, S. M., Couto, C. A., Costa-Silva, L., Armiliato, G. N., Pereira, C. A., Martins, F. S., et al. (2016). Beneficial effect of synbiotic supplementation on hepatic steatosis and anthropometric parameters, but not on gut permeability in a population with nonalcoholic steatohepatitis. *Nutrients* 8 (7). doi: 10.3390/nu8070397

Flemming, J. A., Djerboua, M., Groome, P. A., Booth, C. M., and Terrault, N. A. (2021). NAFLD and alcohol-associated liver disease will be responsible for almost all new diagnoses of cirrhosis in Canada by 2040. *Hepatology* 74 (6), 3330–3344. doi: 10.1002/hep.32032

Forooghi Nia, F., Rahmati, A., Ariamanesh, M., Saeidi, J., Ghasemi, A., and Mohtashami, M. (2023). The anti-Helicobacter pylori effects of Limosilactobacillus reuteri strain 2892 isolated from camel milk in C57BL/6 mice. *World J. Microbiol. Biotechnol.* 39 (5), 119. doi: 10.1007/s11274-023-03555-x

Francavilla, R., Lionetti, E., Castellaneta, S., Ciruzzi, F., Indrio, F., Masciale, A., et al. (2012). Randomised clinical trial: Lactobacillus reuteri DSM 17938 vs. placebo in children with acute diarrhoea-a double-blind study. *Aliment Pharmacol. Ther.* 36 (4), 363–369. doi: 10.1111/j.1365-2036.2012.05180.x

Francavilla, R., Lionetti, E., Castellaneta, S. P., Magista, A. M., Maurogiovanni, G., Bucci, N., et al. (2008). Inhibition of Helicobacter pylori infection in humans by Lactobacillus reuteri ATCC 55730 and effect on eradication therapy: a pilot study. *Helicobacter.* 13 (2), 127–134. doi: 10.1111/j.1523-5378.2008.00593.x

Francavilla, R., Polimeno, L., DemiChina, A., Maurogiovanni, G., Principi, B., Scaccianoce, G., et al. (2014). Lactobacillus reuteri strain combination in Helicobacter pylori infection: a randomized, double-blind, placebo-controlled study. J. Clin. Gastroenterol. 48 (5), 407–413. doi: 10.1097/MCG.000000000000007

Gao, J., Cao, S., Xiao, H., Hu, S., Yao, K., Huang, K., et al. (2022). Lactobacillus reuteri 1 enhances intestinal epithelial barrier function and alleviates the inflammatory response induced by enterotoxigenic escherichia coli K88 via suppressing the MLCK signaling pathway in IPEC-J2 cells. *Front. Immunol.* 13. doi: 10.3389/fmmu.2022.897395

Gao, C., Ganesh, B. P., Shi, Z., Shah, R. R., Fultz, R., Major, A., et al. (2017). Gut microbe-mediated suppression of inflammation-associated colon carcinogenesis by luminal histamine production. *Am. J. Pathol.* 187 (10), 2323–2336. doi: 10.1016/j.ajpath.2017.06.011

Gao, K., Liu, L., Dou, X., Wang, C., Liu, J., Zhang, W., et al. (2016). Doses Lactobacillus reuteri depend on adhesive ability to modulate the intestinal immune response and metabolism in mice challenged with lipopolysaccharide. *Sci. Rep.* 6, 28332. doi: 10.1038/srep28332

Gao, C., Major, A., Rendon, D., Lugo, M., Jackson, V., Shi, Z., et al. (2015). Histamine H2 receptor-mediated suppression of intestinal inflammation by probiotic Lactobacillus reuteri. *mBio* 6 (6), e01358–e01315. doi: 10.1128/mBio.01358-15

Garcia Contreras, A. A., Vasquez Garibay, E. M., Sanchez Ramirez, C. A., Fafutis Morris, M., and Delgado Rizo, V. (2020). Lactobacillus reuteri DSM 17938 and agave inulin in children with cerebral palsy and chronic constipation: a double-blind randomized placebo controlled clinical trial. *Nutrients* 12 (10). doi: 10.3390/nu12102971

Garg, S., Singh, T. P., and Malik, R. K. (2020). *In vivo* implications of potential probiotic Lactobacillus reuteri LR6 on the gut and immunological parameters as an adjuvant against protein energy malnutrition. *Probiotics Antimicrob. Proteins* 12 (2), 517–534. doi: 10.1007/s12602-019-09563-4

Ge, X., Zhao, W., Ding, C., Tian, H., Xu, L., Wang, H., et al. (2017). Potential role of fecal microbiota from patients with slow transit constipation in the regulation of gastrointestinal motility. *Sci. Rep.* 7 (1), 441. doi: 10.1038/s41598-017-00612-y

Gordon, I. O., Abushamma, S., Kurowski, J. A., Holubar, S. D., Kou, L., Lyu, R., et al. (2022). Paediatric ulcerative colitis is a fibrotic disease and is linked with chronicity of inflammation. J. Crohns Colitis 16 (5), 804–821. doi: 10.1093/ecco-jcc/jjab216 Gordon, M., Biagioli, E., Sorrenti, M., Lingua, C., Moja, L., Banks, S. S., et al. (2018). Dietary modifications for infantile colic. *Cochrane Database Syst. Rev.* 10 (10), CD011029. doi: 10.1002/14651858.CD011029.pub2

Grander, C., Adolph, T. E., Wieser, V., Lowe, P., Wrzosek, L., Gyongyosi, B., et al. (2018). Recovery of ethanol-induced Akkermansia muciniphila depletion ameliorates alcoholic liver disease. *Gut* 67 (5), 891–901. doi: 10.1136/gutjnl-2016-313432

Gu, M. J., Song, S. K., Lee, I. K., Ko, S., Han, S. E., Bae, S., et al. (2016). Barrier protection via Toll-like receptor 2 signaling in porcine intestinal epithelial cells damaged by deoxynivalnol. *Vet. Res.* 47, 25. doi: 10.1186/s13567-016-0309-1

Gutierrez-Castrellon, P., Lopez-Velazquez, G., Diaz-Garcia, L., Jimenez-Gutierrez, C., Mancilla-Ramirez, J., Estevez-Jimenez, J., et al. (2014). Diarrhea in preschool children and Lactobacillus reuteri: a randomized controlled trial. *Pediatrics* 133 (4), e904–e909. doi: 10.1542/peds.2013-0652

Haileselassie, Y., Navis, M., Vu, N., Qazi, K. R., Rethi, B., and Sverremark-Ekstrom, E. (2016). Lactobacillus reuteri and Staphylococcus aureus differentially influence the generation of monocyte-derived dendritic cells and subsequent autologous T cell responses. *Immun. Inflammation Dis.* 4 (3), 315–326. doi: 10.1002/iid3.115

Han, J. X., Tao, Z. H., Wang, J. L., Zhang, L., Yu, C. Y., Kang, Z. R., et al. (2023). Microbiota-derived tryptophan catabolites mediate the chemopreventive effects of statins on colorectal cancer. *Nat. Microbiol.* 8 (5), 919–933. doi: 10.1038/s41564-023-01363-5

Happy Tummy, C., Lavalle, L., Sauvageot, N., Cercamondi, C. I., Jankovic, I., Egli, D., et al. (2023). Limosilactobacillus reuteri DSM 17938-containing infant formulas and the associations with gastrointestinal tolerance: a cross-sectional observational study. *Nutrients* 15 (3). doi: 10.3390/nu15030530

Harada, Y., Miyamoto, K., Chida, A., Okuzawa, A. T., Yoshimatsu, Y., Kudo, Y., et al. (2022). Localization and movement of Tregs in gastrointestinal tract: a systematic review. *Inflammation Regener.* 42 (1), 47. doi: 10.1186/s41232-022-00232-8

