



mTORC1 Overactivation as a Key Aging Factor in the Progression to Type 2 Diabetes Mellitus

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OPEN ACCESS

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Specialty section:

This article was submitted to Endocrinology of Aging, a section of the journal Frontiers in Endocrinology

Received: 04 July 2018 Accepted: 27 September 2018 Published: 16 October 2018

Citation:

Guillén C and Benito M (2018) mTORC1 Overactivation as a Key Aging Factor in the Progression to Type 2 Diabetes Mellitus. Front. Endocrinol. 9:621. doi: 10.3389/fendo.2018.00621

Type 2 Diabetes Mellitus (T2DM), a worldwide epidemics, is a progressive disease initially developing an insulin resistant state, with manifest pancreatic beta islet overwork and hyperinsulinemia. As the disease progresses, pancreatic β cells are overwhelmed and fails in their capacity to compensate insulin resistance. In addition, it is usually associated with other metabolic diseases such as hyperlipidemia, obesity and the metabolic syndrome. During the progression to T2DM there is a chronic activation of mTORC1 signaling pathway, which induces aging and acts as an endogenous inhibitor of autophagy. The complex 1 of mTOR (mTORC1) controls cell proliferation, cell growth as well as metabolism in a variety of cell types through a complex signaling network. Autophagy is involved in the recycling of cellular components for energy generation under nutrient deprivation, and serves as a complementary degradation system to the ubiquitin-proteasome pathway. Autophagy represents a protective mechanism for different cell types, including pancreatic β cells, and potentiates β cell survival across the progression to T2DM. Here, we focus our attention on the chronic overactivation of mTORC1 signaling pathway in β islets from prediabetics patients, making these cells more prone to trigger apoptosis upon several cellular stressors and allowing the progression from prediabetes to type 2 diabetes status.

Keywords: T2DM, autophagy, amylin, mitophagy, mTORC1, TSC2

mTORC1 SIGNALING NETWORK: UPSTREAM AND DOWNSTREAM EFFECTORS

Tuberous sclerosis complex (TSC) is an essential regulator of mTORC1 signaling pathway. TSC is a heterotrimer protein complex formed by the association of TSC1 (hamartin), TSC2 (tuberin) and Tre2-Bub2-Cdc16-1 domain family member 7 (TDC1D7) (1). This complex is the major inhibitory node of the mechanistic target of rapamycin complex 1 (mTORC1) (2). TSC1-TSC2-TBC1D7 complex exerts its inhibitory effect on mTORC1 by a member of the Ras superfamily, called Rheb. TSC2 presents a GTPase activating domain (GAP) favoring the transformation of GTP-Rheb (ras homology enriched in brain) into GDP-Rheb, avoiding mTORC1 activation (2–5). TSC1 and TSC2 are modulated by multiple mechanisms, including phosphorylation. TSC1/TSC2 integrates signals coming from the energetic status of the cell and nutritional availability, with those from extracellular signaling coming from hormones or growth factors such as insulin or IGF1 respectively (2, 6).

mTOR (mechanistic target of rapamycin) is a serinethreonine kinase, belonging to the phosphatidyl-inositol-3 kinase related kinases (PIKK) (7). Depending on the proteins that are associated with mTOR, it can be found into two different complexes, mTORC1 and mTORC2. mTORC1 is the "rapamycin-sensitive" complex and mTORC2 is the "rapamycin-independent" complex. There are two specific components of mTORC1, the regulatory-associated protein of mTOR (RAPTOR) and the 40-Kda proline-rich Akt substrate (PRAS40) (8-10). However, the rapamycin-independent companion of mTOR, known as RICTOR, the mammalian stress-activated MAP kinase-interacting protein 1 (mSIN1) and the protein observed with RICTOR (PROTOR) are specific members of mTORC2 complex (11). The best characterized substrates of mTORC1 are S6 kinase 1 (S6K1) and eIF4Ebinding protein 1 (4E-BP1), controlling protein synthesis and ribosome biogenesis (12). Activation of mTORC2 leads to phosphorylation and activation Akt at serine 473. mTORC2 it is considered as the PDK2 required for the full activation of Akt (13 - 15).

In the last years, it has been determined that mTORC1 activation is produced on the lysosomal membrane through a complex mechanism involving different proteins (16). Phosphorylation events coming from growth factors, through the Akt and MAPK kinases, directly phosphorylates TSC2 and dissociates it from the surface of the lysosome, avoiding GAP activity toward Rheb and mTORC1 activation (17). Furthermore, nutrients such as aminoacids are capable to activate mTORC1 signaling in a Rag-dependent manner, another group of small G-proteins, acting as heterodimers for mTORC1 activation (18, 19). The presence of aminoacids affects RagA/B-GTP ratio vs. RagA/B-GDP through GAP activity of GATOR1 (20) and the guanine exchange factor activity (GEF) of the complex called "Ragulator" (21). Then, mTORC1 is regulated by two different systems of small G proteins, named Rag and Rheb, on the surface of the lysosome (16). Our group has demonstrated that TSC2, apart from its phosphorylation state, can be regulated by acetylation in different lysine residues (22). Now, we are analyzing the relevance of acetylation status of TSC2 for its recruitment to the membrane of the lysosome.

