



Platelet-Derived Microvesicles in Cardiovascular Diseases

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Microvesicles (MVs) circulating in the blood are small vesicles (100–1,000 nm in diameter) derived from membrane blebs of cells such as activated platelets, endothelial cells, and leukocytes. A growing body of evidence now supports the concept that platelet-derived microvesicles (PMVs), the most abundant MVs in the circulation, are important regulators of hemostasis, inflammation, and angiogenesis. Compared with healthy individuals, a large increase of circulating PMVs has been observed, particularly in patients with cardiovascular diseases. As observed in MVs from other parent cells, PMVs exert their biological effects in multiple ways, such as triggering various intercellular signaling cascades and by participating in transcellular communication by the transfer of their "cargo" of cytoplasmic components and surface receptors to other cell types. This review describes our current understanding of the potential role of PMVs in mediating hemostasis, inflammation, and angiogenesis and their consequences on the pathogenesis of cardiovascular diseases, such as atherosclerosis, myocardial infarction, and venous thrombosis. Furthermore, new developments of the therapeutic potential of PMVs for the treatment of cardiovascular diseases will be discussed.

Keywords: microvesicles, platelet-derived microvesicles, cardiovascular disease, therapeutic potential, hemostasis, inflammation, angiogenesis

INTRODUCTION

Extracellular vesicles (EVs) encompass a broad range of vesicles released from cells (1). EVs can be classified into different subsets according to their size, cellular origin, content or the mechanism leading to their formation (**Table 1**). Microvesicles (MVs)—also referred to as microparticles— are vesicles typically around 100–1,000 nm in size. By contrast, smaller vesicles (30–100 nm) are referred to as exosomes, while larger vesicles containing nuclear materials are referred to as apoptotic bodies. Although there is a general consensus in most studies that apoptotic bodies are particles >1 μ m (2, 3), there are several studies that describe apoptotic bodies to have a smaller size range of 0.5 μ m (4, 5).

In the context of platelet biology, the plasma membrane fragments shed from activated platelets initially observed to possess procoagulant function were described as "platelet dust" by Wolf (17). Subsequent studies employing electron microscopy demonstrated the budding of vesicles from the platelet plasma membrane (18) thus confirming the cellular origin of the fragments detected by Wolf (17). In fact, 60–90% of EVs have been shown to be derived from platelets as indicated by positive CD41 staining (19). Since then, the role of EVs in the field of cardiovascular research has garnered a huge amount of interest due to their putative role in various pathological conditions.

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	Exosome	Microvesicle	Apoptotic body
Size	≈20–100 nm	≈0.1–1 µm	>1 µm
Origin	Multivesicular bodies, internal compartments	Plasma membrane	Cellular fragments
Markers	- Tetraspanins (CD63, CD9, and CD81)	 Phosphatidylserine (PS) 	– Histones
	 ALG-2-interacting protein X 	 Integrins 	 Fragmented DNA
	 Tumor susceptibility gene 101 protein 	- Selectins	– PS
	 Heat shock 70-kDa proteins 	 CD40 ligands 	
	 Major histocompatibility complex class I and class II 	 Other antigens of parental cell 	
Reference	- (6-9)	- (9-14)	- (9, 15, 16)

TABLE 1 | Characteristics of extracellular vesicles

Elevated levels of platelet-derived microvesicles (PMVs) are observed in diabetes mellitus, sepsis, rheumatoid arthritis, vascular inflammation, and cardiovascular diseases (20–31). Indeed, the pathological events associated with these diseases activate platelets (32–35), which have been demonstrated to increase PMV release, while at the same time, a subpopulation of PMVs coming from agonist-activated platelets have been demonstrated to contribute to pathological events (36). Thus, PMVs may well be both, one of the causes and a consequence of the pathophysiology that drives various diseases.

This review will focus on our current understanding of PMVs in mediating hemostasis, inflammation, and angiogenesis, which are all factors contributing to the pathogenesis of cardiovascular diseases. Furthermore, the clinical relevance of PMVs will be discussed in the context of their therapeutic potential in the treatment of cardiovascular diseases.

FORMATION AND CLEARANCE OF PMVs

Platelet-derived microvesicle formation is complex, and the exact mechanisms involved in the generation of PMVs to date remains to be clearly defined. However, it has been demonstrated that the generation of PMVs can be triggered by various mechanisms: (1) via platelet activation by soluble agonists or (2) shear stress or (3) glycoprotein (GP) IIb/IIIa outside-in signaling. In the case of platelet activation in response to soluble agonist stimulation or in response to high shear stress in the vasculature (37-39), sustained elevation of intracellular calcium has been observed, which initiates the loss of lipid asymmetry of the plasma membrane and cytoskeletal reorganization, ultimately leading to PMV generation (Figure 1) (40). The exposure of negatively charged phospholipids, such as phosphatidylserine (PS), on the outer leaflet of the plasma membrane is regulated by the calcium-dependent scramblase transmembrane protein (TMEM16F) (41). Likewise, calpain, a calcium-dependent protease, is central to regulating cytoskeletal reorganization thus facilitating PMV shedding (42). By contrast, unstimulated platelets have been demonstrated to generate PMVs via GPIIb/ IIIa signaling, which destabilizes the actin cytoskeleton, resulting in shedding of PMVs in the absence of soluble agonist stimulation (43). Once generated, PMVs have generally been observed to have a relatively short lifespan with studies demonstrating PMVs to have half-lives of 30 min in mice (44), 10 min in rabbits (45), and approximately 5.5 h in apheresis-derived

PMVs (46). Active endocytosis has been demonstrated to be involved in the clearance of MVs (47) including those of PMVs (48). Indeed, several studies demonstrated the involvement of PS-dependent phagocytic processes in the clearance of PMVs in the circulation (44, 49-52). Several opsonins such as complement component C3b, \u03b32-glycoprotein-1, lactadherin, and developmental endothelial locus-1 facilitate this PS-dependent phagocytosis (44, 49-51, 53). Upon engulfment of EVs by phagocytic cells such as macrophages and dendritic cells, liver X receptor (LXR) and peroxisome proliferator-activated receptor pathways are activated, which both are known to be induced by lipid derivatives (54). Indeed, the activation of the cholesterol derivative-sensitive pathway LXR by PMVs has been previously demonstrated in plasmacytoid dendritic cells (pDCs) (52), thereby highlighting the importance of lipid composition of EVs on the target cell responses after engulfment. The internalization of PMVs is not only essential for the clearance of PMVs but also ensures the delivery of the MV content into the target cell and thereby exerting their effector functions. Although studies have shown PS-dependent phagocytosis to be involved in the clearance of PMVs, other mechanisms are also involved in this process, which have previously been reviewed in detail by Mulcahy et al. (55).

COMPOSITION OF PMVs

From the humble origin of being just "platelet dust," it is now apparent that PMVs can mediate a diverse range of physiological responses. Their capacity to exert their biological role is attributed to their cell membrane composition and molecular cargo. The phospholipid composition of PMVs is a composite of the platelet plasma and granule membranes with high cholesterol content, also indicating an enrichment of lipid rafts (35). PMVs share many of the antigens as their parental cells that regulate cell adhesion, activation, and coagulation reactions (Figure 1) (10-13). These include various GPs, tissue factors (TF), selectins, and coagulation factors V and VIII (10-13). PMVs are packed with numerous biological molecules, which facilitate the transportation and delivery of bioactive mediators that can modulate the function of target cells. PMVs carry cytokines and chemokines such as interleukin (IL)-1β, CXCL4, CXCL7, and CCL5 (11, 56). In addition, a vast amount of proteins, growth factors, and genetic material such as ribonucleic acid (RNA), messenger RNA, and microRNA can be packaged into PMVs (12, 57-60). Moreover,



scramblase (TMEM16F), mediating the externalization of negatively charged PS (indicated as purple phospholipid). Furthermore, increased intracellular calcium leads to reorganization of the cytoskeleton by activating calpain, thereby cleaving PMVs and releasing them into the circulation. The size, physical characteristics, and cargo of PMVs depend on the environment and agonist(s) causing PMV generation. PMVs are packed with proteins including growth factors, cytokines/chemokines, and apoptotic proteins. PMVs also carry nucleic acids (mRNA, miRNA, and RNA) and mitochondria. PS, phosphatidylserine; GP, glycoprotein; vWF, von Willebrand factor; RNA, ribonucleic acid; mRNA, messenger RNA; miRNA, microRNA; PMV, platelet-derived microvesicle.

PMVs have recently been described to carry mitochondria, which can influence inflammatory responses (61).

The mechanisms of selective packaging have been demonstrated widely in EVs (62–65). Albeit not fully characterized in PMVs, the difference in lipid composition between PMVs and their parental cell, platelets (35), provide evidence for selective membrane assembly. Interestingly, it has been observed that the physical and biological components of PMVs are influenced by the stimulus used to generate PMVs (66–69). The heterogeneity observed in PMVs may explain why they have a diverse range of biological roles. For instance, larger PMVs may be enriched with more receptors and contents within, and thus can exert more effects, while smaller PMVs can deliver their biological cargo to areas that are otherwise difficult to enter, such as tumors, or to cross, such as the blood–brain barrier. Overall, it is important to understand that various pathological conditions will produce different types of PMVs carrying unique biological cargos that exert specific effects on targets cells. However, the precise mechanism by which PMVs selectively package and release their biological cargo to influence cellular function needs to be carefully determined in the future.