He, Q., Han, C., Huang, L., Yang, H., Hu, J., Chen, H., et al. (2020). Astragaloside IV alleviates mouse slow transit constipation by modulating gut microbiota profile and promoting butyric acid generation. *J. Cell Mol. Med.* 24 (16), 9349–9361. doi: 10.1111/jcmm.15586

Hegde, S., Lin, Y. M., Fu, Y., Savidge, T., and Shi, X. Z. (2020). Precision Lactobacillus reuteri therapy attenuates luminal distension-associated visceral hypersensitivity by inducing peripheral opioid receptors in the colon. *Pain* 161 (12), 2737–2749. doi: 10.1097/j.pain.000000000001967

Hoang, T. K., He, B., Wang, T., Tran, D. Q., Rhoads, J. M., and Liu, Y. (2018). Protective effect of Lactobacillus reuteri DSM 17938 against experimental necrotizing enterocolitis is mediated by Toll-like receptor 2. *Am. J. Physiol. Gastrointest. Liver Physiol.* 315 (2), G231–G240. doi: 10.1152/ajpgi.00084.2017

Holz, C., Busjahn, A., Mehling, H., Arya, S., Boettner, M., Habibi, H., et al. (2015). Significant reduction in Helicobacter pylori load in humans with non-viable Lactobacillus reuteri DSM17648: a pilot study. *Probiotics Antimicrob. Proteins* 7 (2), 91–100. doi: 10.1007/s12602-014-9181-3

Hrdy, J., Alard, J., Couturier-Maillard, A., Boulard, O., Boutillier, D., Delacre, M., et al. (2020). Lactobacillus reuteri 5454 and Bifidobacterium aniMalis ssp. lactis 5764 improve colitis while differentially impacting dendritic cells maturation and antimicrobial responses. *Sci. Rep.* 10 (1), 5345. doi: 10.1038/s41598-020-62161-1

Hu, C., Xu, B., Wang, X., Wan, W. H., Lu, J., Kong, D., et al. (2023). Gut microbiotaderived short-chain fatty acids regulate group 3 innate lymphoid cells in HCC. *Hepatology* 77 (1), 48–64. doi: 10.1002/hep.32449

Hurst, N. R., Kendig, D. M., Murthy, K. S., and Grider, J. R. (2014). The short chain fatty acids, butyrate and propionate, have differential effects on the motility of the Guinea pig colon. *Neurogastroenterol. Motil.* 26 (11), 1586–1596. doi: 10.1111/ nmo.12425

Imase, K., Tanaka, A., Tokunaga, K., Sugano, H., Ishida, H., and Takahashi, S. (2007). Lactobacillus reuteri tablets suppress Helicobacter pylori infection-a doubleblind randomised placebo-controlled cross-over clinical study. *Kansenshogaku Zasshi* 81 (4), 387–393. doi: 10.11150/kansenshogakuzasshi1970.81.387

Indrio, F., Dargenio, V. N., Francavilla, R., Szajewska, H., and Vandenplas, Y. (2023). Infantile colic and long-term outcomes in childhood: a narrative synthesis of the evidence. *Nutrients* 15 (3). doi: 10.3390/nu15030615

Indrio, F., Di Mauro, A., Riezzo, G., Civardi, E., Intini, C., Corvaglia, L., et al. (2014). Prophylactic use of a probiotic in the prevention of colic, regurgitation, and functional constipation: a randomized clinical trial. *JAMA Pediatr*. 168 (3), 228–233. doi: 10.1001/ jamapediatrics.2013.4367

Jadresin, O., Hojsak, I., Misak, Z., Kekez, A. J., Trbojevic, T., Ivkovic, L., et al. (2017). Lactobacillus reuteri DSM 17938 in the treatment of functional abdominal pain in children: RCT Study. *J. Pediatr. Gastroenterol. Nutr.* 64 (6), 925–929. doi: 10.1097/ MPG.000000000001478

Jadresin, O., Sila, S., Trivic, I., Misak, Z., Hojsak, I., and Kolacek, S. (2018). Lack of benefit of Lactobacillus reuteri DSM 17938 as an addition to the treatment of functional constipation. *J. Pediatr. Gastroenterol. Nutr.* 67 (6), 763–766. doi: 10.1097/ MPG.00000000002134

Jadresin, O., Sila, S., Trivic, I., Misak, Z., Kolacek, S., and Hojsak, I. (2020). Lactobacillus reuteri DSM 17938 is effective in the treatment of functional abdominal pain in children: results of the double-blind randomized study. *Clin. Nutr.* 39 (12), 3645–3651. doi: 10.1016/j.clnu.2020.04.019 Jarade, A., Di Santo, J. P., and Serafini, N. (2021). Group 3 innate lymphoid cells mediate host defense against attaching and effacing pathogens. *Curr. Opin. Microbiol.* 63, 83–91. doi: 10.1016/j.mib.2021.06.005

Jones, M. L., Martoni, C. J., Ganopolsky, J. G., Sulemankhil, I., Ghali, P., and Prakash, S. (2013). Improvement of gastrointestinal health status in subjects consuming Lactobacillus reuteri NCIMB 30242 capsules: a *post-hoc* analysis of a randomized controlled trial. *Expert Opin. Biol. Ther.* 13 (12), 1643–1651. doi: 10.1517/14712598.2013.833601

Jones, R. M., and Neish, A. S. (2021). Gut microbiota in intestinal and liver disease. Annu. Rev. Pathol. 16, 251–275. doi: 10.1146/annurev-pathol-030320-095722

Kambale, R. M., Ntagazibwa, J. N., Kasengi, J. B., Zigashane, A. B., Francisca, I. N., Mashukano, B. N., et al. (2023). Probiotics for children with uncomplicated severe acute malnutrition (PruSAM study): a randomized controlled trial in the Democratic Republic of Congo. *Am. J. Clin. Nutr.* 117 (5), 976–984. doi: 10.1016/j.ajcnut.2023.01.019

Kao, L., Liu, T. H., Tsai, T. Y., and Pan, T. M. (2020). Beneficial effects of the commercial lactic acid bacteria product, Vigiis 101, on gastric mucosa and intestinal bacterial flora in rats. *J. Microbiol. Immunol. Infect.* 53 (2), 266–273. doi: 10.1016/j.jmii.2018.06.002

Kaplan, G. G. (2015). The global burden of IBD: from 2015 to 2025. Nat. Rev. Gastroenterol. Hepatol. 12 (12), 720-727. doi: 10.1038/nrgastro.2015.150

Kapourchali, F. R., and Cresci, G. A. M. (2020). Early-Life gut microbiome-the importance of maternal and infant factors in its establishment. *Nutr. Clin. Pract.* 35 (3), 386–405. doi: 10.1002/ncp.10490

Karimi, S., Jonsson, H., Lundh, T., and Roos, S. (2018). Lactobacillus reuteri strains protect epithelial barrier integrity of IPEC-J2 monolayers from the detrimental effect of enterotoxigenic Escherichia coli. *Physiol. Rep.* 6 (2). doi: 10.14814/phy2.13514

Khalif, I. L., Quigley, E. M., Konovitch, E. A., and Maximova, I. D. (2005). Alterations in the colonic flora and intestinal permeability and evidence of immune activation in chronic constipation. *Dig Liver Dis.* 37 (11), 838–849. doi: 10.1016/j.dld.2005.06.008

Kim, S. J., Kang, C. H., Kim, G. H., and Cho, H. (2022b). Anti-tumor effects of heatkilled L. reuteri MG5346 and L. casei MG4584 against human colorectal carcinoma through caspase-9-dependent apoptosis in xenograft model. *Microorganisms* 10 (3). doi: 10.3390/microorganisms10030533