Another important effector of mTORC1 signaling is AMPactivated protein kinase (AMPK). AMPK is highly conserved kinase in eukaryotes. AMPK acts as an allosteric sensor of cellular energy status and regulates anabolic and catabolic signaling pathways for the maintenance of energy homeostasis (23, 24). In addition, AMPK inhibits mTORC1 signaling directly by TSC2 phosphorylation, favoring TSC1-TSC2 association (25). Furthermore, AMPK modulates mTORC1, independently from TSC2 by raptor phosphorylation and inactivation of mTORC1 (26). Moreover, AMPK and autophagy are connected directly by phosphorylation of ULK1 that leads to autophagy induction. In fact, several residues of ULK1 protein are directly phosphorylated by AMPK (27, 28). It has recently been proposed that ULK1 acetylation is essential for autophagy activation (29, 30).

AUTOPHAGY AND ITS RELEVANCE IN PANCREATIC $\boldsymbol{\beta}$ CELLS

Autophagy is a process that controls cytoplasm quality by the elimination of protein aggregates or altered organelles (31, 32). Autophagy is involved in the shutdown of energyconsuming pathways and the stimulation of catabolic processes through the degradation of cellular components under nutrient deprivation (33). Alternatively, to the ubiquitin-proteasome system (UPS), cells depend on autophagy as an alternative degradation system. Furthermore, autophagy it is involved in the elimination of misfolded proteins or altered organelles. The term autophagy encompasses several conserved mechanisms in eukaryotes that can be classified as: macroautophagy, chaperonemediated autophagy and microautophagy. Chaperone-mediated autophagy consists in the recognition of a specific sequence by a chaperone (hsc70) and then migration of the complex to the lysosome. There, the complex interacts with a receptor in the lysosomal membrane, called LAMP-2A, translocates inside and it is finally degraded (34). In microautophagy, the lysosome directly engulfs the components that are going to be degraded (35). As a general process, macroautophagy it is characterized by the production of a double membrane compartment, termed autophagosome, which surrounds cytoplasmic components and then, fuses with lysosomes where proteolytic activity of the later will degrade the engulfed components (36). Autophagy protects pancreatic β cells, increasing pancreatic β cell survival in the progression to T2DM (37, 38). Using a mouse model Atg-7 deletion specifically in pancreatic β cells, it has been determined the essential implication of autophagy in the survival of pancreatic β cells (37). Furthermore, our group has defined this protection of autophagy in ER-stress using a model of insulin secretion-deficient β cells (39). The protective role of autophagy has been determined for avoiding the toxicity of human amylin (hIAPP) (40, 41). In fact, a mouse model with a chronic overactivation of mTORC1 (β-TSC2-/-) showed an increase in pancreatic β cell death and an impairment in autophagy, as we demonstrated in collaboration with Yoshiaki Kido's lab in Kobe University (42). It has recently been observed in human pancreatic β cells an association between dysregulation in autophagy with an increased in cell death (43, 44). Accordingly, this data point to mTORC1 as a signaling pathway that is overactivated in diabetic patients and, at least partially responsible of pancreatic beta cell failure. All the data exposed in the present paper are derived from animal models and not from pre-diabetic patients. There is a clear effect of mTORC1 hyperactivation and the progression to T2DM, but in animal models. The extrapolation to the possible effect in humans has to be assured.

mTORC1 AND MITOPHAGY. CONSEQUENCES ON T2DM

mTORC1 it is an endogenous inhibitor of autophagy. Apart from the bulk autophagy, a non-specific mechanism, all the organelles are submitted to a specific degradation. In the case

of mitochondria, this process is named mitophagy. Additionally, a mitochondrial unfolded protein response (mtUPR) for the degradation of protein aggregates in different locations inside the mitochondria, such as the matrix and in the inner membrane, occurs (45, 46). The inhibition of mTORC1 slows aging by an increased in autophagy, favoring the elimination of misfolded proteins and impaired organelles such as mitochondria, avoiding its accumulation, and associated with aging and different aging-related diseases such as T2DM, or Parkinson disease, or Alzheimer disease (47). In fact, in the progression to T2DM exists an increasing alteration in mitochondrial morphology and function. Then, an increased in mitophagy could be a potential mechanism for delaying the progression to the disease and maintain a preserved β -cell function (48). Mitophagy is a complex process and there are several possible mechanisms. During the development and maturation of red blood cells, it is necessary a removal of mitochondria in the precursor cell or reticulocyte through a NIP3-like protein X (NIX, also known as BNIP3L) mechanism (49). However, in mammals exist another mechanism with involves the system PTEN-induced putative kinase 1 (PINK1)-Parkin pathway. PINK1 is a serinethreonine kinase recruited to the healthy mitochondria, which is imported into the TIM complex of the inner membrane, where it is cleaved by the mitochondrial processing peptidase (MPP) (50) and presenilin-associated rhomboid-like protein (PARL) (51, 52). PINK1 accumulates specifically in damaged mitochondria, because the processing system is inhibited. Then, the E3-ubiquitin ligase Parkin it is recruited to the outer membrane (OM) of the mitochondria, ubiquitinates different substrates, driving the mitophagic process (53). Our group has determined that in MEF TSC2 KO, which presents a hyperactivation of mTORC1, there is an impairment in the PINK1 recruitment in response to a mitophagic inducer such as carbonyl cyanide m-chlorophenylhydrazone (CCCP). Furthermore, in TSC2-/- cells there is a reduction in Pink1 expression as well as PINK1 protein production. Then, in cells lacking TSC2, an accumulation of damaged and aged mitochondria occurs (54). This situation contributes to the accumulation of reactive species of oxygen (ROS) as occurred in diabetics (48). In addition, amylin accumulates and forms cytotoxic oligomers, which damage the cell membrane and directly affects to the functionality and viability of β cells (55). We have recently published that INS1E overexpressing human amylin (INS1E-hIAPP) presents a hyperactivation of mTORC1 and a diminished level of TSC2. Furthermore, these cells possess an impairment in mitophagic flux with the concomitant accumulation of damaged mitochondria (56). However, the mechanism underlying pancreatic β cell toxicity during the formation of amyloid remains unknown (57). In Figure 1, it is depicted the main players in the regulation of pancreatic β cell fate and different drugs acting at different levels.