BIOLOGICAL FUNCTION OF PMVs

Hemostasis

It is perhaps not surprising that PMVs are most widely recognized for their role in mediating hemostasis given the resemblance in lipid composition and biological cargo they share with their parent cell—the platelet. Diseases that affect PMV shedding in the circulation have provided insight into their ability to regulate hemostasis. For example, patients with Castaman's defect, where platelets have an isolated inability to generate MVs display a bleeding phenotype (70). Similarly, Scott syndrome, where platelets cannot externalize PS and generate PMVs, is associated with a marked bleeding diathesis (41, 70–72). These disorders highlight the importance of PMVs in mediating hemostasis.

In accordance with their ability to regulate hemostasis, PMVs display both pro- and anticoagulant properties. The assembly of the tenase and prothrombinase complexes, and thus thrombin generation, is increased in the presence of PMVs due to the PS in the outer leaflet of the cell membrane (73). PMVs also express TF, which can initiate the extrinsic coagulation pathway by activating Factor VII (73, 74). On the contrary, PMVs have anticoagulant activity, which is associated with the binding of the natural anticoagulant, protein S and activation of protein C (75–79). Given the potential pro- and anticoagulant effects of PMVs, the tight regulation of PMV levels is likely an important factor regulating the hemostatic response.

Inflammation

Akin to the wealth of literature demonstrating an important pro-inflammatory role for platelets, there is now a growing body of evidence demonstrating that PMVs can regulate inflammatory responses. The immunoglobulins, antigens, cytokines, and chemokines that PMVs carry can directly regulate immune responses (11, 56, 80). The pro-inflammatory effects of PMVs can be demonstrated through their interactions with monocytes and neutrophils. Mechanistically, PMVs bind to leukocytes and form aggregates and can induce monocytes to release inflammatory mediators including IL-1β, tumor necrosis factor (TNF)- α , monocyte chemoattractant protein-1, and matrix metalloproteinase (MMP)-9, which enhance monocyte migration (81, 82). Lipopolysaccharide-induced PMVs carry IL-1β in its mature form, which can activate endothelial cells and induce leukocyte adhesion thus promoting endothelial inflammation (56). Furthermore, mitochondria that are released in vivo in sterile inflammatory diseases, such as rheumatoid arthritis, have been observed to be packaged within PMVs, which can be hydrolyzed by phospholipase A2 IIA to generate bioactive mediators which promote neutrophil pro-inflammatory responses (61).

Intriguingly, there are reports that PMVs can also act as antiinflammatory mediators. Recently, Dinkla and colleagues (83) have shown that PMVs prevent the differentiation of regulatory T cells into a pro-inflammatory phenotype. PMVs can bind to CCR6-HLA-DR⁺ regulatory T cell subsets via P-selectin and inhibit the production of IL-17 and interferon- γ (83). In accordance, PMVs from platelet concentrates can modify innate immune cells such as macrophages and dendritic cells. Macrophage activation is attenuated in the presence of PMVs as indicated by reduced production of TNF- α and IL-10 (84). PMVs also alter the function of monocyte-derived dendritic cells as demonstrated by their reduced capacity to present antigens, diminished production of pro-inflammatory cytokines and decreased phagocytic activity (84). PMVs can also modify inflammatory effects of the target cell. For instance, PMVs have been demonstrated to regulate the inflammatory responses of mast cells by the transfer of Lipoxygenase 12 (85). This leads to the production of Lipoxin A4, which induces anti-inflammatory and anti-angiogenic responses on endothelial cells by suppressing the generation of pro-inflammatory cytokines (85, 86). Furthermore, pDCs, a subset of dendritic cells that augment inflammatory processes by producing a large amount of pro-atherogenic type 1 interferons were observed to engulf PMVs in a PS-dependent manner (52). PMVs were observed to inhibit pDCs pro-inflammatory response by reducing the secretion of TNF- α and IL-8, signifying an antiinflammatory mechanism of PMVs (52).

In addition, our group has demonstrated that pentameric protein C-reactive protein (pCRP) binds to different MVs including PMVs (87). These pCRP-MVs, albeit not pro-inflammatory in healthy individuals, can aggravate existing tissue injury by activating the classical complement pathway and enhancing leukocyte recruitment to inflamed tissues (87). MVs not only bind pCRP but also convert pCRP to a highly pro-inflammatory monomer of C-reactive protein (mCRP), which can bind to endothelial cells and generate pro-inflammatory signals (88-91). In addition, autoantigen proteinase 3, an elastin degrading protease, binds to PS expressing MVs, thereby promoting inflammation via the generation of reactive oxygen species in neutrophils (92). Thus, these studies further highlight the ability of PMVs to partner with proteins to induce a pro-inflammatory phenotype. Therefore, PMVs may play a dual role in inflammation as they may instigate either pro- or anti-inflammatory responses depending on the cell membrane composition and biological cargo transported by the PMV.

Angiogenesis

In addition to harboring a number of pro-inflammatory cytokines, PMVs may carry growth factors such as vascular endothelial growth factor (VEGF), fibroblast growth factor 2, and lipid growth factors suggesting PMVs may play an important role in regulating angiogenesis (93). In accordance, Kim and colleagues have demonstrated that PMVs can inhibit apoptosis while enhancing cell migration, proliferation, survival, and tube formation in human umbilical vein endothelial cells (93). PMVs may also enhance pro-angiogenic MMP-2 and MMP-9 expression in endothelial cells *in vitro* and *in vivo* (94). Furthermore, PMVs stimulate the growth of endothelial

progenitor cells, thus contributing to the formation of new blood vessels (93).

The role of PMVs in regulating angiogenesis in the context of cardiovascular diseases has been highlighted by Brill and colleagues (95). The authors established that PMVs can induce angiogenesis in vitro caused by cytokines, VEGF, basic fibroblast growth factor, and platelet-derived growth factor packaged within PMVs in a process linked to Src, PI-3K, and ERK signaling (95). Moreover, the injection of PMVs in ischemic heart muscle induces the formation of blood vessels in a murine model of myocardial infarction (MI), signifying PMVs capability to induce myocardial angiogenesis in the setting of ischemia (95). PMVs have also been shown to facilitate endothelial repair after arterial injury by enhancing the vasoregenerative capacity of early outgrowth cells (EOCs) (96). PMVs enhance the recruitment, migration, and differentiation of EOCs at the site of injury by enhancing angiogenic growth factors that stimulate resident mature endothelial cells (96). The ability of PMVs to induce angiogenesis has also been demonstrated in the context of neurogenesis following brain injury and tumor progression (97, 98) thus, highlighting the potentially broad role of PMVs in endothelial repair and angiogenesis.

PMVs IN CARDIOVASCULAR DISEASES

Atherosclerosis

The rupturing of an atherosclerotic plaque can lead to MI and stroke, which are leading causes of death and disability globally. Indeed, excessive amounts of PMVs have been observed in patients with atherosclerosis (23-28). The increase in PMV numbers was found to correlate with multiple parameters including carotid artery intima media thickness, lipid-rich atherosclerotic plaques, and plaque burden (23-27). In the setting of atherosclerosis, increased hemodynamic shear stress due to plaque-associated luminal stenosis as well as the accumulation of oxidized low-density lipoprotein can activate platelets and stimulate generation of pro-inflammatory PMVs (33, 99). In atherosclerosis, monocytes adhere to activated endothelial cells, infiltrate the intima, and differentiate to tissue macrophages, which then engulf lipids and form foam cells (100). Smooth muscle cells migrate from the media to the intima and produce extracellular matrix, forming the fibrous cap (100). Macrophages and smooth muscle cells can undergo apoptosis leading to accumulation of extracellular lipid which forms the necrotic core (100). Indeed, PMVs have been implicated with these different stages of atherogenesis. For instance, PMVs encapsulate and transport miR-223 to endothelial cells, which can trigger endothelial apoptosis via the insulin-like growth factor-1 receptor (60). PMVs, together with endothelial MVs, increase endothelial permeability thereby influencing vascular endothelial dysfunction, an early step in the development of atherosclerosis (101). The presence of P-selectin expressed by PMVs allows them to interact with leukocytes via P-selectin GP ligand-1 thereby facilitating leukocyte accumulation at the site of endothelial injury and enhancing leukocyte infiltration from the blood vessel to the intima (82). Further to this,

PMVs can transfer the pro-atherogenic cytokine RANTES to endothelial cells and induce monocytes and endothelial cells to release pro-inflammatory cytokines such as IL-8, IL-1β, TNF- α , and IL-6, further enhancing leukocyte adhesion and infiltration (102–104). PMVs encapsulate active caspase-3 that can induce macrophage apoptosis (105). As a consequence in atherosclerosis, lipids derived from dead macrophages can accumulate and can contribute to the formation of the necrotic core. Furthermore, PMVs have been demonstrated to stimulate smooth muscle cells, leading to the migration of smooth muscle from the media to the intima thereby enhancing lesion progression (106, 107).