Kim, S., Lee, H. H., Choi, W., Kang, C. H., Kim, G. H., and Cho, H. (2022a). Antitumor effect of heat-killed Bifidobacterium bifidum on human gastric cancer through Akt-p53-dependent mitochondrial apoptosis in xenograft models. *Int. J. Mol. Sci.* 23 (17). doi: 10.3390/ijms23179788

Kissova, Z., Tkacikova, L., Mudronova, D., and Bhide, M. R. (2022). Immunomodulatory effect of Lactobacillus reuteri (Limosilactobacillus reuteri) and its exopolysaccharides investigated on epithelial cell line IPEC-J2 challenged with Salmonella Typhimurium. *Life (Basel)* 12 (12). doi: 10.3390/life12121955

Kolodziej, M., and Szajewska, H. (2019). Lactobacillus reuteri DSM 17938 in the prevention of antibiotic-associated diarrhoea in children: a randomized clinical trial. *Clin. Microbiol. Infect.* 25 (6), 699–704. doi: 10.1016/j.cmi.2018.08.017

Korpela, K., Renko, M., Paalanne, N., Vanni, P., Salo, J., Tejesvi, M., et al. (2020). Microbiome of the first stool after birth and infantile colic. *Pediatr. Res.* 88 (5), 776– 783. doi: 10.1038/s41390-020-0804-y

Kotloff, K. L. (2022). Bacterial diarrhoea. Curr. Opin. Pediatr. 34 (2), 147-155. doi: 10.1097/MOP.00000000001107

Kozhakhmetov, S., Meiirmanova, Z., Mukhanbetzhanov, N., Jarmukhanov, Z., Vinogradova, E., Mureyev, S., et al. (2023). Compositional and functional variability of the gut microbiome in children with infantile colic. *Sci. Rep.* 13 (1), 9530. doi: 10.1038/s41598-023-36641-z

Ksonzekova, P., Bystricky, P., Vlckova, S., Patoprsty, V., Pulzova, L., Mudronova, D., et al. (2016). Exopolysaccharides of Lactobacillus reuteri: their influence on adherence of E. coli to epithelial cells and inflammatory response. *Carbohydr Polym.* 141, 10–19. doi: 10.1016/j.carbpol.2015.12.037

Kubota, M., Ito, K., Tomimoto, K., Kanazaki, M., Tsukiyama, K., Kubota, A., et al. (2020). Lactobacillus reuteri DSM 17938 and magnesium oxide in children with functional chronic constipation: a double-blind and randomized clinical trial. *Nutrients* 12 (1). doi: 10.3390/nu12010225

Kuo, W. T., Zuo, L., Odenwald, M. A., Madha, S., Singh, G., Gurniak, C. B., et al. (2021). The tight junction protein ZO-1 is dispensable for barrier function but critical for effective mucosal repair. *Gastroenterology* 161 (6), 1924–1939. doi: 10.1053/j.gastro.2021.08.047

Laosee, W., Kantachote, D., Chansuwan, W., and Sirinupong, N. (2022). Effects of probiotic fermented fruit juice-based biotransformation by lactic acid bacteria and Saccharomyces boulardii CNCM I-745 on anti-Salmonella and antioxidative properties. J. Microbiol. Biotechnol. 32 (10), 1315–1324. doi: 10.4014/jmb.2206.06012

Larabi, A., Barnich, N., and Nguyen, H. T. T. (2020). New insights into the interplay between autophagy, gut microbiota and inflammatory responses in IBD. *Autophagy* 16 (1), 38–51. doi: 10.1080/15548627.2019.1635384

Lasaviciute, G., Barz, M., van der Heiden, M., Arasa, C., Tariq, K., Quin, J., et al. (2022). Gut commensal Limosilactobacillus reuteri induces atypical memory-like phenotype in human dendritic cells *in vitro. Gut Microbes* 14 (1), 2045046. doi: 10.1080/19490976.2022.2045046

Leclercq, S., Matamoros, S., Cani, P. D., Neyrinck, A. M., Jamar, F., Starkel, P., et al. (2014). Intestinal permeability, gut-bacterial dysbiosis, and behavioral markers of alcohol-dependence severity. *Proc. Natl. Acad. Sci. U.S.A.* 111 (42), E4485–E4493. doi: 10.1073/pnas.1415174111

Lee, H. L., Kim, J. M., Moon, J. H., Kim, M. J., Jeong, H. R., Go, M. J., et al. (2022). Anti-amnesic effect of synbiotic supplementation containing corni fructus and Limosilactobacillus reuteri in DSS-induced colitis mice. *Int. J. Mol. Sci.* 24 (1). doi: 10.3390/ijms24010090

Leibovitzh, H., Lee, S. H., Raygoza Garay, J. A., Espin-Garcia, O., Xue, M., Neustaeter, A., et al. (2023). Immune response and barrier dysfunction-related proteomic signatures in preclinical phase of Crohn's disease highlight earliest events of pathogenesis. *Gut* 72 (8), 1462–1471. doi: 10.1136/gutjnl-2022-328421

Li, J., Feng, S., Wang, Z., He, J., Zhang, Z., Zou, H., et al. (2023b). Limosilactobacillus mucosae-derived extracellular vesicles modulates macrophage phenotype and orchestrates gut homeostasis in a diarrheal piglet model. *NPJ Biofilms Microbiomes* 9 (1), 33. doi: 10.1038/s41522-023-00403-6

Li, C., Su, Z., Chen, Z., Cao, J., Liu, X., and Xu, F. (2023a). Lactobacillus reuteri strain 8008 attenuated the aggravation of depressive-like behavior induced by CUMS in high-fat diet-fed mice through regulating the gut microbiota. *Front. Pharmacol.* 14. doi: 10.3389/fphar.2023.1149185

Liang, B., Yuan, Y., Peng, X. J., Liu, X. L., Hu, X. K., and Xing, D. M. (2022). Current and future perspectives for Helicobacter pylori treatment and management: from antibiotics to probiotics. *Front. Cell Infect. Microbiol.* 12. doi: 10.3389/ fcimb.2022.1042070

Lin, Y. P., Thibodeaux, C. H., Pena, J. A., Ferry, G. D., and Versalovic, J. (2008). Probiotic Lactobacillus reuteri suppress proinflammatory cytokines via c-Jun. *Inflammation Bowel Dis.* 14 (8), 1068–1083. doi: 10.1002/ibd.20448

Lin, C., Zheng, Y., Lu, J., Zhang, H., Wang, G., and Chen, W. (2023). Differential reinforcement of intestinal barrier function by various Lactobacillus reuteri strains in mice with DSS-induced acute colitis. *Life Sci.* 314, 121309. doi: 10.1016/j.lfs.2022.121309

Lionetti, E., Miniello, V. L., Castellaneta, S. P., Magista, A. M., de Canio, A., Maurogiovanni, G., et al. (2006). Lactobacillus reuteri therapy to reduce side-effects during anti-Helicobacter pylori treatment in children: a randomized placebo controlled trial. *Aliment Pharmacol. Ther.* 24 (10), 1461–1468. doi: 10.1111/j.1365-2036.2006.03145.x

Liu, Y., Armbrister, S. A., Okeugo, B., Mills, T. W., Daniel, R. C., Oh, J. H., et al. (2023a). Probiotic-derived ecto-5'-nucleotidase produces anti-Inflammatory adenosine metabolites in Treg-deficient scurfy mice. *Probiotics Antimicrob. Proteins* 15, 1001–1013. doi: 10.1007/s12602-023-10089-z

Liu, H. Y., Giraud, A., Seignez, C., Ahl, D., Guo, F., Sedin, J., et al. (2021a). Distinct B cell subsets in Peyer's patches convey probiotic effects by Limosilactobacillus reuteri. *Microbiome* 9 (1), 198. doi: 10.1186/s40168-021-01128-4