mTORC1, AGING AND ITS EFFECTS ON T2DM

It is well known that insulin signaling is involved in the control of longevity in a wide spectrum of organisms including worms, flies, and mice (58-62). In addition, the use of rapamycin or knocking-down mTOR or S6K1 can promote life extension in several species (63-66). In addition, TSC1 and TSC2 activation, which negatively controls mTORC1, prolongs life span in Drosophila (67). During aging or under a hypercaloric diet exists an mTORC1 hyperactivity, which derives into a disruption in autophagy and, concomitantly an increase in ER stress (16). In this regard, nicotinamide adenine dinucleotide (NAD⁺) as well as sirtuins levels can be modulated during caloric restriction, activating autophagy, and directly affecting mammalian longevity (68). Mammalian sirtuins are composed of seven members (SIRT1-7) and can be activated by several stimuli including energy deprivation, caloric restriction and resveratrol (69, 70). Sirtuins are a group of NAD⁺-dependent histone deacetylases with homology to the yeast silent information regulator 2 (Sir2) proteins (71). SIRT1-3 catalyzed deacetylation reaction producing a by-product, nicotinamide and O-acetyl ADP-ribose (OAADPr), along with the deacetylated lysine as it has been reviewed in (72). NAD+ levels declines in different tissues with aging, altering the functionality of sirtuins and hence plays a key role in multiple diseases, such as metabolic disorders, neurodegenerative diseases and many others (73). Then, NAD⁺ plays a central role in aging process and longevity (74).

Sirtuins can be activated by different treatments, such as caloric restriction, rapamycin, resveratrol and many others. In fact, all of these treatments are capable to modulate mTORC1 signaling (75). The overactivation of mTORC1 signaling specifically in pancreatic β cells leads to an augmented in β cell mass, which are related to hyperinsulinemia and hypoglycemia (76–79). However, chronic overactivation of mTORC1 signaling pathway develops a progressive hyperglycemia and a diminished islet mass (76). In this regard, we recently published that overactivation of mTORC1 in beta islets from a β -cell specific deletion of Tsc2 (β Tsc2 (–/–) mice) induces an impairment in autophagy with an accumulation of damaged mitochondria. Those deleterious effects were partially reverted by the use of rapamycin (42).

In pancreatic β cells, SIRT1 represses the expression of uncoupling protein 2 (UCP2), leading to an increase in mitochondrial ATP production, thus enhancing insulin secretion (80). Accordingly, a β cell-specific SIRT1-overexpressing transgenic mice (BESTO) has a better glucose tolerance and a boost in insulin secretion (81). Nowadays, SIRT1 it is considered a regulator of different metabolic pathways. There are multiple SIRT1-regulated targets, modulating transcriptional activity of different transcription factors, such as peroxisome proliferatoractivated receptor gamma (PPAR γ), PPAR α , PPAR gamma coactivator 1 alpha (PGC-1 α), and the forkhead box, subgroup O (FOXO) family. Then, SIRT1 is capable to regulate different processes like insulin secretion, gluconeogenesis and fatty acid oxidation (82).

TYPE 2 DIABETES MELLITUS AND mTORC1

Type 2 diabetes mellitus (T2DM) is a very complicated disorder and it is considered epidemic in the world



(83). T2DM it is a progressive disease including insulin resistance, β -cell hyperplasia and/or β cell hypertrophy, that mediates a compensatory insulin secretion and subsequently hyperinsulinemia, pancreatic β cell dysfunction and a loss of cell identity or de-differentiation (84). However, the underlying mechanism mediating pancreatic β cell fate in type 2 diabetics is not completely known. T2DM is associated with higher production of glucose and lipids facilitating β cell death. Furthermore, advanced glycation end-products (AGEs) have been proposed as pancreatic β cell death inducers (85). The hyperamylinemia found in obese people and in insulin-resistant patients, contributes to its oligomerization inside pancreatic β cells, being deleterious for pancreatic β cells (86). From the whole amount of autopsy from patients with T2DM, around 80% present amyloid deposits in the pancreas. However, the importance of amylin it is not yet completely understood. Thus, at the insulin resistant prediabetic stage, mTORC1 is a key effector for the growth and survival of pancreatic β cells. However, if mTORC1 remains chronically overactivated, pancreatic beta cell death occurs and the compensatory insulin secretion mechanism it is compromised. Then, mTORC1 is a double-edged sword in the progression to T2DM (87). The location where protein synthesis occurs is in the endoplasmic reticulum (ER). When misfolded proteins are accumulated into the ER, the unfolded protein response (UPR) is activated. UPR is a protective mechanism that alleviates the cell from that overload. However, if UPR activation it is maintained for a long period of time, pancreatic β cell death occurs (88). T2DM alters the capacity of ER to manage the increased demand of protein synthesis of pancreatic β cells (89). Increasing the expression of endogenous chaperones such as Bip (90), the use of chemical chaperones, such as taurine-conjugated ursodeoxycholic acid (TUDCA) or 4-Phenyl butyric acid (4-PBA) (91), or increasing the ER protein folding capacity by the use of azoramide (92), diminish β cell failure and facilitate the proper folding and avoiding protein aggregation and accumulation of damaged organelles.