However, the main caveat of these studies is the fact that PMVs are only detected in the circulating blood of patients with atherosclerosis but are not found in the atherosclerotic plaque itself (108). This raises the question as to whether PMVs are merely associated with atherogenesis or play an active role in disease pathogenesis. The absence of PMVs in the plaque, while intriguing because of the evidence of PMVs' infiltration in other inflamed tissues, such as arthritic joints (36), could suggest selective removal of PMVs as they engage with other cells and exert their effects at the intraluminal area of the vessel in the setting of atherosclerosis. PMVs are highly subjected to endocytosis/phagocytosis, due to the high expression of adhesion molecules and PS (109). This potentially enhances the clearance of PMVs in comparison with other blood streamderived MVs, which may possibly be one of the mechanisms contributing to the absence of PMVs in the atherosclerotic plaque. These are important questions for further studies.

Acute Coronary Syndromes (ACS)

High levels of circulatory PMVs have been observed in patients with ACS and are also associated with the degree of elevation of cardiac enzymes, IL-6, and CRP levels (29-31). The elevated levels of PMVs in the plasma of patients with ACS persisted for up to 4 years after MI and is linked with markers of coagulation activation and soluble CD40L (110). In accordance with these findings, PMVs have been demonstrated to correlate with the size of myocardium at risk and microvascular dysfunction after ST elevation MI (STEMI) (111, 112). Also, it appears likely that MVs play an active role in promoting vascular inflammation and cardiac damage in patients after an ACS since MVs, including PMVs containing pro-inflammatory isoforms of CRP, have been demonstrated to be elevated in these patients (87, 88). Furthermore, PMVs independently predict future admission for major bleeding in non-STEMI patients (113). Taken together, these data suggest that PMVs detected in patients with ACS may also act as reporters of vascular inflammation, microvascular obstruction and myocardial damage in cardiovascular diseases (29, 30, 111, 112). While elevated levels of PMVs are often observed in patients with ACS, there are few studies that have reported variances in the levels of PMVs and the lack of association of PMVs with the severity of coronary artery disease (114, 115). The discordance in the levels of PMVs in ACS may be due to the variability of inclusion criteria and medication of the patients enrolled in each study. Also, the lack of consensus in the characteristic and definition of MVs may account for the variations observed in the literature.

Indeed, further studies are required, with establish common protocols and clear MVs nomenclature, to fully elucidate the role of PMVs in ACS.

Thrombotic Disorders

Thrombotic complications are often observed in patients with cardiovascular diseases. Augmented shedding of PMVs is deleterious and may contribute to thrombosis. For instance, in patients with immune thrombocytopenia, there are high levels of PMVs despite low platelet counts, which have been linked to a paradoxical increased risk of thrombotic events (116, 117). Increased PMVs are also detected in other thrombotic diseases such as acute pulmonary embolism and deep vein thrombosis (DVT) (118-121). Likewise, PMVs have been detected to be elevated in the context of thrombophilic states such antithrombin deficiency, protein C deficiency, and the Factor V Leiden mutation—all predispositions to venous thrombosis (122, 123). Increased levels of PMVs are observed to be associated with a heightened risk of venous thromboembolism, and PMVs have been proposed as a biomarker to help diagnose patients with DVT (119, 121). Similar to ACS, variation in plasma levels of PMVs are observed in thrombotic disorders (122, 124-126). Inconsistencies in processing blood samples such as handling, storage and methodology used in isolating PMVs may cause artifactual generation of PMVs and represent one of the underlying reasons for the discrepancies observed in the level of PMVs. Therefore, further studies are needed to fully unravel the role of PMVs in thrombotic diseases.

CURRENT CHALLENGES IN EV RESEARCH

Despite the remarkable progress in the field of EVs, clinical translation of EVs as a diagnostic or prognostic marker of pathological states remains a challenge (127). To fully unravel the potential of EVs in the diagnosis and therapy of cardiovascular diseases, it is imperative to understand the biological roles of PMVs in vivo. One of the drawbacks in the field of EVs is that most of the experiments demonstrating the physiological effects of EVs on target cells, such as endothelial cells, have been done in vitro in culture. The key weakness of this approach is that endothelial cells under in vivo conditions are under constant steady-state exposure to EVs. It is not clear how the response of cells in the in vitro, which is typically EV-free, to the sudden exposure of EVs is related to the in vivo setting. Furthermore, due to the lack of common practice in sample preparation and analysis, EV counts and phenotypes may vary dramatically between laboratories, making data analyses and clinical translation difficult. Indeed, several studies have highlighted the effects of pre-analytical variables on EV measurements. These include the type of anticoagulant used in collecting blood samples, centrifugation protocol, the storage of samples and staining protocols used for surface membrane antigens for determining the cellular origin of EVs (128-131). In regard to PMVs, preventing platelet activation and ensuring complete removal of platelets during processing of samples are crucial as this may result in

inaccurate findings. It is also essential to be mindful of the storage and thawing conditions of samples as this leads to changes in the number of PMVs and Annexin V binding (128–130). In addition to this variability, sample handling, isolation protocol, different antigens used to determine cellular origin, inclusion criteria for patients, and their clinical characteristics may account for the qualitative and quantitative variations observed in the literature regarding the characterization of EVs.

The technological advancements have facilitated new methods to improve the purification and detection of EVs from biological fluids (1, 127, 132–135). These include the following: sensitive single particle detection devices (tunable resistive pulse sensing; nanoparticle tracking analysis; and dynamic light scattering), flow cytometry (conventional; imaging; and impedance based), proteomics, and atomic force microscopy (1, 127, 130, 134, 136-138). However, a combination of multiple methods is still necessary to assess both physical and biological properties of EVs (1, 135). In light of the current limitation of EV studies, the International Society of Extracellular Vesicles endeavors to provide guidelines, harmonizing nomenclatures and practices in an effort to improve reproducibility of EV experiments and to eliminate ambiguity in the field of EVs (139-141). Furthermore, this society has already published several position papers in the Journal of Extracellular Vesicles (139-142) and provides public online databases that catalog EV-associated components, thereby assisting researchers to optimize their practices (EV-TRACK, ExoCarta, Vesiclepedia, and EVpedia).

THERAPEUTIC POTENTIAL OF PMVs

Given the role of PMVs in cardiovascular diseases, this raises the question as to whether MVs can be exploited for therapeutic benefit. Indeed, pharmacological studies that alter the levels of PMVs in the circulation have shown encouraging results (143–146). Also, the physical and biochemical properties of PMVs are advantages that can be utilized for the purpose of developing a therapy for cardiovascular diseases (**Table 2**). For

TABLE 2 | Advantages and disadvantages of extracellular vesicles (EVs) as emerging therapy for cardiovascular diseases.

Advantages	Disadvantages	
Natural homing ability and specific transfer of bioactive molecules (152, 153)	Lack of standardization of pre-analytical variables (127, 140)	
Highly hydrophobic drugs and hydrophilic drugs can be packaged (152, 154)	No clear nomenclature leading to variable qualitative and quantitative analysis (1, 155)	
Good delivery vehicle for drugs based on the ability to cross blood- brain barrier (153, 156)	No recommended isolation protocol for clinical grade production and quality contro of EV-based therapeutics (68, 142)	
Easy to adapt/optimize content and surface receptors for site- specific delivery (147, 154, 156)	Comprised of heterogeneous components depending on the isolation, handling, and agonist(s) used (67, 69, 128–130)	
More stable upon freezing and thawing compared with cells, biocompatible, and non-cytotoxic (142, 156)	The need to establish techniques and methodologies to rigorously quantify and characterized the molecular and physical aspects of EVs (142)	



instance, PMV-inspired nanovesicles have been engineered to deliver thrombolysis specifically to sites of clot formation (147). The design of the therapeutic nano vesicle was based on the biological aspects of PMVs that are relevant to thrombus formation. Surface receptors GPIIb/IIIa and P-selectin were used to target the site of clot formation, and the enzyme phospholipase A2 was employed to rupture the vesicle and release the lytic drug, thus allowing the targeting of thrombolysis only at sites of clot formation (147). Thus, this study highlights the feasibility of altering the cargo of EVs and surface receptors for site-specific delivery (Table 2). Also, natural PMVs have showed therapeutic potential to treat cardiovascular diseases. Pretreating circulating-angiogenic cells (CACs) with PMVs derived from atherosclerotic patients (PMV-CACs) enhances the re-endothelization capacity of these cells (148). PMV-CACs enhance blood flow and increase capillary density in rats suffering from hind limb ischemia via PMV release of RANTES (148). Furthermore, a cardioprotective role of MVs, specifically PMVs, has been demonstrated in a study by Ma and colleagues (149). In this study, the transfusion of PMVs from rats that underwent hind limb ischemia-reperfusion preconditioning resulted in increased levels of PMVs and a reduction of infarct size, indicative of a protective role of PMVs in the context of cardiac ischemia-reperfusion injury. This was further supported by data showing that PMVs transfused into rats with middle cerebral artery occlusion reduced infarct area (150). Similarly, other types of EVs have shown promising therapeutic potential. EVs, mainly MVs and exosomes, displayed a cardioprotective role by enhancing the recovery of cardiac function in a postinfarct heart failure animal model (151). Although the precise mechanisms involved in the protective role of EVs in cardiovascular diseases is still incompletely understood, the capacity of EVs to carry vast biological cargos and their ability to transfer a wide array of bioactive molecules to target cells may explain the beneficial effect of EVs as a therapy for cardiovascular diseases. Furthermore, EVs can be used as a drug delivery system to increase solubility, stability, and bioavailability of hydrophobic drug in the blood circulation (Table 2). Sun et al. encapsulated the drug curcumin in exosomes and successfully delivered the drug to activated monocytes, thus inducing an anti-inflammatory response in a preclinical model of septic shock (152). Despite the early promise from these studies involving PMVs, further studies are needed to delineate the potential utility of PMVs in cardiovascular diseases. Further to this, a number of challenges remain in translating therapeutic EVs to the clinic (Table 2).