Liu, H. Y., Gu, F., Zhu, C., Yuan, L., Zhu, C., Zhu, M., et al. (2022a). Epithelial heat shock proteins mediate the protective effects of Limosilactobacillus reuteri in dextran sulfate sodium-induced colitis. *Front. Immunol.* 13. doi: 10.3389/fimmu.2022.865982

Liu, Y., Hoang, T. K., Park, E. S., Freeborn, J., Okeugo, B., Tran, D. Q., et al. (2023b). Probiotic-educated Tregs are more potent than naive Tregs for immune tolerance in stressed new-born mice. *Benef. Microbes* 14 (1), 73–84. doi: 10.3920/BM2022.0095

Liu, Y., Hoang, T. K., Taylor, C. M., Park, E. S., Freeborn, J., Luo, M., et al. (2021b). Limosilactobacillus reuteri and Lacticaseibacillus rhamnosus GG differentially affect gut microbes and metabolites in mice with Treg deficiency. *Am. J. Physiol. Gastrointest. Liver Physiol.* 320 (6), G969–G981. doi: 10.1152/ajpgi.00072.2021

Liu, Y., Tian, X., He, B., Hoang, T. K., Taylor, C. M., Blanchard, E., et al. (2019). Lactobacillus reuteri DSM 17938 feeding of healthy newborn mice regulates immune responses while modulating gut microbiota and boosting beneficial metabolites. *Am. J. Physiol. Gastrointest. Liver Physiol.* 317 (6), G824–G838. doi: 10.1152/ajpgi.00107.2019

Liu, Y., Zhong, X., Lin, S., Xu, H., Liang, X., Wang, Y., et al. (2022c). Limosilactobacillus reuteri and caffeoylquinic acid synergistically promote adipose browning and ameliorate obesity-associated disorders. *Microbiome* 10 (1), 226. doi: 10.1186/s40168-022-01430-9

Liu, J., Zhu, W., Qin, N., Ren, X., and Xia, X. (2022b). Propionate and butyrate inhibit biofilm formation of Salmonella Typhimurium grown in laboratory media and food models. *Foods* 11 (21). doi: 10.3390/foods11213493

Lu, X., Zhang, M., Ma, Y., Li, G., Zhao, X., and Qian, W. (2023). Protective effect of Limosilactobacillus reuteri-fermented yogurt on mouse intestinal barrier injury induced by enterotoxigenic Escherichia coli. *J. Sci. Food Agric.* doi: 10.1002/jsfa.12836

Lukasik, J., Dierikx, T., Besseling-van der Vaart, I., de Meij, T., Szajewska, H., and Multispecies Probiotic in, A.A.D.S.G. (2022). Multispecies probiotic for the prevention of antibiotic-associated diarrhea in children: a randomized clinical trial. *JAMA Pediatr.* 176 (9), 860–866. doi: 10.1001/jamapediatrics.2022.1973

Mancabelli, L., Milani, C., Lugli, G. A., Turroni, F., Mangifesta, M., Viappiani, A., et al. (2017). Unveiling the gut microbiota composition and functionality associated with constipation through metagenomic analyses. *Sci. Rep.* 7 (1), 9879. doi: 10.1038/s41598-017-10663-w

Maragkoudaki, M., Chouliaras, G., Moutafi, A., Thomas, A., Orfanakou, A., and Papadopoulou, A. (2018). Efficacy of an oral rehydration solution enriched with Lactobacillus reuteri DSM 17938 and Zinc in the management of acute diarrhoea in infants: a randomized, double-blind, placebo-controlled trial. *Nutrients* 10 (9). doi: 10.3390/nu10091189 Maragkoudaki, M., Chouliaras, G., Orel, R., Horvath, A., Szajewska, H., and Papadopoulou, A. (2017). Lactobacillus reuteri DSM 17938 and a placebo both significantly reduced symptoms in children with functional abdominal pain. *Acta Paediatr.* 106 (11), 1857–1862. doi: 10.1111/apa.13992

Mathey, M. D., Pennella, C. L., and Zubizarreta, P. (2021). Colorectal carcinoma in children and adolescents. *Arch. Argent Pediatr.* 119 (5), e487–e498. doi: 10.5546/ aap.2021.eng.e487

Mehling, H., and Busjahn, A. (2013). Non-viable Lactobacillus reuteri DSMZ 17648 (PylopassTM) as a new approach to Helicobacter pylori control in humans. *Nutrients* 5 (8), 3062–3073. doi: 10.3390/nu5083062

Meisenheimer Es Md, M. B. A., Epstein, C. D., and Thiel D Md, M. P. H. (2022). Acute diarrhea in adults. *Am. Fam. Physician* 106 (1), 72–80.

Mercuri, M., Stack, D. M., Mantis, I., Moszkowski, R., and Field, T. M. (2023). Maternal and infant touching behaviours during perturbed interactions: Associations with maternal depressive symptomatology and infant crying. *Infant Behav. Dev.* 71, 101821. doi: 10.1016/j.infbeh.2023.101821

Mi, G. L., Zhao, L., Qiao, D. D., Kang, W. Q., Tang, M. Q., and Xu, J. K. (2015). Effectiveness of Lactobacillus reuteri in infantile colic and colicky induced maternal depression: a prospective single blind randomized trial. *Antonie Van Leeuwenhoek* 107 (6), 1547–1553. doi: 10.1007/s10482-015-0448-9

Mobini, R., Tremaroli, V., Stahlman, M., Karlsson, F., Levin, M., Ljungberg, M., et al. (2017). Metabolic effects of Lactobacillus reuteri DSM 17938 in people with type 2 diabetes: a randomized controlled trial. *Diabetes Obes. Metab.* 19 (4), 579–589. doi: 10.1111/dom.12861

Montgomery, T. L., Eckstrom, K., Lile, K. H., Caldwell, S., Heney, E. R., Lahue, K. G., et al. (2022). Lactobacillus reuteri tryptophan metabolism promotes host susceptibility to CNS autoimmunity. *Microbiome* 10 (1), 198. doi: 10.1186/s40168-022-01408-7

Moreno Marquez, C., Fernandez Alvarez, P., Valdes Delgado, T., Castro Laria, L., Arguelles Arias, F., Caunedo Alvarez, A., et al. (2022). Randomized, double-blind, placebo-controlled clinical trial on the usefulness of probiotic Lactobacillus reuteri in bismuth-containing quadruple eradication therapy for infection with Helicobacter pylori. *Rev. Esp. Enferm. Dig* 114 (2), 89–95. doi: 10.17235/reed.2021.7931/2021

Morgan, E., Arnold, M., Gini, A., Lorenzoni, V., Cabasag, C. J., Laversanne, M., et al. (2023). Global burden of colorectal cancer in 2020 and 2040: incidence and mortality estimates from GLOBOCAN. *Gut* 72 (2), 338–344. doi: 10.1136/gutjnl-2022-327736

Mu, Q., Tavella, V. J., and Luo, X. M. (2018). Role of Lactobacillus reuteri in human health and diseases. *Front. Microbiol.* 9. doi: 10.3389/fmicb.2018.00757

Mulhem, E., Khondoker, F., and Kandiah, S. (2022). Constipation in children and adolescents: evaluation and treatment. Am. Fam. Physician 105 (5), 469-478.