However, mTORC1 hyperactivation is observed in other tissues, such as in the heart muscle contributing to the dysfunction and complication of T2DM (93). Furthermore, not only in cardiac tissue, in the liver, apart from being involved in the regulation of lipid homeostasis, facilitating lipogenesis and inhibiting lipolysis and lipophagy (94), is upregulated in insulin resistant states, such as T2DM contributing to the dysregulation of glucose as well as lipid homeostasis (95). In adipose tissues (96), mTORC1 is also involved in the generation of insulin resistance as it has been reviewed in (16).

CONCLUDING REMARKS

Diabetes is a multifactorial and progressive disease with two phases; firstly, a prediabetic stage, with an insulin resistance and hyperinsulinemia, and secondly as manifest diabetes associated with hypoinsulinemia and hyperglycemia. Then, it is crucial to understand the transition from prediabetes to type 2-diabetes status and the underlying molecular mechanisms of disease. At this stage, chronic overactivation of mTORC1 signaling pathway in β islets from prediabetics patients leads to on one hand to the expansion of the pancreatic beta cell mass and, on the other to the inhibition of autophagy as protective mechanism of beta cells against the attack of several stressors, making these cells more prone to trigger apoptosis. Thus, the maintenance of a functional autophagy it is an essential component to protect and prolong pancreatic β cell life span precluding chronic hyperglycemia.

REFERENCES

- Dibble CC, Elis W, Menon S, Qin W, Klekota J, Asara JM, et al. TBC1D7 is a third subunit of the TSC1-TSC2 complex upstream of mTORC1. *Mol Cell* (2012) 47:535–46. doi: 10.1016/j.molcel.2012.06.009
- Huang J, Manning BD. The TSC1-TSC2 complex: a molecular switchboard controlling cell growth. *Biochem J.* (2008) 412:179–90. doi: 10.1042/BJ20080281
- Zhang Y, Gao X, Saucedo LJ, Ru B, Edgar BA, Pan D. Rheb is a direct target of the tuberous sclerosis tumour suppressor proteins. *Nat Cell Biol.* (2003) 5:578–81. doi: 10.1038/ncb999
- Tee AR, Manning BD, Roux PP, Cantley LC, Blenis J. Tuberous sclerosis complex gene products, tuberin and hamartin, control mTOR signaling by acting as a GTPase-activating protein complex toward *Rheb. Curr Biol.* (2003) 13:1259–68. doi: 10.1016/S0960-9822(03)00506-2
- Inoki K, Li Y, Xu T, Guan KL. Rheb GTPase is a direct target of TSC2 GAP activity and regulates mTOR signaling. *Genes Dev.* (2003) 17:1829–34. doi: 10.1101/gad.1110003
- Orlova KA, Crino PB. The tuberous sclerosis complex. Ann N Y Acad Sci. (2010) 1184:87–105. doi: 10.1111/j.1749-6632.2009.05117.x
- Lempiäinen H, Halazonetis TD. Emerging common themes in regulation of PIKKs and PI3Ks. EMBO J. (2009) 28:3067–73. doi: 10.1038/emboj.2009.281
- Sabatini DM. mTOR and cancer: insights into a complex relationship. Nat Rev cancer (2006) 6:729–34. doi: 10.1038/nrc1974
- Sancak Y, Thoreen CC, Peterson TR, Lindquist RA, Kang SA, Spooner E, et al. PRAS40 is an insulin-regulated inhibitor of the mTORC1 protein kinase. *Mol Cell* (2007) 25:903–15. doi: 10.1016/j.molcel.2007.03.003
- Vander Haar E, Lee SI, Bandhakavi S, Griffin TJ, Kim DH. Insulin signalling to mTOR mediated by the Akt/PKB substrate PRAS40. *Nat Cell Biol.* (2007) 9:316–23. doi: 10.1038/ncb1547
- Zoncu R, Efekan A, Sabatini DM. mTOR: from growth signal integration to cancer, diabetes and ageing. *Nat Rev Mol Cell Biol.* (2011) 12:21–35. doi: 10.1038/nrm3025
- 12. Ma XM, Blenis J. Molecular mechanisms of mTOR-mediated translational control. *Nat Rev Mol Cell Biol.* (2009) 10:307–18. doi: 10.1038/nrm2672
- Garcia-Martínez JM, Moran J, Clarke RG, Gray A, Cosulich SH, Chresta CM, et al. Ku-0063794 is a specific inhibitor of the mammalian target of rapamycin (mTOR). *Biochem J*. (2009) 421:29–42. doi: 10.1042/BJ20090489
- Ikenoue T, Inoki K, Yang Q, Zhou X, Guan KL. Essential function of TORC2 in PKC and Akt turn motif phosphorylation, maturation and signalling. *EMBO J.* (2008) 27:1919–31. doi: 10.1038/emboj.2008.119
- Sarbassov DD, Guertin DA, Ali SM, Sabatini DM. Phosphorylation and regulation of Akt/PKB by the rictor-mTOR complex. *Science* (2005) 307:1098– 101. doi: 10.1126/science.1106148
- Laplante M, Sabatini DM. mTOR signaling in growth control and disease. *Cell* (2012) 149:274–93. doi: 10.1016/j.cell.2012.03.017

AUTHOR CONTRIBUTIONS

All authors listed have made a substantial, direct and intellectual contribution to the work, and approved it for publication.