REFERENCES

- van der Pol E, Böing AN, Gool EL, Nieuwland R. Recent developments in the nomenclature, presence, isolation, detection and clinical impact of extracellular vesicles. *J Thromb Haemost* (2016) 14(1):48–56. doi:10.1111/ jth.13190
- Boulanger CM, Loyer X, Rautou PE, Amabile N. Extracellular vesicles in coronary artery disease. *Nat Rev Cardiol* (2017) 14(5):259–72. doi:10.1038/ nrcardio.2017.7
- Aikawa E. Extracellular vesicles in cardiovascular disease: focus on vascular calcification. J Physiol (2016) 594(11):2877–80. doi:10.1113/JP272112

CONCLUSION

Platelet-derived microvesicles have indeed come a long way from their initial descriptions as "platelet dust" to now being considered as major mediators of intercellular communication. Their role in hemostasis, inflammation, angiogenesis, and wound healing may be beneficial or deleterious and can contribute to the pathogenesis of cardiovascular diseases (Figure 2). PMVs represent a heterogeneous population of EVs derived from platelets. This heterogenicity is mainly due to the variability of stimuli capable of inducing platelet activation and PMV release. Indeed, the mode of platelet activation seems to define the size, content, and amount of PMVs, which together dictate their fate (cell targeting or not) and their effects and potential involvement in diseases. However, despite the undoubted progress in our understanding of the biological roles PMVs, there remains a pressing need to establish common protocols, analysis, and nomenclature in the field of EV research. Advancing our knowledge of the biological function of EVs holds promise to influence the treatment of cardiovascular diseases. Most interestingly, EV-based therapy represents an emerging and novel biological therapeutic approach for cardiovascular diseases. Therefore, harnessing our knowledge of EV biology may indeed unlock the full potential of EVs for the diagnosis and therapy of cardiovascular diseases and beyond.

AUTHOR CONTRIBUTIONS

All authors listed have made a substantial, direct, and intellectual contribution to the work and approved it for publication.

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- Crescitelli R, Lässer C, Szabó TG, Kittel A, Eldh M, Dianzani I, et al. Distinct RNA profiles in subpopulations of extracellular vesicles: apoptotic bodies, microvesicles and exosomes. *J Extracell Vesicles* (2013) 2(1):20677. doi:10.3402/jev.v2i0.20677
- Akers JC, Gonda D, Kim R, Carter BS, Chen CC. Biogenesis of extracellular vesicles (EV): exosomes, microvesicles, retrovirus-like vesicles, and apoptotic bodies. *J Neurooncol* (2013) 113(1):1–11. doi:10.1007/s11060-013-1084-8
- Willms E, Johansson HJ, Mäger I, Lee Y, Blomberg KE, Sadik M, et al. Cells release subpopulations of exosomes with distinct molecular and biological properties. *Sci Rep* (2016) 6:22519. doi:10.1038/srep22519

- Kowal J, Arras G, Colombo M, Jouve M, Morath JP, Primdal-Bengtson B, et al. Proteomic comparison defines novel markers to characterize heterogeneous populations of extracellular vesicle subtypes. *Proc Natl Acad Sci* USA (2016) 113(8):8. doi:10.1073/pnas.1521230113
- Poe AJ, Knowlton AA. Exosomes as agents of change in the cardiovascular system. J Mol Cell Cardiol (2017) 111:40–50. doi:10.1016/j.yjmcc.2017. 08.002
- Thery C, Ostrowski M, Segura E. Membrane vesicles as conveyors of immune responses. Nat Rev Immunol (2009) 9(8):581–93. doi:10.1038/ nri2567
- Müller I, Klocke A, Alex M, Kotzsch M, Luther T, Morgenstern E, et al. Intravascular tissue factor initiates coagulation via circulating microvesicles and platelets. *FASEB J* (2003) 17(3):476–8. doi:10.1096/fj.02-0574fje
- Garcia BA, Smalley DM, Cho H, Shabanowitz J, Ley K, Hunt DF. The platelet microparticle proteome. J Proteome Res (2005) 4(5):1516–21. doi:10.1021/ pr0500760
- Vélez P, Izquierdo I, Rosa I, García Á. A 2D-DIGE-based proteomic analysis reveals differences in the platelet releasate composition when comparing thrombin and collagen stimulations. *Sci Rep* (2015) 5:8198. doi:10.1038/ srep08198
- Mobarrez F, Sjövik C, Soop A, Hållström L, Frostell C, Pisetsky DS, et al. CD40L expression in plasma of volunteers following LPS administration: a comparison between assay of CD40L on platelet microvesicles and soluble CD40L. *Platelets* (2015) 26(5):486–90. doi:10.3109/09537104. 2014.932339
- Mause SF, Weber C. Microparticles: protagonists of a novel communication network for intercellular information exchange. *Circ Res* (2010) 107(9): 1047–57. doi:10.1161/CIRCRESAHA.110.226456
- Hristov M, Erl W, Linder S, Weber PC. Apoptotic bodies from endothelial cells enhance the number and initiate the differentiation of human endothelial progenitor cells in vitro. *Blood* (2004) 104(9):2761–6. doi:10.1182/ blood-2003-10-3614
- Kerr JF, Wyllie AH, Currie AR. Apoptosis: a basic biological phenomenon with wide-ranging implications in tissue kinetics. *Br J Cancer* (1972) 26(4):239–57. doi:10.1038/bjc.1972.33
- Wolf P. The nature and significance of platelet products in human plasma. Br J Haematol (1967) 13(3):269–88. doi:10.1111/j.1365-2141.1967.tb08741.x
- Sandberg H, Bode AP, Dombrose FA, Hoechli M, Lentz BR. Expression of coagulant activity in human platelets: release of membranous vesicles providing platelet factor 1 and platelet factor 3. *Thromb Res* (1985) 39(1): 63–79. doi:10.1016/0049-3848(85)90122-7
- Brisson AR, Tan S, Linares R, Gounou C, Arraud N. Extracellular vesicles from activated platelets: a semiquantitative cryo-electron microscopy and immuno-gold labeling study. *Platelets* (2017) 28(3):263–71. doi:10.1080/ 09537104.2016.1268255
- Duchez AC, Boudreau LH, Naika GS, Bollinger J, Belleannée C, Cloutier N, et al. Platelet microparticles are internalized in neutrophils via the concerted activity of 12-lipoxygenase and secreted phospholipase A2-IIA. *Proc Natl Acad Sci U S A* (2015) 112(27):E3564–73. doi:10.1073/pnas. 1507905112
- Zhang Y, Meng H, Ma R, He Z, Wu X, Cao M, et al. Circulating microparticles, blood cells, and endothelium induce procoagulant activity in sepsis through phosphatidyleserine exposure. *Shock* (2016) 45(3):299–307. doi:10.1097/ SHK.000000000000509
- Li S, Wei J, Zhang C, Li X, Meng W, Mo X, et al. Cell-derived microparticles in patients with type 2 diabetes mellitus: a systematic review and metaanalysis. *Cell Physiol Biochem* (2016) 39(6):2439–50. doi:10.1159/000452512
- 23. Fortin PR, Cloutier N, Bissonnette V, Aghdassi E, Eder L, Simonyan D, et al. Distinct subtypes of microparticle-containing immune complexes are associated with disease activity, damage, and carotid intima-media thickness in systemic lupus erythematosus. *J Rheumatol* (2016) 43(11):2019–25. doi:10.3899/jrheum.160050
- Suades R, Padró T, Alonso R, Mata P, Badimon L. High levels of TSP1+/ CD142+ platelet-derived microparticles characterise young patients with high cardiovascular risk and subclinical atherosclerosis. *Thromb Haemost* (2015) 114(6):1310–21. doi:10.1160/TH15-04-0325
- 25. Lukasik M, Rozalski M, Luzak B, Michalak M, Ambrosius W, Watala C, et al. Enhanced platelet-derived microparticle formation is associated with

carotid atherosclerosis in convalescent stroke patients. *Platelets* (2013) 24(1):63-70. doi:10.3109/09537104.2011.654292