Naghibzadeh, N., Salmani, F., Nomiri, S., and Tavakoli, T. (2022). Investigating the effect of quadruple therapy with Saccharomyces boulardii or Lactobacillus reuteri strain (DSMZ 17648) supplements on eradication of Helicobacter pylori and treatments adverse effects: a double-blind placebo-controlled randomized clinical trial. *BMC Gastroenterol.* 22 (1), 107. doi: 10.1186/s12876-022-02187-z

Nishikawa, H., Nakamura, S., Miyazaki, T., Kakimoto, K., Fukunishi, S., Asai, A., et al. (2021). Inflammatory bowel disease and sarcopenia: its mechanism and clinical importance. *J. Clin. Med.* 10 (18). doi: 10.3390/jcm10184214

Nishino, K., Nishida, A., Inoue, R., Kawada, Y., Ohno, M., Sakai, S., et al. (2018). Analysis of endoscopic brush samples identified mucosa-associated dysbiosis in inflammatory bowel disease. *J. Gastroenterol.* 53 (1), 95–106. doi: 10.1007/s00535-017-1384-4

Oh, J. K., Vasquez, R., Kim, S. H., Hwang, I. C., Song, J. H., Park, J. H., et al. (2021). Multispecies probiotics alter fecal short-chain fatty acids and lactate levels in weaned pigs by modulating gut microbiota. *J. Anim. Sci. Technol.* 63 (5), 1142–1158. doi: 10.5187/jast.2021.e94

Ojetti, V., Ianiro, G., Tortora, A., D'Angelo, G., Di Rienzo, T. A., Bibbo, S., et al. (2014). The effect of Lactobacillus reuteri supplementation in adults with chronic functional constipation: a randomized, double-blind, placebo-controlled trial. *J. Gastrointest. Liver Dis.* 23 (4), 387–391. doi: 10.15403/jgld.2014.1121.234.elr

Ojetti, V., Saviano, A., Brigida, M., Petruzziello, C., Caronna, M., Gayani, G., et al. (2022). Randomized control trial on the efficacy of Limosilactobacillus reuteri ATCC PTA 4659 in reducing inflammatory markers in acute uncomplicated diverticulitis. *Eur. J. Gastroenterol. Hepatol.* 34 (5), 496–502. doi: 10.1097/MEG.00000000002342

Oliva, S., Di Nardo, G., Ferrari, F., Mallardo, S., Rossi, P., Patrizi, G., et al. (2012). Randomised clinical trial: the effectiveness of Lactobacillus reuteri ATCC 55730 rectal enema in children with active distal ulcerative colitis. *Aliment Pharmacol. Ther.* 35 (3), 327–334. doi: 10.1111/j.1365-2036.2011.04939.x

Pang, Y., Ermann Lundberg, L., Mata Forsberg, M., Ahl, D., Bysell, H., Pallin, A., et al. (2022). Extracellular membrane vesicles from Limosilactobacillus reuteri strengthen the intestinal epithelial integrity, modulate cytokine responses and antagonize activation of TRPV1. *Front. Microbiol.* 13. doi: 10.3389/fmicb.2022.1032202

Partty, A., Kalliomaki, M., Salminen, S., and Isolauri, E. (2017). Infantile colic is associated with low-grade systemic inflammation. *J. Pediatr. Gastroenterol. Nutr.* 64 (5), 691–695. doi: 10.1097/MPG.00000000001340

Peery, A. F., Crockett, S. D., Murphy, C. C., Jensen, E. T., Kim, H. P., Egberg, M. D., et al. (2022). Burden and cost of gastrointestinal, liver, and pancreatic diseases in the United States: Update 2021. *Gastroenterology* 162 (2), 621–644. doi: 10.1053/j.gastro.2021.10.017

Pernica, J. M., Steenhoff, A. P., MokOmane, M., Moorad, B., Lechile, K., Smieja, M., et al. (2017). Rapid enteric testing to permit targeted antimicrobial therapy, with and without Lactobacillus reuteri probiotics, for paediatric acute diarrhoeal disease in Botswana: a pilot, randomized, factorial, controlled trial. *PloS One* 12 (10), e0185177. doi: 10.1371/journal.pone.0185177

Pernomian, L., Duarte-Silva, M., and de Barros Cardoso, C. R. (2020). The aryl hydrocarbon receptor (AHR) as a potential target for the control of intestinal inflammation: insights from an immune and bacteria sensor receptor. *Clin. Rev. Allergy Immunol.* 59 (3), 382–390. doi: 10.1007/s12016-020-08789-3

Petruzziello, C., Migneco, A., Cardone, S., Covino, M., Saviano, A., Franceschi, F., et al. (2019). Supplementation with Lactobacillus reuteri ATCC PTA 4659 in patients affected by acute uncomplicated diverticulitis: a randomized double-blind placebo controlled trial. *Int. J. Colorectal Dis.* 34 (6), 1087–1094. doi: 10.1007/s00384-019-03295-1

Piccioni, A., Franza, L., Vaccaro, V., Saviano, A., Zanza, C., Candelli, M., et al. (2021). Microbiota and probiotics: the role of Limosilactobacillus Reuteri in diverticulitis. *Medicina (Kaunas)* 57 (8). doi: 10.3390/medicina57080802

Poonyam, P., Chotivitayatarakorn, P., and Vilaichone, R. K. (2019). High effective of 14-day high-dose PPI- bismuth-containing quadruple therapy with probiotics supplement for Helicobacter pylori eradication: a double blinded-randomized placebo-controlled study. *Asian Pac. J. Cancer Prev.* 20 (9), 2859–2864. doi: 10.31557/APJCP.2019.20.9.2859

Pourmirzaiee, M. A., Famouri, F., Moazeni, W., Hassanzadeh, A., and Hajihashemi, M. (2020). The efficacy of the prenatal administration of Lactobacillus reuteri LR92 DSM 26866 on the prevention of infantile colic: a randomized control trial. *Eur. J. Pediatr.* 179 (10), 1619–1626. doi: 10.1007/s00431-020-03641-4

Preidis, G. A., Saulnier, D. M., Blutt, S. E., Mistretta, T. A., Riehle, K. P., Major, A. M., et al. (2012). Host response to probiotics determined by nutritional status of rotavirusinfected neonatal mice. *J. Pediatr. Gastroenterol. Nutr.* 55 (3), 299–307. doi: 10.1097/ MPG.0b013e31824d2548

Qin, W., Ren, Z., Xu, C., Cao, Y. N., Sun, M. A., Huang, R., et al. (2023a). Chromatin accessibility and transcriptional landscape during inhibition of Salmonella enterica by Lactobacillus reuteri in IPEC-J2 cells. *Cells* 12 (6). doi: 10.3390/cells12060968

Qin, W., Xie, Y., Ren, Z., Xu, C., Sun, M. A., Yin, Z., et al. (2023b). Integrative ATACseq and RNA-seq analyses of IPEC-J2 cells reveals porcine transcription and chromatin accessibility changes associated with Escherichia coli F18ac inhibited by Lactobacillus reuteri. *Front. Microbiol.* 14. doi: 10.3389/fmicb.2023.1101111

Quaglio, A. E. V., Grillo, T. G., De Oliveira, E. C. S., Di Stasi, L. C., and Sassaki, L. Y. (2022). Gut microbiota, inflammatory bowel disease and colorectal cancer. *World J. Gastroenterol.* 28 (30), 4053–4060. doi: 10.3748/wjg.v28.i30.4053

Rezasoltani, S., Asadzadeh Aghdaei, H., Dabiri, H., Akhavan Sepahi, A., Modarressi, M. H., and Nazemalhosseini Mojarad, E. (2018). The association between fecal microbiota and different types of colorectal polyp as precursors of colorectal cancer. *Microb. Pathog.* 124, 244–249. doi: 10.1016/j.micpath.2018.08.035

Ricciardiello, L. (2022). Digestive diseases: Big burden, low funding? Results of the new United European gastroenterology white book on digestive diseases. *United Eur. Gastroenterol. J.* 10 (7), 627–628. doi: 10.1002/ueg2.12297

Riezzo, G., Chimienti, G., Orlando, A., D'Attoma, B., Clemente, C., and Russo, F. (2019). Effects of long-term administration of Lactobacillus reuteri DSM-17938 on circulating levels of 5-HT and BDNF in adults with functional constipation. *Benef. Microbes* 10 (2), 137–147. doi: 10.3920/BM2018.0050