FUNDING

Our research it is supported by grant SAF2017-82133R from M.C.T., Spain.

ACKNOWLEDGMENTS

We thank all the current and former members of the lab, which directly or indirectly, has contributed to this mini-review. We apologize to all not cite authors, whose work was excluded owing to the space limitation.

- Menon S, Dibble CC, Talbott G, Hoxha G, Valvezan AJ, Takahashi H, et al. Spatial control of the TSC complex integrates insulin and nutrient regulation of mTORC1 at the lysosome. *Cell* (2014) 156:771–85. doi: 10.1016/j.cell.2013.11.049
- Kim E, Goraksha-Hicks P, Li L, Neufeld TP, Guan KL. Regulation of TORC1 by Rag GTPases in nutrient response. *Nat Cell Biol.* (2008) 10:935–45. doi: 10.1038/ncb1753
- Sancak Y, Peterson TR, Shaul YD, Lindquist RA, Thoreen CC, Bar-Peled L, et al. The Rag GTPases bind raptor and mediate amino acid signaling to mTORC1. *Science* (2008) 320:1496–501. doi: 10.1126/science.1157535
- Bar-Peled L, Chantranupong L, Cherniack AD, Chen WW, Ottina KA, Grabiner BC, et al. A Tumor suppressor complex with GAP activity for the Rag GTPases that signal amino acid sufficiency to mTORC1. *Science* (2013) 340:1100–6. doi: 10.1126/science.1232044
- Bar-Peled L, Schweitzer LD, Zoncu R, Sabatini DM. Ragulator is a GEF for the rag GTPases that signal amino acid levels to mTORC1. *Cell* (2012) 150:1196–208. doi: 10.1016/j.cell.2012.07.032
- García Aguilar A, Guillén C, Nellist M, Bartolomé A, Benito M. TSC2 N-terminal lysine acetylation status affects to its stability modulating mTORC1 signaling and autophagy. BBA. *Mol Cell Res.* (2016) 1863:2658–67. doi: 10.1016/j.bbamcr.2016.08.006
- Hardie DG. Minireview: the AMP-activated protein kinase cascade: the key sensor of cellular energy status. *Endocrinology* (2003) 144:5179–83. doi: 10.1210/en.2003-0982
- 24. Hardie DG. AMP-activated protein kinase: a cellular energy sensor with a key role in metabolic disorders and in cancer. *Biochem Soc Trans.* (2011) 39:1–13. doi: 10.1042/BST0390001
- Inoki K, Zhu T, Guan KL. TSC2 mediates cellular energy response to control cell growth and survival. *Cell* (2003) 115:577–90. doi: 10.1016/S0092-8674(03)00929-2
- Gwinn DM, Shackelford DB, Egan DF, Mihaylova MM, Mery A, Vasquez DS et al. AMPK phosphorylation of raptor mediates a metabolic checkpoint. *Mol Cell* (2008) 30:214–26. doi: 10.1016/j.molcel.2008.03.003
- Egan DF, Shackelford DB, Mihaylova MM, Melino G, Kohnz RA, Mair W, et al. Phosphorylation of ULK1 (hATG1) by AMP-activated protein kinase connects energy sensing to mitophagy. *Science* (2011) 331:456–61. doi: 10.1126/science.1196371
- Kim J, Kundu M, Viollet B, Guan KL. AMPK and mTOR regulate autophagy through direct phosphorylation of Ulk1. *Nature Cell Biol.* (2011) 13:132–41. doi: 10.1038/ncb2152
- Lin SY, Li TY, Liu Q, Zhang C, Li X, Chen Y, et al. GSK3-TIP60-ULK1 signaling pathway links growth factor deprivation to autophagy. *Science* (2012) 336:477–81. doi: 10.1126/science.1217032
- Lin SY, Li TY, Liu Q, Zhang C, Li X, Chen Y, et al. Protein phosphorylationacetylation cascade connects growth factor deprivation to autophagy. *Autophagy* (2012) 8:1385–6. doi: 10.4161/auto.20959