- 26. Csongrádi É, Nagy B Jr, Fulop T, Varga Z, Karányi Z, Magyar MT, et al. Increased levels of platelet activation markers are positively associated with carotid wall thickness and other atherosclerotic risk factors in obese patients. *Thromb Haemost* (2011) 106(4):683–92. doi:10.1160/TH11-01-0030
- Kuriyama N, Nagakane Y, Hosomi A, Ohara T, Kasai T, Harada S, et al. Evaluation of factors associated with elevated levels of platelet-derived microparticles in the acute phase of cerebral infarction. *Clin Appl Thromb Hemost* (2010) 16(1):26–32. doi:10.1177/1076029609338047
- Tan KT, Tayebjee MH, Lim HS, Lip GY. Clinically apparent atherosclerotic disease in diabetes is associated with an increase in platelet microparticle levels. *Diabet Med* (2005) 22(12):1657–62. doi:10.1111/j.1464-5491.2005. 01707.x
- Cui Y, Zheng L, Jiang M, Jia R, Zhang X, Quan Q, et al. Circulating microparticles in patients with coronary heart disease and its correlation with interleukin-6 and C-reactive protein. *Mol Biol Rep* (2013) 40(11):6437–42. doi:10.1007/s11033-013-2758-1
- Stępień E, Stankiewicz E, Zalewski J, Godlewski J, Zmudka K, Wybrańska I. Number of microparticles generated during acute myocardial infarction and stable angina correlates with platelet activation. *Arch Med Res* (2012) 43(1):31–5. doi:10.1016/j.arcmed.2012.01.006
- Hartopo AB, Puspitawati I, Gharini PP, Setianto BY. Platelet microparticle number is associated with the extent of myocardial damage in acute myocardial infarction. Arch Med Sci (2016) 12(3):529–37. doi:10.5114/ aoms.2016.59926
- Webber AJ, Johnson SA. Platelet participation in blood coagulation aspects of hemostasis. Am J Pathol (1970) 60(1):19–42.
- 33. Holme PA, Orvim U, Hamers MJ, Solum NO, Brosstad FR, Barstad RM, et al. Shear-induced platelet activation and platelet microparticle formation at blood flow conditions as in arteries with a severe stenosis. *Arterioscler Thromb Vasc Biol* (1997) 17(4):646–53. doi:10.1161/01.ATV. 17.4.646
- Sakariassen KS, Holme PA, Orvim U, Barstad RM, Solum NO, Brosstad FR. Shear-induced platelet activation and platelet microparticle formation in native human blood. *Thromb Res* (1998) 92(6 Suppl 2):S33–41. doi:10.1016/ S0049-3848(98)00158-3
- 35. Biró E, Akkerman JW, Hoek FJ, Gorter G, Pronk LM, Sturk A, et al. The phospholipid composition and cholesterol content of platelet-derived microparticles: a comparison with platelet membrane fractions. J Thromb Haemost (2005) 3(12):2754–63. doi:10.1111/j.1538-7836.2005.01646.x
- Boilard E, Nigrovic PA, Larabee K, Watts GF, Coblyn JS, Weinblatt ME, et al. Platelets amplify inflammation in arthritis via collagen-dependent microparticle production. *Science* (2010) 327(5965):580–3. doi:10.1126/ science.1181928
- Connor DE, Exner T, Ma DD, Joseph JE. The majority of circulating platelet-derived microparticles fail to bind annexin V, lack phospholipiddependent procoagulant activity and demonstrate greater expression of glycoprotein Ib. *Thromb Haemost* (2010) 103(5):1044–52. doi:10.1160/ TH09-09-0644
- Diehl P, Aleker M, Helbing T, Sossong V, Germann M, Sorichter S, et al. Increased platelet, leukocyte and endothelial microparticles predict enhanced coagulation and vascular inflammation in pulmonary hypertension. J Thromb Thrombolysis (2011) 31(2):173–9. doi:10.1007/s11239-010-0507-z
- Diehl P, Nagy F, Sossong V, Helbing T, Beyersdorf F, Olschewski M, et al. Increased levels of circulating microparticles in patients with severe aortic valve stenosis. *Thromb Haemost* (2008) 99(4):711–9. doi:10.1160/ TH07-05-0334
- Pasquet JM, Dachary-Prigent J, Nurden AT. Calcium influx is a determining factor of calpain activation and microparticle formation in platelets. *Eur J Biochem* (1996) 239(3):647–54. doi:10.1111/j.1432-1033.1996. 0647u.x
- Fujii T, Sakata A, Nishimura S, Eto K, Nagata S. TMEM16F is required for phosphatidylserine exposure and microparticle release in activated mouse platelets. *Proc Natl Acad Sci U S A* (2015) 112(41):12800–5. doi:10.1073/ pnas.1516594112

- Crespin M, Vidal C, Picard F, Lacombe C, Fontenay M. Activation of PAK1/2 during the shedding of platelet microvesicles. *Blood Coagul Fibrinolysis* (2009) 20(1):63–70. doi:10.1097/MBC.0b013e32831bc310
- Cauwenberghs S, Feijge MA, Harper AG, Sage SO, Curvers J, Heemskerk JW. Shedding of procoagulant microparticles from unstimulated platelets by integrin-mediated destabilization of actin cytoskeleton. *FEBS Lett* (2006) 580(22):5313–20. doi:10.1016/j.febslet.2006.08.082
- Flaumenhaft R. Formation and fate of platelet microparticles. Blood Cells Mol Dis (2006) 36(2):182–7. doi:10.1016/j.bcmd.2005.12.019
- Rand ML, Wang H, Bang KW, Packham MA, Freedman J. Rapid clearance of procoagulant platelet-derived microparticles from the circulation of rabbits. *J Thromb Haemost* (2006) 4(7):1621–3. doi:10.1111/j.1538-7836.2006. 02011.x
- Rank A, Nieuwland R, Crispin A, Grützner S, Iberer M, Toth B, et al. Clearance of platelet microparticles in vivo. *Platelets* (2011) 22(2):111–6. doi:10.3109/09537104.2010.520373
- 47. Angelot F, Seillès E, Biichlé S, Berda Y, Gaugler B, Plumas J, et al. Endothelial cell-derived microparticles induce plasmacytoid dendritic cell maturation: potential implications in inflammatory diseases. *Haematologica* (2009) 94(11):1502–12. doi:10.3324/haematol.2009.010934
- Faille D, El-Assaad F, Mitchell AJ, Alessi MC, Chimini G, Fusai T, et al. Endocytosis and intracellular processing of platelet microparticles by brain endothelial cells. *J Cell Mol Med* (2012) 16(8):1731–8. doi:10.1111/j. 1582-4934.2011.01434.x
- 49. Dasgupta SK, Abdel-Monem H, Niravath P, Le A, Bellera RV, Langlois K, et al. Lactadherin and clearance of platelet-derived microvesicles. *Blood* (2009) 113(6):1332–9. doi:10.1182/blood-2008-07-167148
- Abdel-Monem H, Dasgupta SK, Le A, Prakasam A, Thiagarajan P. Phagocytosis of platelet microvesicles and beta2- glycoprotein I. *Thromb Haemost* (2010) 104(2):335–41. doi:10.1160/TH09-12-0849
- Dasgupta SK, Le A, Chavakis T, Rumbaut RE, Thiagarajan P. Developmental endothelial locus-1 (Del-1) mediates clearance of platelet microparticles by the endothelium. *Circulation* (2012) 125(13):1664–72. doi:10.1161/ CIRCULATIONAHA.111.068833
- Ceroi A, Delettre FA, Marotel C, Gauthier T, Asgarova A, Biichlé S, et al. The anti-inflammatory effects of platelet-derived microparticles in human plasmacytoid dendritic cells involve liver X receptor activation. *Haema-tologica* (2016) 101(3):e72–6. doi:10.3324/haematol.2015.135459
- Köppler B, Cohen C, Schlöndorff D, Mack M. Differential mechanisms of microparticle transfer to B cells and monocytes: anti-inflammatory properties of microparticles. *Eur J Immunol* (2006) 36(3):648–60. doi:10.1002/ eji.200535435
- A-Gonzalez N, Hidalgo A. Nuclear receptors and clearance of apoptotic cells: stimulating the macrophage's appetite. *Front Immunol* (2014) 5:211. doi:10.3389/fimmu.2014.00211
- Mulcahy LA, Pink RC, Carter DR. Routes and mechanisms of extracellular vesicle uptake. *J Extracell Vesicles* (2014) 3(1):24641. doi:10.3402/jev.v3. 24641
- Lindemann S, Tolley ND, Dixon DA, McIntyre TM, Prescott SM, Zimmerman GA, et al. Activated platelets mediate inflammatory signaling by regulated interleukin 1beta synthesis. *J Cell Biol* (2001) 154(3):485–90. doi:10.1083/jcb.200105058
- Diehl P, Fricke A, Sander L, Stamm J, Bassler N, Htun N, et al. Microparticles: major transport vehicles for distinct microRNAs in circulation. *Cardiovasc Res* (2012) 93(4):633–44. doi:10.1093/cvr/cvs007
- Liang H, Yan X, Pan Y, Wang Y, Wang N, Li L, et al. MicroRNA-223 delivered by platelet-derived microvesicles promotes lung cancer cell invasion via targeting tumor suppressor EPB41L3. *Mol Cancer* (2015) 14:58. doi:10.1186/ s12943-015-0327-z
- Jansen F, Yang X, Proebsting S, Hoelscher M, Przybilla D, Baumann K, et al. MicroRNA expression in circulating microvesicles predicts cardiovascular events in patients with coronary artery disease. *J Am Heart Assoc* (2014) 3(6):e001249. doi:10.1161/JAHA.114.001249
- Pan Y, Liang H, Liu H, Li D, Chen X, Li L, et al. Platelet-secreted microRNA-223 promotes endothelial cell apoptosis induced by advanced glycation end products via targeting the insulin-like growth factor 1 receptor. *J Immunol* (2014) 192(1):437–46. doi:10.4049/jimmunol.1301790
- 61. Boudreau LH, Duchez AC, Cloutier N, Soulet D, Martin N, Bollinger J, et al. Platelets release mitochondria serving as substrate for bactericidal group