Riezzo, G., Orlando, A., D'Attoma, B., Linsalata, M., Martulli, M., and Russo, F. (2018). Randomised double blind placebo controlled trial on Lactobacillus reuteri DSM 17938: improvement in symptoms and bowel habit in functional constipation. *Benef. Microbes* 9 (1), 51–60. doi: 10.3920/BM2017.0049

Romano, C., Ferrau, V., Cavataio, F., Iacono, G., Spina, M., Lionetti, E., et al. (2014). Lactobacillus reuteri in children with functional abdominal pain (FAP). *J. Paediatr. Child Health* 50 (10), E68–E71. doi: 10.1111/j.1440-1754.2010.01797.x

Saeed, N. K., Al-Beltagi, M., Bediwy, A. S., El-Sawaf, Y., and Toema, O. (2022). Gut microbiota in various childhood disorders: Implication and indications. *World J. Gastroenterol.* 28 (18), 1875–1901. doi: 10.3748/wjg.v28.i18.1875

Saez, A., Herrero-Fernandez, B., Gomez-Bris, R., Sanchez-Martinez, H., and Gonzalez-Granado, J. M. (2023). Pathophysiology of inflammatory bowel disease: innate immune system. *Int. J. Mol. Sci.* 24 (2). doi: 10.3390/ijms24021526

Sander, L. E., Lorentz, A., Sellge, G., Coeffier, M., Neipp, M., Veres, T., et al. (2006). Selective expression of histamine receptors H1R, H2R, and H4R, but not H3R, in the human intestinal tract. *Gut* 55 (4), 498–504. doi: 10.1136/gut.2004.061762

Saviano, A., Brigida, M., Migneco, A., Gunawardena, G., Zanza, C., Candelli, M., et al. (2021). Lactobacillus Reuteri DSM 17938 (Limosilactobacillus reuteri) in diarrhea and constipation: two sides of the same coin? *Medicina (Kaunas)* 57 (7). doi: 10.3390/ medicina57070643

Savino, F., Ceratto, S., Poggi, E., Cartosio, M. E., Cordero di Montezemolo, L., and Giannattasio, A. (2015). Preventive effects of oral probiotic on infantile colic: a prospective, randomised, blinded, controlled trial using Lactobacillus reuteri DSM 17938. *Benef. Microbes* 6 (3), 245–251. doi: 10.3920/BM2014.0090

Savino, F., Cordisco, L., Tarasco, V., Calabrese, R., Palumeri, E., and Matteuzzi, D. (2009). Molecular identification of coliform bacteria from colicky breastfed infants. *Acta Paediatr.* 98 (10), 1582–1588. doi: 10.1111/j.1651-2227.2009.01419.x

Savino, F., Cordisco, L., Tarasco, V., Palumeri, E., Calabrese, R., Oggero, R., et al. (2010). Lactobacillus reuteri DSM 17938 in infantile colic: a randomized, double-blind, placebo-controlled trial. *Pediatrics* 126 (3), e526–e533. doi: 10.1542/peds.2010-0433

Savino, F., Galliano, I., Garro, M., Savino, A., Dapra, V., Montanari, P., et al. (2018a). Regulatory T cells and Toll-like receptor 2 and 4 mRNA expression in infants with colic treated with Lactobacillus reuteri DSM17938. *Benef. Microbes* 9 (6), 917–925. doi: 10.3920/BM2017.0194

Savino, F., Garro, M., Montanari, P., Galliano, I., and Bergallo, M. (2018b). Crying time and RORgamma/FOXP3 expression in Lactobacillus reuteri DSM17938-treated infants with colic: a randomized trial. J. Pediatr. 192, 171–177 e171. doi: 10.1016/j.jpeds.2017.08.062

Savino, F., Pelle, E., Palumeri, E., Oggero, R., and Miniero, R. (2007). Lactobacillus reuteri (American type culture collection strain 55730) versus simethicone in the treatment of infantile colic: a prospective randomized study. *Pediatrics* 119 (1), e124–e130. doi: 10.1542/peds.2006-1222

Savino, F., Quartieri, A., De Marco, A., Garro, M., Amaretti, A., Raimondi, S., et al. (2017). Comparison of formula-fed infants with and without colic revealed significant differences in total bacteria, Enterobacteriaceae and faecal ammonia. *Acta Paediatr.* 106 (4), 573–578. doi: 10.1111/apa.13642

Schiering, C., Wincent, E., Metidji, A., Iseppon, A., Li, Y., Potocnik, A. J., et al. (2017). Feedback control of AHR signalling regulates intestinal immunity. *Nature* 542 (7640), 242–245. doi: 10.1038/nature21080

Schnupf, P., Gaboriau-Routhiau, V., and Cerf-Bensussan, N. (2018). Modulation of the gut microbiota to improve innate resistance. *Curr. Opin. Immunol.* 54, 137–144. doi: 10.1016/j.coi.2018.08.003

Sgritta, M., Dooling, S. W., Buffington, S. A., Momin, E. N., Francis, M. B., Britton, R. A., et al. (2019). Mechanisms underlying microbial-mediated changes in social behavior in mouse models of autism spectrum disorder. *Neuron* 101 (2), 246-259.e246. doi: 10.1016/j.neuron.2018.11.018

Shi, Z., Fultz, R. S., Engevik, M. A., Gao, C., Hall, A., Major, A., et al. (2019). Distinct roles of histamine H1- and H2-receptor signaling pathways in inflammation-associated colonic tumorigenesis. *Am. J. Physiol. Gastrointest. Liver Physiol.* 316 (1), G205–G216. doi: 10.1152/ajpgi.00212.2018

Shornikova, A. V., Casas, I. A., Isolauri, E., Mykkanen, H., and Vesikari, T. (1997a). Lactobacillus reuteri as a therapeutic agent in acute diarrhea in young children. J. Pediatr. Gastroenterol. Nutr. 24 (4), 399-404. doi: 10.1097/00005176-199704000-00008

Shornikova, A. V., Casas, I. A., Mykkanen, H., Salo, E., and Vesikari, T. (1997b). Bacteriotherapy with Lactobacillus reuteri in rotavirus gastroenteritis. *Pediatr. Infect. Dis. J.* 16 (12), 1103–1107. doi: 10.1097/00006454-199712000-00002

Shulpekova, Y. O., Nechaev, V. M., Popova, I. R., Deeva, T. A., Kopylov, A. T., Malsagova, K. A., et al. (2021). Food intolerance: the role of histamine. *Nutrients* 13 (9). doi: 10.3390/nu13093207

Sitkin, S., Lazebnik, L., Avalueva, E., Kononova, S., and Vakhitov, T. (2022). Gastrointestinal microbiome and Helicobacter pylori: eradicate, leave it as it is, or take a personalized benefit-risk approach? *World J. Gastroenterol.* 28 (7), 766–774. doi: 10.3748/wjg.v28.i7.766

Skrzydlo-Radomanska, B., Prozorow-Krol, B., Cichoz-Lach, H., Majsiak, E., Bierla, J. B., Kosikowski, W., et al. (2020). The effectiveness of synbiotic preparation containing Lactobacillus and Bifidobacterium probiotic strains and short chain fructooligosaccharides in patients with diarrhea predominant irritable bowel syndrome-a randomized double-blind, placebo-controlled study. *Nutrients* 12 (7). doi: 10.3390/nu12071999

Smith, P. M., Howitt, M. R., Panikov, N., Michaud, M., Gallini, C. A., Bohlooly, Y. M., et al. (2013). The microbial metabolites, short-chain fatty acids, regulate colonic Treg cell homeostasis. *Science* 341 (6145), 569–573. doi: 10.1126/science.1241165