- Menzies FM, Moreau K, Rubinsztein DC. Protein misfolding disorders and macroautophagy. *Curr Opin Cell Biol.* (2011) 23:190–7. doi: 10.1016/j.ceb.2010.10.010
- Masiero E, Sandri M. Autophagy inhibition induces atrophy and myopathy in adult skeletal muscles. *Autophagy* (2010) 6:307–9. doi: 10.4161/auto.6.2. 11137
- Stipanuk MH. Macroautophagy and its role in nutrient homeostasis. *Nutr Rev.* (2009) 67:677–89. doi: 10.1111/j.1753-4887.2009.00252.x
- Kaushik S, Cuervo AM. Chaperone-mediated autophagy: a unique way to enter the lysosome world. *Trends Cell Biol.* (2012) 22:407–17. doi: 10.1016/j.tcb.2012.05.006
- Li WW, Li J, Bao JK. Microautophagy: lesser-known self-eating. Cell Mol Life Sci. (2012) 69:1125–36. doi: 10.1007/s00018-011-0865-5
- Chen Y, Klionsky DJ. The regulation of autophagy unanswered questions. J Cell Sci. (2011) 124:161–70. doi: 10.1242/jcs.064576
- 37. Ebato C, Uchida T, Arakawa M, Komatsu M, Ueno T, Komiya K et al. Autophagy is important in islet homeostasis and compensatory increase of beta cell mass in response to high-fat diet. *Cell Metab.* (2008) 8:325–32. doi: 10.1016/j.cmet.2008.08.009
- Jung HS, Chung KW, Won Kim J, Kim J, Komatsu M, Tanaka K, et al. Loss of autophagy diminishes pancreatic beta cell mass and function with resultant hyperglycemia. *Cell Metab.* (2008) 8:318–24. doi: 10.1016/j.cmet.2008.08.013
- Bartolomé A, Guillén C, Benito M. Autophagy plays a protective role in endoplasmic reticulum stress-mediated pancreatic β cell death. *Autophagy* (2012) 8:1757–68. doi: 10.4161/auto.21994
- Shigihara N, Fukunaka A, Hara A, Komiya K, Honda A, Uchida T, et al. Human IAPP-induced pancreatic β cell toxicity and its regulation by autophagy. *J Clin Invest.* (2014) 124:3634–44. doi: 10.1172/JCI69866
- Rivera JF, Costes S, Gurlo T, Glabe CG, Butler PC. Autophagy defends pancreatic β cells from human islet amyloid polypeptide-induced toxicity. J Clin Invest. (2014) 124:3489–500. doi: 10.1172/JCI71981
- 42. Bartolomé A, Kimura-Koyanagi M, Asahara S, Guillén C, Inoue H, Teruyama K, et al. Pancreatic β -cell failure mediated by mTORC1 hyperactivity and autophagic impairment. *Diabetes* (2014) 63:2996–3008. doi: 10.2337/db13-0970
- 43. Yuan T, Rafizadeh S, Gorrepati KD, Lupse B, Oberholzer J, Maedler K, et al. Reciprocal regulation of mTOR complexes in pancreatic islets from humans with type 2 diabetes. *Diabetologia* (2017) 60:668–78. doi: 10.1007/s00125-016-4188-9
- Masini M, Bugliani M, Lupi R, Del Guerra S, Boggi U, Filipponi F, et al. Autophagy in human type 2 diabetes pancreatic beta cells. *Diabetologia* (2009) 52:1083–6. doi: 10.1007/s00125-009-1347-2
- Herst PM, Rowe MR, Carson GM, Berridge MV. Functional mitochondria in health and disease. *Front Endocrinol.* (2017) 8:296. doi: 10.3389/fendo.2017.00296
- Baker MJ, Tatsuta T, Langer T. Quality control of mitochondrial proteostasis. *Cold Spring Harb Perspect Biol.* (2011) 3:a007559. doi: 10.1101/cshperspect.a007559
- 47. Saxton RA, Sabatini DM. mTOR Signaling in growth, metabolism, and disease. *Cell* (2017) 168:969–76. doi: 10.1016/j.cell.2017.02.004
- Bhansali S, Bhansali A, Walia R, Saikia UN, Dhawan V. Alterations in mitochondrial oxidative stress and mitophagy in subjects with prediabetes and type 2 diabetes mellitus. *Front Endocrinol.* (2017) 8:347. doi: 10.3389/fendo.2017.00347
- Youle RJ, Narendra DP. Mechanisms of mitophagy. Nat Rev Mol Cell Biol. (2011) 12:9–14. doi: 10.1038/nrm3028
- Greene AW, Grenier K, Aguileta MA, Muise S, Farazifard R, Haque ME, et al. Mitochondrial processing peptidase regulates PINK1 processing, import and Parkin recruitment. *EMBO Rep.* (2012) 13:378–85. doi: 10.1038/embor.2012.14
- Jin SM, Lazarou M, Wang C, Kane LA, Narendra DP, Youle RJ. Mitochondrial membrane potential regulates PINK1 import and proteolytic destabilization by PARL. J Cell Biol. (2010) 191:933–42. doi: 10.1083/jcb.201008084
- Meissner C, Lorenz H, Weihofen A, Selke DJ, Lemberg MK. The mitochondrial intramembrane protease PARL cleaves human Pink1 to regulate Pink1 trafficking. J Neurochem. (2011) 117:856–67. doi: 10.1111/j.1471-4159.2011.07253.x