IIA-secreted phospholipase A2 to promote inflammation. *Blood* (2014) 124(14):2173-83. doi:10.1182/blood-2014-05-573543

- Iavello A, Frech VS, Gai C, Deregibus MC, Quesenberry PJ, Camussi G. Role of Alix in miRNA packaging during extracellular vesicle biogenesis. *Int J Mol Med* (2016) 37(4):958–66. doi:10.3892/ijmm.2016.2488
- Janas T, Janas MM, Sapoń K, Janas T. Mechanisms of RNA loading into exosomes. FEBS Lett (2015) 589(13):1391–8. doi:10.1016/j.febslet.2015. 04.036
- Zhang J, Li S, Li L, Li M, Guo C, Yao J, et al. Exosome and exosomal microRNA: trafficking, sorting, and function. *Genomics Proteomics Bioinformatics* (2015) 13(1):17–24. doi:10.1016/j.gpb.2015.02.001
- 65. Klein-Scory S, Tehrani MM, Eilert-Micus C, Adamczyk KA, Wojtalewicz N, Schnölzer M, et al. New insights in the composition of extracellular vesicles from pancreatic cancer cells: implications for biomarkers and functions. *Proteome Sci* (2014) 12(1):50. doi:10.1186/s12953-014-0050-5
- Dean WL, Lee MJ, Cummins TD, Schultz DJ, Powell DW. Proteomic and functional characterisation of platelet microparticle size classes. *Thromb Haemost* (2009) 102(4):711–8. doi:10.1160/TH09-04-243
- Milioli M, Ibáñez-Vea M, Sidoli S, Palmisano G, Careri M, Larsen MR. Quantitative proteomics analysis of platelet-derived microparticles reveals distinct protein signatures when stimulated by different physiological agonists. J Proteomics (2015) 121:56–66. doi:10.1016/j.jprot.2015.03.013
- Shai E, Rosa I, Parguiña AF, Motahedeh S, Varon D, García Á. Comparative analysis of platelet-derived microparticles reveals differences in their amount and proteome depending on the platelet stimulus. *J Proteomics* (2012) 76:287–96. doi:10.1016/j.jprot.2012.02.030
- Perez-Pujol S, Marker PH, Key NS. Platelet microparticles are heterogeneous and highly dependent on the activation mechanism: studies using a new digital flow cytometer. *Cytometry A* (2007) 71(1):38–45. doi:10.1002/ cyto.a.20354
- Castaman G, Yu-Feng L, Rodeghiero F. A bleeding disorder characterised by isolated deficiency of platelet microvesicle generation. *Lancet* (1996) 347(9002):700–1. doi:10.1016/S0140-6736(96)91259-3
- Mattheij NJ, Braun A, van Kruchten R, Castoldi E, Pircher J, Baaten CC, et al. Survival protein anoctamin-6 controls multiple platelet responses including phospholipid scrambling, swelling, and protein cleavage. *FASEB J* (2016) 30(2):727–37. doi:10.1096/fj.15-280446
- 72. Sims PJ, Wiedmer T, Esmon CT, Weiss HJ, Shattil SJ. Assembly of the platelet prothrombinase complex is linked to vesiculation of the platelet plasma membrane. Studies in Scott syndrome: an isolated defect in platelet procoagulant activity. J Biol Chem (1989) 264(29):17049–57.
- Owens AP, Mackman N. Microparticles in hemostasis and thrombosis. Circ Res (2011) 108(10):1284–97. doi:10.1161/CIRCRESAHA.110.233056
- Khan MM, Hattori T, Niewiarowski S, Edmunds LH Jr, Colman RW. Truncated and microparticle-free soluble tissue factor bound to peripheral monocytes preferentially activate factor VII. *Thromb Haemost* (2006) 95(3):462–8. doi:10.1160/TH05-06-0432
- Dahlback B, Wiedmer T, Sims PJ. Binding of anticoagulant vitamin K-dependent protein S to platelet-derived microparticles. *Biochemistry* (1992) 31(51):12769–77. doi:10.1021/bi00166a009
- Somajo S, Koshiar RL, Norström E, Dahlbäck B. Protein S and factor V in regulation of coagulation on platelet microparticles by activated protein C. *Thromb Res* (2014) 134(1):144–52. doi:10.1016/j.thromres.2014.04.031
- Stavenuiter F, Davis NF, Duan E, Gale AJ, Heeb MJ. Platelet protein S directly inhibits procoagulant activity on platelets and microparticles. *Thromb Haemost* (2013) 109(2):229–37. doi:10.1160/TH12-08-0622
- Berckmans RJ, Nieuwland R, Böing AN, Romijn FP, Hack CE, Sturk A. Cell-derived microparticles circulate in healthy humans and support low grade thrombin generation. *Thromb Haemost* (2001) 85(4):639–46.
- 79. Tans G, Rosing J, Thomassen MC, Heeb MJ, Zwaal RF, Griffin JH. Comparison of anticoagulant and procoagulant activities of stimulated platelets and platelet-derived microparticles. *Blood* (1991) 77(12):2641–8.
- Cloutier N, Tan S, Boudreau LH, Cramb C, Subbaiah R, Lahey L, et al. The exposure of autoantigens by microparticles underlies the formation of potent inflammatory components: the microparticle-associated immune complexes. *EMBO Mol Med* (2013) 5(2):235–49. doi:10.1002/emmm. 201201846
- 81. Bei JJ, Liu C, Peng S, Liu CH, Zhao WB, Qu XL, et al. Staphylococcal SSL5-induced platelet microparticles provoke proinflammatory responses

via the CD40/TRAF6/NFkappaB signalling pathway in monocytes. *Thromb Haemost* (2016) 115(3):632–45. doi:10.1160/TH15-04-0322

- Forlow SB, McEver RP, Nollert MU. Leukocyte-leukocyte interactions mediated by platelet microparticles under flow. *Blood* (2000) 95(4): 1317–23.
- Dinkla S, van Cranenbroek B, van der Heijden WA, He X, Wallbrecher R, Dumitriu IE, et al. Platelet microparticles inhibit IL-17 production by regulatory T cells through P-selectin. *Blood* (2016) 127(16):1976–86. doi:10.1182/ blood-2015-04-640300
- Sadallah S, Eken C, Martin PJ, Schifferli JA. Microparticles (ectosomes) shed by stored human platelets downregulate macrophages and modify the development of dendritic cells. *J Immunol* (2011) 186(11):6543–52. doi:10.4049/jimmunol.1002788
- Tang K, Liu J, Yang Z, Zhang B, Zhang H, Huang C, et al. Microparticles mediate enzyme transfer from platelets to mast cells: a new pathway for lipoxin A4 biosynthesis. *Biochem Biophys Res Commun* (2010) 400(3):432–6. doi:10.1016/j.bbrc.2010.08.095
- Baker N, O'Meara SJ, Scannell M, Maderna P, Godson C. Lipoxin A4: antiinflammatory and anti-angiogenic impact on endothelial cells. *J Immunol* (2009) 182(6):3819–26. doi:10.4049/jimmunol.0803175
- Braig D, Nero TL, Koch HG, Kaiser B, Wang X, Thiele JR, et al. Transitional changes in the CRP structure lead to the exposure of proinflammatory binding sites. *Nat Commun* (2017) 8:14188. doi:10.1038/ ncomms14188
- Habersberger J, Strang F, Scheichl A, Htun N, Bassler N, Merivirta RM, et al. Circulating microparticles generate and transport monomeric C-reactive protein in patients with myocardial infarction. *Cardiovasc Res* (2012) 96(1):64–72. doi:10.1093/cvr/cvs237
- Thiele JR, Habersberger J, Braig D, Schmidt Y, Goerendt K, Maurer V, et al. Dissociation of pentameric to monomeric C-reactive protein localizes and aggravates inflammation: in vivo proof of a powerful proinflammatory mechanism and a new anti-inflammatory strategy. *Circulation* (2014) 130(1):35–50. doi:10.1161/CIRCULATIONAHA.113.007124
- Eisenhardt SU, Thiele JR, Bannasch H, Stark GB, Peter K. C-reactive protein: how conformational changes influence inflammatory properties. *Cell Cycle* (2009) 8(23):3885–92. doi:10.4161/cc.8.23.10068
- Eisenhardt SU, Habersberger J, Murphy A, Chen YC, Woollard KJ, Bassler N, et al. Dissociation of pentameric to monomeric C-reactive protein on activated platelets localizes inflammation to atherosclerotic plaques. *Circ Res* (2009) 105(2):128–37. doi:10.1161/CIRCRESAHA.108. 190611
- Martin KR, Kantari-Mimoun C, Yin M, Pederzoli-Ribeil M, Angelot-Delettre F, Ceroi A, et al. Proteinase 3 is a phosphatidylserine-binding protein that affects the production and function of microvesicles. *J Biol Chem* (2016) 291(20):10476–89. doi:10.1074/jbc.M115.698639
- Kim HK, Song KS, Chung JH, Lee KR, Lee SN. Platelet microparticles induce angiogenesis in vitro. *Br J Haematol* (2004) 124(3):376–84. doi:10.1046/j.1365-2141.2003.04773.x
- 94. Sun C, Feng SB, Cao ZW, Bei JJ, Chen Q, Zhao WB, et al. Up-regulated expression of matrix metalloproteinases in endothelial cells mediates platelet microvesicle-induced angiogenesis. *Cell Physiol Biochem* (2017) 41(6):2319–32. doi:10.1159/000475651
- Brill A, Dashevsky O, Rivo J, Gozal Y, Varon D. Platelet-derived microparticles induce angiogenesis and stimulate post-ischemic revascularization. *Cardiovasc Res* (2005) 67(1):30–8. doi:10.1016/j.cardiores.2005. 04.007
- Mause SF, Ritzel E, Liehn EA, Hristov M, Bidzhekov K, Müller-Newen G, et al. Platelet microparticles enhance the vasoregenerative potential of angiogenic early outgrowth cells after vascular injury. *Circulation* (2010) 122(5):495–506. doi:10.1161/CIRCULATIONAHA.109.909473
- Hayon Y, Dashevsky O, Shai E, Varon D, Leker RR. Platelet microparticles promote neural stem cell proliferation, survival and differentiation. *J Mol Neurosci* (2012) 47(3):659–65. doi:10.1007/s12031-012-9711-y
- Janowska-Wieczorek A, Wysoczynski M, Kijowski J, Marquez-Curtis L, Machalinski B, Ratajczak J, et al. Microvesicles derived from activated platelets induce metastasis and angiogenesis in lung cancer. *Int J Cancer* (2005) 113(5):752–60. doi:10.1002/ijc.20657
- 99. Wang H, Wang ZH, Kong J, Yang MY, Jiang GH, Wang XP, et al. Oxidized low-density lipoprotein-dependent platelet-derived microvesicles