Smythe, P., and Efthimiou, G. (2022). In silico genomic and metabolic atlas of Limosilactobacillus reuteri DSM 20016: an Insight into human health. *Microorganisms* 10 (7). doi: 10.3390/microorganisms10071341

Socala, K., Doboszewska, U., Szopa, A., Serefko, A., Włodarczyk, M., Zielinska, A., et al. (2021). The role of microbiota-gut-brain axis in neuropsychiatric and neurological disorders. *Pharmacol. Res.* 172, 105840. doi: 10.1016/j.phrs.2021.105840

Sommermeyer, H., Bernatek, M., Pszczola, M., Krauss, H., and Piatek, J. (2022). Supporting the diagnosis of infantile colic by a point of care measurement of fecal calprotectin. *Front. Pediatr.* 10. doi: 10.3389/fped.2022.978545

Sun, M. C., Zhang, F. C., Yin, X., Cheng, B. J., Zhao, C. H., Wang, Y. L., et al. (2018). Lactobacillus reuteri F-9-35 prevents DSS-induced colitis by inhibiting proinflammatory gene expression and restoring the gut microbiota in mice. *J. Food Sci.* 83 (10), 2645–2652. doi: 10.1111/1750-3841.14326

Sung, V., Hiscock, H., Tang, M. L., Mensah, F. K., Nation, M. L., Satzke, C., et al. (2014). Treating infant colic with the probiotic Lactobacillus reuteri: double blind, placebo controlled randomised trial. *BMJ* 348, g2107. doi: 10.1136/bmj.g2107

Szajewska, H., Gyrczuk, E., and Horvath, A. (2013). Lactobacillus reuteri DSM 17938 for the management of infantile colic in breastfed infants: a randomized, double-blind, placebo-controlled trial. *J. Pediatr.* 162 (2), 257–262. doi: 10.1016/j.jpeds.2012.08.004

Szymanski, H., and Szajewska, H. (2019). Lack of efficacy of Lactobacillus reuteri DSM 17938 for the treatment of acute gastroenteritis: a randomized controlled trial. *Pediatr. Infect. Dis. J.* 38 (10), e237–e242. doi: 10.1097/INF.000000000002355

Tezuka, H., and Ohteki, T. (2019). Regulation of IgA production by intestinal dendritic cells and related cells. *Front. Immunol.* 10. doi: 10.3389/fimmu.2019.01891

Thapar, N., Benninga, M. A., Crowell, M. D., Di Lorenzo, C., Mack, I., Nurko, S., et al. (2020). Paediatric functional abdominal pain disorders. *Nat. Rev. Dis. Primers* 6 (1), 89. doi: 10.1038/s41572-020-00222-5

Thomas, C. M., Hong, T., van Pijkeren, J. P., Hemarajata, P., Trinh, D. V., Hu, W., et al. (2012). Histamine derived from probiotic Lactobacillus reuteri suppresses TNF via modulation of PKA and ERK signaling. *PloS One* 7 (2), e31951. doi: 10.1371/journal.pone.0031951

Tiberio, L., Del Prete, A., Schioppa, T., Sozio, F., Bosisio, D., and Sozzani, S. (2018). Chemokine and chemotactic signals in dendritic cell migration. *Cell Mol. Immunol.* 15 (4), 346–352. doi: 10.1038/s41423-018-0005-3

Trivic, I., Niseteo, T., Jadresin, O., and Hojsak, I. (2021). Use of probiotics in the treatment of functional abdominal pain in children-systematic review and metaanalysis. *Eur. J. Pediatr.* 180 (2), 339–351. doi: 10.1007/s00431-020-03809-y

Turco, R., Russo, M., Bruzzese, D., and Staiano, A. (2021). Efficacy of a partially hydrolysed formula, with reduced lactose content and with Lactobacillus reuteri DSM 17938 in infant colic: a double blind, randomised clinical trial. *Clin. Nutr.* 40 (2), 412–419. doi: 10.1016/j.clnu.2020.05.048

Urbanska, M., Gieruszczak-Bialek, D., Szymanski, H., and Szajewska, H. (2016). Effectiveness of Lactobacillus reuteri DSM 17938 for the prevention of nosocomial diarrhea in children: a randomized, double-blind, placebo-controlled trial. *Pediatr. Infect. Dis. J.* 35 (2), 142–145. doi: 10.1097/INF.00000000000948

Urrutia-Baca, V. H., Escamilla-Garcia, E., de la Garza-Ramos, M. A., Tamez-Guerra, P., Gomez-Flores, R., and Urbina-Rios, C. S. (2018). *In vitro* antimicrobial activity and downregulation of virulence gene expression on Helicobacter pylori by reuterin. *Probiotics Antimicrob. Proteins* 10 (2), 168–175. doi: 10.1007/s12602-017-9342-2

Van den Abbeele, P., Goggans, M., Deyaert, S., Baudot, A., Van de Vliet, M., Calatayud Arroyo, M., et al. (2023). Lacticaseibacillus rhamnosus ATCC 53103 and Limosilactobacillus reuteri ATCC 53608 synergistically boost butyrate levels upon tributyrin administration ex vivo. *Int. J. Mol. Sci.* 24 (6). doi: 10.3390/ijms24065859

Vester-Andersen, M. K., Mirsepasi-Lauridsen, H. C., Prosberg, M. V., Mortensen, C. O., Trager, C., Skovsen, K., et al. (2019). Increased abundance of proteobacteria in aggressive Crohn's disease seven years after diagnosis. *Sci. Rep.* 9 (1), 13473. doi: 10.1038/s41598-019-49833-3

Vitale, I., Spano, M., Puca, V., Carradori, S., Cesa, S., Marinacci, B., et al. (2023). Antibiofilm activity and NMR-based metabolomic characterization of cell-free supernatant of Limosilactobacillus reuteri DSM 17938. *Front. Microbiol.* 14. doi: 10.3389/fmicb.2023.1128275

Walsham, A. D., MacKenzie, D. A., Cook, V., Wemyss-Holden, S., Hews, C. L., Juge, N., et al. (2016). Lactobacillus reuteri inhibition of enteropathogenic Escherichia coli adherence to human intestinal epithelium. *Front. Microbiol.* 7. doi: 10.3389/fmicb.2016.00244

Wang, W., Ma, H., Zhu, Y., Ni, K., Qin, G., Tan, Z., et al. (2021b). Screening of lactic acid bacteria with inhibitory activity against ETEC K88 as feed additive and the effects on sows and piglets. *Anim. (Basel)* 11 (6). doi: 10.3390/ani11061719

Wang, R., Tang, R., Li, B., Ma, X., Schnabl, B., and Tilg, H. (2021a). Gut microbiome, liver immunology, and liver diseases. *Cell Mol. Immunol.* 18 (1), 4–17. doi: 10.1038/s41423-020-00592-6

Wang, T., Teng, K., Liu, G., Liu, Y., Zhang, J., Zhang, X., et al. (2018). Lactobacillus reuteri HCM2 protects mice against enterotoxigenic Escherichia coli through modulation of gut microbiota. *Sci. Rep.* 8 (1), 17485. doi: 10.1038/s41598-018-35702-y

Wang, T., Zheng, N., Luo, Q., Jiang, L., He, B., Yuan, X., et al. (2019). Probiotics Lactobacillus reuteri abrogates immune checkpoint blockade-associated colitis by inhibiting group 3 innate lymphoid cells. *Front. Immunol.* 10. doi: 10.3389/fimmu.2019.01235

Wanke, M., and Szajewska, H. (2012). Lack of an effect of Lactobacillus reuteri DSM 17938 in preventing nosocomial diarrhea in children: a randomized, double-blind, placebo-controlled trial. J. Pediatr. 161 (1), 40–43.e41. doi: 10.1016/j.jpeds.2011.12.049