- Pickrell AM, Youle RJ. The roles of PINK1, parkin, and mitochondrial fidelity in Parkinson's disease. *Neuron* (2015) 85:257–73. doi: 10.1016/j.neuron.2014.12.007
- Bartolomé A, García-Aguilar A, Asahara SI, Kido Y, Guillén C, Pajvani UB, et al. MTORC1 regulates both general autophagy and mitophagy induction after oxidative phosphorylation uncoupling. *Mol Cell Biol.* (2017) 37:e00441– 17. doi: 10.1128/MCB.00441-17
- Caillon L, Hoffmann AR, Botz A, Khemtemourian L. Molecular structure, membrane interactions, and toxicity of the islet amyloid polypeptide in type 2 diabetes mellitus. J Diabetes Res. (2016) 2016:5639875. doi: 10.1155/2016/5639875
- 56. García-Hernández M, García Aguilar A, Burillo J, Gómez Oca R, Manca MA, Novials A, et al. Pancreatic β cells overexpressing hIAPP impaired mitophagy and unbalanced mitochondrial dynamics. *Cell Death Dis.* (2018) 9:481. doi: 10.1038/s41419-018-0533-x
- 57. Zraika S, Hull RL, Verchere CB, Clark A, Potter KJ, Fraser PE, et al. Toxic oligomers and islet beta cell death: guilty by association or convicted by circumstantial evidence? *Diabetologia* (2010) 53:1046–56. doi: 10.1007/s00125-010-1671-6
- Guarente L, Kenyon C. Genetic pathways that regulate ageing in model organisms. *Nature* (2000) 408:255-62. doi: 10.1038/35041700
- Kaeberlein M. Longevity and aging. F1000prime Rep. (2013) 5:5. doi: 10.12703/P5-5
- Sun X, Chen WD, Wang YD. DAF-16/FOXO transcription factor in aging and longevity. Front Pharmacol. (2017) 8:548. doi: 10.3389/fphar.2017.00548
- Tatar M, Post S, Yu K. Nutrient control of drosophila longevity. *Trends* Endocrinol Metab. (2014) 25:509–17. doi: 10.1016/j.tem.2014.02.006
- 62. Fadini GP, Ceolotto G, Pagnin E, de Kreutzenberg S, Avogaro A. At the crossroads of longevity and metabolism: the metabolic syndrome and lifespan determinant pathways. *Aging Cell* (2011) 10:10–7. doi: 10.1111/j.1474-9726.2010.00642.x
- Bjedov I, Toivonen JM, Kerr F, Slack C, Jacobson J, Foley A, et al. Mechanisms of life span extension by rapamycin in the fruit fly Drosophila melanogaster. *Cell Metab.* (2010) 11:35–46. doi: 10.1016/j.cmet.2009.11.010
- Zhang Y, Bokov A, Gelfond J, Soto V, Ikeno Y, Hubbard G, et al. Rapamycin extends life and health in C57BL/6 mice. *J Gerontol A Biol Sci Med Sci.* (2014) 69:119–30. doi: 10.1093/gerona/glt056
- Harrison DE, Strong R, Sharp ZD, Nelson JF, Astle CM, Flurkey K, et al. Rapamycin fed late in life extends lifespan in genetically heterogeneous mice. *Nature* (2009) 460:392–5. doi: 10.1038/nature08221
- Selman C, Tullet JM, Wieser D, Irvine E, Lingard SJ, Choudhury AI, et al. Ribosomal protein S6 kinase 1 signaling regulates mammalian life span. *Science* (2009) 326:140–4. doi: 10.1126/science.1177221
- Kapahi P, Zid BM, Harper T, Koslover D, Sapin V, Benzer S. Regulation of lifespan in Drosophila by modulation of genes in the TOR signaling pathway. *Curr Biol.* (2004) 14:885–90. doi: 10.1016/j.cub.2004.03.059
- Morselli E, Maiuri MC, Markaki M, Megalou E, Pasparaki A, Palikaras K, et al. Caloric restriction and resveratrol promote longevity through the Sirtuin-1-dependent induction of autophagy. *Cell Death Dis.* (2010) 1:e10. doi: 10.1038/cddis.2009.8
- Lam YY, Peterson CM, Ravussin E. Resveratrol vs. calorie restriction: data from rodents to humans. *Exp Gerontol.* (2013) 48:1018–24. doi: 10.1016/j.exger.2013.04.005
- Greer EL, Brunet A. Different dietary restriction regimens extend lifespan by both independent and overlapping genetic pathways in *C. elegans. Aging Cell* (2009) 8:113–27. doi: 10.1111/j.1474-9726.2009.00459.x
- Imai S, Armstrong CM, Kaeberlein M, Guarente L. Transcriptional silencing and longevity protein Sir2 is an NAD-dependent histone deacetylase. *Nature* (2000) 403:795–800. doi: 10.1038/35001622
- Knyphausen P, de Boor S, Kuhlmann N, Scislowski L, Extra A, Baldus L, et al. Insights into lysine deacetylation of natively folded substrate proteins by sirtuins. *J Biol Chem.* (2016) 291:14677–94. doi: 10.1074/jbc.M116.726307
- Johnson S, Imai SI. NAD ⁺ biosynthesis, aging, and disease. *F1000Res.* (2018) 7:132. doi: 10.12688/f1000research.12120.1
- 74. Imai SI, Guarente L. It takes two to tango: NAD⁺ and sirtuins in aging/longevity control. NPJ Aging Mech Dis. (2016) 2:16017. doi: 10.1038/npjamd.2016.17