trigger procoagulant effects and amplify oxidative stress. *Mol Med* (2012) 18:159-66. doi:10.2119/molmed.2011.00295

- Libby P, Ridker PM, Hansson GK. Progress and challenges in translating the biology of atherosclerosis. *Nature* (2011) 473(7347):317–25. doi:10.1038/ nature10146
- 101. Marcos-Ramiro B, Oliva Nacarino P, Serrano-Pertierra E, Blanco-Gelaz MA, Weksler BB, Romero IA, et al. Microparticles in multiple sclerosis and clinically isolated syndrome: effect on endothelial barrier function. BMC Neurosci (2014) 15:110. doi:10.1186/1471-2202-15-110
- 102. Mause SF, von Hundelshausen P, Zernecke A, Koenen RR, Weber C. Platelet microparticles: a transcellular delivery system for RANTES promoting monocyte recruitment on endothelium. *Arterioscler Thromb Vasc Biol* (2005) 25(7):1512–8. doi:10.1161/01.ATV.0000170133.43608.37
- 103. Nomura S, Tandon NN, Nakamura T, Cone J, Fukuhara S, Kambayashi J. High-shear-stress-induced activation of platelets and microparticles enhances expression of cell adhesion molecules in THP-1 and endothelial cells. *Atherosclerosis* (2001) 158(2):277–87. doi:10.1016/S0021-9150(01) 00433-6
- 104. Vasina EM, Cauwenberghs S, Staudt M, Feijge MA, Weber C, Koenen RR, et al. Aging- and activation-induced platelet microparticles suppress apoptosis in monocytic cells and differentially signal to proinflammatory mediator release. *Am J Blood Res* (2013) 3(2):107–23.
- 105. Böing AN, Hau CM, Sturk A, Nieuwland R. Platelet microparticles contain active caspase 3. *Platelets* (2008) 19(2):96–103. doi:10.1080/ 09537100701777295
- 106. Pakala R. Serotonin and thromboxane A2 stimulate platelet-derived microparticle-induced smooth muscle cell proliferation. *Cardiovasc Radiat Med* (2004) 5(1):20–6. doi:10.1016/j.carrad.2003.12.002
- 107. Weber A, Koppen HO, Schror K. Platelet-derived microparticles stimulate coronary artery smooth muscle cell mitogenesis by a PDGFindependent mechanism. *Thromb Res* (2000) 98(5):461–6. doi:10.1016/ S0049-3848(00)00192-4
- Leroyer AS, Isobe H, Lesèche G, Castier Y, Wassef M, Mallat Z, et al. Cellular origins and thrombogenic activity of microparticles isolated from human atherosclerotic plaques. *J Am Coll Cardiol* (2007) 49(7):772–7. doi:10.1016/j.jacc.2006.10.053
- 109. Badlou BA, Wu YP, Smid WM, Akkerman JW. Platelet binding and phagocytosis by macrophages. *Transfusion* (2006) 46(8):1432–43. doi:10.1111/ j.1537-2995.2006.00913.x
- 110. Michelsen AE, Brodin E, Brosstad F, Hansen JB. Increased level of platelet microparticles in survivors of myocardial infarction. *Scand J Clin Lab Invest* (2008) 68(5):386–92. doi:10.1080/00365510701794957
- 111. Porto I, Biasucci LM, De Maria GL, Leone AM, Niccoli G, Burzotta F, et al. Intracoronary microparticles and microvascular obstruction in patients with ST elevation myocardial infarction undergoing primary percutaneous intervention. *Eur Heart J* (2012) 33(23):2928–38. doi:10.1093/ eurheartj/ehs065
- 112. Jung C, Sörensson P, Saleh N, Arheden H, Rydén L, Pernow J. Circulating endothelial and platelet derived microparticles reflect the size of myocardium at risk in patients with ST-elevation myocardial infarction. *Atherosclerosis* (2012) 221(1):226–31. doi:10.1016/j.atherosclerosis.2011.12.025
- 113. Montoro-García S, Shantsila E, Tapp LD, López-Cuenca A, Romero AI, Hernández-Romero D, et al. Small-size circulating microparticles in acute coronary syndromes: relevance to fibrinolytic status, reparative markers and outcomes. *Atherosclerosis* (2013) 227(2):313–22. doi:10.1016/j. atherosclerosis.2013.01.028
- 114. Giannopoulos G, Oudatzis G, Paterakis G, Synetos A, Tampaki E, Bouras G, et al. Red blood cell and platelet microparticles in myocardial infarction patients treated with primary angioplasty. *Int J Cardiol* (2014) 176(1):145–50. doi:10.1016/j.ijcard.2014.07.022
- 115. Tan KT, Tayebjee MH, Macfadyen RJ, Lip GY, Blann AD. Elevated platelet microparticles in stable coronary artery disease are unrelated to disease severity or to indices of inflammation. *Platelets* (2005) 16(6):368–71. doi:10.1080/00207230500120401
- Galli M, Grassi A, Barbui T. Platelet-derived microvesicles in thrombotic thrombocytopenic purpura and hemolytic uremic syndrome. *Thromb Haemost* (1996) 75(3):427–31.
- 117. Sewify EM, Sayed D, Abdel Aal RF, Ahmad HM, Abdou MA. Increased circulating red cell microparticles (RMP) and platelet microparticles (PMP)

in immune thrombocytopenic purpura. *Thromb Res* (2013) 131(2):e59–63. doi:10.1016/j.thromres.2012.11.021