Wegner, A., Banaszkiewicz, A., Kierkus, J., Landowski, P., Korlatowicz-Bilar, A., Wiecek, S., et al. (2018). The effectiveness of Lactobacillus reuteri DSM 17938 as an adjunct to macrogol in the treatment of functional constipation in children. A randomized, double-blind, placebo-controlled, multicentre trial. *Clin. Res. Hepatol. Gastroenterol.* 42 (5), 494–500. doi: 10.1016/j.clinre.2018.03.008

Wei, W., Mu, S., Han, Y., Chen, Y., Kuang, Z., Wu, X., et al. (2022). Gpr174 knockout alleviates DSS-induced colitis via regulating the immune function of dendritic cells. *Front. Immunol.* 13. doi: 10.3389/fimmu.2022.841254

Weizman, Z., Abu-Abed, J., and Binsztok, M. (2016). Lactobacillus reuteri DSM 17938 for the management of functional abdominal pain in childhood: a randomized, double-blind, placebo-controlled trial. *J. Pediatr.* 174, 160–164.e161. doi: 10.1016/j.jpeds.2016.04.003

Wen, S., He, L., Zhong, Z., Zhao, R., Weng, S., Mi, H., et al. (2021). Stigmasterol restores the balance of Treg/Th17 cells by activating the butyrate-PPARgamma axis in colitis. *Front. Immunol.* 12. doi: 10.3389/fimmu.2021.741934

Werlinger, P., Nguyen, H. T., Gu, M., Cho, J. H., Cheng, J., and Suh, J. W. (2022). Lactobacillus reuteri MJM60668 prevent progression of non-alcoholic fatty liver disease through anti-adipogenesis and anti-inflammatory pathway. *Microorganisms* 10 (11). doi: 10.3390/microorganisms10112203

Wu, R. Y., Pasyk, M., Wang, B., Forsythe, P., Bienenstock, J., Mao, Y. K., et al. (2013). Spatiotemporal maps reveal regional differences in the effects on gut motility for Lactobacillus reuteri and rhamnosus strains. *Neurogastroenterol. Motil.* 25 (3), e205– e214. doi: 10.1111/nmo.12072

Wu, H., Xie, S., Miao, J., Li, Y., Wang, Z., Wang, M., et al. (2020). Lactobacillus reuteri maintains intestinal epithelial regeneration and repairs damaged intestinal mucosa. *Gut Microbes* 11 (4), 997–1014. doi: 10.1080/19490976.2020.1734423

Xu, Y., Ding, X., Wang, Y., Li, D., Xie, L., Liang, S., et al. (2022). Bacterial metabolite reuterin attenuated LPS-induced oxidative stress and inflammation response in HD11 macrophages. *Antioxid. (Basel)* 11 (9). doi: 10.3390/antiox11091662

Yang, C., Liang, L., Lv, P., Liu, L., Wang, S., Wang, Z., et al. (2021). Effects of nonviable Lactobacillus reuteri combining with 14-day standard triple therapy on Helicobacter pylori eradication: A randomized double-blind placebo-controlled trial. *Helicobacter* 26 (6), e12856. doi: 10.1111/hel.12856

Yang, J., Wang, C., Liu, L., and Zhang, M. (2020). Lactobacillus reuteri KT260178 supplementation reduced morbidity of piglets through its targeted colonization, improvement of cecal microbiota profile, and immune functions. *Probiotics Antimicrob. Proteins* 12 (1), 194–203. doi: 10.1007/s12602-019-9514-3

Yi, H., Wang, L., Xiong, Y., Wang, Z., Qiu, Y., Wen, X., et al. (2018). Lactobacillus reuteri LR1 improved expression of genes of tight junction proteins via the MLCK pathway in IPEC-1 Cells during infection with enterotoxigenic Escherichia coli K88. *Mediators Inflammation* 2018, 6434910. doi: 10.1155/2018/6434910

Yu, Z., Chen, J., Liu, Y., Meng, Q., Liu, H., Yao, Q., et al. (2023b). The role of potential probiotic strains Lactobacillus reuteri in various intestinal diseases: New roles for an old player. *Front. Microbiol.* 14. doi: 10.3389/fmicb.2023.1095555

Yu, T., Ding, Y., Qian, D., Lin, L., and Tang, Y. (2023a). Characteristics of fecal microbiota in different constipation subtypes and association with colon physiology, lifestyle factors, and psychological status. *Therap. Adv. Gastroenterol.* 16, 17562848231154101. doi: 10.1177/17562848231154101

Zeevenhooven, J., Browne, P. D., L'Hoir, M. P., de Weerth, C., and Benninga, M. A. (2018). Infant colic: mechanisms and management. *Nat. Rev. Gastroenterol. Hepatol.* 15 (8), 479–496. doi: 10.1038/s41575-018-0008-7

Zelante, T., Iannitti, R. G., Cunha, C., De Luca, A., Giovannini, G., Pieraccini, G., et al. (2013). Tryptophan catabolites from microbiota engage aryl hydrocarbon receptor and balance mucosal reactivity via interleukin-22. *Immunity* 39 (2), 372–385. doi: 10.1016/j.immuni.2013.08.003

Zhang, S., Wang, R., Li, D., Zhao, L., and Zhu, L. (2021). Role of gut microbiota in functional constipation. *Gastroenterol. Rep. (Oxf)* 9 (5), 392–401. doi: 10.1093/gastro/goab035

Zhang, Y., Wang, X. L., Zhou, M., Kang, C., Lang, H. D., Chen, M. T., et al. (2018). Crosstalk between gut microbiota and sirtuin-3 in colonic inflammation and tumorigenesis. *Exp. Mol. Med.* 50 (4), 1–11. doi: 10.1038/s12276-017-0002-0

Zhao, T., Wang, H., Liu, Z., Liu, Y., De, J., Li, B., et al. (2023). Recent perspective of Lactobacillus in reducing oxidative stress to prevent disease. *Antioxid. (Basel)* 12 (3). doi: 10.3390/antiox12030769

Zheng, T. X., Pu, S. L., Tan, P., Du, Y. C., Qian, B. L., Chen, H., et al. (2020). Liver metabolomics reveals the effect of Lactobacillus reuteri on alcoholic liver disease. *Front. Physiol.* 11. doi: 10.3389/fphys.2020.595382

Zhou, Q., Wu, F., Chen, S., Cen, P., Yang, Q., Guan, J., et al. (2022b). Lactobacillus reuteri improves function of the intestinal barrier in rats with acute liver failure through Nrf-2/HO-1 pathway. *Nutrition* 99-100, 111673. doi: 10.1016/j.nut.2022.111673

Zhou, L., Zeng, Y., Zhang, H., and Ma, Y. (2022a). The role of gastrointestinal microbiota in functional dyspepsia: A Review. *Front. Physiol.* 13. doi: 10.3389/ fphys.2022.910568

Zhu, L., Liu, W., Alkhouri, R., Baker, R. D., Bard, J. E., Quigley, E. M., et al. (2014). Structural changes in the gut microbiome of constipated patients. *Physiol. Genomics* 46 (18), 679–686. doi: 10.1152/physiolgenomics.00082.2014

Zong, M., Chang, C., Anjum, R., Xu, H., Guo, Y., Pan, D., et al. (2023). Multifunctional LPXTG-motif surface protein derived from Limosilactobacillus reuteri SH 23 in DSS-induced ulcerative colitis of mice. *FASEB J.* 37 (5), e22895. doi: 10.1096/fj.2022-0252-RR

Zoppi, G., Cinquetti, M., Luciano, A., Benini, A., Muner, A., and Bertazzoni Minelli, E. (1998). The intestinal ecosystem in chronic functional constipation. *Acta Paediatr.* 87 (8), 836–841. doi: 10.1080/080352598750013590