- Kitada M, Koya D. The use of calorie restriction mimetics to study aging. Methods Mol Biol. (2013) 1048:95–107. doi: 10.1007/978-1-62703-556-9_8
- 76. Shigeyama Y, Kobayashi T, Ido Y, Hashimoto N, Asahara S, Matsuda T, et al. Biphasic response of pancreatic beta-cell mass to ablation of tuberous sclerosis complex 2 in mice. *Mol Cell Biol.* (2008) 28:2971–9. doi: 10.1128/MCB.01695-07
- Rachdi L, Balcazar N, Osorio-Duque F, Elghazi L, Weiss A, Gould A, et al. Disruption of Tsc2 in pancreatic beta cells induces beta cell mass expansion and improved glucose tolerance in a TORC1-dependent manner. *Proc Natl Acad Sci USA*. (2008) 105:9250–5. doi: 10.1073/pnas.0803047105
- Mori H, Inoki K, Opland D, Münzberg H, Villanueva EC, Faouzi M, et al. Critical roles for the TSC-mTOR pathway in β-cell function. Am J Physiol Endocrinol Metab. (2009) 297:E1013–22. doi: 10.1152/ajpendo.00262.2009
- 79. Hamada S, Hara K, Hamada T, Yasuda H, Moriyama H, Nakayama R, et al. Upregulation of the mammalian target of rapamycin complex 1 pathway by Ras homolog enriched in brain in pancreatic beta-cells leads to increased beta-cell mass and prevention of hyperglycemia. *Diabetes* (2009) 58:1321–32. doi: 10.2337/db08-0519
- Bordone L, Motta MC, Picard F, Robinson A, Jhala US, Apfeld J, et al. Sirt1 Regulates Insulin Secretion by Repressing UCP2 in Pancreatic β Cells. Plos Biol. (2006) 4:e31. doi: 10.1371/journal.pbio.0040031
- Moynihan KA, Grimm AA, Plueger MM, Bernal-Mizrachi E, Ford E, Cras-Méneur C, et al. Increased dosage of mammalian Sir2 in pancreatic beta cells enhances glucose-stimulated insulin secretion in mice. *Cell Metab.* (2005) 2:105–17. doi: 10.1016/j.cmet.2005.07.001
- Haigis MC, Sinclair DA. Mammalian sirtuins: biological insights and disease relevance. Annu Rev Pathol. (2010)5:293–95. doi: 10.1146/annurev.pathol.4.110807.092250
- Hu FB. Globalization of diabetes: the role of diet, lifestyle, and genes. *Diabetes Care* (2011) 34:1249–57. doi: 10.2337/dc11-0442
- Remedi MS, Emfinger C. Pancreatic β-cell identity in diabetes. *Diabetes Obes* Metab. (2016) 18:110–6. doi: 10.1111/dom.12727
- Nowotny K, Jung T, Höhn A, Weber D, Grune T. Advanced glycation end products and oxidative stress in type 2 diabetes mellitus *Biomolecules* (2015) 5:194–222. doi: 10.3390/biom5010194
- Akter R, Cao P, Noor H, Ridgway Z, Tu LH, Wang H, et al. Islet amyloid polypeptide: structure, function, and pathophysiology. J Diabetes Res. (2016) 2016:1–18. doi: 10.1155/2016/2798269
- 87. Ardestani A, Lupse B, Kido Y, Leibowitz G, Maedler K. mTORC1 Signaling: a double-edged sword in diabetic β cells. *Cell Metab.* (2018) 27:314–31. doi: 10.1016/j.cmet.2017.11.004

- Back SH, Kaufman RJ. Endoplasmic reticulum stress and type 2 diabetes. Annu Rev Biochem. (2012) 81:767–93. doi: 10.1146/annurev-biochem-072909-095555
- Mukherjee A, Morales-Scheihing D, Butler PC, Soto C. Type 2 diabetes as a protein misfolding disease. *Trends Mol Med.* (2015) 21:439–49. doi: 10.1016/j.molmed.2015.04.005
- Fritz JM, Dong M, Apsley KS, Marin EP, Na CL, Sitamaran S, et al. Deficiency of the BiP cochaperone ERdj4 causes constitutive endoplasmic reticulum stress and metabolic defects. *Mol Biol Cell* (2014) 25:431–40. doi: 10.1091/mbc.e13-06-0319
- Ozcan U, Yilmaz E, Ozcan L, Furuhashi M, Vaillancourt E, Smith RO, et al. Chemical chaperones reduce ER stress and restore glucose homeostasis in a mouse model of type 2 diabetes. *Science* (2006) 313:1137–40. doi: 10.1126/science.1128294
- 92. Fu S, Yalcin A, Lee GY, Li P, Fan J, Arruda AP, et al. Phenotypic assays identify azoramide as a small-molecule modulator of the unfolded protein response with antidiabetic activity. *Sci Transl Med.* (2015) 7:292ra98. doi: 10.1126/scitranslmed.aaa9134.
- 93. Suhara T, Baba Y, Shimada BK, Higa JK, Matsui T. The mTOR signaling pathway in myocardial dysfunction in type 2 diabetes mellitus. *Curr Diab Rep.* (2017) 17:38. doi: 10.1007/s11892-017-0865-4
- Han J, Wang Y. mTORC1 signaling in hepatic lipid metabolism. Protein Cell (2018)c 9:145–51. doi: 10.1007/s13238-017-0409-3
- Titchenell PM, Lazar MA, Birnbaum MJ. Unraveling the regulation of hepatic metabolism by insulin. *Trends Endocrinol Metab.* (2017) 28:497–505. doi: 10.1016/j.tem.2017.03.003
- Lee P, Jung SM, Guertin DA. The complex roles of mTOR in adipocytes and beyond. *Trends Endocrinol Metab.* (2017):28:319–39. doi: 10.1016/j.tem.2017.01.004

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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