- 118. Bal L, Ederhy S, Di Angelantonio E, Toti F, Zobairi F, Dufaitre G, et al. Circulating procoagulant microparticles in acute pulmonary embolism: a case-control study. *Int J Cardiol* (2010) 145(2):321–2. doi:10.1016/j. ijcard.2009.11.048
- Bucciarelli P, Martinelli I, Artoni A, Passamonti SM, Previtali E, Merati G, et al. Circulating microparticles and risk of venous thromboembolism. *Thromb Res* (2012) 129(5):591–7. doi:10.1016/j.thromres.2011.08.020
- 120. Campello E, Spiezia L, Radu CM, Bulato C, Castelli M, Gavasso S, et al. Endothelial, platelet, and tissue factor-bearing microparticles in cancer patients with and without venous thromboembolism. *Thromb Res* (2011) 127(5):473–7. doi:10.1016/j.thromres.2011.01.002
- 121. Ramacciotti E, Blackburn S, Hawley AE, Vandy F, Ballard-Lipka N, Stabler C, et al. Evaluation of soluble P-selectin as a marker for the diagnosis of deep venous thrombosis. *Clin Appl Thromb Hemost* (2011) 17(4):425–31. doi:10.1177/1076029611405032
- 122. Campello E, Spiezia L, Radu CM, Bulato C, Gavasso S, Tormene D, et al. Circulating microparticles and the risk of thrombosis in inherited deficiencies of antithrombin, protein C and protein S. *Thromb Haemost* (2016) 115(1):81–8. doi:10.1160/TH15-04-0286
- 123. Lincz LF, Scorgie FE, Enjeti A, Seldon M. Variable plasma levels of factor V Leiden correlate with circulating platelet microparticles in carriers of factor V Leiden. *Thromb Res* (2012) 129(2):192–6. doi:10.1016/j.thromres.2011. 06.018
- 124. Ye R, Ye C, Huang Y, Liu L, Wang S. Circulating tissue factor positive microparticles in patients with acute recurrent deep venous thrombosis. *Thromb Res* (2012) 130(2):253–8. doi:10.1016/j.thromres.2011.10.014
- 125. Chirinos JA, Heresi GA, Velasquez H, Jy W, Jimenez JJ, Ahn E, et al. Elevation of endothelial microparticles, platelets, and leukocyte activation in patients with venous thromboembolism. *J Am Coll Cardiol* (2005) 45(9):1467–71. doi:10.1016/j.jacc.2004.12.075
- 126. Martinez M, Ricart JM, Ruiz-Aja S, Rus A, Todolí J, Calvo J, et al. Platelet activation and red blood cell phosphatidylserine exposure evaluated by flow cytometry in patients with Behcet's disease: are they related to thrombotic events? *Pathophysiol Haemost Thromb* (2007) 36(1):18–22. doi:10.1159/000112635
- Yuana Y, Bertina RM, Osanto S. Pre-analytical and analytical issues in the analysis of blood microparticles. *Thromb Haemost* (2011) 105(3):396–408. doi:10.1160/TH10-09-0595
- Kong F, Zhang L, Wang H, Yuan G, Guo A, Li Q, et al. Impact of collection, isolation and storage methodology of circulating microvesicles on flow cytometric analysis. *Exp Ther Med* (2015) 10(6):2093–101. doi:10.3892/ etm.2015.2780
- 129. Trummer A, De Rop C, Tiede A, Ganser A, Eisert R. Recovery and composition of microparticles after snap-freezing depends on thawing temperature. *Blood Coagul Fibrinolysis* (2009) 20(1):52–6. doi:10.1097/ MBC.0b013e32831be9c5
- Chandler WL. Microparticle counts in platelet-rich and platelet-free plasma, effect of centrifugation and sample-processing protocols. *Blood Coagul Fibrinolysis* (2013) 24(2):125–32. doi:10.1097/MBC.0b013e32835a0824
- Jayachandran M, Miller VM, Heit JA, Owen WG. Methodology for isolation, identification and characterization of microvesicles in peripheral blood. *J Immunol Methods* (2012) 375(1–2):207–14. doi:10.1016/j. jim.2011.10.012
- 132. Pospichalova V, Svoboda J, Dave Z, Kotrbova A, Kaiser K, Klemova D, et al. Simplified protocol for flow cytometry analysis of fluorescently labeled exosomes and microvesicles using dedicated flow cytometer. *J Extracell Vesicles* (2015) 4:25530. doi:10.3402/jev.v4.25530
- Li P, Kaslan M, Lee SH, Yao J, Gao Z. Progress in exosome isolation techniques. *Theranostics* (2017) 7(3):789–804. doi:10.7150/thno.18133
- Lobb RJ, Becker M, Wen SW, Wong CS, Wiegmans AP, Leimgruber A, et al. Optimized exosome isolation protocol for cell culture supernatant and human plasma. *J Extracell Vesicles* (2015) 4:27031. doi:10.3402/jev. v4.27031
- 135. Gardiner C, Di Vizio D, Sahoo S, Théry C, Witwer KW, Wauben M, et al. Techniques used for the isolation and characterization of extracellular vesicles: results of a worldwide survey. *J Extracell Vesicles* (2016) 5:32945. doi:10.3402/jev.v5.32945

- Poncelet P, Robert S, Bailly N, Garnache-Ottou F, Bouriche T, Devalet B, et al. Tips and tricks for flow cytometry-based analysis and counting of microparticles. *Transfus Apher Sci* (2015) 53(2):110–26. doi:10.1016/j. transci.2015.10.008
- Headland SE, Jones HR, D'Sa AS, Perretti M, Norling LV. Cuttingedge analysis of extracellular microparticles using ImageStream(X) imaging flow cytometry. *Sci Rep* (2014) 4:5237. doi:10.1038/srep05237
- Chandler WL. Measurement of microvesicle levels in human blood using flow cytometry. *Cytometry B Clin Cytom* (2016) 90(4):326–36. doi:10.1002/ cyto.b.21343
- 139. EV-TRACK Consortium, Van Deun J, Mestdagh P, Agostinis P, Akay Ö, Anand S, et al. EV-TRACK: transparent reporting and centralizing knowledge in extracellular vesicle research. *Nat Methods* (2017) 14(3):228–32. doi:10.1038/nmeth.4185
- 140. Witwer KW, Buzás EI, Bemis LT, Bora A, Lässer C, Lötvall J, et al. Standardization of sample collection, isolation and analysis methods in extracellular vesicle research. J Extracell Vesicles (2013) 27:2. doi:10.3402/jev. v2i0.20360
- 141. Lötvall J, Hill AF, Hochberg F, Buzás EI, Di Vizio D, Gardiner C, et al. Minimal experimental requirements for definition of extracellular vesicles and their functions: a position statement from the International Society for Extracellular Vesicles. *J Extracell Vesicles* (2014) 3:26913. doi:10.3402/ jev.v3.26913
- 142. Lener T, Gimona M, Aigner L, Börger V, Buzas E, Camussi G, et al. Applying extracellular vesicles based therapeutics in clinical trials – an ISEV position paper. *J Extracell Vesicles* (2015) 4:30087. doi:10.3402/jev. v4.30087
- 143. Giacomazzi A, Degan M, Calabria S, Meneguzzi A, Minuz P. Antiplatelet agents inhibit the generation of platelet-derived microparticles. *Front Pharmacol* (2016) 7:314. doi:10.3389/fphar.2016.00314
- 144. Connor DE, Ly K, Aslam A, Boland J, Low J, Jarvis S, et al. Effects of antiplatelet therapy on platelet extracellular vesicle release and procoagulant activity in health and in cardiovascular disease. *Platelets* (2016) 27(8):805–11. doi:10.1080/09537104.2016.1190008
- 145. Bulut D, Becker V, Mugge A. Acetylsalicylate reduces endothelial and platelet-derived microparticles in patients with coronary artery disease. *Can J Physiol Pharmacol* (2011) 89(4):239–44. doi:10.1139/y11-013
- 146. Skeppholm M, Mobarrez F, Malmqvist K, Wallén H. Platelet-derived microparticles during and after acute coronary syndrome. *Thromb Haemost* (2012) 107(6):1122–9. doi:10.1160/TH11-11-0779
- 147. Pawlowski CL, Li W, Sun M, Ravichandran K, Hickman D, Kos C, et al. Platelet microparticle-inspired clot-responsive nanomedicine for targeted fibrinolysis. *Biomaterials* (2017) 128:94–108. doi:10.1016/j.biomaterials. 2017.03.012
- 148. Ohtsuka M, Sasaki K, Ueno T, Seki R, Nakayoshi T, Koiwaya H, et al. Platelet-derived microparticles augment the adhesion and neovascularization capacities of circulating angiogenic cells obtained from atherosclerotic patients. *Atherosclerosis* (2013) 227(2):275–82. doi:10.1016/j. atherosclerosis.2013.01.040
- Ma F, Liu H, Shen Y, Zhang Y, Pan S. Platelet-derived microvesicles are involved in cardio-protective effects of remote preconditioning. *Int J Clin Exp Pathol* (2015) 8(9):10832–9.
- 150. Shan LY, Li JZ, Zu LY, Niu CG, Ferro A, Zhang YD, et al. Plateletderived microparticles are implicated in remote ischemia conditioning in a rat model of cerebral infarction. CNS Neurosci Ther (2013) 19(12):917–25. doi:10.1111/cns.12199
- 151. Kervadec A, Bellamy V, El Harane N, Arakélian L, Vanneaux V, Cacciapuoti I, et al. Cardiovascular progenitor-derived extracellular vesicles recapitulate the beneficial effects of their parent cells in the treatment of chronic heart failure. *J Heart Lung Transplant* (2016) 35(6):795–807. doi:10.1016/j. healun.2016.01.013
- 152. Sun D, Zhuang X, Xiang X, Liu Y, Zhang S, Liu C, et al. A novel nanoparticle drug delivery system: the anti-inflammatory activity of curcumin is enhanced when encapsulated in exosomes. *Mol Ther* (2010) 18(9):1606–14. doi:10.1038/mt.2010.105
- 153. Zhuang X, Xiang X, Grizzle W, Sun D, Zhang S, Axtell RC, et al. Treatment of brain inflammatory diseases by delivering exosome encapsulated anti-inflammatory drugs from the nasal region to the brain. *Mol Ther* (2011) 19(10):1769–79. doi:10.1038/mt.2011.164

- 154. Tian Y, Li S, Song J, Ji T, Zhu M, Anderson GJ, et al. A doxorubicin delivery platform using engineered natural membrane vesicle exosomes for targeted tumor therapy. *Biomaterials* (2014) 35(7):2383–90. doi:10.1016/j. biomaterials.2013.11.083
- Gould SJ, Raposo G. As we wait: coping with an imperfect nomenclature for extracellular vesicles. J Extracell Vesicles (2013) 2(1):20389. doi:10.3402/ jev.v2i0.20389
- Alvarez-Erviti L, Seow Y, Yin H, Betts C, Lakhal S, Wood MJ. Delivery of siRNA to the mouse brain by systemic injection of targeted exosomes. *Nat Biotechnol* (2011) 29(4):341–5. doi:10.1038/nbt.1807

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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